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Insects (*Thrips hawaiiensis* (Morgan)) change the stereochemical configuration of 1-phenylethanol emitted from tea (*Camellia sinensis*) flowers[†]

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Insect attacks change the quantity of volatiles emitted from plants, but little is known about the effects on the volatiles' stereochemical configurations. 1-Phenylethanol (1PE) is a major internal volatile of tea (*Camellia sinensis*) flowers. In the present study, insect-damaged *C. sinensis* flowers in the field emitted a different (*R*)-/(*S*)-1PE pattern than that emitted from undamaged flowers. Laboratory experiments showed that changes in the (*R*)-/(*S*)-1PE ratio were caused by treatments with *Thrips hawaiiensis* (Morgan), a pest of cut flowers and vegetable crops. These treatments led to an increase in jasmonic acid (JA) through the activation of allene oxide synthase, which is involved in JA biosynthesis. Exposing flowers to exogenous JA induced an (*R*)-/(*S*)-1PE emission pattern that was similar to that induced by a *T. hawaiiensis* attack, suggesting that JA was involved in the changes in the stereochemical configuration of 1PE emitted from the flowers after insect treatments. This study provides new insights into the involvement of volatiles in interactions between plants and insects, showing that not only the quantity but also the stereochemical configuration of plant volatiles is affected by insects.

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Introduction

Plant volatile compounds are low molecular weight metabolites (<300 Da) with relatively low boiling points (<260 °C) that can be emitted from flowers, leaves, fruits, and roots into the atmosphere or soil, allowing the plants to interact with other organisms.¹ The volatiles emitted from plant vegetative parts are involved in interactions between plants and herbivores, and have several positive functions, including direct and indirect defenses against herbivores, and within-plant and plant-plant signaling.^{2,3} In general, herbivore attacks induce increases in the level of volatiles emitted from plant vegetative parts.

However, less is known about changes in the composition of the volatiles induced by herbivore attack. Allmann and Baldwin reported that attacks by the specialist herbivore *Manduca sexta* induced a rapid (*Z*)/(*E*) isomeric change in the green leaf volatiles released by *Nicotiana* plants.⁴ This change lowered the (*Z*)/(*E*) ratio of the green leaf volatiles blend, which increased the predation rate of predators that fed on the herbivore's eggs.

Floral volatiles are generally believed to play roles in attracting pollinators. In contrast to volatiles emitted from vegetative parts, floral volatiles are reduced markedly soon after pollination.⁵ Such quantitative postpollination changes in floral bouquets decreases the attractiveness of these flowers and increases the overall reproductive success of the plant by directing pollinators to unpollinated flowers. This is particularly important for plants with a low visitation rate, in which reproductive success is mostly pollinator limited.⁶ The direct effects of floral volatiles on pollinator attraction have been observed by many researchers. However, attracting pollinators may increase the probability of attracting herbivores, and thus, fragrance bouquets may also be defensive. Junker and Bluthgen demonstrated that floral volatiles repel potentially nectar-thieving ants and suggested that floral volatiles may function as allomones against enemies, as well as synomones to attract mutualists.⁷ There is a growing appreciation for the indirect effects of floral volatiles, particularly for its unexpected impact on the visual discrimination by color-sensitive foragers, such as bumble bees and diurnal hawkmoths.^{8–10} Moreover, floral volatiles may play a key role as a selective attractant in plants

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with specialized pollination systems, particularly in cases where floral morphology does not function as a filter of flower visitors.¹¹ The indirect effects of floral volatiles, including flower or fruit destruction and modified pollinator behavior,¹² also result from their potential to attract floral enemies.

Insect visits/attacks change the quantity of volatiles emitted from either vegetative or floral plant parts, but less is known about the effects of insect visits/attacks on the stereochemical configurations of volatiles emitted from these parts. Our previous studies found that *Camellia sinensis* (tea) flowers contained phenylpropanoid/benzenoid compounds as their major internal volatiles,^{13–16} which are mostly involved in interactions between insects and flowers. 1-Phenylethanol (1PE) was identified as a major volatile phenylpropanoid/benzenoid compound in *C. sinensis* flowers. 1PE and its precursor acetophenone (AP) accounted for more than 60% of internal volatiles of *C. sinensis* flowers.¹³ In *C. sinensis* flowers, 1PE was the most major volatile compound with stereochemical configuration, while other volatiles with stereochemical configuration, for example linalool, occurred with little or trace amounts. Therefore, our study aimed to discover the mechanism of change of (R)-/(S)-1PE ratio after insect attacks. In the present study, we

found that an attack by *Thrips hawaiiensis* (Morgan) induced changes in the (R)-/(S)-1PE ratio emitted from tea flowers. *T. hawaiiensis* attack led to an increase in jasmonic acid (JA). Moreover, exposing flowers to exogenous JA induced a (R)-/(S)-1PE pattern that was similar to that emitted from flowers after an *T. hawaiiensis* attack. This study provides new insights into the involvement of volatiles in interactions between plants and insects, showing that not only the quantity but also the stereochemical configurations of plant volatiles could be affected by insects.

Results and discussion

T. hawaiiensis treatments induced significant changes in the (R)-/(S)-1PE ratio emitted from *C. Sinensis* flowers

To investigate the influences of insect attacks on the volatiles emitted by *C. sinensis* flowers, undamaged flowers and insect-damaged flowers (Fig. 1A) were collected from field experiments. The internal 1PE was reduced in insect-damaged flowers in contrast to undamaged flowers (Fig. 1B). However, insect attacks did not induce significant changes in the (R)-/(S) ratio of the internal 1PE. Interestingly, after insect attacks, the flowers

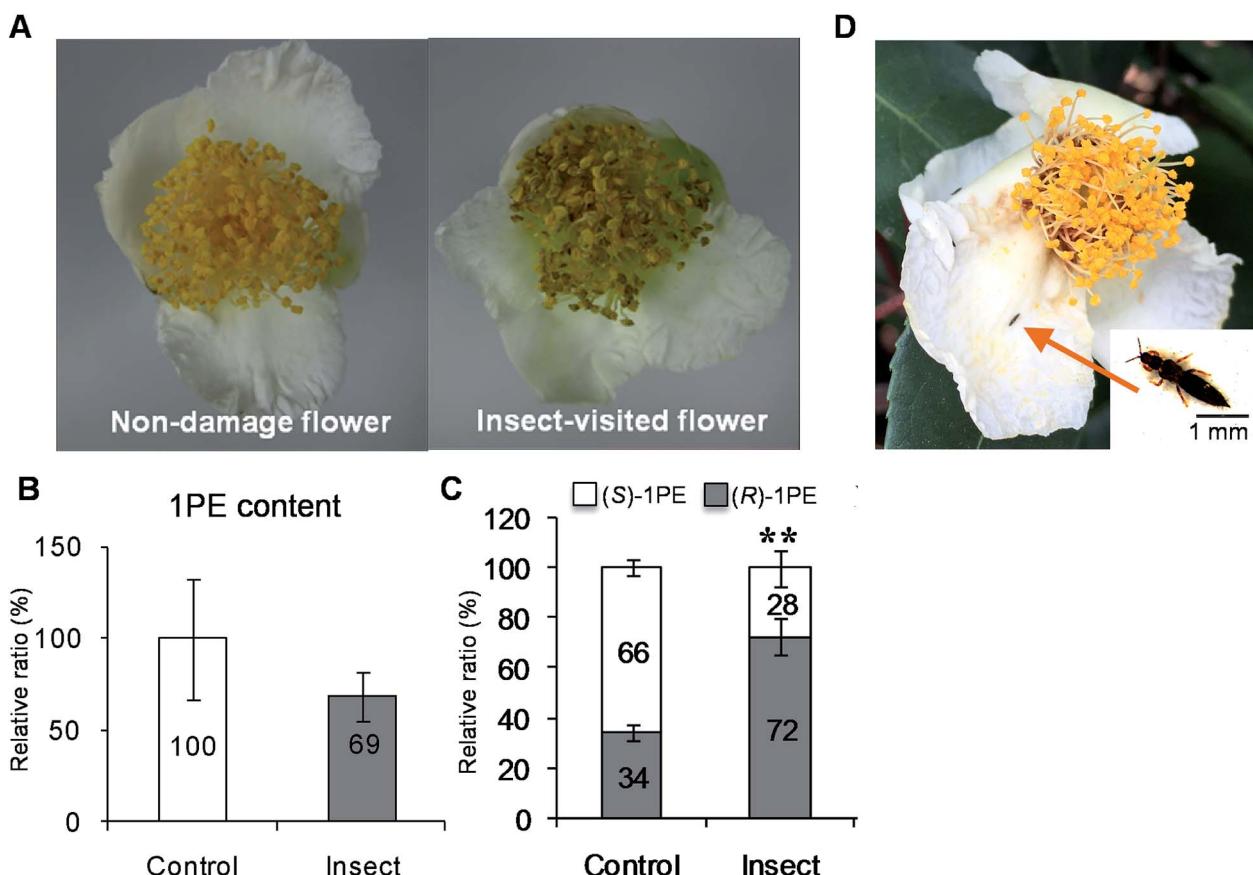


Fig. 1 Effect of insect attacks on phenotype and 1PE in *C. sinensis* flowers in field experiments. (A) The non-damage and insect-visited/damaged *C. sinensis* flowers. (B and C) Effects of insect attacks on internal 1PE content (B), and emitted (R)-/(S)-1PE ratio (C) in *C. sinensis* flowers. 1PE, 1-phenylethanol. Control, undamaged flowers. Insect, insect-damaged flowers. (B) The concentration of 1PE in control was set as 100%. (C) Significant differences between control and insect are indicated (** $p \leq 0.01$). (D) *Thrips hawaiiensis* (Morgan) was found to be major insect that attacked *C. sinensis* flowers in tea field. Data represent the mean value \pm standard deviation of three independent experiments performed in triplicate.

emitted a different (R) -/ (S) -1PE pattern compared with that of undamaged flowers (control) (Fig. 1C).

While investigating which insect attacks induced changes in the (R) -/ (S) -1PE ratio emitted from *C. sinensis* flowers, we found that *T. hawaiiensis* was the major insect attacking *C. sinensis* flowers in the field experiments (Fig. 1D). In laboratory experiments, adult *T. hawaiiensis* were used to treat *C. sinensis* flowers (Fig. 2A). After 6 h, the (R) -1PE level emitted from *C. sinensis* flowers increased (Fig. 2B), and the pattern was similar to that from insect-damaged flowers collected in the field experiments (Fig. 1C).

(A) Treatment of *T. hawaiiensis* on *C. sinensis* flowers. Control, undamaged flowers. Insect, *T. hawaiiensis*-damaged flowers. GC-MS, gas chromatography-mass spectrometry; UPLC-QTOF-MS, ultra-performance liquid chromatography quadrupole-time-of-flight mass spectrometry; SPME, solid-phase microextraction. (B) Effect of *T. hawaiiensis* attacks on emitted (R) -/ $(R + S)$ -1PE ratio in *C. sinensis* flowers. 1PE, 1-phenylethanol. The ratio of (R) -1PE to $(R + S)$ -1PE in control was set as 1. Significant differences between control and insect are indicated (* $p \leq 0.05$). (C) UPLC-QTOF-MS (negative) of phytohormones from authentic standards and extracts of *C. sinensis* flower. JA, jasmonic acid; ABA, abscisic acid; SA, salicylic acid. (D) Effect of *T. hawaiiensis* attacks on phytohormones contents in *C. sinensis* flowers. The contents of JA, ABA, and SA were calculated based on peak area of m/z 209.118 ($[\text{M} - \text{H}]^-$ of JA), m/z 263.128 ($[\text{M} - \text{H}]^-$ of ABA), and m/z 137.024 ($[\text{M} - \text{H}]^-$ of SA), respectively. The concentrations of phytohormones in control were set as 1. (E) Treatment of *C. sinensis* flowers using JA. JA, jasmonic acid. JA, 0.5 mM jasmonic acid treatment. (F) Effects of jasmonic acid treatment on emitted (R) -/ $(R + S)$ -1PE in *C. sinensis* flowers. The ratio of (R) -1PE to $(R + S)$ -1PE in control was set as 1. Significant differences between control and JA are indicated (** $p \leq 0.01$). Data represent the mean value \pm standard deviation of three independent experiments performed in triplicate.

In nature, floral structures, colors, and volatiles are critical factors in interactions between insects and flowers. Although floral structure and color provide important information for insects at a close range, floral volatiles can function as both long- and short-distance attractants and as a nectar guide to a variety of insects.¹⁸ The pollination of flowers by insects leads to changes in scent emission. These changes in scent emission after pollination include a general decrease in all of the scent compounds, as found in *Platanthera bifolia* (Orchidaceae),¹⁹ or a decrease in some compounds, while others increase or remain unchanged, as was discovered in *Ophrys* orchids.²⁰ However, it has not been reported that the stereochemical configurations of floral volatiles could be affected by insects, and there is only one report on vegetative volatiles. Allmann *et al.* reported that attacks by the specialist herbivore *M. sexta* larvae and the addition of their oral secretions to mechanical wounds induced a rapid (Z) / (E) isomeric change in the green leaf volatiles released from *Nicotiana* plants.⁴ This change lowered the (Z) / (E) ratio of the green leaf volatiles blend, resulting in an increase in the predation rate of predators that fed on the herbivore's eggs. These findings suggest that, in some cases, insect responses are dependent on the content of the volatile emission rather than merely the quantity. Our study

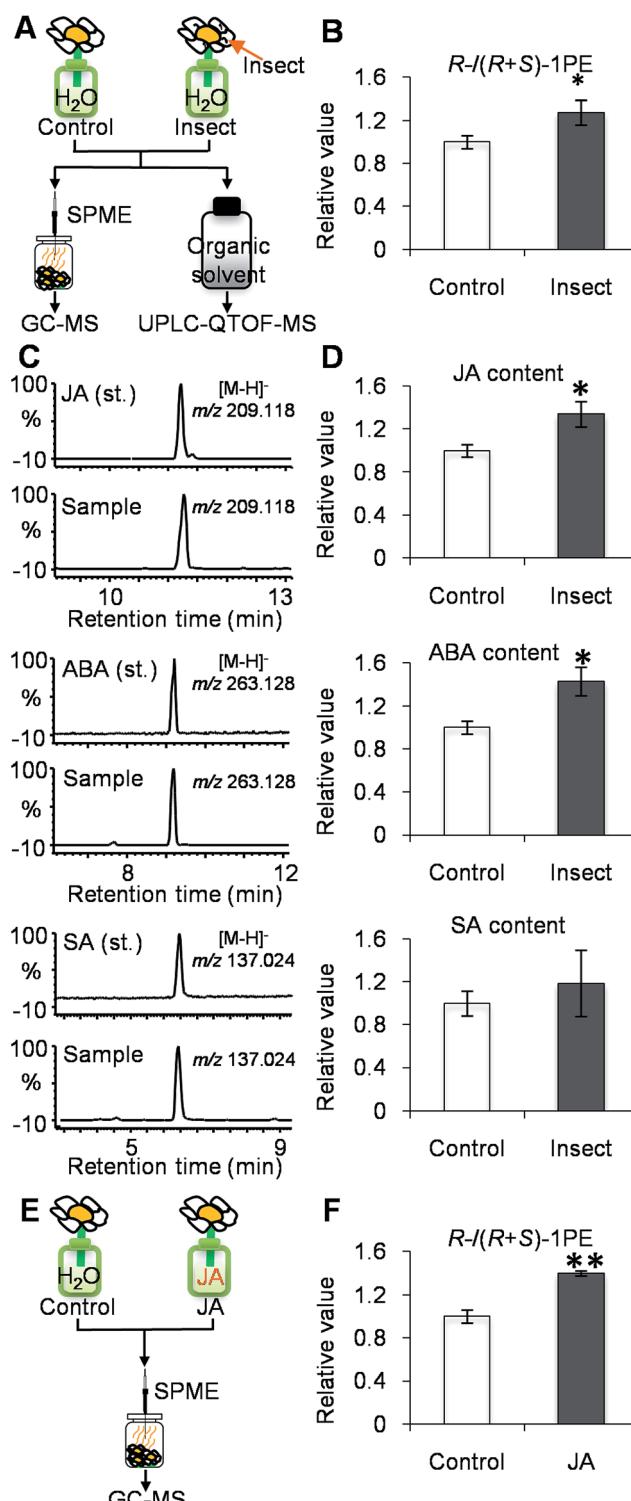


Fig. 2 Effect of *Thrips hawaiiensis* (Morgan) attacks on phytohormones and emitted (R) -/ (S) -1PE ratio in *C. sinensis* flowers in lab experiments.

indicated that such insect-induced changes in the stereochemical configuration of the volatiles occurred in not only vegetative parts but also in floral parts. Further studies on the effects of changes in the stereochemical configurations of floral volatiles on insects will help us understand the need for these flower responses.



JA was involved in changes in the (R)-/(S)-1PE ratio emitted from *C. sinensis* flowers induced by *T. hawaiiensis* treatment

To investigate if phytohormones are involved in changes in the (R)-/(S)-1PE ratio emitted from *C. sinensis* flowers induced

by *T. hawaiiensis* treatments, we first analyzed the levels of phytohormones, including JA, abscisic acid (ABA), and salicylic acid, in *C. sinensis* flowers 6 h after a *T. hawaiiensis* treatment. Using the UPLC-QTOF-MS technique, the three

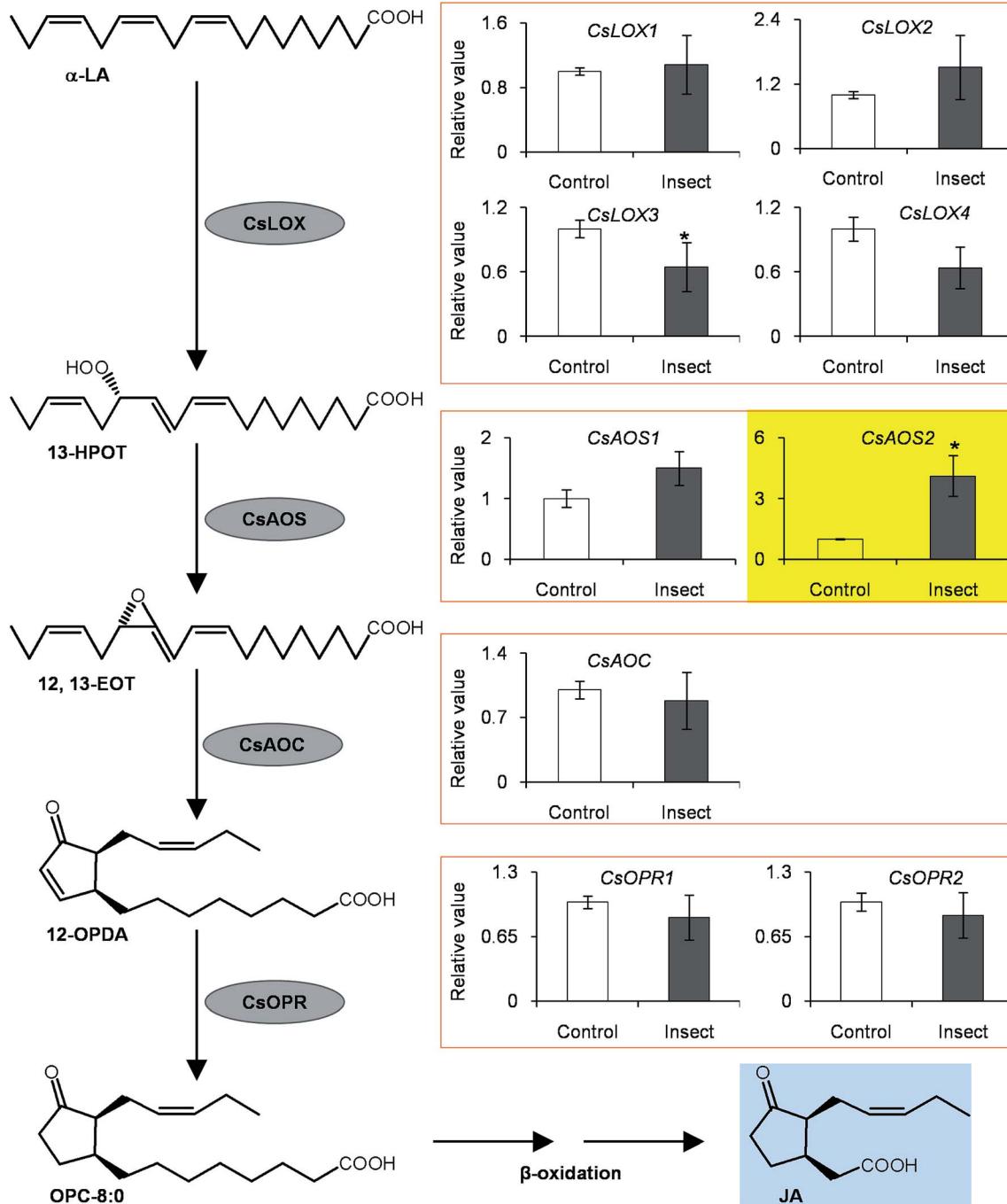


Fig. 3 Effect of *Thrips hawaiiensis* (Morgan) attacks on expression levels of jasmonic acid synthesis related genes in *C. sinensis* flowers. AOC, allene oxide cyclase; AOS, allene oxide synthase; 12,13-EOT, 12,13(S)-epoxy-9(Z),11,15(Z)-octa-decatrienoic acid; 13-HPOT, 13(S)-hydroperoxylinolenic acid; JA, jasmonic acid; LA, linolenic acid; LOX, 9/13-lipoxygenase; OPC-8:0, 3-oxo-2-[2'(Z)-pentenyl]-cyclopentane-1-octanoic acid; 12-OPDA, 12-oxo-phytodienoic acid; OPR, 12-oxo-phytodienoic acid reductase. Control, undamaged flowers. Insect, *T. hawaiiensis* -damaged flowers. Transcript abundance was calculated based on the difference in cycle threshold (C_t) values between target gene and internal reference gene transcripts by the normalized relative quantitation $2^{-\Delta\Delta C_t}$ method. The expression level in control was set as 1. Significant differences between control and insect are indicated (* $p \leq 0.05$). Data represent the mean value \pm standard deviation of three independent experiments performed in triplicate.

phytohormones were detected in *C. sinensis* flowers (Fig. 2C). The JA and ABA contents were significantly increased 6 h after the *T. hawaiiensis* treatment, while the SA content did not show a significant change (Fig. 2D). To further investigate if the phytohormones were involved in changes in the (*R*)-/(*S*)-1PE ratio emitted from *C. sinensis* flowers induced by the *T. hawaiiensis* treatment, JA and ABA, which were increased by the *T. hawaiiensis* treatment (Fig. 2D), were independently supplied to *C. sinensis* flowers. The exogenous ABA treatment did not significantly affect the (*R*)-/(*S*)-1PE ratio emitted from flowers (Fig. S1†), but the exogenous application of JA to the flowers induced an increase in the level of (*R*)-1PE, resulting in a (*R*)-/(*S*)-1PE pattern that was similar to that emitted from the insect-damaged flowers (Fig. 2E and F). We also investigated whether ethylene, which is generally reported to be involved in insect and floral volatiles, affected the (*R*)-/(*S*)-1PE ratio emitted from flowers. Like the ABA treatments, ethylene did not significantly affect the (*R*)-/(*S*)-1PE ratio emitted from flowers (Fig. S1†). Thus, JA was involved in the altered (*R*)-/(*S*)-1PE ratio emitted from *C. sinensis* flowers induced by the *T. hawaiiensis* treatments. We also investigated the effects of insect treatments on the expression levels of genes involved in JA biosynthesis, including 9/13-lipoxygenase, allene oxide synthase, allene oxide cyclase, 12-oxo-phytodienoic acid reductase. Only the *C. sinensis* allene oxide synthase 2 gene's expression was significantly activated 6 h after the *T. hawaiiensis* treatment (Fig. 3).

To investigate how *T. hawaiiensis* treatments affected the (*R*)-/(*S*)-1PE ratio emitted from *C. sinensis* flowers, we investigated the effects of *T. hawaiiensis* treatments on the formation of 1PE pathways. There are two pathways for the formation of 1PE. One pathway leads from the glycosides of 1PE to 1PE.¹⁴ 1PE β -primeverosides are the main glycosides of 1PE. β -primeverosidase is involved in the hydrolysis of 1PE β -primeverosides to form free 1PE, while glycosyl transferases are responsible for the transformation from free 1PE to 1PE β -primeverosides (Fig. 4A).^{14,21,22} The (*R*)-/(*S*)-1PE β -primeveroside ratio was not affected by *T. hawaiiensis* treatments (Fig. 4B). In addition, the β -primeverosidase (*CsPri*) and glycosyl transferases (*CsGT1* and *CsGT2*) expression levels were not significantly increased by *T. hawaiiensis* treatments (Fig. 4C–E). The other pathway leads from acetophenone to 1PE.¹³ Because the enzymes in this pathway are still unknown in plants, we used stable isotope-labeling acetophenone as a supplement given to the flowers. Then, *T. hawaiiensis* was co-treated with the [²H₅]AP supplement (Fig. 5A). The emitted [²H₅]1PE was collected, and (*R*)-[²H₅]1PE was increased by *T. hawaiiensis* attack (Fig. 5B), suggesting that the *T. hawaiiensis* treatment mainly affected the pathway leading from AP to 1PE, thus leading to a change in the emitted (*R*)-/(*S*)-1PE ratio.

Phytohormones are key links between insects and the formation of plant volatiles. JA is mostly reported to be involved in interactions between insects and volatiles from plant vegetative parts, while ethylene is mainly responsible for interactions between insects and volatiles from plant floral parts. There is considerable evidence that many postpollination processes in flowers, such as corolla abscission and/or

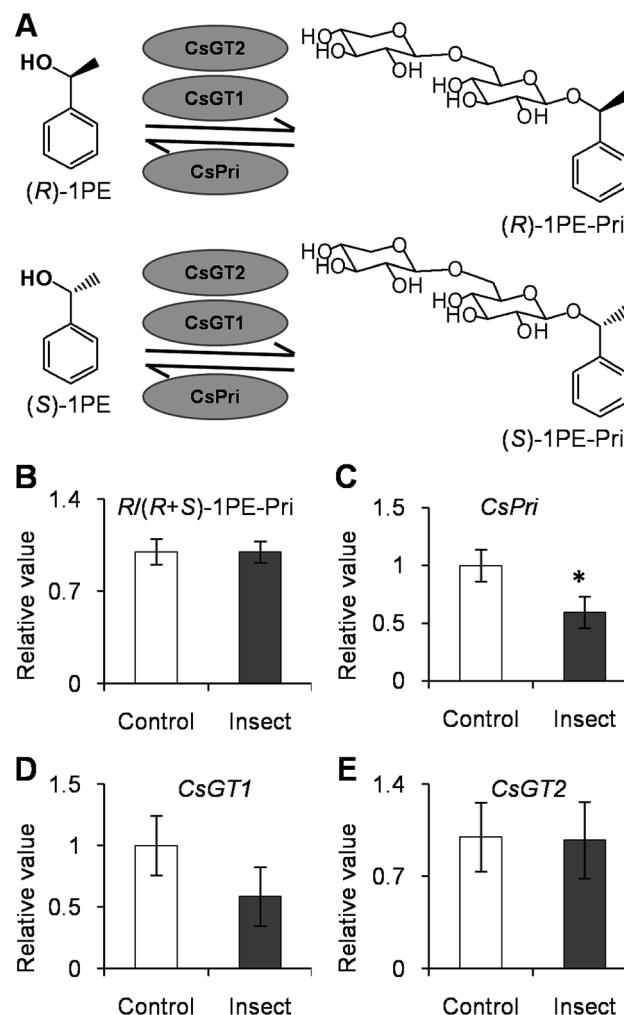


Fig. 4 Effect of *Thrips hawaiiensis* (Morgan) attacks on (*R*)-/(*R* + *S*)-1PE-Pri ratio and expression levels of *CsGT1*, *CsGT2*, and *CsPri* in *C. sinensis* flowers. (A) Schemes of transformation between (*R*)-/(*S*)-1PE-Pri and (*R*)-/(*S*)-1PE. 1PE, 1-phenylethanol; 1PE-Pri, 1PE- β -primeveroside; GT, glycosyltransferases; Pri, β -primeverosidase. (B) Effect of *T. hawaiiensis* attacks on (*R*)-/(*R* + *S*)-1PE-Pri in *C. sinensis* flowers. Control, undamaged flowers. Insect, *T. hawaiiensis*-damaged flowers. The ratio of (*R*)-1PE-Pri to (*R* + *S*)-1PE-Pri in control was set as 1. (C–E) Effect of *T. hawaiiensis* attacks on expression levels of *CsPri*, *CsGT1*, and *CsGT2* in *C. sinensis* flowers. Transcript abundance was calculated based on the difference in cycle threshold (*C_t*) values between target gene and internal reference gene transcripts by the normalized relative quantitation $2^{-\Delta\Delta C_t}$ method. The expression level in control was set as 1. Significant differences between control and insect are indicated (* $p \leq 0.05$). Data represent the mean value \pm standard deviation of three independent experiments performed in triplicate.

senescence, pigmentation changes in the perianth, and ovary maturation, are mediated by endogenous ethylene that is produced as a consequence of pollination.^{23–25} In petunia, pollination leads to an increase in ethylene synthesis.⁵ Pollination also affects floral scent emission.⁵ Moreover, there is growing evidence that in many cases, plant responses to ethylene are associated with changes in gene expression.^{5,26} In the present study, change in the (*R*)-/(*S*)-1PE ratio emitted from flowers was not due to ethylene (Fig. S1†), but resulted from JA



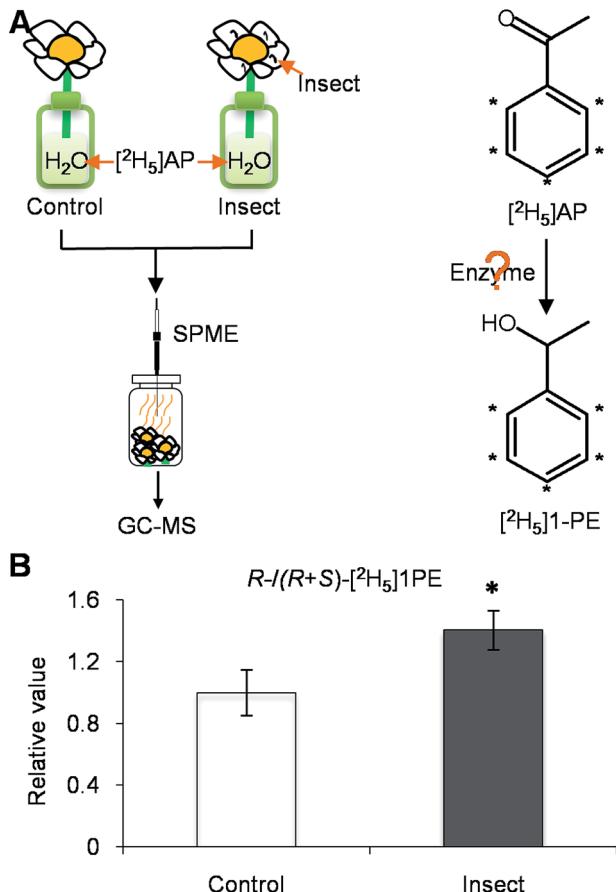


Fig. 5 Effect of *Thrips hawaiiensis* (Morgan) attacks on conversion of emitted labelled 1PE from labelled AP in *C. sinensis* flowers. (A) Treatment of *C. sinensis* flowers using insect. Control, undamaged flowers. Insect, *T. hawaiiensis*-damaged flowers. AP, acetophenone; GC-MS, gas chromatography-mass spectrometry; 1PE, 1-phenylethanol; SPME, solid-phase microextraction. (B) Effect of *T. hawaiiensis* attacks on emitted $(R)-/(R+S)-[\text{H}_5]\text{1PE}$ in *C. sinensis* flowers. Control, undamaged flowers. Insect, *T. hawaiiensis*-damaged flowers. The ratio of $(R)-[\text{H}_5]\text{1PE}$ to $(R+S)-[\text{H}_5]\text{1PE}$ in control was set as 1. Significant differences between control and insect are indicated (* $p \leq 0.05$). Data represent the mean value \pm standard deviation of three independent experiments performed in triplicate.

(Fig. 2F), suggesting that JA is another important phytohormone involved in volatile-mediated interactions between flowers and insects.

Conclusions

Insect attacks can lead to changes in the quantities of emitted plant volatiles, but the present study provides evidence that the stereochemical configurations of plant volatiles are also affected by insects. Moreover, JA, which is generally involved in relationships between insects and volatiles from vegetative plant parts, was found to also be responsible for changes in the stereochemical configuration of floral volatiles induced by insects (Fig. 6). The results obtained from this study will advance our understanding of the interactions between insects and plants mediated by volatiles.

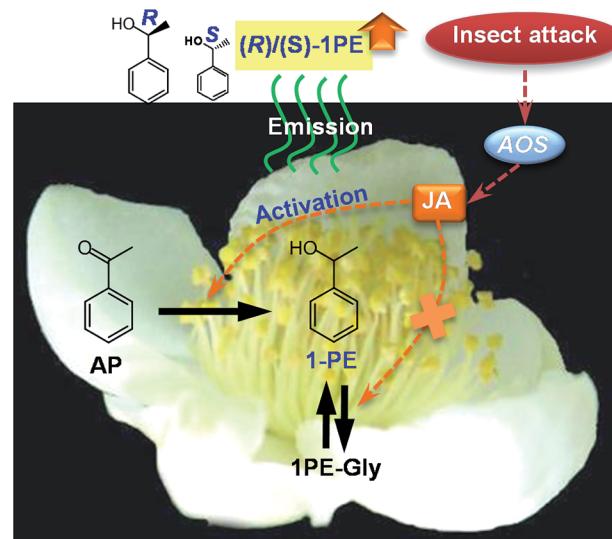


Fig. 6 Proposed schematic model of change in ratio of $(R)\text{-1PE}$ to $(S)\text{-1PE}$ emitted from *C. sinensis* flowers exposed to insect attacks. AP, acetophenone; 1PE, 1-phenylethanol; 1PE-Gly, glycosides of 1-phenylethanol; JA, jasmonic acid; AOS, allene oxide synthase.

Materials and methods

Chemicals

ABA was purchased from Aladdin Industrial Co. Shanghai, China. $[\text{H}_5]\text{AP}$ was purchased from Cambridge Isotope Laboratories Inc., Cambridge, MA. Amberlite XAD-2 column and polyvinylpolypyrrolidone (PVPP) were purchased from Sigma-Aldrich Co. Ltd., USA. Ethepron was purchased from Fluka Analytical, Shanghai, China. Ethyl *n*-decanoate, 1PE, $(R)\text{-1PE}$ and $(S)\text{-1PE}$ were purchased from Wako Pure Chemical Industries Ltd., Japan. Formic acid was purchased from CNW Technologies, Germany. JA was purchased from Tokyo Chemical Industry Co., Ltd., Japan. Methanol was purchased from EMD Millipore Corporation, Germany. *N*-Methyl-*N*-(trimethylsilyl)-trifluoroacetamide (MSTFA) was purchased from Regis Chemical Co., USA. Quick RNA Isolation Kit was purchased from Huayueyang Biotechnology Co., Ltd., China. $2\times$ SYBR Green Universal PCR Mastermix were purchased from Bio-Rad Laboratories, USA.

Plants and treatments

The nearly open flowers of *C. sinensis* var. Jinxuan were obtained from tea fields at the South China Agricultural University (Guangzhou, China) with stalks. The harvested flowers were reared in the laboratory to prevent insect attack for 16 h until fully-opened. *T. hawaiiensis* is a major pest founded in *C. sinensis* flowers in tea fields at the South China Agricultural University. They damage the flowers and suck the juice with rasping-mouthparts. In the laboratory experiments, the following three treatments were used: (1) insect treatments: five insects (adult *T. hawaiiensis*) were placed on a fully-opened flower that was in water. After treatment, the flowers were checked to make sure that the insects existed. The control was

an individual fully-opened flower that was in water. (2) Phytohormone treatments: the individual fully-opened flower was placed in a solution containing 0.5 mM JA, ABA, ethylene. The control was an individual fully-opened flower that was in water. (3) Stable labeling [$^2\text{H}_5$]AP, the precursor of 1PE,¹³ feeding treatment: five insects (adult *T. hawaiiensis*) were placed on a fully-opened flower that was in the 12 mM [$^2\text{H}_5$]AP solution. The control was an individual fully-opened flower that was in the 12 mM [$^2\text{H}_5$]AP solution. All of the treatments were performed for 6 h at room temperature. Every three flowers were used as one replicate for the collection of volatiles emitted from the flowers. Every 10 flowers were collected as one replicate for metabolite, phytohormone, and gene expression analyses. Each treatment was performed in triplicate.

Extraction of internal volatiles and collection of flower-emitted volatiles

To extract internal volatiles, 0.3 g finely powdered flowers was extracted with 2 mL CH_2Cl_2 containing 5 nmol ethyl *n*-decanoate as an internal standard. The extract was dried over anhydrous sodium sulfate. Then, 1 μL of the extract was subjected to a gas chromatography-mass spectrometry (GC-MS) analysis. To collect the emitted volatile of flowers, 250 mL beaker which contained three flowers were sealed with silver paper. The solid phase microextraction (divinylbenzene/carboxen/polydimethylsiloxane) was inserted into the beaker and the volatiles emitted from the flowers were collected by solid phase microextraction for 1 h at room temperature. The volatiles absorbed by the solid phase microextraction were analyzed using GC-MS.

Analysis of 1PE, (R)-1PE, and (S)-1PE by GC-MS

Samples were analyzed using GC-MS equipped with a SUPELCOWAX™ 10 column (Supelco Inc., 30 m \times 0.25 mm \times 0.25 μm). The injector temperature was 240 °C, the splitless mode was used with a splitless time of 1 min, and helium was the carrier gas at a flow rate of 1.0 mL min⁻¹ throughout the program. The column was used at an initial temperature of 40 °C, a ramp of 5 °C min⁻¹ to 250 °C, and then a hold at 250 °C for 15 min. The MS analyses were carried out in full scan (mass range *m/z* 70–200) and selected ion-monitoring mode (SIM) (for 1PE: *m/z* 77, 79, and 107). 1PE was identified by comparing their retention times and mass spectra with those of authentic compounds.

To determine the stereochemistry of 1PE [(*R*)-1PE, and (*S*)-1PE], a GC-MS equipped with an InertCap™ CHIRAMIX column (GL Sciences, 30 m \times 0.25 mm \times 0.25 μm) was employed. The injector temperature was 180 °C, the splitless mode was used with a splitless time of 1 min, and helium was the carrier gas at a flow rate of 1.0 mL min⁻¹ throughout the program. The GC temperatures were 60 °C for 2 min, ramp of 40 °C min⁻¹ to 105 °C, followed by 2 °C min⁻¹ to 137 °C, then 80 °C min⁻¹ to 180 °C, and finally 180 °C for 10 min. The MS was operated in SIM mode [for (*R*) or (*S*)-1PE: *m/z* 77, 79, 107, and 122; for [$^2\text{H}_5$] (*R*) or (*S*)-1PE: *m/z* 82, 84, 112, and 127]. (*R*)-1PE and (*S*)-1PE were

identified by comparing their retention times and mass spectra with those of authentic compounds.

Extraction and analysis of glycosides of 1PE

The methods used to extract and analyze the glycosides of 1PE were described previously.¹⁴ Briefly, 1 g of finely powdered flower was extracted with 6 mL methanol by vortexing for 30 s, followed by an ultrasonic extraction for 10 min. The extracts were mixed with 6 mL CHCl_3 and 2.4 mL H_2O for phase separation. After centrifugation for 5 min at 3500g, the upper phase was transferred to a new tube and dried under vacuum. The resultant extract was re-dissolved with 10 mL H_2O . The resulting solution was mixed with 600 mg PVPP, left standing for 60 min, and then centrifuged (16 400g, 4 °C, 10 min). The resulting supernatant was loaded on an Amberlite XAD-2 column and eluted successively with 15 mL water, 15 mL pentane : dichloromethane (2 : 1), and 10 mL methanol. The methanol eluate was concentrated in a vacuum, dried under nitrogen gas, and derivatized with 70 μL of MSTFA at 37 °C for 60 min. The MSTFA derivatives were analyzed by GC-MS equipped with an HP-5 column (30 m \times 0.25 mm \times 0.25 μm , Agilent Technologies, Germany). The injector temperature was 240 °C, the splitless mode was used with a splitless time of 1 min, and helium was the carrier gas with a velocity of 1 mL min⁻¹. The GC temperature was 80 °C for 2 min, then 15 °C min⁻¹ to 300 °C, followed by a hold at 300 °C for 10 min. The MS was operated in full-scan mode (*m/z* 150–550). The authentic standards (*R*)-1PE β -primeveroside and (*S*)-1PE β -primeveroside were obtained by chemical synthesis.¹⁴

Analysis of phytohormones

To analyze the phytohormones, 1 g of finely powdered flowers was extracted with 10 mL of 80% methanol overnight. The supernatant was dried under nitrogen gas to remove methanol. The residual water was adjusted to pH 2.8 with acetic acid and extracted with an equal volume of ethyl ether twice. The upper phase was combined and dried under nitrogen gas. The residual was dissolved in methanol, and phytohormones, including JA, ABA, and salicylic acid, were analyzed by an ultra-performance liquid chromatography/quadrupole time-of-flight mass spectrometry (UPLC-QTOF-MS) (Acquity UPLC I-Class/Xevo® G2-XS QTOF, Waters Corporation, Milford, MA, USA) equipped with an AQUITY UPLC BEH C18 column (Waters Corporation, 2.1 mm \times 100 mm \times 1.7 μm). Solvent A was distilled water with 0.1% (v/v) formic acid. Solvent B was methanol with 0.1% (v/v) formic acid. Elution was started using 30% solvent B for 4 min, then linearly increased to 65% within 15 min. The flow rate was 0.25 mL min⁻¹. The column temperature was 40 °C. The MS conditions were capillary voltage: 2.5 kV; source temperature: 100 °C; desolvation temperature: 350 °C; cone gas flow: 50 L h⁻¹; and desolvation gas flow: 600 L h⁻¹.

Transcript expression analysis of related genes

RNA was isolated from 500 mg flowers using a Quick RNA Isolation Kit (Huayueyang, Beijing, China). Then, 20 μL of cDNA



was synthesized from 500 ng of total RNA using PrimeScript™ RT reagent Kit (Takara, Dalian, China) according to the manufacturer's instructions. The reverse transcription condition was 37 °C for 15 min, followed by 85 °C for 5 s. The primers used in the quantitative real-time PCR (qRT-PCR) are listed in Table S1.† Elongation factor-1 α (EF-1 α) was used as an internal reference. Reactions were performed using SYBR® PrimeScript™ RT-PCR Kit (Perfect Real Time) (Takara, Dalian, China) in a 20 μ L volume containing 10 μ L SYBR® Premix Ex Taq™ (2 \times), 0.4 μ M each of the specific sense and antisense primers, and 2 μ L of 50 \times diluted template. The qRT-PCR was carried out on a Roche LightCycler 480 under conditions of 95 °C for 30 s, followed by 40 cycles of 95 °C for 3 s and 60 °C for 34 s. A melt curve was performed at the end of each reaction to verify PCR product specificity. The 2 $^{-\Delta\Delta C_t}$ method was used to calculate the relative expression levels.¹⁷

Statistical analysis

Statistical analysis was performed using the SPSS Ver. 19.0 software. The paired and independent samples *t*-tests were used to determine differences between two groups. A probability level of 5% ($p \leq 0.05$) was considered significant.

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Notes and references

- 1 F. Negre-Zakharov, M. C. Long and N. Dudareva, in *Plant-Derived Natural Products*, ed. A. E. Osbourn and V. Lanzotti, Springer Berlin, Heidelberg, 2009, vol. 19, pp. 405–431.
- 2 E. Pichersky and J. Gershenson, *Curr. Opin. Plant Biol.*, 2002, 5, 237–243.
- 3 F. Dong, X. M. Fu, N. Watanabe, X. G. Su and Z. Y. Yang, *Molecules*, 2016, 21, 124, DOI: 10.3390/molecules21020124.
- 4 S. Allmann and I. T. Baldwin, *Science*, 2010, 329, 1075–1078.
- 5 F. Negre, C. M. Kish, J. Boatright, B. Underwood, K. Shibuya, C. Wagner, D. G. Clark and N. Dudareva, *Plant Cell*, 2003, 15, 2992–3006.
- 6 M. R. M. Neiland and C. C. Wilcock, *Am. J. Bot.*, 1998, 85, 1657–1671.
- 7 R. R. Junker and N. Bluthgen, *Evolutionary Ecology Research*, 2008, 10, 295–308.
- 8 J. Kunze and A. Gumbert, *Behav. Chem. Ecol.*, 2001, 12, 447–456.
- 9 R. J. Gegear and T. M. Laverty, *Anim. Behav.*, 2005, 69, 939–949.
- 10 A. Balkenius and A. Kelber, *Naturwissenschaften*, 2006, 93, 255–258.
- 11 A. Shuttleworth and S. D. Johnson, *Ann. Bot.*, 2009, 103, 715–725.
- 12 N. B. Theis, M. Lerdau and R. A. Raguso, *Int. J. Plant Sci.*, 2007, 168, 587–601.
- 13 F. Dong, Z. Y. Yang, S. Baldermann, Y. Kajitani, S. Ota, H. Kasuga, Y. Imazeki, T. Ohnishi and N. Watanabe, *J. Plant Physiol.*, 2012, 169, 217–225.
- 14 Y. Zhou, F. Dong, A. Kunimasa, Y. Q. Zhang, S. H. Cheng, J. M. Lu, L. Zhang, A. Murata, F. Mayer, P. Fleischmann, N. Watanabe and Z. Y. Yang, *J. Agric. Food Chem.*, 2014, 62, 8042–8050.
- 15 Y. Zhou, L. Zhang, J. D. Gui, F. Dong, S. H. Cheng, X. Mei, L. Y. Zhang, Y. Q. Li, X. G. Su, S. Baldermann, N. Watanabe and Z. Y. Yang, *Plant Mol. Biol. Rep.*, 2015, 33, 253–263.
- 16 F. Dong, Y. Zhou, L. T. Zeng, Q. Y. Peng, L. Zhang, X. G. Su, N. Watanabe and Z. Y. Yang, *Molecules*, 2016, 21, 1106, DOI: 10.3390/molecules21091106.
- 17 K. J. Livak and T. D. Schmittgen, *Methods*, 2001, 25, 402–408.
- 18 H. E. M. Dobson, in *Insect-Plant Interactions*, ed. E. A. Bernays, CRC Press, Boston, 1994, vol. 5, pp. 46–81.
- 19 L. Tollsten, *Nord J. Bot.*, 1993, 13, 495–499.
- 20 F. P. Schiestl and M. Ayasse, *Oecologia*, 2001, 126, 531–534.
- 21 J. D. Gui, X. M. Fu, Y. Zhou, T. Katsuno, X. Mei, R. F. Deng, X. L. Xu, L. Zhang, F. Dong, N. Watanabe and Z. Y. Yang, *J. Agr. Food Chem.*, 2015, 63, 6905–6914.
- 22 S. Ohgami, E. Ono, M. Horikawa, J. Murata, K. Totsuka, H. Toyonaga, Y. Ohba, H. Dohra, T. Asai, K. Matsui, M. Mizutani, N. Watanabe and T. Ohnishi, *Plant Physiol.*, 2015, 168, 464–477.
- 23 D. G. Clark, C. Richards, Z. Hilioti, S. Lindversen and K. Brown, *Plant Mol. Biol.*, 1997, 34, 855–865.
- 24 J. Q. Wilkinson, M. B. Lanahan, D. G. Clark, A. B. Bleecker, C. Chang, E. M. Meyerowitz and H. J. Klee, *Nat. Biotechnol.*, 1997, 15, 444–447.
- 25 M. Kato, H. Shimizu, T. Onozaki, N. Tanikawa, H. Ikeda, T. Hisamatsu and K. Ichimura, *J. Jpn. Soc. Hortic. Sci.*, 2002, 71, 385–387.
- 26 K. Weterings, M. Pezzotti, M. Cornelissen and C. Mariani, *Plant Physiol.*, 2002, 130, 1190–1200.

