

Cite this: *RSC Adv.*, 2017, 7, 24129

Received 9th March 2017

Accepted 26th April 2017

DOI: 10.1039/c7ra02869e

rsc.li/rsc-advances

# Grandiflodines A and B, two novel diterpenoid alkaloids from *Delphinium grandiflorum*†

Neng-Hua Chen,<sup>ab</sup> Yu-Bo Zhang,<sup>ab</sup> Wen Li,<sup>ab</sup> Pan Li,<sup>ab</sup> Li-Feng Chen,<sup>ab</sup> Yao-Lan Li,<sup>ab</sup> Guo-Qiang Li<sup>\*c</sup> and Guo-Cai Wang<sup>\*ab</sup>

Two novel diterpenoid alkaloids, grandiflodines A and B (**1** and **2**), were isolated from *Delphinium grandiflorum*. Compound **1** represents a rare hetisine-type C<sub>20</sub>-diterpenoid alkaloid in which the bond between the atoms of N and C-17 is broken. Compound **2** features an unusual lycotonine-type C<sub>19</sub>-diterpenoid alkaloid skeleton with the cleavage of N–C<sub>19</sub> and C<sub>7</sub>–C<sub>17</sub> bonds, and the construction of the N–C<sub>7</sub> bond. Structural elucidations of the isolates were performed by spectroscopic analysis, X-ray diffraction and comparison with the literature. These compounds were tested for their antiviral and anti-inflammatory activities.

## Introduction

The genus *Delphinium* belongs to the family Ranunculaceae and consists of about 300 species distributed throughout the northern hemisphere.<sup>1,2</sup> Among the 300 species, more than 113 ones are endemic to China and about 18 ones are used as folk medicines.<sup>1,2</sup> As an important medicinal plant, *Delphinium* plants are used to treat traumatic injury, analgesia and rheumatism, *etc.*<sup>1</sup> Recent investigations showed that the diterpenoid alkaloids are the main components of *Delphinium* plants, and the alkaloids possess complex structure skeletons and exhibit a wide spectrum of pharmacological activities.<sup>3–7</sup> Thus, the diterpenoid alkaloids have become an increasing, attractive target for medicinal chemists.<sup>8</sup>

*Delphinium grandiflorum* is a perennial herb mainly distributed in the Northwest of China and some regions of Siberia and

People's Republic of Mongolia.<sup>9</sup> As a folk medicine, the *D. grandiflorum* is applied for the treatment of toothache, and used as native pesticide as well.<sup>9</sup> As part of our ongoing research on the bioactive natural products from *Delphinium* plants,<sup>10</sup> an extensive phytochemical investigation on *D. grandiflorum* was undertaken, leading to the isolation of two novel diterpenoid alkaloids, grandiflodines A and B (**1** and **2**). Compound **1** is a rare hetisine-type C<sub>20</sub>-diterpenoid alkaloid with the cleavage of the bond between the atoms of N and C-17. Compound **2** features an unusual lycotonine-type C<sub>19</sub>-diterpenoid alkaloid skeleton with the cleavage of N–C<sub>19</sub> and C<sub>7</sub>–C<sub>17</sub> bonds, and construction of the N–C<sub>7</sub> bond. Herein, we report the isolation, structure elucidation and biological activities of **1** and **2** (Fig. 1).

## Results and discussion

Grandiflodine A (**1**) was isolated as colorless block crystal. The molecular formula of **1** was established as C<sub>22</sub>H<sub>28</sub>N<sub>2</sub>O<sub>3</sub> by its HR-ESI-MS ( $m/z$  369.2175 [M + H]<sup>+</sup>, calcd for C<sub>22</sub>H<sub>29</sub>N<sub>2</sub>O<sub>3</sub>: 369.2173). The UV spectrum of **1** displayed the absorption maxima at 208 nm, and its IR spectrum showed the characteristic absorptions for hydroxyl groups (3479, 3423 cm<sup>−1</sup>), cyanogroup (2228 cm<sup>−1</sup>) and carbonyl group (1673 cm<sup>−1</sup>). The <sup>1</sup>H NMR and HSQC spectroscopic data of **1** provided the resonances for two methyls [ $\delta_{\text{H}}$  1.18, 2.26 (each 3H, s);  $\delta_{\text{C}}$  25.6, 33.4], an olefinic methylene [ $\delta_{\text{H}}$  4.51, 4.67 (each 1H, d,  $J$  = 1.8 Hz);  $\delta_{\text{C}}$  103.4] and an oxygenated methine [ $\delta_{\text{H}}$  3.36 (1H, t,  $J$  = 5.8 Hz);  $\delta_{\text{C}}$  72.9]. The <sup>13</sup>C and DEPT NMR data exhibited 22 signals of two methyls, seven methylenes, six methines and seven quaternary carbons, including a cyanogroup ( $\delta_{\text{C}}$  117.6), a pair of double bond ( $\delta_{\text{C}}$  103.4, 150.3) and a carbonyl group ( $\delta_{\text{C}}$  216.9). Detailed comparison of the <sup>1</sup>D NMR data of **1** (Table 1) with those of anhydroignavinol<sup>11</sup> showed that they were similar except for the presence of additional carbonyl and cyanogroup, and the

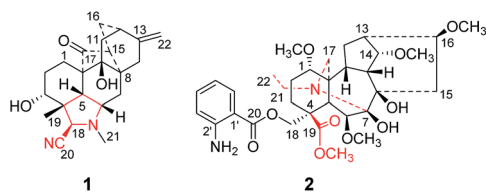


Fig. 1 Chemical structures of compounds **1** and **2**.

<sup>a</sup>Institute of Traditional Chinese Medicine & Natural Products, College of Pharmacy, Jinan University, Guangzhou 510632, P. R. China. E-mail: twangguocai@jnu.edu.cn

<sup>b</sup>Guangdong Province Key Laboratory of Pharmacodynamic Constituents of TCM and New Drugs Research, Jinan University, Guangzhou 510632, P. R. China

<sup>c</sup>Foshan University, Foshan, 528000, P. R. China. E-mail: liguogiang@jnu.edu.cn

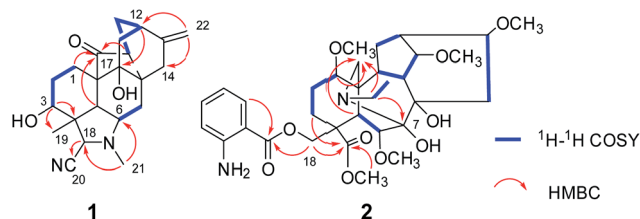
† Electronic supplementary information (ESI) available: UV, IR, HRESIMS and NMR spectra of compounds **1** and **2**. CCDC 1517870. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c7ra02869e



**Table 1** NMR spectroscopic data for grandiflodines A (**1**) and B (**2**) ( $\delta$  in ppm)

Position	Grandiflodine A ( <b>1</b> ) <sup>a</sup>		Grandiflodine A ( <b>2</b> ) <sup>b</sup>	
	$\delta_C$	$\delta_H$ ( <i>J</i> in Hz)	$\delta_C$	$\delta_H$ ( <i>J</i> in Hz)
1	23.2	1.62	88.9	2.88 dd (10.8, 4.1)
2	26.6	1.06 m	21.7	1.98 m
3	72.9	2.60 m	30.1	1.75 m
4	47.7	1.62	—	2.34 m
5	53.5	3.36 t (5.8)	—	1.44 m
6	58.6	—	49.8	—
7	31.2	2.30	49.9	2.02 d (6.7)
8	—	1.75 dd (14.8, 4.5)	90.5	3.71 m
9	39.6	—	87.8	—
10	76.5	—	—	—
11	51.7	—	82.0	—
12	38.7	1.86	39.9	2.30 m
13	—	1.38 dd (14.1, 2.6)	51.3	1.89 m
14	35.9	2.16	42.7	—
15	150.3	—	29.6	1.83 m
16	32.7	1.62	—	1.35 m
17	—	1.56 dd (12.8, 4.3)	45.3	2.43 m
18	49.2	2.30	85.3	3.66 t (3.9)
19	—	2.37 m	33.2	2.37 m
20	31.2	1.70 d (14.5)	—	1.57 dd (13.8, 8.1)
21	216.9	2.16	84.2	3.12 m
22	55.3	3.81 s	42.3	2.99 d (11.0)
3-OH	25.6	1.18 s	70.5	2.65 d (11.0)
9-OH	117.6	—	4.66 d (10.9)	4.66 d (10.9)
1-OCH <sub>3</sub>	33.4	2.26 s	43.7	4.15 d (10.9)
6-OCH <sub>3</sub>	—	—	—	—
14-OCH <sub>3</sub>	—	—	—	—
16-OCH <sub>3</sub>	—	—	—	—
19-OCH <sub>3</sub>	—	—	—	—
1'	—	—	174.9	—
2'	—	—	167.7	—
3'	—	—	—	—
4'	—	—	—	—
5'	—	—	—	—
6'	—	—	—	—
1-OCH <sub>3</sub>	—	—	56.8	3.24, s
6-OCH <sub>3</sub>	—	—	61.3	3.62, s
14-OCH <sub>3</sub>	—	—	57.9	3.38, s
16-OCH <sub>3</sub>	—	—	56.4	3.28, s
19-OCH <sub>3</sub>	—	—	52.1	3.72, s
1'	—	—	110.7	—
2'	—	—	150.8	—
3'	—	—	117.0	6.64, m
4'	—	—	134.4	7.24, m
5'	—	—	116.5	6.58, m
6'	—	—	131.0	7.71, dd (8.0, 1.4)

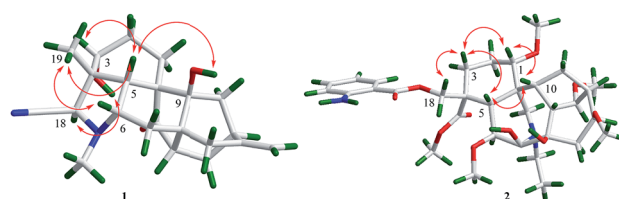
<sup>a</sup> Measured at 500/125 MHz in DMSO-*d*<sub>6</sub>. <sup>b</sup> Measured at 300/75 MHz in CDCl<sub>3</sub>. Overlapped signals are reported without designating multiplicity.

**Fig. 2** Key <sup>1</sup>H–<sup>1</sup>H COSY and HMBC correlations of **1** and **2**.

absence of two oxygenated methines in **1**. In the HMBC spectrum, the correlations (Fig. 2) between H-1/H-5/H-15 and the carbonyl group ( $\delta_C$  216.9) revealed that the carbonyl group was located at C-17. Moreover, the HMBC correlations between H-21 [ $\delta_H$  2.26, (3H, s)] and C-6/C-18 suggested that the methyl ( $\delta_C$  33.4, C-21) was connected to the nitrogen atom. The above information implied that the N–C<sub>17</sub> bond was broken to form a unique hetidines-type C<sub>20</sub>-diterpenoid alkaloid skeleton as depicted. In addition, the cyanogroup ( $\delta_C$  117.6) was located at C-18 based upon the HMBC correlation between H-18 and C-20 ( $\delta_C$  117.6). And the HMBC correlations between H-3 ( $\delta_H$  3.36, 1H, t, *J* = 5.8 Hz) and C-4/C-18/C-19, between H-11/H-12/H-15/H-16 and C-9 ( $\delta_C$  76.5) indicated that the carbons at C-3 and C-9 were substituted by hydroxyls, respectively. In light of the evidences mentioned above, the planar structure of **1** was finally established.

The relative configuration of **1** could be elucidated by the NOESY experiment. The correlations (Fig. 3) between 3-OH and H-18, between H-19 and H-3/H-5/H-6, as well as between 9-OH and H-5 established the relative configuration of **1**. Finally, the structure and configuration were further elucidated by an X-ray diffraction analysis (Fig. 4). The final refinement of the Cu K $\alpha$  data resulted in a small flack parameter of –0.05 (6) allowing the assignment of the absolute configuration of **1** as 3*R*, 4*R*, 5*R*, 6*S*, 8*S*, 9*S*, 10*S*, 12*S*, 15*S*, 18*R*.

The molecular formula of **2** was deduced as C<sub>33</sub>H<sub>48</sub>N<sub>2</sub>O<sub>10</sub> by HR-ESI-MS at *m/z* 633.3387 [*M* + H]<sup>+</sup> (calcd for C<sub>33</sub>H<sub>49</sub>N<sub>2</sub>O<sub>10</sub>: 633.3382). The <sup>1</sup>H NMR spectrum of **2** displayed the signals of one *ortho*-substituted benzene ring at  $\delta_H$  7.71 (1H, dd, *J* = 8.0, 1.4 Hz), 7.24 (1H, m), 6.64 (1H, m), 6.58 (1H, m), and five methoxys at  $\delta_H$  3.72, 3.62, 3.38, 3.28, 3.24 (each 3H, s). The <sup>13</sup>C and DEPT NMR data displayed thirty-three carbon signals including six methyls, seven methylenes, thirteen methines and seven quaternary carbons. Detailed analysis of the <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) of **2** showed a number of similarities to those of anthranoyllycoctonine.<sup>12</sup> The most notable differences were the existence of an additional carbonyl ( $\delta_C$  174.9) and an

**Fig. 3** NOESY correlations of **1** and **2**.

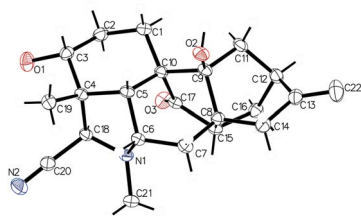


Fig. 4 Perspective drawing of the X-ray structure of 1.

additional methoxyl ( $\delta_C$  52.1) in 2. The HMBC correlations between H-3/H-18 and C-19 ( $\delta_C$  174.9), and between 19- $\text{OCH}_3$  ( $\delta_C$  52.1) and C-19 indicated that the N-C<sub>19</sub> bond was broken, and the carbon at C-19 was oxidized to be carbonyl. Furthermore, the HMBC correlations between H-1/H-5/H-21 and the methylene at C-17 ( $\delta_C$  42.3) revealed that the C<sub>7</sub>-C<sub>17</sub> linkage was broken. In addition, the correlation from H-21 to C-7 suggested that a new bond was constructed between the nitrogen atom and C-7. Hence, the planar structure of 2 was established. The relative configuration of 2 was the same as that of anthra-noyllycoctonine by interpretation of the NOESY data (Fig. 3).<sup>12</sup>

Compounds 1 and 2 were tested for their antiviral effect against the respiratory syncytial virus (RSV), and anti-inflammatory activity on Nitric Oxide (NO) production. Both the two compounds showed no cell cytotoxicity towards the tested cells with the  $\text{CC}_{50}$  values more than 100  $\mu\text{M}$ . Compound 2 displayed weak inhibitory effect on the growth of RSV and the production of NO in tested cells with the  $\text{IC}_{50}$  values of 75.3 and 72.7  $\mu\text{M}$ , respectively, and 1 was virtually inactive with  $\text{IC}_{50}$  values more than 100  $\mu\text{M}$ .

## Conclusions

In summary, compounds 1 and 2, two novel diterpenoid alkaloids were isolated from *D. grandiflorum*. Compound 1 represents a rare hetisine-type C<sub>20</sub>-diterpenoid alkaloid, and 2 features an unusual lycoctonine-type C<sub>19</sub>-diterpenoid alkaloid skeleton, revealing that the alkaloids in *Delphinium* plants possess complex structure skeletons and adding the diversity of alkaloid compositions isolated from *Delphinium* plants. Moreover, the assays of anti-RSV and anti-inflammatory activities showed that these two compounds had little cytotoxicity towards the tested cells, providing more potentiality for further pharmacologic study.

## Experimental section

### General

Melting point was obtained on an X-5 microscopic melting point apparatus. Optical rotations were recorded on a digital JASCO P-2000 polarimeter. UV spectra were obtained using a JASCO V-550 UV/VIS spectrophotometer. IR spectra were measured on a JASCO FT/IR-480 plus FT-IR spectrometer. NMR spectra were obtained by Bruker AV-500/300 spectrometers, with TMS as an internal standard. The chemical shifts ( $\delta$ ) were expressed in ppm and coupling constants ( $J$ ) in Hz. HR-ESI-MS

data was recorded on an Agilent 6210 ESI/TOF mass spectrometer. Analytical HPLC was performed using a Dionex ultimate 3000 system with a Cosmosil C<sub>18</sub> analytical column (5  $\mu\text{m}$ ,  $4.6 \times 250$  mm). Preparative HPLC was performed using an Agilent 1100 liquid chromatograph with a Cosmosil C<sub>18</sub> preparative column (5  $\mu\text{m}$ ,  $20 \times 250$  mm). Column chromatographies were performed with silica gel (80–100, 200–300, 300–400 mesh; Qingdao Marine Chemical Group Co. Ltd, Qingdao, China), ODS (50  $\mu\text{m}$ , 120 Å; YMC) and Sephadex LH-20 (Pharmacia Biotech, Uppsala, Sweden). Silica gel GF<sub>254</sub> plates (Yantai Chemical Industry Research Institute, Yantai, China) were used for thin-layer chromatography (TLC). Fractions were monitored by TLC, and spots were detected with modified Dragendorff's reagent.

### Plant material

The dried rhizomes of *D. grandiflorum* were purchased in Guangzhou, Guangdong Province of China, in July, 2015. The plant was authenticated by Prof. Guang-Xiong Zhou (College of Pharmacy, Jinan University). A voucher specimen (no. 150713) was deposited in the Institute of Traditional Chinese Medicine and Natural Products, Jinan University, Guangzhou, P. R. China.

### Extraction and isolation

The air-dried and powdered rhizome (10.0 kg) was extracted four times with 95% alcohol ( $4 \times 35$  L) at room temperature. After evaporation of alcohol, the crude extract (492.2 g) was suspended in water (2 L) and acidified with HCl to pH = 4–5, then partitioned with  $\text{CHCl}_3$  ( $3 \times 4$  L) to give a water-soluble fraction. The water-soluble fraction was basified with  $\text{NH}_3 \cdot \text{H}_2\text{O}$  to pH = 9–10 and then partitioned with a  $\text{H}_2\text{O}/\text{CHCl}_3$  mixture to give a  $\text{CHCl}_3$ -soluble fraction (63.0 g). The  $\text{CHCl}_3$ -soluble fraction was chromatographed on silica gel column (300–400 mesh, 1000 g) eluted with a solvent system of  $\text{CHCl}_3/\text{CH}_3\text{OH}$  (100 : 0 to 0 : 100, v/v), yielding six fractions (Fr.A–F). Fr.B (9.2 g) was further separated on an ODS column (200 g) eluted with  $\text{MeOH}/\text{H}_2\text{O}$  (30 : 70 to 100 : 0, v/v) to afford 9 sub-fractions (Fr.B1–B9). Fr.B5 (1.2 g) were purified by Sephadex LH-20 ( $\text{MeOH}/\text{CHCl}_3$ , 1 : 1, v/v) and compound 1 (15.0 mg) was crystallized from the eluent. Then 2 (12.3 mg) was obtained by the preparative HPLC with  $\text{MeOH}/\text{H}_2\text{O}$  (68 : 32, v/v) from Fr.B5.

**Grandiflodine A (1).** Colorless and block crystals ( $\text{MeOH}$ ); mp 273–274 °C;  $[\alpha]_D^{19} +6.8$  ( $c$  0.6, DMSO); UV ( $\text{MeOH}$ )  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 208.6 (3.57) nm; IR (KBr)  $\nu_{\text{max}}$  3480, 3423, 2933, 2876, 2228, 1674, 1462, 1057, 1054, 894  $\text{cm}^{-1}$ ;  $^1\text{H}$  and  $^{13}\text{C}$  NMR data see Table 1; HRESIMS  $m/z$  369.2175 (calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_3$ , 369.2173).

**Grandiflodine B (2).** White powder;  $[\alpha]_D^{25} +10.7$  ( $c$  0.96,  $\text{MeOH}$ ); UV ( $\text{MeOH}$ )  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 219.5 (3.78), 250.2 (3.30), 340.5 (3.18) nm; IR (KBr)  $\nu_{\text{max}}$  3455, 2931, 2874, 1677, 1453, 1360, 1189, 1055, 893  $\text{cm}^{-1}$ ;  $^1\text{H}$  and  $^{13}\text{C}$  NMR data see Table 1; HRESIMS  $m/z$  633.3382 (calcd for  $\text{C}_{33}\text{H}_{49}\text{N}_2\text{O}_{10}$ , 633.3387).

**X-ray crystallographic analysis of 1.** Colorless blocks,  $\text{C}_{22}\text{H}_{28}\text{N}_2\text{O}_3$ ,  $M_r = 368.46$ ; monoclinic, space group  $P2_1$ ;  $a = 9.7305$  (2) Å,  $b = 8.76406$  (16) Å,  $c = 11.0765$  (2) Å,  $\alpha = 90^\circ$ ,  $\beta = 109.45$  (2)°,  $\gamma = 90^\circ$ ;  $V = 890.68$  (3) Å<sup>3</sup>,  $Z = 2$ ,  $d_x = 1.374$  Mg  $\text{m}^{-3}$ ,



$F(000) = 396.0$ ,  $\mu(\text{Cu K}\alpha) = 0.731 \text{ mm}^{-1}$ . Data collection was performed on a Gemini S Ultra using graphite-monochromated radiation ( $\lambda = 1.54184 \text{ \AA}$ ); 2829 unique reflections were collected to  $\theta_{\text{max}} = 125.536^\circ$ , where 14 056 reflections were observed [ $F_2 > 2\sigma(F_2)$ ]. The structure was solved by direct methods (SHELXS 97)<sup>13</sup> and refined by full-matrix least-squares on  $F_2$ . Final  $R = 0.0297$ ,  $R_w = 0.0840$ , and  $S = 1.111$ . Crystallographic data for these structures have been deposited with the Cambridge Crystallographic Data Center as CCDC 1517870 for compound **1**.

### Assay of anti-RSV activities on Hep-2 cells

The human larynx epidermoid carcinoma (Hep-2, ATCC CCL-23) cells and RSV A2 (ATCC VR-1540) strains were purchased from Medicinal Virology Institute, Wuhan University, China. Hep-2 cells were cultured in DMEM (Gibco) supplemented with  $100 \text{ U mL}^{-1}$  penicillin and streptomycin solution, and virus was propagated in Hep-2 cells and incubated in DMEM with  $2 \text{ mM}$  L-glutamine, 2% FBS, and  $100 \text{ U mL}^{-1}$  penicillin and streptomycin solution. All of the cells were cultured in a 95% humidified atmosphere supplied with 5%  $\text{CO}_2$  at  $37^\circ\text{C}$ , and the ribavirin (Sigma, purity of 99%) was used as the positive control. The cytotoxicity of the compounds toward Hep-2 cells was detected by the MTT assay in 96-well plates (Corning) with the optical density (OD) values measured in an enzyme immunoassay reader (Thermo Labsystems Multiskan MK3) at  $570 \text{ nm}$ , and the 50% cytotoxic concentration ( $\text{CC}_{50}$ ) was estimated by regression analysis. The antiviral activities of the isolates against the RSV-A2 strain were assessed by the CPE reduction assay as reported in previous paper.<sup>14</sup> The concentration that reduces 50% of CPE with respect to the virus control was estimated from the plots of the data and was defined as the 50% inhibitory concentration ( $\text{IC}_{50}$ ) of the tested compounds.

### Assay of anti-inflammatory activities on NO production toward RAW 264.7 cells

RAW 264.7 cells were provided by the Medicinal Virology Institute of Wuhan University and maintained in DMEM (Gibco) containing 10% FBS (Gibco), and supplemented with  $100 \text{ U mL}^{-1}$  penicillin and streptomycin solution. Cells were cultured at  $37^\circ\text{C}$  in a 95% humidified atmosphere supplied with 5%  $\text{CO}_2$ . The cytotoxicity of the compounds on RAW 264.7 cells was detected by the MTT assay in 96-well plates with the OD values measured at  $570 \text{ nm}$ , and the  $\text{CC}_{50}$  was estimated by regression analysis. The anti-inflammatory activities of the compounds were evaluated by the inhibitory effect on NO production. RAW 264.7 cells ( $4 \times 10^4$  cells per well) were incubated in a 96-well plate for 14 h and then pretreated with  $100 \text{ ng mL}^{-1}$  LPS and different concentrations of compounds ( $6.25$ –

$100 \text{ }\mu\text{M}$ ) for 24 h. Then, the Griess reagent ( $100 \text{ }\mu\text{L}$ ) was added and blended with the supernatant ( $100 \text{ }\mu\text{L}$ ), and the absorbance was measured at  $540 \text{ nm}$  with an enzyme immunoassay reader. NO levels were determined *via* a calibration curve constructed with  $\text{NaNO}_2$  concentrations of  $3.12$ – $100 \text{ }\mu\text{M}$ . Inhibitory effects of compounds on NO production ( $\text{IC}_{50}$ ) were calculated by regression analysis of the dose–response curve generated from the data.

## Acknowledgements

This work was supported by the National Natural Science Foundation (No. 81473116, 81673319), and Science and Technology Planning Project of Guangdong Province (No. 2016B030301004, 2016A030303011).

## Notes and references

- 1 K. J. Guan, *Editorial committee of Flora of China*, Science Press, Flora of China, Beijing, 1979, p. 326.
- 2 Y. Q. He, Z. Y. Ma, X. M. Wei, D. J. Liu, B. Z. Du, B. H. Yao and L. M. Gao, *Chem. Biodiversity*, 2011, **8**, 2104–2109.
- 3 X. L. Zhou, D. L. Chen, Q. H. Chen and F. P. Wang, *J. Nat. Prod.*, 2005, **68**, 1076–1079.
- 4 J. G. Diaz, J. G. Ruiz and G. D. Fuente, *J. Nat. Prod.*, 2000, **63**, 1136–1139.
- 5 J. Li, D. L. Chen, X. X. Jian and F. P. Wang, *Molecules*, 2007, **12**, 353–360.
- 6 T. F. Xu, S. Liu, L. L. Meng, Z. F. Pi, F. R. Song and Z. Q. Liu, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2016, **1026**, 56–66.
- 7 F. P. Wang, Q. H. Chen and X. Y. Liu, *Nat. Prod. Rep.*, 2010, **27**, 529–570.
- 8 F. Z. Chen, D. L. Chen, Q. H. Chen and F. P. Wang, *J. Nat. Prod.*, 2009, **72**, 18–23.
- 9 K. J. Guan, *Editorial committee of Flora of China*, Science Press, Flora of China, Beijing, 1979, p. 445.
- 10 L. Yang, Y. B. Zhang, L. Zhuang, T. Li, N. H. Chen, Z. N. Wu, P. Li, Y. L. Li and G. C. Wang, *Planta Med.*, 2017, **83**, 111–116.
- 11 S. W. Pelletier, S. W. Page and M. G. Newton, *Tetrahedron Lett.*, 1970, **55**, 4825–4827.
- 12 J. Lu, H. K. Desai, S. A. Ross, H. M. Sayed and S. W. Pelletier, *J. Nat. Prod.*, 1993, **56**, 2098–2103.
- 13 G. M. Sheldrick, *SHELXS-97: Manual of Program for the Refinement of Crystal Structures*, University of Göttingen, Germany, 1997.
- 14 H. W. Geng, X. L. Zhang, G. C. Wang, X. X. Yang, X. Wu, Y. F. Wang, W. C. Ye and Y. L. Li, *J. Asian Nat. Prod. Res.*, 2011, **13**, 665–669.

