



Cite this: RSC Adv., 2017, 7, 22699

Design and synthesis of uracil urea derivatives as potent and selective fatty acid amide hydrolase inhibitors[†]

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Fatty acid amide hydrolase (FAAH) is one of the key enzymes involved in the biological degradation of endocannabinoids, especially anandamide. Pharmacological blockage of FAAH restores the levels of endocannabinoids, providing therapeutic benefits in the management of inflammation, depression and multiple sclerosis. In this study, a series of uracil urea derivatives as FAAH inhibitors were designed and synthesized. Structural modifications at the C5 position and side chain of *N*-hexyl-2,4-dioxo-3,4-dihydropyrimidine-1(2*H*)-carboxamide (**1a**) led to FAAH inhibitors with improved potency and selectivity. Structure–activity relationship (SAR) studies indicated that C5 electron-withdrawing substituents were preferred for optimal potency but not for selectivity, whereas replacement of the alkyl chain with phenylalkyl moieties or biphenyl groups significantly improved both inhibitory potency and selectivity towards FAAH. Two highly potent picomolar FAAH inhibitors (**4c**, $IC_{50} = 0.3 \pm 0.05$ nM; **4d**, $IC_{50} = 0.8 \pm 0.1$ nM) were developed. Compound **4c** inhibited FAAH in a rapid, selective, noncompetitive, and irreversible pattern. This study provides several highly potent and selective FAAH inhibitors and an optimized chemical scaffold for the development of FAAH inhibitors. We anticipate that these FAAH inhibitors will enable new possibilities in understanding FAAH functions and development of therapeutics for pain and inflammatory diseases.

Received 23rd February 2017
 Accepted 17th April 2017

DOI: 10.1039/c7ra02237a
rsc.li/rsc-advances

1. Introduction

Fatty acid amide hydrolase (FAAH) is a member of the amidase signature family, responsible for the inactivation of endogenous cannabinoids, especially anandamide (AEA).^{1–3} AEA modulates a wide range of physiological processes through the interaction with G protein-coupled CB1 and CB2 cannabinoid receptors. Genetic and pharmacological blocking of FAAH leads to increased endogenous AEA levels without the undesirable “cannabinoid effects” caused by cannabinoid receptor agonists.^{4–6} Therefore, restoring the endogenous AEA by FAAH inhibition has become an alternative therapeutic strategy for inflammation, cancer, and neuropathic diseases.^{5,7–11}

So far, there are two main categories of FAAH inhibitors that have been developed.^{12–14} First, reversible inhibitors, such as the α -ketoheterocycle-based inhibitors, bind to the active serine residue of FAAH with a reversible hemiketal bond. OL-135 is the most well-studied potent and reversible FAAH inhibitor.^{4,15,16} Second, irreversible inhibitors, including aryl carbamates,^{17–20} ureas^{21–25} and sulfonyl fluoride analogues,²⁶ which form covalent binding with the catalytic site of FAAH, *e.g.*, URB597.²⁷ Administration of URB597 to rats elevated endogenous AEA levels and exhibited anxiolytic effects in animal models.^{5,28} Additionally, benzothiazole,²⁹ β -lactam,³⁰ (thio)hydantoins³¹ and some nonsteroidal anti-inflammatory drugs (NSAIDs)^{32–34} also have been reported to inhibit FAAH.

Although the overall physiological role of FAAH has been reported, it is difficult to distinguish the function of FAAH in each specific tissue and organ. The reason is the wide distribution of FAAH in the whole body and lack of suitable topical FAAH inhibitors. Most of current FAAH inhibitors have low blood clearance and readily distributed and inhibit FAAH throughout the body after treatment. They are suitable for the systemic inhibition of FAAH, but it's difficult to obtain targeted FAAH inhibition in the specific tissues without inhibiting FAAH in other non-targeted sites. Given the critical pharmacological properties of FAAH, understanding the function of FAAH in different tissues is of great significance. FAAH inhibitors

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† Electronic supplementary information (ESI) available. See DOI: [10.1039/c7ra02237a](https://doi.org/10.1039/c7ra02237a)

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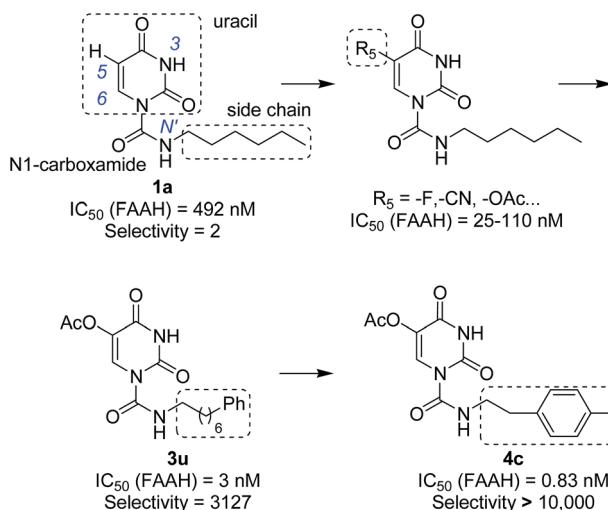


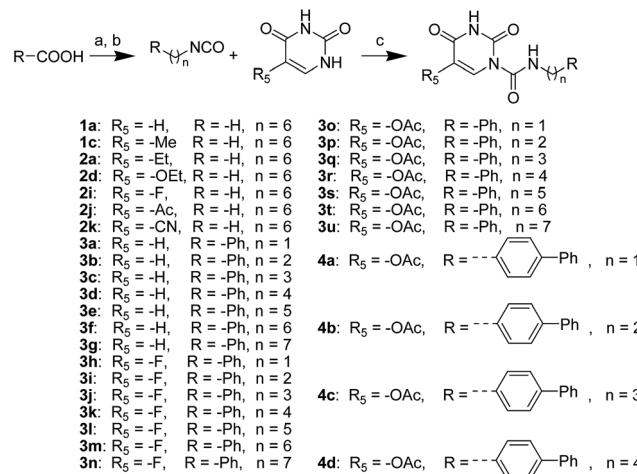
Fig. 1 The design strategies of uracil urea derivatives as highly potent and selective FAAH inhibitors. The selectivity represents the selectivity of uracil urea derivatives towards FAAH over NAAA.

suitable for the topical administration is highly desired to explore functions of FAAH in different tissues and organs.

In our previous study,³⁵ we found that uracil urea **1a** (Fig. 1), an acid ceramidase (AC) inhibitor ($IC_{50} = 426 \pm 104$ nM),³⁶ was also a potent FAAH ($IC_{50} = 492 \pm 53$ nM) and *N*-acylethanolamine-hydrolysing acid amidase (NAAA) inhibitor ($IC_{50} = 990 \pm 170$ nM). To develop FAAH inhibitors with properties suitable for topical FAAH inhibition and improve the selectivity of **1a** towards FAAH, we modified the uracil electronic properties and the side chain conformation of **1a**. A series of uracil urea derivatives designed as selective FAAH inhibitors were synthesized and characterized. The results showed that modification of **1a** targeting the uracil ring improved the inhibition potency towards FAAH, and the side chain replacement with phenylalkyl moieties or biphenyl groups enhanced both potency and selectivity (Fig. 1). Several highly potent and selective FAAH inhibitors were reported, including two pico-molar inhibitor, **4c** ($IC_{50} = 0.3 \pm 0.05$ nM) and **4d** ($IC_{50} = 0.8 \pm 0.1$ nM), a series of nanomolar inhibitors, *e.g.*, **3f** ($IC_{50} = 4 \pm 0.9$ nM), **3u** ($IC_{50} = 3 \pm 0.4$ nM), **4a** ($IC_{50} = 1.1 \pm 0.2$ nM) and **4b** ($IC_{50} = 8.3 \pm 1.2$ nM).

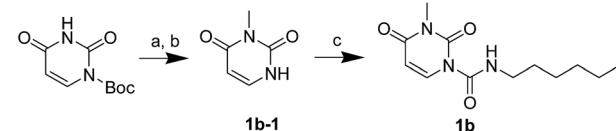
2. Chemistry

The uracil ureas **1a**, **1c**, **1d**, **2a**, **2d**, **2i-2k**, **3a-3u**, **4a-4d** were prepared by the reaction of isocyanates with 5-substituted uracils following a reported procedures with minor modification.³⁶ Isocyanates were synthesized *via* Curtius rearrangement of appropriate acylazides prepared from successive reactions of the corresponding acid with oxalyl chloride ($(COCl)_2$) and sodium azide (NaN_3) (Scheme 1).²⁰ **1b** and **1e** were synthesized following a literature procedure.³⁶ **1b** was prepared by the reaction of hexyl isocyanate with 3-methyl substituted uracil **1b-1**. **1b-1** was obtained by reaction of *N*-Boc uracil and iodomethane (MeI), and then deprotection with HCl in MeOH

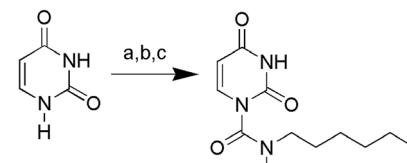


Scheme 1 The synthesis of compounds **1a**, **1c**, **2a**, **2d**, **2i-2k**, **3a-3u**, **4b-4d**. Reagents and conditions: (a) $(COCl)_2$, cat. DMF, CH_2Cl_2 , then NaN_3 in H_2O , 0 °C; (b) toluene, reflux; (c) DMAP, pyridine, 60 °C.

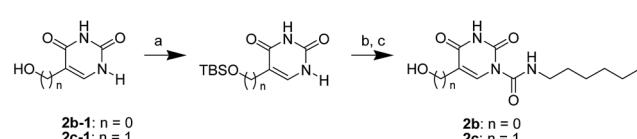
(Scheme 2). **1e** was synthesized by reaction of uracil with triphosgene followed by treatment with *N*-methylhexanamine (Scheme 3). **2b** and **2c** were prepared with 3-step procedure starting from **2b-1** and **2c-1**. Compound **2b-1** and **2c-1** were protected by *t*-butyl di-methylsilyl chloride (TBSCl) and directly used in the carbamoylation reaction, followed by deprotection



Scheme 2 The synthesis of compound **1b**. Reagents and conditions: (a) Mel, Cs_2CO_3 , THF, room temperature; (b) HCl, CH_3OH , room temperature; (c) hexyl isocyanate, DMAP, pyridine, 60 °C.

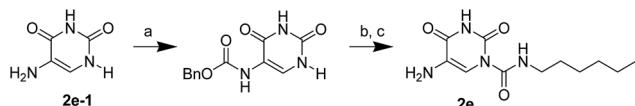


Scheme 3 The synthesis of compound **1e**. Reagents and conditions: (a) NaH , triphosgene, THF, room temperature; (b) HCl, CH_3OH , room temperature; (c) DMAP, pyridine, *N*-methylhexylamine, room temperature.

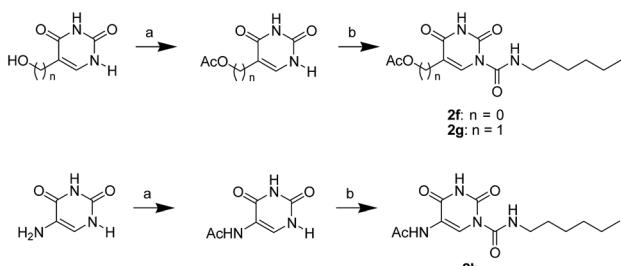


Scheme 4 The synthesis of compounds **2b** and **2c**. Reagents and conditions: (a) pyridine, TBSCl, room temperature; (b) hexyl isocyanate, DMAP, pyridine, 60 °C; (c) TBAF, THF , room temperature.

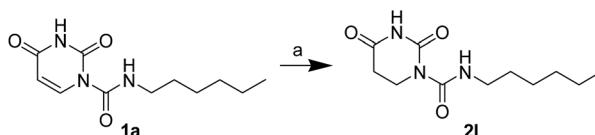




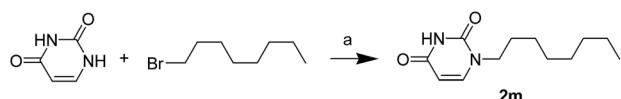
Scheme 5 The synthesis of compound **2e**. Reagents and conditions: (a) pyridine, CbzCl, room temperature; (b) hexyl isocyanate, DMAP, pyridine, 60 °C; (c) TMSI, THF, room temperature.



Scheme 6 The synthesis of compounds **2f**, **2g** and **2h**. Reagents and conditions: (a) pyridine, Ac₂O, room temperature; (b) hexyl isocyanate, DMAP, pyridine, 60 °C.



Scheme 7 The synthesis of compound **2l**. Reagents and conditions: (a) 10% Pd/C, H₂, CH₃OH, room temperature.



Scheme 8 The synthesis of compound **2m**. Reagents and conditions: (a) K₂CO₃, DMF, room temperature.

with tetrabutylammonium fluoride (TBAF) (Scheme 4). 5-Amino uracil (**2e-1**) protected with benzyl carbonochloride (CbzCl) in pyridine was directly used in the carbamoylation reaction, and followed by deprotection with trimethylsilyl iodide (TMSI) to afford **2e** (Scheme 5). **2f–2h** were synthesized by treatment of the uracil derivatives with acetic anhydride (Ac₂O) followed by reaction with isocyanates (Scheme 6). **2l** was prepared by catalytic hydrogenation of **1a** (Scheme 7). **2m** was obtained by reaction of 1-bromo-octane with uracil (Scheme 8).

3. Enzyme assay

The inhibition activity of new compounds against rat FAAH (rFAAH) and rat NAAA (rNAAA) was tested, using [³H]-AEA and d4-PEA, respectively, as substrates. The residual hydrolysis products of the substrates were determined by liquid chromatography-mass spectrometer (LC-MS).

4. Results and discussion

4.1. SAR study

It has been reported that carbamate and urea FAAH inhibitors interact with the nucleophilic Ser241 of FAAH through the electrophilic carbonyl group, and most of these inhibitors contain O-aryl or N-aryl moiety as leaving group. We speculated that the activity of uracil urea inhibitors may depend on the presence of *N*1-carboxamide as the electrophilic carbonyl group and uracil as leaving group. Accordingly, two different chemical modifications were used to optimize the potency and selectivity of uracil urea derivatives. First, inspired by existed urea FAAH inhibitors, we replaced the uracil ring with chemical groups with different electronic properties to increase the leaving group ability of uracil. Second, we modified the side chain with phenyl-containing groups that can mimic the conformation and π -unsaturation of double bonds of anandamide, and provided better drug-like characteristics and the opportunity for structure exploration (Fig. 1).

In preliminary SAR study, analogues (**1a–1e**, Table 1) bearing a methyl were prepared to define positions available for substitution without adversely affecting the inhibitor potency. The results showed that substitutions of position N3 (**1b**), C6 (**1d**) and N' (**1e**) of **1a** resulted in a greatly diminished or complete loss in inhibition toward FAAH and NAAA, while substitution of position C5 (**1c**) slightly decreased the inhibition against both FAAH and NAAA. Therefore, we focused on the modification of C5 in the following studies.

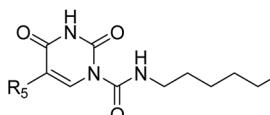
Next, we did systematic substitution of **1a** at C5 (**1c**, **2a–2k**) to investigate the effect of electronic properties of ring substitution on the inhibitory activity of uracil urea derivatives toward FAAH. Some analogues (**2a**, **2d**, **2i**, **2k**) have been described by other groups were also synthesized and tested as control compounds.^{36,37} The results suggested that the electronic properties of C5 substituents were of great significance for the inhibition towards both FAAH and NAAA (Table 2). Compared to parent inhibitor **1a**, the electron-withdrawing substituents (**2i–2k**) exhibited 2–5 folds enhanced inhibition against FAAH.

Table 1 Inhibitory effects of compounds **1a–1e** on rFAAH and rNAAA activities^a

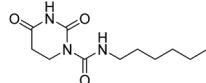
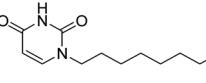
	R ₃	R ₅	R ₆	R _N	IC ₅₀ of FAAH (nM)	IC ₅₀ of NAAA (nM)
1a	–H	–H	–H	–H	492 \pm 53	990 \pm 170
1b	–Me	–H	–H	–H	7811 \pm 1563	>10 000
1c	–H	–Me	–H	–H	1026 \pm 205	3650 \pm 840
1d	–H	–H	–Me	–H	>10 000	>10 000
1e	–H	–H	–H	–Me	>10 000	>10 000

^a Data are presented as IC₅₀ \pm standard error of the mean. All experiments were performed in triplicate.

Table 2 Inhibitory effects of compounds **1a**, **1c**, **2a**–**2m** on rFAAH and rNAAA activities^a

				
	R ₅	IC ₅₀ of FAAH (nM)	IC ₅₀ of NAAA (nM)	Selectivity
1a	–H	492 ± 53	990 ± 170	2
1c	–Me	1026 ± 205	3650 ± 840	3.6
2a	–Et	543 ± 94	1837 ± 428	3.4
2b	–OH	693 ± 117	5049 ± 1103	7.3
2c	–CH ₂ OH	425 ± 75	3265 ± 519	7.6
2d	–OEt	703 ± 136	1048 ± 168	1.5
2e	–NH ₂	1520 ± 280	5265 ± 864	3.5
2f	–OAc	110 ± 10	190 ± 20	1.7
2g	–CH ₂ OAc	120 ± 31	271 ± 25	2.2
2h	–NHAc	221 ± 44	423 ± 78	1.9
2i	–F	38 ± 9	250 ± 50	6.6
2j	–Ac	42 ± 11	486 ± 89	11.5
2k	–CN	25 ± 5	560 ± 42	22.4
URB597		26 ± 5	>10 000	

^a Data are presented as IC₅₀ ± standard error of the mean. All experiments were performed in triplicate.

Structure	IC ₅₀ of FAAH (nM)	IC ₅₀ of NAAA (nM)
2l 	>10 000	>10 000
2m 	>10 000	>10 000

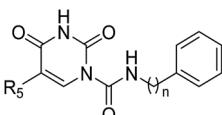
Among compounds **2a**–**2k**, **2i**–**2k** were the three most potent inhibitors, being equipotent with URB597 tested under the same conditions. On the contrary, the electron-donating substituents (**1c**, **2a**–**2e**) showed slightly decreased inhibitory activity against FAAH. We then tested whether further modulation of the electron-donating C5 substituents with electron-withdrawing group can restore the potency. The results showed that the addition of acetyl to the compounds **2f**–**2h** resulted in a modest (3–7 folds) increase of inhibitory potency compared to corresponding derivatives (**2b**, **2c** and **2e**). These results suggested that the electron-withdrawing C5 substitution was critical for potent FAAH inhibition. The reason might be such modulation increases the leaving group ability of uracil, and makes molecules more sensitive to nucleophilic attack of Ser241 of FAAH. To examine this hypothesis, two **1a** derivative was synthesized and tested. Derivative **2l** is the saturated derivative of **1a** without the leaving group uracil. Derivative **2m** without electrophilic carbonyl group was obtained by replacement of N1-carboxamide with methylene. The data showed that inhibitory activity of **2l** and **2m** against FAAH was completely eliminated. These results confirmed that uracil leaving group and N1-carboxamide group are critical for FAAH inhibition.

Except from enhancing inhibition potency towards FAAH, electron-withdrawing C5 substitution also increased the inhibition potency of uracil urea derivatives toward NAAA. For instance, acetyl substituents **2f**–**2h** enhanced NAAA inhibition by 12–27 folds compared to the unsubstituted derivatives (**2b**, **2c** and **2e**). Electron-withdrawing C5 substituents exhibited weak selectivity towards FAAH over NAAA. The electron-withdrawing substituents failed to improve the selectivity towards FAAH over acid ceramidase. The IC₅₀ values of electron-withdrawing substituents towards acid ceramidase (**2i**–**2k**) was an order of magnitude lower than **1a**.³⁶ These results suggested that electron-withdrawing substituents of uracil ring exhibited enhanced the inhibition potency, but their selectivity towards FAAH over acid ceramidase and NAAA was still not ideal. Further modification was required to enable high selectivity for FAAH over other competitive enzymes.

Finally, we further modified the side chain to improve the selectivity towards FAAH. FAAH hydrolyzes unsaturated substrates (e.g., anandamide) faster than saturated FAEs, while NAAA and acid ceramidase strongly prefer saturated substrates (e.g., PEA or ceramide).³⁸ Replacement of the hexyl chain with an unsaturated group might mimic the π -characteristics and conformation of the anandamide, leading to binding affinity preference towards FAAH. Therefore, we optimized the chain terminus of C5 uracil substituents (–H, –F, –OAc) with a phenyl ring, a chemical moiety that can mimic the conformation and π -unsaturation of the $\Delta^{8,9}/\Delta^{11,12}$ double bonds of anandamide. Three representative series where R₅ = H, F, OAc were designed and synthesized. As shown in Table 3, most of the analogous with an alkylphenyl group exhibited dramatically increased inhibitory potency against FAAH. The inhibitory potency was progressively improved by prolonging the linker length of the alkylphenyl. Interestingly, the inhibition activity of these substituents was independent of electron effect of C5 group, which was previously expected to significantly modulate the binding affinity of these derivatives. The results suggested that van der Waals interactions, strongly related to the closeness between inhibitors and protein surfaces, might be the most significant contribution to binding affinity.

These uracil urea derivatives exhibited different inhibition pattern against NAAA (Table 3). Most compounds with alkylphenyl replacement were less active than the parent hexyl derivatives. The optimal potency was observed with the linker length of C5 (for 5-F series) or C4 (for 5-H and 5-OAc series) methylene spacer, and the inhibition potency progressively decreased with lengthened or shortened linker. The tight binding pocket of NAAA, may explain the loss of NAAA inhibitory and gain in FAAH selectivity by bulkier structures. Most of the alkylphenyl derivatives were more selective for FAAH compared to parent inhibitors (**1a**, **2f** and **2i**). Among these alkylphenyl derivatives **3g**, **3t** and **3u** were the most promising inhibitors with high potency and selectivity towards FAAH. We further substituted the phenylalkyl chain of **3t** and **3u** with biphenyl group and tested whether biphenyl modification will further enhance the potency. As showed in Table 4, the biphenyl derivatives (**4a**, **4c** and **4d**) were 3–16 folds more potent or similar potent (**4b**) compared to the parent inhibitors (**3t** and

Table 3 Inhibitory effects of compounds **3a**–**3u** on rFAAH and rNAAA activities^a

					
R ₅	n	IC ₅₀ of rFAAH (nM)	IC ₅₀ of rNAAA (nM)	Selectivity	
3a	–H	1	155 ± 23	8020 ± 1290	52
3b	2	333 ± 105	6340 ± 1080	19	
3c	3	19 ± 7	1572 ± 326	83	
3d	4	13 ± 4	305 ± 57	23	
3e	5	11 ± 1	435 ± 63	40	
3f	6	4 ± 0.9	2490 ± 372	623	
3g	7	3 ± 0.8	4768 ± 541	1589	
3h	–F	1	122 ± 10	4655 ± 482	38
3i	2	98 ± 12	4510 ± 371	9.4	
3j	3	21 ± 3	440 ± 96	20	
3k	4	16 ± 2	481 ± 35	30	
3l	5	13 ± 3	105 ± 11	8	
3m	6	4 ± 1	290 ± 30	73	
3n	7	3 ± 0.8	822 ± 85	274	
3o	–OAc	1	51 ± 10	5880 ± 647	115
3p	2	96 ± 12	5440 ± 1023	57	
3q	3	32 ± 6	810 ± 100	25	
3r	4	18 ± 3	220 ± 25	12	
3s	5	10 ± 1	3545 ± 641	355	
3t	6	5 ± 1.1	7908 ± 855	1648	
3u	7	3 ± 0.4	9382 ± 1425	3127	

^a Data are presented as IC₅₀ ± standard error of the mean. All experiments were performed in triplicate.

Table 4 Inhibitory effects of compounds **4a**–**4d** on rFAAH and rNAAA activities^a

n	IC ₅₀ of rFAAH (nM)	IC ₅₀ of rNAAA (nM)
4a	1	1.1 ± 0.2
4b	2	8.3 ± 1.2
4c	3	0.3 ± 0.05
4d	4	0.8 ± 0.1

^a Data are presented as IC₅₀ ± standard error of the mean. All experiments were performed in triplicate.

3u). Moreover, the selectivity of biphenyl substituents towards FAAH was significantly enhanced. **4a**–**4d** exhibited profound selectivity (>3000 folds) towards FAAH over NAAA.

4.2. Selectivity evaluation

NAAA, acid ceramidase (AC) and endocannabinoid hydrolase monoacylglycerol lipase (MAGL) are three competitive targets

for the uracil urea derivatives. Representative inhibitors (**2f**, **2i**, **3u** and **4c**) were selected and their selectivity towards FAAH was examined. As shown in Table 5, the inhibitors were highly selective for FAAH over MAGL. Hexyl derivatives **2f** and **2i** exhibited inhibitory effects against all the three enzymes. Phenyl ring substitutions at the terminus of the hexyl side chain enhanced the binding affinity of the inhibitors to FAAH over NAAA (**2f**, 1.7 folds; **2i**, 6.5 folds; **3u** and **4c**, >3000 folds), but it did not decrease their inhibition potency towards AC. Notably, biphenyl group substitution dramatically increased both potency and selectivity of the derivatives towards FAAH. The most potent inhibitor **4c** was more than 3000 folds selective for FAAH over NAAA, and modestly selective for FAAH over AC (Table 5).

The potency and selectivity of inhibitors were depended on the compound reactivity as well as the molecular shape and flexibility. The inhibition profile of uracil urea derivatives was modulated by electron properties of the C5 substituents and the conformation of the side chain. Electron-withdrawing substituent enhances both potency and selectivity for FAAH over NAAA, but failed to improve the selectivity towards FAAH over AC.³⁶ Electron-withdrawing substitution turning **1a** to a dual inhibitor of FAAH–AC. On the other hand, the side chain is the major factor contributing to the selectivity of uracil urea derivatives. Incorporation of phenyl-containing group into the side chain significantly enhanced their inhibition potency and selectivity towards FAAH over NAAA and AC, turning the uracil urea scaffold from a multi-target inhibitor to a FAAH-selective inhibitor.

4.3. Investigations of the inhibition mechanism

To explore the pharmacological properties of uracil urea derivatives, we analysed their interaction pattern with FAAH. Representative inhibitor **4c** was selected in these studies. As well as its significant inhibition towards recombinant rat FAAH protein (IC₅₀ = 0.3 ± 0.05 nM, Table 4), **4c** displayed a profound inhibition toward recombinant human FAAH (IC₅₀ = 0.6 ± 0.1 nM) (Fig. 2A). Kinetic analysis revealed that **4c** rapidly inhibited rFAAH ($t_{1/2} < 1$ minute) (Fig. 2B), and changed the maximal catalytic velocity (V_{max}) of rFAAH (V_{max} in pmol per min per mg protein, vehicle: 2056 ± 107; and **4c**, 50 nM: 698 ± 84), but it did not affect Michaelis–Menten constant K_m (K_m in μ M, vehicle: 9.8 ± 0.3; and **4c**, 50 nM: 10.4 ± 0.6) (Fig. 2C). No recovery of the rFAAH activity was observed by dialysis of the **4c**–rFAAH

Table 5 Selectivity screening of compounds **2f**, **2i**, **3u** and **4c**^a

	IC ₅₀ of rFAAH (nM)	IC ₅₀ of rNAAA (nM)	IC ₅₀ of rAC (nM)	IC ₅₀ of rMAGL (nM)
2f	110 ± 10	190 ± 20	102 ± 23	>10 000
2i	38 ± 9	250 ± 50	45 ± 9	>10 000
3u	3 ± 0.4	9382 ± 1425	68 ± 10	>10 000
4c	0.3 ± 0.05	>10 000	360 ± 74	>10 000

^a Data are presented as IC₅₀ ± standard error of the mean. All experiments were performed in triplicate.

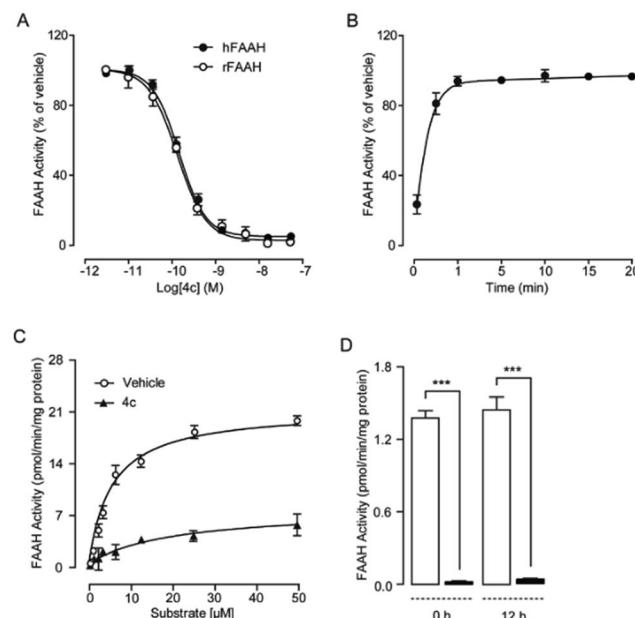


Fig. 2 Effect of **4c** on FAAH activity *in vitro*. (A) Dose-dependent inhibition of recombinant rat FAAH (rFAAH) and recombinant human FAAH (hFAAH) activity by **4c**. (B) Time course of rFAAH inhibition by **4c** (50 nM). (C) Michaelis–Menten analysis of the FAAH reaction in the presence of vehicle (open circles, 1% DMSO) or **4c** (closed triangles, 50 nM). (D) FAAH activity in the presence of vehicle (open bars 1% DMSO) or **4c** (closed bars, 50 nM) before dialysis and 12 hours after dialysis. Results are expressed as mean \pm SEM ($n = 4$ –6). *** $P < 0.001$.

interaction complex even after 12 hours (Fig. 2C). These results indicated that **4c** was an irreversible and non-competitive FAAH inhibitor, thereby providing strong evidence for the formation of covalent bonds between compound and enzyme.

4.4. Biological stability evaluation

The hydrolysis of compounds (**2f**, **2i** and **4c**) was studied in 80% (v/v) rat plasma at 37 °C. The results showed that more than 60% of compounds were hydrolyzed after 10 min incubation (Table 6). Analogues **2f** and **2i** containing less sterically hindered hexyl chain were cleaved more rapidly than the bulkier compound **4c**. LC-MS analysis of degradation products of inhibitors (**2f**, **2i** and **4c**) showed the formation of hexylamine and 4-biphenylpropylamine, which resulted from the hydrolysis of the *N*1-carboxamide (data not shown). FAAH is broadly

Table 6 Percent of compounds **2f**, **2i** and **4c** remaining after incubation in 80% (v/v) rat plasma^a

	2f	2i	4c
Remain inhibitors	0 min	100%	100%
	5 min	15%	21%
	10 min	7%	11%
	15 min	n.a.	8%

^a Compounds were measured at 10 μ M in 80% (v/v) rat plasma with 2.5% DMSO.

distributed throughout the body, involved in a complex anandamide-based signalling system. Most of current FAAH inhibitors, *e.g.* PF-04457845, are distributed throughout all tissues and organs after administration. They are suitable for the systemic application, but probably not optimal agents to investigate the function of FAAH in specific organ or tissue. The inhibitors we developed exert acute FAAH inhibitory in the target site and rapidly inactivated to prevent unwanted systemic effects. Our findings may offer a new topically applied tool to investigate the function of FAAH at a selected tissue or organ.

5. Conclusions

Here, we report the synthesis and characterization of a series of uracil urea derivatives, designed as FAAH inhibitors by progressive modification of **1a**. SAR studies suggested that electron-withdrawing substituents in the C5 position enhanced the potency of FAAH inhibition and the replacement of the side chain with phenylalkyl moieties or biphenyl groups significantly improved both the inhibitory potency and selectivity towards FAAH. The two most potent inhibitors, **4c** ($IC_{50} = 0.3 \pm 0.05$ nM) and **4d** ($IC_{50} = 0.8 \pm 0.1$ nM) showed moderate to high selectivity towards FAAH over other endocannabinoid-metabolizing enzymes. Rapid dilution and kinetic analysis indicate that **4c** is a selective, irreversible and non-competitive FAAH inhibitor. This study provides us some promising FAAH inhibitors and an optimized scaffold for the development of strategies to investigate tropical FAAH function, and to explore new therapeutics for inflammation, cancer, and neuropathic diseases.

Acknowledgements

We thank Dr Daniele Piomelli at the University of California, Irvine for the kind gifts of the HEK293-rNAAA cells and the HEK293-rFAAH cells. This study was supported by grants from National Natural Sciences Foundation of China (No. 81373273 to QY, No. 81602974 to YL), Key Research Program of Frontier Science, CAS, Grant No. QYZDJ-SSW-SLH033, The Major State Basic Research Development Program of China (grant number 2014CB965101), and China Postdoctoral Science Foundation Funded Project (2016M592876XB) to YL.

References

- 1 D. K. Giang and B. F. Cravatt, *Proc. Natl. Acad. Sci. U. S. A.*, 1997, **94**, 2238.
- 2 S. Shin, T. H. Lee, N. C. Ha, H. M. Koo, S. Y. Kim, H. S. Lee, Y. S. Kim and B. H. Oh, *EMBO J.*, 2002, **21**, 2509.
- 3 M. H. Bracey, M. A. Hanson, K. R. Masuda, R. C. Stevens and B. F. Cravatt, *Science*, 2002, **298**, 1793.
- 4 A. H. Lichtman, D. Leung, C. C. Shelton, A. Saghatelian, C. Hardouin, D. L. Boger and B. F. Cravatt, *J. Pharmacol. Exp. Ther.*, 2004, **311**, 441.
- 5 S. Kathuria, S. Gaetani, D. Fegley, F. Valino, A. Duranti, A. Tontini, M. Mor, G. Tarzia, G. La Rana, A. Calignano,



A. Giustino, M. Tattoli, M. Palmery, V. Cuomo and D. Piomelli, *Nat. Med.*, 2003, **9**, 76.

6 B. F. Cravatt, K. Demarest, M. P. Patricelli, M. H. Brace, D. K. Giang, B. R. Martin and A. H. Lichtman, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 9371.

7 C. J. Fowler, K. O. Jonsson and G. Tiger, *Biochem. Pharmacol.*, 2001, **62**, 517.

8 H. S. Hansen, B. Moesgaard, H. H. Hansen and G. Petersen, *Chem. Phys. Lipids*, 2000, **108**, 135.

9 H. S. Hansen, *Exp. Neurol.*, 2010, **224**, 48.

10 H. H. Schmid, P. C. Schmid and V. Natarajan, *Prog. Lipid Res.*, 1990, **29**, 1.

11 J. M. Walker, S. M. Huang, N. M. Strangman, K. Tsou and M. C. Sanudo-Pena, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, **96**, 12198.

12 M. Salaga, M. Sobczak and J. Fichna, *Eur. J. Pharm. Sci.*, 2014, **52**, 173.

13 K. Otrubova, C. Ezzili and D. L. Boger, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 4674.

14 M. Seierstad and J. G. Breitenbacher, *J. Med. Chem.*, 2008, **51**, 7327.

15 D. L. Boger, H. Miyauchi, W. Du, C. Hardouin, R. A. Fecik, H. Cheng, I. Hwang, M. P. Hedrick, D. Leung, O. Acevedo, C. R. Guimaraes, W. L. Jorgensen and B. F. Cravatt, *J. Med. Chem.*, 2005, **48**, 1849.

16 L. Chang, L. Luo, J. A. Palmer, S. Sutton, S. J. Wilson, A. J. Barbier, J. G. Breitenbacher, S. R. Chapman and M. Webb, *Br. J. Pharmacol.*, 2006, **148**, 102.

17 G. Tarzia, A. Duranti, A. Tontini, G. Piersanti, M. Mor, S. Rivara, P. V. Plazzi, C. Park, S. Kathuria and D. Piomelli, *J. Med. Chem.*, 2003, **46**, 2352.

18 G. Moreno-Sanz, A. Duranti, L. Melzig, C. Fiorelli, G. F. Ruda, G. Colombano, P. Mestichelli, S. Sanchini, A. Tontini, M. Mor, T. Bandiera, R. Scarpelli, G. Tarzia and D. Piomelli, *J. Med. Chem.*, 2013, **56**, 5917.

19 M. Mor, S. Rivara, A. Lodola, P. V. Plazzi, G. Tarzia, A. Duranti, A. Tontini, G. Piersanti, S. Kathuria and D. Piomelli, *J. Med. Chem.*, 2004, **47**, 4998.

20 M. Mor, A. Lodola, S. Rivara, F. Vacondio, A. Duranti, A. Tontini, S. Sanchini, G. Piersanti, J. R. Clapper, A. R. King, G. Tarzia and D. Piomelli, *J. Med. Chem.*, 2008, **51**, 3487.

21 K. Ahn, S. E. Smith, M. B. Liimatta, D. Beidler, N. Sadagopan, D. T. Dudley, T. Young, P. Wren, Y. Zhang, S. Swaney, K. Van Becelaere, J. L. Blankman, D. K. Nomura, S. N. Bhattachar, C. Stiff, T. K. Nomanbhoy, E. Weerapana, D. S. Johnson and B. F. Cravatt, *J. Pharmacol. Exp. Ther.*, 2011, **338**, 114.

22 M. Kono, T. Matsumoto, T. Kawamura, A. Nishimura, Y. Kiyota, H. Oki, J. Miyazaki, S. Igaki, C. A. Behnke, M. Shimojo and M. Kori, *Bioorg. Med. Chem.*, 2013, **21**, 28.

23 G. L. Li, H. Winter, R. Arends, G. W. Jay, V. Le, T. Young and J. P. Huggins, *Br. J. Clin. Pharmacol.*, 2012, **73**, 706.

24 J. M. Keith, R. Apodaca, M. Tichenor, W. Xiao, W. Jones, J. Pierce, M. Seierstad, J. Palmer, M. Webb, M. Karbarz, B. Scott, S. Wilson, L. Luo, M. Wennerholm, L. Chang, S. Brown, M. Rizzolio, R. Rynberg, S. Chapman and J. G. Breitenbacher, *ACS Med. Chem. Lett.*, 2012, **3**, 823.

25 D. S. Johnson, C. Stiff, S. E. Lazerwith, S. R. Kesten, L. K. Fay, M. Morris, D. Beidler, M. B. Liimatta, S. E. Smith, D. T. Dudley, N. Sadagopan, S. N. Bhattachar, S. J. Kesten, T. K. Nomanbhoy, B. F. Cravatt and K. Ahn, *ACS Med. Chem. Lett.*, 2011, **2**, 91.

26 S. O. Alapafuja, S. P. Nikas, I. T. Bharathan, V. G. Shukla, M. L. Nasr, A. L. Bowman, N. Zvonok, J. Li, X. Shi, J. R. Engen and A. Makriyannis, *J. Med. Chem.*, 2012, **55**, 10074.

27 A. Lodola, M. Mor, S. Rivara, C. Christov, G. Tarzia, D. Piomelli and A. J. Mulholland, *Chem. Commun.*, 2008, 214.

28 C. J. Fowler, *Eur. Neuropsychopharmacol.*, 2015, **25**, 749.

29 X. Wang, K. Sarris, K. Kage, D. Zhang, S. P. Brown, T. Kolasa, C. Surowy, O. F. El Kouhen, S. W. Muchmore, J. D. Brioni and A. O. Stewart, *J. Med. Chem.*, 2009, **52**, 170.

30 M. Feledziak, C. Michaux, A. Urbach, G. Labar, G. G. Muccioli, D. M. Lambert and J. Marchand-Brynaert, *J. Med. Chem.*, 2009, **52**, 7054.

31 G. G. Muccioli, N. Fazio, G. K. Scriba, W. Poppitz, F. Cannata, J. H. Poupaert, J. Wouters and D. M. Lambert, *J. Med. Chem.*, 2006, **49**, 417.

32 G. Palermo, A. D. Favia, M. Convertino and M. De Vivo, *ChemMedChem*, 2016, **11**, 1252.

33 A. D. Favia, D. Habrant, R. Scarpelli, M. Migliore, C. Albani, S. M. Bertozi, M. Dionisi, G. Tarozzo, D. Piomelli, A. Cavalli and M. De Vivo, *J. Med. Chem.*, 2012, **55**, 8807.

34 L. Bertolacci, E. Romeo, M. Veronesi, P. Magotti, C. Albani, M. Dionisi, C. Lambruschini, R. Scarpelli, A. Cavalli, M. De Vivo, D. Piomelli and G. Garau, *J. Am. Chem. Soc.*, 2013, **135**, 22.

35 Y. Qiu, Y. Zhang, Y. Li and J. Ren, *Molecules*, 2016, **21**, 229.

36 D. Pizzirani, C. Pagliuca, N. Realini, D. Branduardi, G. Bottegoni, M. Mor, F. Bertozi, R. Scarpelli, D. Piomelli and T. Bandiera, *J. Med. Chem.*, 2013, **56**, 3518.

37 N. Realini, C. Solorzano, C. Pagliuca, D. Pizzirani, A. Armirotti, R. Luciani, M. P. Costi, T. Bandiera and D. Piomelli, *Sci. Rep.*, 2013, **3**, 1035.

38 N. Ueda, K. Yamanaka and S. Yamamoto, *J. Biol. Chem.*, 2001, **276**, 35552.

