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Adamantyl derivatives and rearranged benzophenones from *Garcinia xanthochymus* fruits†

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Garcinia xanthochymus is a tropical fruit yielding tree native to South East Asia. Its fruit is used to treat bilious conditions, diarrhea and dysentery. In this study, three new adamantyl derivatives, two new rearranged benzophenones, named as garcixanthochymones A–E (1–5), together with 12 known compounds including 7 xanthonones (6–12) and 5 flavonoids (13–17) were isolated from the fruits of *G. xanthochymus*. Their structures were elucidated by detailed spectroscopic analyses. All isolated compounds were evaluated for their anti-proliferative activities against four human tumor cells (HepG2, A549, SGC7901, MCF-7). Adamantyl derivatives (1–3) and rearranged benzophenones (4–5) displayed potential inhibitory activity against four human cancer cell lines with IC₅₀ values of 5.16–16.45 μ M. These data suggested that the extracts of the fruits of *G. xanthochymus* are potent candidates for cancer prevention.

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1. Introduction

Fruits contain high levels of biologically active components that impart health benefits beyond their basic nutritional value. Non-nutritive components in fruits such as phenolics, flavonoids, and in particular anthocyanins contribute in varying degrees to the antioxidant activity of individual fruits. Antioxidants play a crucial role in the prevention of chronic ailments such as heart attacks, cancer, diabetes, hypertension, stroke and Alzheimer's disease by combating oxidative stress. This suggests that a change in dietary behavior, such as increasing the consumption of fruits, may provide desirable health benefits for reducing the incidence of chronic diseases.^{1–3} The genus *Garcinia* belongs to Guttiferae (*Clusiaceae*) family and is a rich source of secondary metabolites including xanthonones, flavonoids, benzophenones, lactones and phenolic acids. Several species have been documented as ethnobotanically important plants used traditionally as food and medicine.⁴ *Garcinia xanthochymus* is a fruit yielding tree native to South East Asia. Its fruits are edible and also used in jams preserves and vinegar. A preparation of the fruit of *G. xanthochymus* is given to treat

bilious conditions, diarrhea and dysentery.^{5,6} Phytochemical studies have revealed that the fruits of *G. xanthochymus* contain various chemical constituents, including 7 polyisoprenylated benzophenones (PIBs): guttiferone H, gamboginone, aristophenone A, xanthochymol, guttiferone E, cycloxanthochymol, isoxanthochymol, 4 xanthonones: alloathyriol, maclurin, euxanthone, 1,5-dihydroxyxanthone and 8 biflavonoids: amentoflavone, 3,8''-biapigenin, (\pm)-fukugetin, (\pm)-fukugiside, 3,8''-biflavanone, morelloflavone, GB 1a, (\pm)-volkensiflavone.^{6–9} The fruits of *G. xanthochymus* have been reported to show a variety of biological activities such as antioxidant,¹⁰ antidiabetic,¹¹ anti-plasmodial⁷ and cytotoxic activities.¹² It has been reported that the *G. xanthochymus* fruits possess 31.31 mg g⁻¹ and 5.313 mg g⁻¹ of total phenol and flavonoid contents.² PIBs are a major class of compounds isolated from the fruits of *Garcinia* plant, since fruits are the only edible plant part. PIBs with structural diversity have a wide range of biological activities.⁴ Much effort has been devoted to verify the effectiveness of the fruit of *G. xanthochymus* against cancer. A series of studies found that PIBs isolated from the fruit of *G. xanthochymus* showed inhibitory effects on human colon cancer cell lines by regulating the mammalian target of rapamycin (mTOR) cell survival pathway.¹²

As part of our continuing research on bioactive components from nutritional fruits, we have investigated the fruits of *G. xanthochymus*, isolated and identified three new adamantyl derivatives, two new rearranged benzophenones, named as garcixanthochymones A–E (1–5) and 12 known compounds. Cytotoxicities against four human tumor cell lines (HepG2, A549, SGC7901, MCF-7) of the 17 isolated compounds are also reported.

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2. Materials and methods

2.1 General experimental procedures

Optical rotations were determined in MeOH on a P-1020 digital polarimeter. (JASCO Co., Tokyo, Japan). UV spectra were obtained on a 2401PC spectrophotometer (Shimadzu Co., Tokyo, Japan). 1D and 2D NMR spectra were recorded on a Bruker AVANCE III-500 MHz and a Bruker AV-400 MHz spectrometer (Bruker, Ettlingen, Germany) in DMSO- d_6 , C_5D_5N , acetone- d_6 using tetramethylsilane (TMS) as an internal reference standard. Chemical shifts (δ) have been expressed in ppm and the coupling constants (J) have been given in Hz. High-resolution electrospray mass spectroscopy was performed on an Agilent G6230 TOF mass spectrometer (HR-ESI-MS) (Agilent Technologies Co. Ltd., California, America) and a Waters Autospec Premier 776 mass spectrometer (HR-EI-MS) (Waters Technologies Co. Ltd., Massachusetts, America). High-performance liquid chromatography (HPLC) was conducted on an Ultimate 3000 HPLC system (Dionex Co., Sunnyvale, CA, USA) equipped with an Ultimate 3000 pump and Ultimate 3000 variable wavelength detector, as well as a semi-preparative YMC-Pack ODS-A column (250 \times 10 mm, 5 μ m) and a preparative YMC-Pack ODS-A column (250 \times 20 mm, 5 μ m) from YMC Co., Ltd (Kyoto, Japan), column chromatography (CC) was conducted over silica gel (200–300 mesh and 300–400 mesh, Qingdao Haiyang Chemical Industry Co., Ltd., Qingdao, China). Chromatographic grade acetonitrile was purchased from Chang Tech Enterprise Co., Ltd (Taiwan, China). The cell lines were purchased from the cell bank of Chinese Academy of Sciences (Shanghai, China). 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2-*H*-tetrazolium bromide (MTT) was purchased from Biosharp Co. Ltd. (Hefei, Anhui, China). Doxorubicin hydrochloride was purchased from Aladdin Chemical Co. Ltd. (Shanghai, China). Dulbecco modified Eagle medium (DMEM), phosphate buffer saline (PBS) and penicillin-streptomycin solution were purchased from Hyclone Laboratories, GE healthcare life science (Logan, UT, USA). Fetal bovine serum (FBS) was purchased from Gibco, Life technologies (Grand Island, NY, USA). Reagent grade dimethyl sulfoxide (DMSO) was purchased from Vetec, Sigma Chemical Co. (St. Louis, MO, USA). The absorbance was read on a Multiskan GO microplate reader (Thermo Fisher Scientific Inc. Waltham, MA, USA).

2.2 Plant material

The fruits of *G. xanthochymus* were purchased from Xishuangbanna Prefecture, Yunnan Province, P. R. China and identified by Prof. Ying-hong Zhao, Xishuangbanna Prefecture National Medicine Research Institute. The voucher specimen (2013 091 201) was deposited in the herbarium of School of Pharmaceutical Sciences, South Central University for Nationalities.

2.3 Extraction and isolation

The dried fruits of *G. xanthochymus* (6.18 kg) were powdered and extracted with 95% EtOH at room temperature for three times

(each time for 24 h) to obtain EtOH extract 2.94 kg, and then successively partitioned with petroleum ether (P. E.), EtOAc and *n*-BuOH to get petroleum ether extract 267 g, EtOAc extract 711 g, *n*-BuOH extract 460 g. The EtOAc extract (332 g) was chromatographed on a silica gel column (200–300 mesh) eluted successively with P. E./EtOAc gradient (19 : 1, 9 : 1, 7 : 3, 6 : 4, 1 : 1, 4 : 6, 3 : 7, 0 : 1) to obtain 10 fractions (Fr. 1–Fr. 10). Fr. 3 (13.3 g) was subjected to octadecylsilane CC eluting with H_2O –MeOH (7 : 3, 6 : 4, 1 : 1, 4 : 6, 3 : 7, 2 : 8, 0 : 1) to obtain 10 major fractions (Fr. 3.1–Fr. 3.10). Fr. 3.2 (145.4 mg) was purified by semipreparative HPLC (MeOH– H_2O , 52 : 48) to afford **11** (7.4 mg) at t_R 14.0 min. Fr. 3.4 (33 mg) was purified by semipreparative HPLC (MeOH– H_2O , 55 : 45) to afford **13** (5.1 mg) at t_R 13.9 min. Fr. 3.5 (287 mg) was purified by recrystallization to afford **10** (6.5 mg), then further subjected to a silica gel column (300–400 mesh) eluted successively with $CHCl_3$ /MeOH gradient (200 : 1, 100 : 1, 50 : 1, 19 : 1, 7 : 3, 1 : 1, 3 : 7, 0 : 1) to obtain 6 fractions (Fr. 3.5.1–Fr. 3.5.6). Fr. 3.5.3 (20 mg) was purified by semipreparative HPLC (CH_3CN – H_2O , 35 : 65) to afford **6** (1.5 mg) at t_R 23.6 min. Fr. 3.5.4 (10.5 mg) was purified by semipreparative HPLC (MeOH– H_2O , 50 : 50) to afford **9** (2.5 mg) at t_R 37.0 min. Fr. 3.5.5 (50 mg) was purified by semipreparative HPLC (MeOH– H_2O , 63 : 37) to afford **7** (4.3 mg) at t_R 12.3 min, **8** (10.2 mg) at t_R 14.1 min respectively. Fr. 3.6 (1.16 g) was subjected to a silica gel column (300–400 mesh) eluted successively with $CHCl_3$ /MeOH gradient (200 : 1, 100 : 1, 50 : 1, 20 : 1, 10 : 1, 8 : 2, 6 : 4, 1 : 1, 2 : 8, 0 : 1) to obtain 7 fractions (Fr. 3.6.1–Fr. 3.6.7). Fr. 3.6.3 (350 mg) was purified by semipreparative HPLC with gradient MeOH– H_2O (73 : 27 to 98 : 2) to afford **14** (23.1 mg) at t_R 25.9 min and the lower polar fractions (61.3 mg) at t_R 41 min, which was further purified by semipreparative HPLC (CH_3CN – H_2O , 64 : 36) to afford **1** (6.6 mg) at t_R 33.5 min, **2** (4.1 mg) at t_R 42.3 min, **3** (4.0 mg) at t_R 37.4 min. Fr. 3.7 (4.3 g) was subjected to a silica gel column (300–400 mesh) eluted successively with P. E./ $CHCl_3$ /MeOH gradient (20 : 5 : 0.1, 10 : 5 : 0.1, 5 : 5 : 0.1, 5 : 5 : 1, 5 : 5 : 2, 0 : 9 : 1, 0 : 0 : 1) to obtain 3 fractions (Fr. 3.7.1–Fr. 3.7.3). Fr. 3.7.3 (1.58 g) was subjected to a silica gel column (300–400 mesh) eluted successively with P. E./ $CHCl_3$ /MeOH gradient (20 : 5 : 0.1, 5 : 5 : 0.1, 5 : 5 : 0.2, 0 : 0 : 1) to obtain 3 fractions (Fr. 3.7.3.1–Fr. 3.7.3.3). Fr. 3.7.3.2 (620 mg) was purified by preparative HPLC (CH_3CN – H_2O , 93 : 7) to obtain 11 fractions (Fr. 3.7.3.2.1–Fr. 3.7.3.2.11). Fr. 3.7.3.2.1 (53.5 mg) was purified by semipreparative HPLC (CH_3CN – H_2O , 75 : 25) to afford **4** (23.2 mg) at t_R 19.5 min. Fr. 3.7.3.2.2 (81.5 mg) was purified by semipreparative HPLC (CH_3CN – H_2O , 70 : 30) to afford **5** (24.9 mg) at t_R 31.1 min. Fraction 5 (9.5 g) was subjected to octadecylsilane CC eluting with H_2O –MeOH (7 : 3, 6 : 4, 1 : 1, 4 : 6, 3 : 7, 2 : 8, 0 : 1) to give 12 major fractions (Fr. 5.1–Fr. 5.12). Fr. 5.6 (30 mg) was purified by semipreparative HPLC (MeOH– H_2O , 82 : 18) to afford **12** (5.4 mg) at t_R 13.1 min. Fraction 8 (49.9 g) was purified by recrystallization to afford **16** (364 mg). Fraction 7 (32.4 g) was subjected to a silica gel column (200–300 mesh) eluted successively with $CHCl_3$ /MeOH gradient (500 : 1, 100 : 1, 20 : 1, 10 : 1, 7 : 3, 0 : 1) to obtain 14 fractions (Fr. 7.1–Fr. 7.14). Fr. 7.12 (2.3 g) was subjected to octadecylsilane CC eluting with H_2O –MeOH (7 : 3, 1 : 1, 3 : 7, 1 : 9, 0 : 1) to obtain **15** (200 mg) and **17** (166 mg).



Garcixanthochymone A (1), white powder. $[\alpha]_D^{21} = -13.8$ ($c = 0.660$, MeOH); UV (MeOH) λ_{\max} nm (log ϵ): 258 (4.07), 309 (3.79), 360 (3.86); ^1H - and ^{13}C -NMR: see Table 1. HR-EI-MS m/z : 618.3550 $[\text{M}]^+$ (calcd for $\text{C}_{38}\text{H}_{50}\text{O}_7$: 618.3557).

Garcixanthochymone B (2), white powder. $[\alpha]_D^{21} = -0.81$ ($c = 0.410$, MeOH); UV (MeOH) λ_{\max} nm (log ϵ): 258 (3.95), 310 (3.69), 363 (3.75); ^1H - and ^{13}C -NMR: see Table 1. HR-ESI-MS (positive ion mode) m/z : 639.3296 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{38}\text{H}_{48}\text{O}_7\text{Na}$: 639.3292).

Garcixanthochymone C (3), white powder. $[\alpha]_D^{21} = +51.0$ ($c = 0.400$, MeOH); UV (MeOH) λ_{\max} nm (log ϵ): 258 (3.93), 309 (3.77), 357 (3.73); ^1H - and ^{13}C -NMR: see Table 1. HR-ESI-MS (positive

ion mode) m/z : 635.3577 $[\text{M} + \text{H}]^+$ (calcd for $\text{C}_{38}\text{H}_{51}\text{O}_8$: 635.3578).

Garcixanthochymone D (4), white powder. $[\alpha]_D^{21} = +9.49$ ($c = 0.604$, MeOH); UV (MeOH) λ_{\max} nm (log ϵ): 260 (3.95), 323 (3.67), 365 (3.77); ^1H - and ^{13}C -NMR: see Table 2. HR-ESI-MS (positive ion mode) m/z : 555.2718 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{33}\text{H}_{40}\text{O}_6\text{Na}$: 555.2723).

Garcixanthochymone E (5), white powder. $[\alpha]_D^{21} = +22.7$ ($c = 0.577$, MeOH); UV (MeOH) λ_{\max} nm (log ϵ): 260 (3.97), 322 (3.71), 365 (3.77); ^1H - and ^{13}C -NMR: see Table 2. HR-ESI-MS (positive ion mode) m/z : 555.2726 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{33}\text{H}_{40}\text{O}_6\text{Na}$: 555.2723).

Table 1 ^1H , ^{13}C NMR data of compounds 1–3 (in acetone- d_6)^a

No.	^{13}C -NMR			^1H -NMR		
	1	2	3	1	2	3
1	77.6	77.6	77.2			
2	202.3	202.6	207.7			
3	80.3	80.2	75.5			
4	202.7	202.5	107.3			
5	69.1	69.3	57.7			
6	45.4	45.0	39.0	2.51 m 2H	2.55 m 2H	3.63 dd (14.8, 4.8) 1.74 m 2.23 m
7	48.3	48.3	43.4	1.81 m	1.83 m	
8	54.5	54.4	57.0			
9	204.3	204.3	208.0			
10	192.3	192.1	198.2			
11	128.9	128.6	133.2			
12	116.8	117.4	119.5	6.91 d (2.0)	6.87 d (1.6)	7.12 d (2.0)
13	145.3	145.1	143.8			
14	150.5	150.8	149.2			
15	115.1	115.3	113.8	6.78 d (8.4)	6.78 m	6.70 d (8.4)
16	124.0	124.1	125.2	6.72 dd (8.4, 2.0)	6.78 m	6.97 dd (8.4, 2.0)
17	34.9	34.4	28.8	1.89 dd (14.4, 6.0) 2.00 m 2.73 m	1.91 dd (14.8, 6.8) 1.98 dd (14.8, 6.8) 3.15 m	2.17 dd (13.2, 3.2) 1.30 (m) 1.56 m
18	43.8	39.9	41.7			
19	149.4	149.0	80.1			
20	113.5	113.9	22.3	4.76 dd (2.4, 1.2) 4.66 d (2.4)	4.79 d (2.4) 4.83 m	1.01 s 3H
21	18.0	18.3	29.2	1.61 s 3H	1.67 s 3H	0.80 s 3H
22	29.2	41.2	29.4	1.46 m 1.30 m 1.30 m 2H	1.62 m 1.37 m 3.89 d (10.0)	1.45 m 1.02 m 2.25 m, 2.02 m
23	42.4	73.4	36.2			
24	70.5	150.1	146.7			
25	29.5	109.7	110.7	1.17 s 3H	4.72 s 4.93 s	4.76 s 4.71 s
26	29.5	18.4	22.8	1.17 s 3H	1.72 s 3H	1.74 s 3H
27	24.0	24.0	24.2	2.39 dd (14.4, 6.2) 2.50 m 4.92 t (6.2)	2.49 dd (14.0, 6.4) 2.38 dd (14.0, 6.8) 4.89 t (6.4)	2.34 m 2H
28	121.5	121.4	122.8			5.06 t (7.0)
29	133.8	133.9	131.9			
30	18.3	18.5	18.2	1.66 s 3H	1.65 s 3H	1.64 s 3H
31	26.1	26.2	26.2	1.60 s 3H	1.58 s 3H	1.64 s 3H
32	23.1	23.1	24.6	1.17 s 3H	1.17 s 3H	1.05 s 3H
33	22.7	22.7	23.5	1.22 s 3H	1.22 s 3H	1.02 s 3H
34	52.0	52.1	52.2	4.15 dt (8.0, 2.4)	4.14 d (7.8)	3.68 t (3.2)
35	122.4	122.4	88.8	4.98 dt (8.0, 1.2)	5.01 d (7.8)	4.26 d (3.2)
36	134.2	134.2	71.7			
37	26.3	26.2	29.5	1.69 s 3H	1.69 s 3H	1.34 s 3H
38	18.5	18.7	28.0	1.74 s 3H	1.75 s 3H	1.27 s 3H

^a The chemical shifts have been expressed in δ ppm. The coupling constants (J) have been expressed in Hz.



Table 2 ^1H , ^{13}C NMR and HMBC data of compounds 4 and 5 (in $\text{C}_5\text{D}_5\text{N}$)

	4			5		
No.	¹³ C-NMR	¹ H-NMR	HMBC	¹³ C-NMR	¹ H-NMR	HMBC
1	64.4			64.3		
2	214.8			214.8		
3	71.2			71.2		
4	33.6	2.28, 1.91 m 2H		33.8	1.97 dd (11, 8.5 Hz), 2.26 m 2H	
5	57.2	1.99 m		57.7	2.07 m	
6	42.8			42.7		
7	93.1			93.2		
8	70.1			70.1		
9	206.6			206.5		
10	48.3	1.91, 2.10 m 2H		48.3	2.09 m, 1.91 d (13.5) 2H	
11	26.7	2.66 m 2H	9, 2, 12, 13, 1, 10	26.7	2.66 m 2H	9, 2, 12, 13, 1
12	120.8	5.54 t (7.5)		120.6	5.54 t (7.5)	14, 15
13	134.2			134.2		
14	26.3	1.62 s 3H	12, 13	26.3	1.62 s 3H	12, 13
15	18.3	1.63 s 3H	12, 13	18.3	1.64 s 3H	12, 13
16	29.9	2.51 dd (11.5, 9.2), 2.24 m 2H	18, 17, 3, 2	30.0	2.50 dd (11.0, 9.5), 2.23 m 2H	18, 17, 3, 2
17	57.6	3.07 dd (9.2, 8.5)	7, 8, 18, 19, 20, 9	57.6	3.06 dd (9.5, 8.0)	7, 8, 18, 19, 20, 9
18	37.4			37.4		
19	27.7	1.24 s 3H	17, 29	27.8	1.22 s 3H	17, 29, 18
20	31.0	1.42 s 3H	17, 29	31.0	1.39 s 3H	17, 29, 18
21	33.8	1.84, 1.91 m 2H		34.1	2.40 m 2H	
22	37.1	2.09, 1.83 m		124.6	5.14 t (7.0)	24, 25
23	146.5			132.1		
24	110.7	4.78 s, 4.80 s	25, 22	26.3	1.68 s 3H	22, 23
25	22.9	1.69 s 3H	24, 23	18.4	1.60 s 3H	22, 23
26	19.7	1.78 s 3H	7, 6, 10	19.7	1.80 s 3H	7, 6, 10, 5
27	196.7			196.7		
28	129.0			129.0		
29	146.1			146.1		
30	112.4	7.23 s	18, 28, 31, 32	112.4	7.22 s	18, 28, 31, 32
31	146.3			146.3		
32	154.0			154.0		
33	115.1	8.06 s	27, 32, 31	115.1	8.06 s	27, 32, 31

2.4 Measurement of inhibition activity on tumor cell proliferation

The antiproliferative activities against four human tumor cell lines (HepG2, A549, SGC7901, MCF-7) of isolated compounds were measured by the MTT assay. The four human tumor cell lines were cultured in 96-well plates at a concentration of 10^4 cells per well in DMEM containing 10% FBS and 1% penicillin-streptomycin at 37 °C in 5% CO_2 atmosphere. After 24 h of growth, washed the cells for three times by PBS, then added 200 μL sample solution or doxorubicin solution (dissolved in DMEM containing 1% penicillin-streptomycin) in each well at a gradient of 50 μM , 25 μM , 12.5 μM , 6.25 μM and 3.125 μM . After 24 h of incubation, removed the cell culture medium and washed the cells for three times by PBS, then added 100 μL MTT (0.5 mg mL^{-1}) solution (dissolved in DMEM containing 1% penicillin-streptomycin) in each well incubated for 4 h. After the incubation, removed the MTT solution and add 100 μL DMSO, read the absorbance at a wavelength of 492 nm by using the microplate reader. The inhibition rate of the compound (%) was calculated with the formula $[\text{A}]_{\text{test}}/[\text{A}]_{\text{control}} \times 100\%$, where $[\text{A}]_{\text{test}}$ is the absorbance of the test group that the cells incubated

in sample solution, and $[\text{A}]_{\text{control}}$ is the absorbance of the control group that the cells incubated only with the cell culture medium. The IC_{50} values was calculated according to the inhibition rate by using GraphPad Prism 6. All the 17 compounds have been tested the antiproliferative activities against four

Table 3 The antiproliferative activities of isolated compounds against four human tumor cells

$(\text{IC}_{50} \text{ in } \mu\text{M})$				
Compounds ^a	HepG2	A549	SGC7901	MCF-7
Doxorubicin ^b	6.52 \pm 0.13	14.03 \pm 0.21	7.54 \pm 1.11	4.40 \pm 1.17
1	8.34 \pm 1.2	15.88 \pm 0.48	7.32 \pm 0.29	5.77 \pm 0.17
2	5.77 \pm 1.43	15.13 \pm 5.99	5.16 \pm 1.16	14.12 \pm 1.29
3	6.64 \pm 0.87	16.25 \pm 3.11	6.80 \pm 0.29	9.75 \pm 0.62
4	9.08 \pm 0.91	7.48 \pm 0.03	10.71 \pm 1.97	15.69 \pm 0.87
5	9.03 \pm 0.36	8.24 \pm 0.16	15.25 \pm 4.79	16.45 \pm 0.23
14	12.82 \pm 2.49	11.29 \pm 1.40	19.30 \pm 3.06	13.55 \pm 1.97

^a Other isolates with $\text{IC}_{50} > 50 \mu\text{M}$ for all cell lines are not listed.

^b Doxorubicin was used as positive control.



human tumor cell lines (HepG2, A549, SGC7901, MCF-7) as summarized in Table 3.

3. Result and discussion

3.1 Structural elucidation of isolated compounds

Garcixanthochymone A (**1**) was isolated as a white powder and gave the molecular formula $C_{38}H_{50}O_7$ with 14 degrees of unsaturation by its HR-EI-MS ($[M]^+$ m/z 618.3550, calcd 618.3557). Its UV spectrum gave its maximum absorption wavelengths at 258 nm, 309 nm and 360 nm which indicated the presence of an aromatic moiety. The 1H -NMR spectrum revealed the presence of a 1,3,4-trisubstituted benzene ring signals [δ_H 6.72 (1H, dd, J = 8.4, 2.0 Hz), 6.78 (1H, d, J = 8.4 Hz) and 6.91 (1H, d, J = 2.0 Hz)], two olefinic proton signals [δ_H 4.92 (1H, t, J = 6.2 Hz) and 4.98 (1H, dt, J = 8.0, 1.2 Hz)], one terminal double bonds [δ_H 4.66 (1H, d, J = 2.4 Hz) and 4.76 (1H, dd, J = 2.4, 1.2 Hz)], nine methyl signals [δ_H 1.17 (3H, s), 1.17 (3H, s), 1.17 (3H, s), 1.22 (3H, s), 1.60 (3H, s), 1.61 (3H, s), 1.66 (3H, s), 1.69 (3H, s) and 1.74 (3H, s)], one methylene signal [δ_H 2.51 (2H, m)], two methine signals [δ_H 1.81 (1H, m), δ_H 4.15 (1H, dt, J = 8.0, 2.4 Hz)]. The ^{13}C -NMR and DEPT spectrum indicated the presence of three nonconjugated carbonyl carbons [δ_C 202.3, 202.7, and 204.3], one conjugated carbonyl carbon [δ_C 192.3], a 3,4-dihydroxybenzoyl group [δ_C 115.1, 116.8, 124.0, 128.9, 145.3 and 150.5], two trisubstituted double bonds [δ_C 121.5, 122.4, 133.8, and 134.2], one terminal double bonds [δ_C 113.5, 149.4], five quaternary carbons [δ_C 54.5, 69.1, 70.5, 77.6 and 80.3], five methylenes [δ_C 24.0, 29.2, 34.9, 42.4 and 45.4], three methines [δ_C 43.8, 48.3 and 52.0], nine methyls [δ_C 18.0, 18.3, 18.5, 22.7, 23.1, 26.1, 26.3, 29.5 and 29.5]. On the basis of these data, **1** has an adamantyl phloroglucinol skeleton.¹³ Comparison of 1H and ^{13}C NMR data of **1** with those of garcinialiptone A¹⁴ indicated that the two compounds were closely related, except for a 3-

hydroxy-3-methylbutyl group at C-18 of **1** [δ_H 1.17 (6H, s), δ_H 1.30 (3H, m), δ_H 1.46 (1H, m); δ_C 29.2 (C-22), 29.5 (C-25, 26), 42.4 (C-23), 70.5 (C-24)] instead of a 3-methylbut-3-enyl group at C-18 of garcinialiptone A.¹⁴ This assumption was further confirmed by the 1H - 1H COSY from H-18 to H₂-22 and H₂-22 to H₂-23 and HMBC correlations (see Fig. 2) from H₃-25, H₃-26 to C-23 (δ_C 42.4) and C-24 (δ_C 70.5). Regarding the relative configuration of **1**, the ROESY spectrum of **1** (see Fig. 3) showed correlations of H-34/H₃-33, H-34/H-7, which indicated that H-34, H-7, H₃-33 are on the same side of molecular plane which was the same as that of garcinialiptone A. Furthermore, the ROESY correlations between H-35/H₂-6, H₂-6/H₃-32 revealed that the H-35, H₃-32 and H₂-6 were on the opposite face to H-34, H-7, H₃-33. As a result of the above information, the structure of **1** was deduced completely as showed in the Fig. 1.

Garcixanthochymone B (**2**) was obtained as a white powder and gave the molecular formula $C_{38}H_{48}O_7$ with 15 degrees of unsaturation by its HR-ESI-MS ($[M + Na]^+$ m/z 639.3296, calcd 639.3292). Both **2** and **1** gave similar 1H and ^{13}C NMR spectra data, except for the signals of a 2-hydroxy-3-methylbut-3-enyl group (C(22)–C(26)) at C-18 in the former rather than 3-hydroxy-3-methylbutyl group in the latter. This finding was also supported by the ^{13}C NMR data of C-23 (δ 73.4), C-24 (δ 150.1), and C-25 (δ 109.7) and HMBC correlations (see Fig. 2) between H-23/C-26, 22, 25, 24. Thus, compound **2** was elucidated as 23-hydroxy garcinialiptone A, named as garcixanthochymone B.

Garcixanthochymone C (**3**) was obtained as a white powder and gave the molecular formula $C_{38}H_{50}O_8$ with 14 degrees of unsaturation by its HR-ESI-MS ($[M + H]^+$ m/z 635.3577, calcd 635.3578). In comparison of the NMR data of **3** with those of **1**, revealed that **3** contained an adamantyl phloroglucinol skeleton as that of **1**, but lacked resonances for an olefinic proton in 1H NMR and one trisubstituted double bond in ^{13}C NMR in **1**. In contrast, four additional carbon resonances, including one

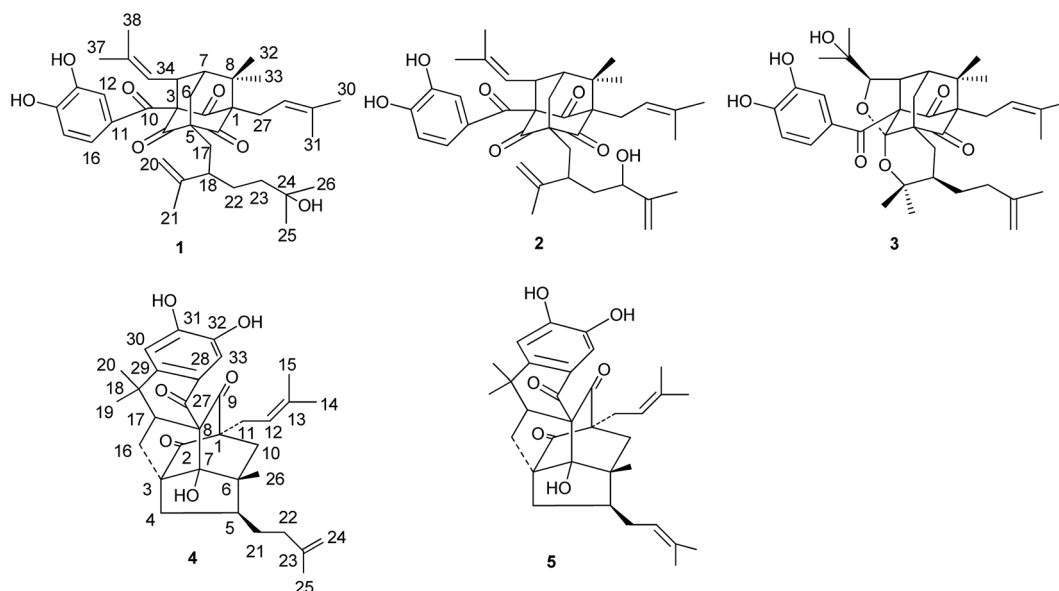


Fig. 1 Structures of compounds 1–5.



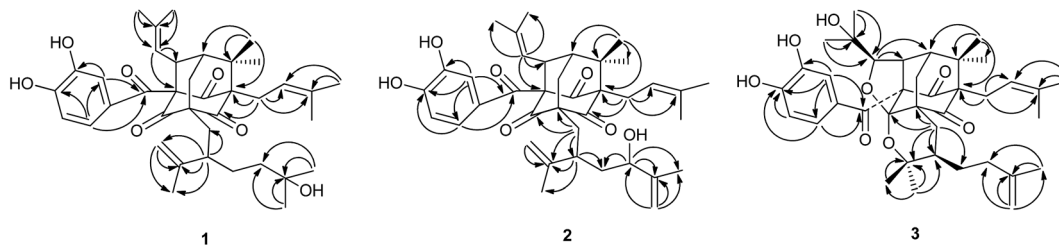


Fig. 2 Selected HMBC for compound 1–3.

oxygenated methine (δ_{C} 88.8), two oxygenated quaternary carbon (δ_{C} 71.7, 80.1) and one ketal carbon (δ_{C} 107.3) were found in 3. It suggested that the 2-methyl-2-propenyl group at C-34 and 2-isopropenyl-5-methyl-5-hydroxyhexyl group at C-5 in 1 had undergone an alternative cyclization. HMBC correlations (see Fig. 2) of δ_{H} 1.27 (CH₃-37) and 1.34 (CH₃-38) to the carbons at δ_{C} 88.8 (C-35) and 71.7 (C-36), of δ_{H} 4.26 (H-35) to the carbons at δ_{C} 52.2 (C-34) and 43.4 (C-7) indicated that a 2,3-dioxygenated 2-methylpropyl group was attached to C-34 instead of 2-methyl-2-propenyl group at C-34 in 1. HMBC correlations of δ_{H} 1.01 (CH₃-20) and 0.80 (CH₃-21) to the carbons at δ_{C} 80.1 (C-19) and 41.7 (C-18) indicated that 2-oxygenated isopropyl group was attached to C-18 instead of isopropenyl group at C-18 in 1. The aforementioned groups accounted for 12 out of 14 degrees of unsaturation in compound 3, indicating the presence of two additional rings. The chemical shifts of C-35 (δ_{C} 88.8) and C-19 (δ_{C} 80.1) and one more ketal carbon (δ_{C} 107.3) implied the ether ring closure between C-35 and C-4, C-19 and C-4 to construct 2-(1-hydroxy-1-methylethyl)-tetrahydrofuran ring fused with the phloroglucinol moiety at C-3 and C-4 and 2,2-dimethyl-3-(3-methylbut-3-enyl)-tetrahydropyrano ring fused with the phloroglucinol moiety at C-5 and C-4. Regarding the relative configuration of 3, the ROESY spectrum of 3 (see Fig. 3) showed correlations of H-34/H₃-33, H-34/H-7, H-34/H-35, H₂-6a/H₃-32 and H₂-6b/H₃-37 which indicated that H-34, H-35, H-7 and H₃-33 are on the same side of molecular plane and H₂-6 and H₃-32 are on the other side of molecular plane. The ROESY correlations between Heq-17 at δ_{H} 2.17 (1H, dd, J = 13.2, 3.2)/H-18 and H-18/H₃-21 revealed that the C-18 side chain on the tetrahydropyran ring should be in an equatorial configuration. Therefore, the structure of 3 was deduced completely as showed in the Fig. 1.

Garcixanthochymone D (4) was obtained as a white powder and gave the molecular formula C₃₃H₄₀O₆ with 14 degrees of unsaturation by its HR-ESI-MS ($[M + Na]^+$ m/z 555.2718, calcd 555.2723). The ¹H NMR spectrum exhibited three olefinic protons [δ_{H} 4.78 (1H, s), 4.80 (1H, s), 5.54 (1H, t, J = 7.5 Hz)], a 1,2,4,5-tetrasubstituted benzene moiety [δ_{H} 7.23 (1H, s), 8.06 (1H, s)], and six singlet methyls [δ_{H} 1.24 (3H, s), 1.42 (3H, s), 1.62 (3H, s), 1.63 (3H, s), 1.69 (3H, s), 1.78 (3H, s)]. The ¹³C NMR spectrum showed 33 carbons attributable to six methyls, seven methylenes, of which one is a terminal olefinic carbon [δ_{C} 110.7 (C-24)], five methines, of which two are aromatic and one olefinic [δ_{C} 112.4 (C-30), 115.1 (C-33), 120.8 (C-12)], 15 quaternary carbons, of which three are keto groups [δ_{C} 196.7 (C-27), 206.6 (C-9), 214.8 (C-2)]. From the above analysis of the ¹H and ¹³C-NMR spectroscopic data, it was implied that compound 4 is a tetracyclo [4.3.2.1^{1,6}.0^{3,7}] decane derivative similar to that of doitunggarcinone A.¹⁵ Further comparison of the NMR spectroscopic data of 4 with those of doitunggarcinone A revealed that a 1,2,4,5-tetrasubstituted benzene moiety fused at C-28 and C-29 in 4 instead of an *ortho*-disubstituted benzene moiety in doitunggarcinone A. This difference was evidenced by the HMBC correlations from H-30 (δ_{H} 7.23) to C-18 (δ_{C} 37.4) and C-28 (δ_{C} 129.0) and from H-33 (δ_{H} 8.06) to C-27 (δ_{C} 196.7). The relative configuration of doitunggarcinone A was previously deduced from ROESY correlations, and 3-methylbut-3-enyl at C-5 was determined as α -orientation which was opposite to CH₃-26 configuration. However, the relative stereochemistry of doitunggarcinone A have been revised by synthesis and comparison of the ¹³C NMR spectral data with those of garcibracteateone and 5-*epi*-garcibracteateone. It was suggested that the stereochemistry at C-5 is a β -orientation.¹⁶ In order to determine the relative stereochemistry of 4, ¹H, ¹³C-NMR and 2D-NMR of 4

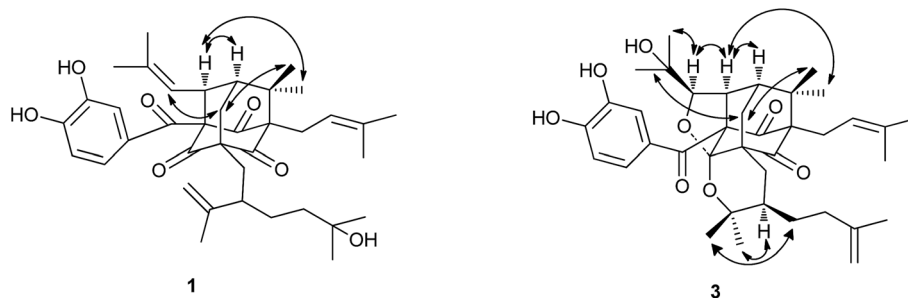
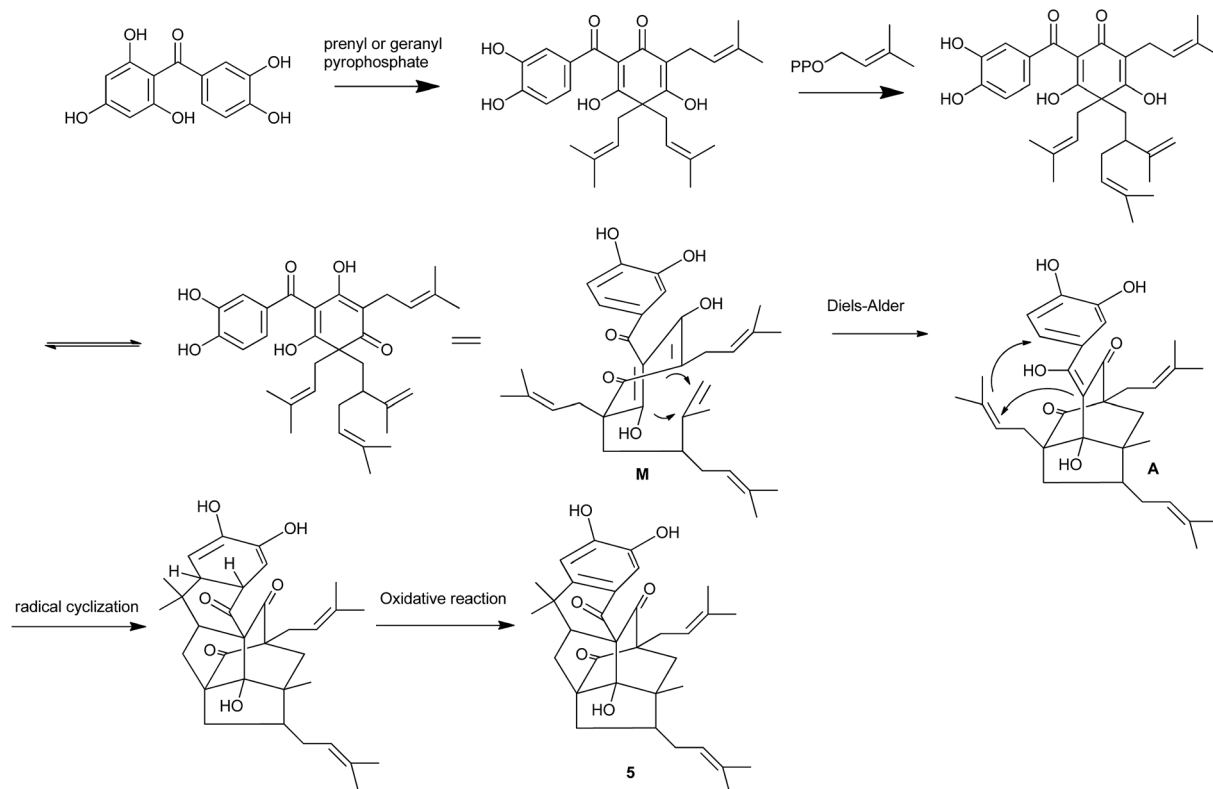


Fig. 3 Selected ROESY of compounds 1 and 3.





Scheme 1 Plausible biogenetic pathway for 5.

were measured again in CDCl_3 . By comparison of the ^{13}C NMR spectral data of **4** with those of doitunggarcinone A in CDCl_3 , it suggested that the relative stereochemistry of **4** is the same as that of doitunggarcinone A. Thus, the structure of **4** was deduced completely as showed in the Fig. 1.

Garcixanthochymone E (**5**) was obtained as a white powder and gave the molecular formula $\text{C}_{33}\text{H}_{40}\text{O}_6$ with 14 degrees of unsaturation by its HR-ESI-MS ($[\text{M} + \text{Na}]^+$ m/z 555.2726, calcd 555.2723), which was the same as that of **4**. By comparing the NMR data of **5** with those of **4**, it indicated that **5** was the isomer of **4**. However, the main difference found was that compound **5** displays ^1H NMR and ^{13}C NMR data for a 3-methylbut-2-enyl group at C-5 instead of a 3-methylbut-3-enyl group in **4**. This deduction was supported by ^1H - ^1H COSY correlations of H_2 -4/ H_5 -21/ H_2 -22. Thus, the structure of **5** was determined and named garcixanthochymone E.

Polycyclic polyprenylated acylphloroglucinols (PPAPs) are a class of secondary metabolites that usually possess bicyclo [3.3.1]nonane-2,4,9-trione, adamantyl, or homoadamantyl-like core structures.¹⁷ Up to now, 7 PPAPs have been isolated and identified from *G. xanthochymus*, such as xanthochymol and isoxanthochymol which belongs to bicycle [3.3.1] nonane-2,4,9-trione type.⁶ To the best of our knowledge, it was firstly time to isolate adamantly PPAPs from this plant. In addition, compound **3** featured two additional furan and pyrano rings incorporated into the tricyclo [3.3.1.1^{3,7}] decane system which are unique in natural adamantly PPAPs. Few adamantly-like PPAPs, such as sampsoniones A, B,¹⁸ hyperisampsins D¹⁹ and xerophenones A

and B²⁰ were found to possess a hemiketal carbon in the core skeleton, therefore, **3** represents the first example of adamantly PPAPs with an unusual ketal carbon in the adamantly skeleton.

Structurally, compound **4** and **5** was elucidated to possess an unprecedented tetracyclo-[4.3.2.1^{1,6}.0^{3,7}] decane core similar to that of garcibracteaton,²¹ doitunggarcinone A,¹⁵ hyperuralone B,²² hyphenrone R and S.²³ Up to now, this type of caged PPAPs is the most structurally complex PPAP natural product with a highly compact polycyclic ring system containing seven stereocenters, five of which are quaternary. The biosynthetic pathway of **5** was proposed as shown in Scheme 1. An intramolecular Diels-Alder reaction of M would form the key intermediate A, then underwent radical cyclization, tautomerization and oxidative rearomatization to give **5**.

The twelve known compounds including 7 xanthenes (**6**–**12**) and 5 flavonoids (**13**–**17**) were identified as 1,4,5-trihydroxyxanthone (**6**),²⁴ 1,3,7-trihydroxyxanthone (**7**),²⁵ 1,3,5-trihydroxyxanthone (**8**),²⁶ 1,5,6-trihydroxy-3-methoxyxanthone (**9**),²⁶ 1,3,6-trihydroxy-7-methoxyxanthone (**10**),^{27,28} 2,5-dihydroxy-1-methoxyxanthone (**11**),²⁹ 1,3,5,6-tetrahydroxy-2-isoprenylxanthone (**12**),³⁰ naringenin (**13**),³¹ 6-prenyl-4',5,7-trihydroxyflavone (**14**),³² GB-2a (**15**),³³ (\pm)-fukugetin (**16**),³⁴ volkensiflavone (**17**)³⁴ by comparison of the spectroscopic data with those reported in the literature.

3.2 The antiproliferative activities of isolated compounds

The antiproliferative activities of 17 compounds were evaluated on four human tumor cell lines (HepG2, A549, SGC7901, MCF-7). As shown in Table 3, only compounds **1**–**5** and **12** could



inhibit cancer proliferation derived from different tissues. PPAP derivatives 1–5 exhibited a stronger inhibitory effect on all four cancer cell, especially more effectively against HepG2 cell lines with IC₅₀ values less than 10 μ M. These results suggested that PPAPs might be responsible for the antitumor effect of *G. xanthochymus*.

4. Conclusions

In this study, chemical investigation on the fruits of *Garcinia xanthochymus* yielded five new PPAPs (1–5) and 12 known compounds. Their structures were identified by extensive spectroscopic analyses. To the best of our knowledge, this study represents the first reported account of the isolation of adamantyl PPAPs from this plant. All isolated compounds were evaluated anti-proliferative activities against four human tumor cells (HepG2, A549, SGC7901, MCF-7). As a result, PPAPs derivatives (1–5) displayed potential inhibitory activity against four human cancer cell lines with IC₅₀ values of 5.16–16.45 μ M. These data suggest that the extracts of the fruits of *G. xanthochymus* are potent candidates for cancer prevention.

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References

- 1 S. Karuppusamy, G. Muthuraja and K. M. Rajasekaran, *Indian J. Nat. Prod. Resour.*, 2011, **2**, 174–178.
- 2 B. Sharma, P. J. Handique and H. S. Devi, *J. Food Sci. Technol.*, 2015, **52**, 894–902.
- 3 K. U. Yilmaz, S. Ercisli, Y. Zengin, M. Sengul and E. Y. Kafkas, *Food Chem.*, 2009, **114**, 408–412.
- 4 S. Kumar, S. Sharma and S. K. Chattopadhyay, *Fitoterapia*, 2013, **89**, 86–125.
- 5 K. Trisuwan, S. Boonyaketgason, V. Rukachaisiriku and S. Phongpaichit, *Tetrahedron Lett.*, 2014, **55**, 3600–3602.
- 6 S. Baggett, P. Protiva, E. P. Mazzola, H. Yang, E. T. Ressler, M. J. Basile, I. B. Weinstein and E. J. Kennelly, *J. Nat. Prod.*, 2005, **68**, 354–360.
- 7 J. T. Lyles, A. Negrin, S. I. Khan, K. He and E. J. Kennelly, *Planta Med.*, 2014, **80**, 676–681.
- 8 R. K. Baslas and P. Kumar, *Acta Cienc. Indica, Chem.*, 1981, **7**, 31–34.
- 9 R. K. Baslas and P. Kumar, *Curr. Sci.*, 1979, **48**, 814–815.
- 10 M. Fu, H. J. Feng, Y. Chen, D. B. Wang and G. Z. Yang, *Chin. J. Nat. Med.*, 2012, **10**, 129–134.
- 11 M. Fu, X. Hu, J. Xu, Y. Chen and G. Z. Yang, *Nat. Prod. Res. Dev.*, 2014, **26**, 255–259.
- 12 P. Protiva, M. E. Hopkins, S. Baggett, H. Yang, M. Lipkin, P. R. Holt, E. J. Kennelly and I. B. Weinstein, *Int. J. Cancer*, 2008, **123**, 687–694.
- 13 N. Tanaka, Y. Takaishi, Y. Shikishima, Y. Nakanishi, K. Bastow, K. H. Lee, G. Honda, M. Ito, Y. Takeda, O. K. Kodzhimatov and O. Ashurmetov, *J. Nat. Prod.*, 2004, **67**, 1870–1875.
- 14 L. J. Zhang, C. T. Chiou, J. J. Cheng, H. C. Huang, L. M. Yang Kuo, C. C. Liao, K. F. Bastow, K. H. Lee and Y. H. Kuo, *J. Nat. Prod.*, 2010, **73**, 557–562.
- 15 C. Tantapakul, W. Phakhodee, T. Ritthiwigrom, S. Cheenpracha, U. Prawat, S. Deachathai and S. Laphookhieo, *J. Nat. Prod.*, 2012, **75**, 1660–1664.
- 16 H. P. Pepper, H. C. Lam, W. M. Bloch and J. H. George, *Org. Lett.*, 2012, **14**, 5162–5164.
- 17 R. Ciochina and R. B. Grossman, *Chem. Rev.*, 2006, **106**, 3963–3986.
- 18 L. H. Hu and K. Y. Sim, *Tetrahedron Lett.*, 1998, **39**, 7999–8002.
- 19 H. Zhu, C. Chen, J. Yang, X. Li, J. Liu, B. Sun, S. Huang, D. Li, G. Yao, Z. Luo, Y. Li, J. Zhang, Y. Xue and Y. Zhang, *Org. Lett.*, 2014, **16**, 6322–6325.
- 20 G. E. Henry, H. Jacobs, S. McLean, W. F. Reynolds and J. Yang, *Tetrahedron Lett.*, 1995, **36**, 4575–4578.
- 21 O. Thoison, D. D. Cuong, A. Gramain, A. Chiaroni, N. V. Hung and T. Séveneta, *Tetrahedron*, 2005, **61**, 8529–8535.
- 22 J. Zhang, J. Yang, Y. Liao, X. Yang, J. Ma, Q. Xiao, L. Yang and G. Xu, *Org. Lett.*, 2014, **16**, 4912–4915.
- 23 Y. Liao, S. Yang, X. Li, X. Yang and G. Xu, *Sci. China: Chem.*, 2016, **59**, 1216–1223.
- 24 M. Iin uma, H. Tosa, T. Tanaka, F. Asai and R. Shimano, *Phytochemistry*, 1995, **38**, 247–249.
- 25 C. N. Lin, M. I. Chung, S. J. Liou, T. H. Lee and J. P. Wang, *J. Pharm. Pharmacol.*, 1996, **48**, 532–538.
- 26 W. Li, Y. N. Sun, X. T. Yan, S. Y. Yang, C. W. Choi, J. W. Hyun, H. K. Kang, K. Y. Paek and Y. H. Kim, *Food Sci. Biotechnol.*, 2013, **22**, 945–949.
- 27 M. Linuma, H. Tosa, T. Ito, T. Tanaka and S. Riswan, *Chem. Pharm. Bull.*, 1996, **44**, 1744–1747.
- 28 H. Tosa, M. Linuma, K. I. Murakami, T. Ito, T. Tanaka, V. Chelladurai and S. Riswan, *Phytochemistry*, 1997, **45**, 133–136.
- 29 H. Minami, E. Takahashi, M. Kodama and Y. Fukuyama, *Phytochemistry*, 1996, **41**, 629–633.
- 30 K. Panthong, W. Pongcharoen, S. Phongpaichit and W. C. Taylor, *Phytochemistry*, 2006, **67**, 999–1004.
- 31 W. Gao, L. Yang, H. H. Li, C. Q. Liu, S. J. Liu and J. M. Hu, *Mod. Chin. Med.*, 2015, **17**, 311–314.
- 32 S. J. Lee, A. R. Wood, C. G. A. Maier, R. A. Dixon and T. J. Mabry, *Phytochemistry*, 1998, **49**, 2573–2577.
- 33 B. M. Abegaz, B. T. Ngadjui, E. Dongo and H. Tamboue, *Phytochemistry*, 1998, **49**, 1147–1150.
- 34 R. S. Compagnone, A. C. Suarez, S. G. Leitao and F. D. Monache, *Rev. Bras. Farmacogn.*, 2008, **18**, 6–10.

