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# CV-APC, a colorimetric and red-emitting fluorescent dual probe for the highly sensitive detection of palladium†

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A highly sensitive colorimetric and red-emitting fluorescent dual probe was synthesized and applied to imaging palladium species based on the Tsuji–Trost reaction. Notably, it displayed specific and turn-on fluorescent response toward Pd<sup>0</sup> with a vivid color change from yellow to purple in aqueous media. The detection limit was calculated to be as low as 0.78 nM. Furthermore, the probe was successfully applied for fluorescence imaging of palladium in living cells. All these properties made this probe a promising useful tool not only in environmental detection but also more widely in bioanalytical fields.

## Introduction

Palladium, one of the platinum-group elements, can catalyze the production of some compounds in fewer steps.<sup>1</sup> As a result of its excellent catalytic performances, palladium has been widely used in automotive, catalytic, electrical, dental, jewelry, chemical investment and some other fields.<sup>2</sup> However, a large amount of palladium is discharged into the environment with its excessive application. Consequently, people are exposed to palladium through inhalation and the food chain.<sup>3,4</sup> A recent study even indicated that palladium nanoparticles could penetrate the skin and reach the dermis through skin contact.<sup>5</sup> In contrast, people only know palladium is a potent sensitizer and may interfere with some cellular functions.<sup>1,6,7</sup> Besides, there are limited data about the effect of palladium on environment and human health. This may partly be attributed to the lack of new methods for palladium determination. Because of traditional analytic methods such as atomic absorption spectroscopy, X-ray fluorescence, *etc.*, which can't analyze palladium *in situ* and suffered from the cost of expensive instrumentation and high skilled professionals.<sup>8–10</sup> Therefore, it is highly urgent to develop new methods for monitoring the environmental and biological level of palladium and researching on its harmful effect on human health and environment.

In this case, the fluorescent probe has attracted a lot of attention because of its simplicity and sensitivity.<sup>11</sup> More importantly, palladium can be real-time monitored in biological samples using fluorescent probe,<sup>12–16</sup> which enable us to trace and study the presence of palladium in the human body better.

There are some drawbacks of current palladium fluorescent probes. Firstly, some palladium probes need to be monitored in organic solvents, are not conducive to the monitoring of palladium in the environment and living cells, which reduced the practicality; secondly, most of the palladium probes are prone to response to the fluorescence but not to the UV, and which means they were difficult to be recognized by the naked eye; thirdly, the sensitivity of the reported probes need to improved, in order to realize the trace analysis of palladium. All of these drawbacks will restrict their practical usage. Therefore, it is of great value and necessity to develop new colorimetric and fluorescent dual probes for the highly sensitive and selective detection of palladium both environment and biological samples.

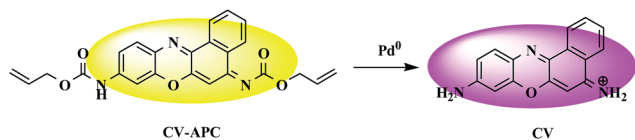
To overcome these drawbacks, a colorimetric and turn-on fluorescent probe, **CV-APC**, has been designed and synthesized. Herein, cresyl violet was chose as a signaling fluorophore because of its good solubility, excellent tissue permeability, tunable electronic system, long analytical wavelengths, high fluorescent quantum yield and efficient fluorescence quenching *via* the substitution of the amino group.<sup>17–20</sup> According to the literature and our previous work, **CV-APC** was designed by incorporation of allyl chloroformate into cresyl violet as the response site to trap Pd<sup>0</sup> according to the Tsuji–Trost reaction.<sup>21,22</sup> We anticipated that the incorporation of allyl chloroformate could induce the fluorescence quenching of the cresyl violet fluorophore as a result of the decreased intramolecular charge transfer (ICT) effect,<sup>21</sup> and the breakage of allyl chloroformate by Pb<sup>0</sup> would result in the releasing of free cresyl violet fluorophore, accompanied with the turn-on red-emitting fluorescence and colorimetric response (Scheme 1).

## Results and discussion

The desired probe **CV-APC** was easily prepared by attaching allyl chloroformate to cresyl violet acetate by one step reaction, and

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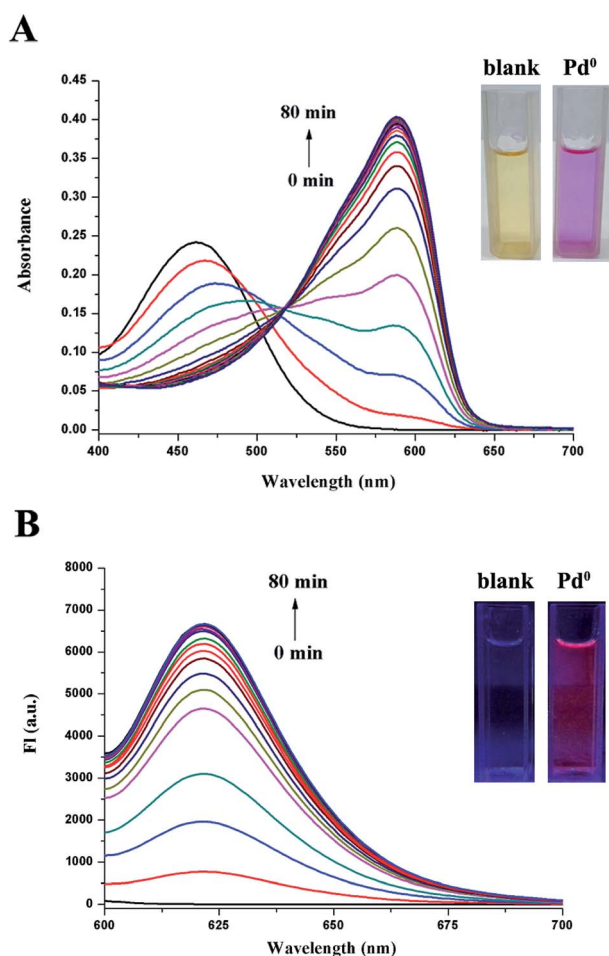


**Scheme 1** Proposed sensing process of CV-APC based on the Tsuji-Trost reaction.

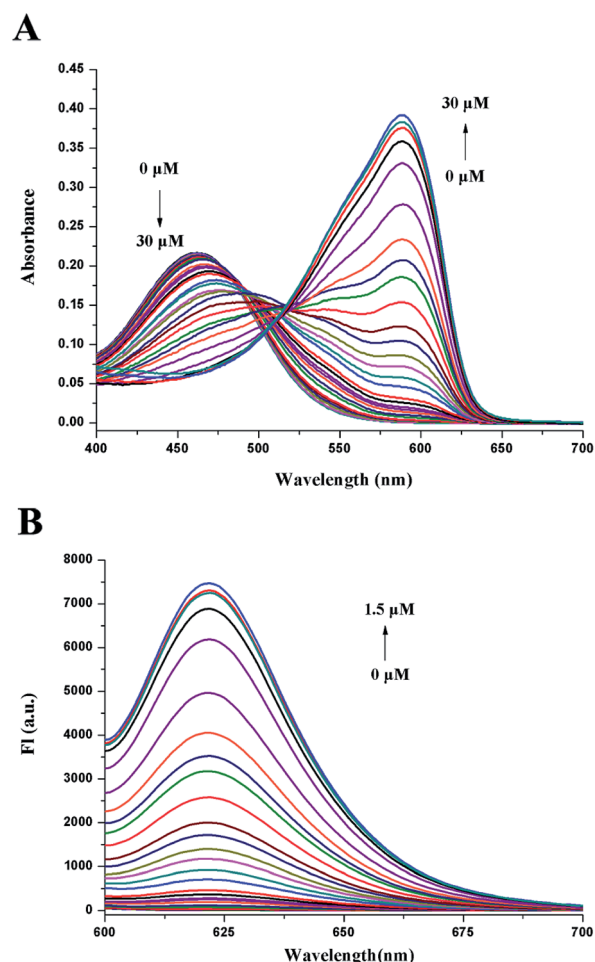
the detailed synthetic procedure is described in Scheme S1 in the ESI.† The time-dependent spectroscopic properties of probe CV-APC (10  $\mu\text{M}$ ) toward  $\text{Pd}^0$  were investigated in  $\text{CH}_3\text{CN}$ -PBS (1 : 1, v/v, Fig. 1) upon addition of  $\text{Pd}(\text{PPh}_3)_4$  (30  $\mu\text{M}$ ). The results indicated that CV-APC displays significant absorption and emission responses to  $\text{Pd}^0$ . In the absence of  $\text{Pd}^0$ , there is a predominant absorbance peak around 460 nm, after adding 3 equiv. of  $\text{Pd}(\text{PPh}_3)_4$ , the absorbance peak at 460 nm decreased gradually and a new absorbance peak sharply appeared at around 588 nm, which increased gradually with the incubation time and reached the maximum values after incubation for about 80 min at room

temperature (Fig. 1A). This large red shift (128 nm) of absorbance was accompanied with a vivid color change from yellow to purple (inset of Fig. 1A). Moreover, almost no emission peak was observed in the fluorescent spectrum of CV-APC alone. Upon the addition of  $\text{Pd}(\text{PPh}_3)_4$ , a dramatic enhancement of fluorescence intensity around 621 nm could be found and the intensity reached a plateau in 80 min at room temperature (Fig. 1B). The significant emission enhancement could be easily observed under the UV light (inset of Fig. 1B).

UV-vis and fluorescence titration experiments were further performed to study the sensing ability of CV-APC to  $\text{Pd}^0$ . As shown in Fig. 2A, the original absorbance peak at 460 nm decreased and a new peak at 588 nm appeared with the increased concentrations of  $\text{Pd}(\text{PPh}_3)_4$  (from 0 to 30  $\mu\text{M}$ ). Similarly, CV-APC had almost no fluorescence in the absence of  $\text{Pd}^0$ , the emission intensity around 621 nm greatly enhanced with the increased concentrations of  $\text{Pd}(\text{PPh}_3)_4$  (from 0 to 1.5  $\mu\text{M}$ , Fig. 1A). The turn-on fluorescence may be attributed to the releasing of free cresyl violet fluorophore through the broken of the carbamate bond, which was confirmed by the HRMS result



**Fig. 1** UV-vis (A) and fluorescence spectral changes (B) of CV-APC against time in the presence of  $\text{Pd}(\text{PPh}_3)_4$  (3 equiv.) in  $\text{CH}_3\text{CN}$ -PBS (1 : 1 v/v) at room temperature. Conditions:  $[\text{CV-APC}] = 10 \mu\text{M}$  and 0.5  $\mu\text{M}$ , respectively;  $\lambda_{\text{ex}} = 588 \text{ nm}$ . Inset: color changes and emission changes of CV-APC upon addition of  $\text{Pd}(\text{PPh}_3)_4$ .



**Fig. 2** UV-vis (A) and fluorescent (B) spectroscopic titration of CV-APC by stepwise addition of  $\text{Pd}(\text{PPh}_3)_4$  in  $\text{CH}_3\text{CN}$ -PBS (1 : 1 v/v) at room temperature. Conditions:  $[\text{CV-APC}] = 10 \mu\text{M}$  and 0.5  $\mu\text{M}$ , respectively;  $\lambda_{\text{ex}} = 588 \text{ nm}$ . The spectra were recorded at 80 min intervals.



(Fig. S4†). The plots of the fluorescence intensity fit linearly with the  $\text{Pd}^0$  concentration range of 0–0.55  $\mu\text{M}$  with a correlation coefficient of 0.99469 (Fig. S5†). The detection limit was calculated to be as low as 0.78 nM, which shows that **CV-APC** is highly sensitive to  $\text{Pd}^0$ , compared with reported probes for  $\text{Pd}^0$  (Table S1†). According to the WHO report, residue threshold of palladium in drugs should be less than 5.0–10.0 ppm (47.0–94.0  $\mu\text{M}$ ).<sup>23</sup> Therefore, **CV-APC** showed great potential in the trace analysis of palladium.

High selectivity is an important parameter to evaluate a newly designed fluorescent probe performance. **CV-APC** was treated with a wide variety of cations to assess its selectivity. As shown in Fig. 3 and S6,† only the addition of  $\text{Pd}^0$  can induced significant change on the absorption and fluorescence spectra of **CV-APC**. In contrast, even with the treatment of 10 equivalents of other cations such as  $\text{Fe}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Ni}^+$ ,  $\text{K}^+$ ,  $\text{Mn}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Zn}^{2+}$  and  $\text{Pb}^{2+}$  (Fig. 3 and S6†), the spectra of **CV-APC** had extremely weak disturbance. All these experimental results indicated that

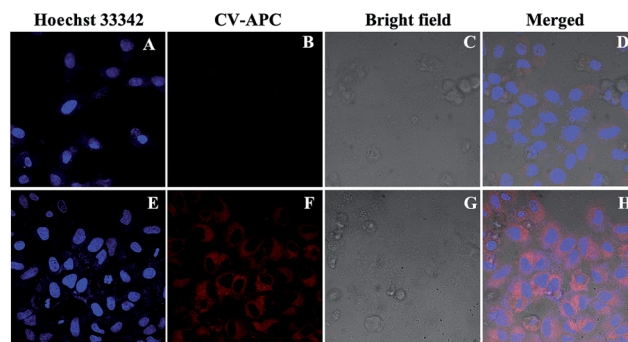


Fig. 4 Confocal fluorescence images of living HeLa cells. HeLa cells were co-stained with Hoechst 33342 (10  $\mu\text{M}$ ) and **CV-APC** (10  $\mu\text{M}$ ), without (A–D) and with (E–H) incubation of  $\text{Pd}^0$  ( $\text{Pd}(\text{PPh}_3)_4$ , 50  $\mu\text{M}$ ).

**CV-APC** exhibited excellent selectivity for  $\text{Pd}^0$  over other metal ions.

Encouraged by the above results, fluorescent imaging of palladium in living cells (HeLa cells) was further carried out using laser scanning confocal microscope to demonstrate the practical applicability of the probe in biological systems. In this regard, a standard MTT assay was performed to evaluate the cytotoxicity of **CV-APC** to HeLa cells. The results indicated that **CV-APC** exhibited very low cytotoxicity to living cells for 48 h even up to 50  $\mu\text{M}$  (Fig. S7†). In the study, Hoechst 33342, a commercial nuclear staining dye, was applied to co-stain cells.

As displayed in Fig. 4B, HeLa cells under normal culture condition showed almost no emission after incubation with 10  $\mu\text{M}$  **CV-APC**. In contrast, after the addition of 50  $\mu\text{M}$   $\text{Pd}^0$ , intense red fluorescence could be observed (Fig. 4F and H). The preliminary results indicated that **CV-APC** is both cell-permeable and capable of sensing palladium in living cells, which makes **CV-APC** a robust and effective tool for monitoring of palladium species in both environmental samples and in living cells.

## Conclusions

In summary, a highly sensitive colorimetric and red-emitting fluorescent dual probe for  $\text{Pd}^0$  has been developed based on the cresyl violet moiety through the  $\text{Pd}^0$ -mediated Tsuji–Trost reaction. The proposed probe, **CV-APC**, exhibits high specificity and sensitivity toward  $\text{Pd}^0$ . The detection limit of the probe is 0.78 nM, which is far less than the threshold of palladium in drugs. Only  $\text{Pd}^0$  can make an obvious color change from yellow to purple, indicating that the probe can be applied in visual detection. Moreover, the live cell imaging results revealed that **CV-APC** holds great potential for tracking intracellular palladium species. All these properties make this probe a promising and robust tool not only in environmental monitoring but also in more widely bioanalytical fields.

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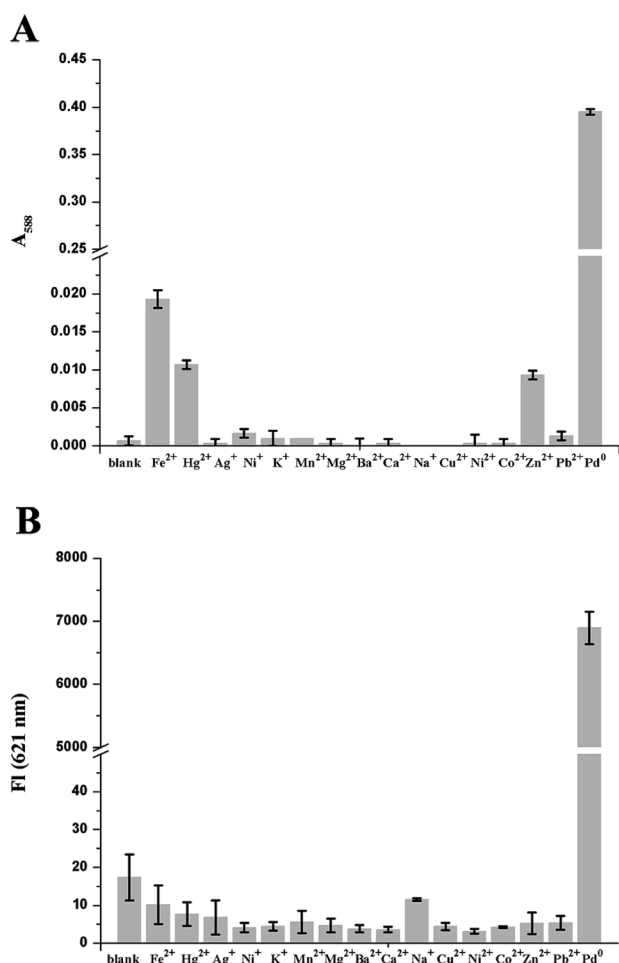


Fig. 3 Absorbance at 588 nm (A) and fluorescence intensity at 621 nm (B) of **CV-APC** in the presence of  $\text{Pd}(\text{PPh}_3)_4$  (3 equiv.), and the excess of representative metal ions (10 equiv.) in  $\text{CH}_3\text{CN}$ –PBS (1 : 1 v/v) at room temperature. Conditions:  $[\text{CV-APC}] = 10 \mu\text{M}$  and 0.5  $\mu\text{M}$ , respectively;  $\text{CH}_3\text{CN}$ –PBS (1 : 1, v/v),  $\lambda_{\text{ex}} = 588 \text{ nm}$ . The spectra were recorded after incubation for 80 min.



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## Notes and references

- 1 C. E. Garrett and K. Prasad, *Adv. Synth. Catal.*, 2004, **346**, 889–900.
- 2 M. Paraskevas, F. Tsopelas and M. Ochsenkühn-Petropoulou, *Microchim. Acta*, 2012, **176**, 235–242.
- 3 J. Kielhorn, C. Melber, D. Keller and I. Mangelsdorf, *Int. J. Hyg. Environ. Health*, 2002, **205**, 417–432.
- 4 K. Ravindra, L. Bencs and R. Van Grieken, *Sci. Total Environ.*, 2004, **318**, 1–43.
- 5 F. Larese Filon, M. Crosera, M. Mauro, E. Baracchini, M. Bovenzi, T. Montini, P. Fornasiero and G. Adami, *Environ. Pollut.*, 2016, **214**, 497–503.
- 6 M. Schmid, S. Zimmermann, H. F. Krug and B. Sures, *Environ. Int.*, 2007, **33**, 385–390.
- 7 E. Orion, H. Matz and R. Wolf, *Contact Dermatitis*, 2003, **49**, 216–217.
- 8 S. H. Fu, Z. F. Liu, S. P. Liu, J. T. Liu and A. E. Yi, *Anal. Chim. Acta*, 2007, **599**, 271–278.
- 9 C. Locatelli, D. Melucci and G. Torsi, *Anal. Bioanal. Chem.*, 2005, **382**, 1567–1573.
- 10 B. Dimitrova, K. Benkhedda, E. Ivanova, F. Adams and J. Anal, *At. Spectrosc.*, 2004, **19**, 1394–1396.
- 11 Y. Yang, Q. Zhao, W. Feng and F. Li, *Chem. Rev.*, 2012, **113**, 192–270.
- 12 J. W. Yan, X. L. Wang, Q. F. Tan, P. F. Yao, J. H. Tan and L. Zhang, *Analyst*, 2016, **141**, 2376–2379.
- 13 F. Liu, J. Du, M. Xu and G. Sun, *Chem.-Asian J.*, 2016, **11**, 43–48.
- 14 Z. Y. Xu, X. L. Wang, J. W. Yan, J. Li, S. Guan and L. Zhang, *RSC Adv.*, 2016, **6**, 43539–43542.
- 15 X. Wang, Z. Guo, S. Zhu, H. Tian and W. Zhu, *Chem. Commun.*, 2014, **50**, 13525–13528.
- 16 M. Wang, X. Liu, H. Lu, H. Wang and Z. Qin, *ACS Appl. Mater. Interfaces*, 2015, **7**, 1284–1289.
- 17 Q. Q. Wan, X. H. Gao, X. Y. He, S. M. Chen, Y. C. Song, Q. Y. Gong, X. H. Li and H. M. Ma, *Chem.-Asian J.*, 2014, **9**, 2058–2062.
- 18 J. Zhang, A. Shibata, M. Ito, S. Shuto, Y. Ito, B. Mannervik, H. Abe and R. Morgenstern, *J. Am. Chem. Soc.*, 2011, **133**, 14109–14119.
- 19 Q. Q. Wan, Y. C. Song, Z. Li, X. H. Gao and H. M. Ma, *Chem. Commun.*, 2013, **49**, 502–504.
- 20 J. Alijan-pour, K. Abrari, T. L. Bluki, M. T. Ghorbanian, I. Goudarzi, M. E. Salmani and M. Mirshekar, *Pharmacol., Biochem. Behav.*, 2012, **102**, 321–328.
- 21 F. Song, A. L. Garner and K. Koide, *J. Am. Chem. Soc.*, 2007, **129**, 12354–12355.
- 22 M. E. Moragues, A. Toscani, F. Sancenón, R. Martínez-Mañez, A. J. P. White and J. D. E. T. Wilton-Ely, *J. Am. Chem. Soc.*, 2014, **136**, 11930–11933.
- 23 M. Wang, X. Liu, H. Lu, H. Wang and Z. Qin, *ACS Appl. Mater. Interfaces*, 2015, **7**, 1284–1289.

