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The reactions of α -amino acids and α -amino acid esters with high valent transition metal halides: synthesis of coordination complexes, activation processes and stabilization of α -ammonium acylchloride cations[†]

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Titanium tetrachloride smoothly reacted with a selection of α -amino acids (aaH) in CH₂Cl₂ affording yellow to orange solid coordination compounds, 1a-d, in 70-78% yields. The salts [NHEt][TiCl₄(aa)], 2a-b, were obtained from TiCl₄/aaH/NEt₃ (aa = L-phenylalanine, N,N-dimethylphenylalanine), in 60–65% yields. The complex $Nb_2Cl_8\{\mu-\kappa^2O,\kappa^2N-iCHCH,CH,CH(N),CH(O)\}$, 3, was isolated from the reaction of L-proline with NbCl₅/NHⁱPr₂, performed in CH₂Cl₂ at room temperature. The X-ray structure of 3 features a bridging (E)-1,2-bis(3,4-dihydro-2H-pyrrol-5-yl)ethene-1,2-diolate ligand, resulting from the unprecedented C-C coupling between two proline units. Unusually stable *a*-ammonium acyl chlorides were prepared by the reactions of PCl_5/MCl_n ($MCl_n = NbCl_5$, WCl_6) with \bot -proline, N,N-dimethylphenylalanine, sarcosine and \bot methionine. MX₅ (M = Nb, Ta; X = F, Cl) reacted with L-leucine methylester and L-proline ethylester to give ionic coordination compounds, $[MX_4L_2][MX_6]$ (M = Nb, L = Me₂CHCH₂CH(NH₂)CO₂Me, X = F, **9**; Cl, **11a**; M = Nb, X = Cl, L = HNCH, CH, CH, CH, CH, CD, Et, **11c**; Ta, **11d**), in moderate to good yields. [NbCl₅(Me₂CHCH₂CHNH₃CO₂Me)][NbCl₆], 12, was isolated as a co-product of the reaction of NbCl₅ with L-leucine isopropylester, and crystallographically characterized. The reaction of NbCl₅ with L-serine isopropylester afforded NbCl₃(OCH₂CHNHCO₂ⁱPr), 13, in 66% yield. The activation of the ester O-R bond was observed in the reactions of L-leucine methyl ester with NbF₅ and L-proline ethyl ester with MBr₅ (M =Nb, Ta), these reactions proceeding with the release of EtF and EtBr, respectively. All the metal products were characterized by analytical and spectroscopic methods, while DFT calculations were carried out in order to provide insight into the structural and mechanistic aspects.

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Introduction

 α -Amino acids constitute a class of organic compounds arousing great interest in synthetic chemistry, in view of their easy availability and low toxicity,¹ the typical presence of a stereogenic centre (making them suitable substrates for asymmetric catalysis)² and the possibility of firmly coordinating metal ions.³

The esterification of the carboxylic acid moiety is one of the most viable modifications of the α -amino acid skeleton, and indeed a good number of α -amino acid esters have been synthesized and employed with reference to several application fields.⁴ Metal complexes containing α -amino acid esters as ligands are especially relevant to bio-inorganic chemistry, being useful to the synthesis of peptides,^{5,6} as biological models,⁷ and as scaffolds for the development of new drugs.⁸ Furthermore, α -amino acids and α -amino acid ester metal complexes, being possible chiral sources,⁹ have found increasing attention as privileged, potential catalysts for environmentally friendly asymmetric syntheses.^{2*h*,10}

It is noteworthy that the large majority of these studies refer to middle to late transition metals, whereas very little is known about the parallel chemistry with early transition metal compounds. In particular, the homoleptic halides of high valent

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elements (oxidation state ≥ 4) belonging to groups 4, 5 and 6, HVTMH, are strongly oxophilic species, usually very air sensitive and incompatible with water. This characteristic has probably discouraged the linking with an "opposite world", *i.e.* the exploration of the reactivity with α -amino acids, which in turn exhibit high water affinity, and their simple derivatives such as α -amino acid esters.

As a matter of fact, the coordination chemistry of HVTMH with α -amino acid esters still remains an unexplored field of research, with the exception of a former synthetic study regarding MoCl₅.¹¹ Similarly, the only information available up to 2014 on the interaction of HVTMH with α -amino acids, in the absence of further reactants, is a note dealing with the reactivity of TiCl₄ with glycine.¹² In all of the cases, the structural characterization of the products relied on limited data.

Recently, in the framework of our interest in the chemistry of HVTMH with naturally occurring compounds,¹³ we have found that $MoCl_5$ and WCl_6 behave as chlorinating agents towards natural α -amino acids, affording fairly stable α -ammonium acylchloride salts.¹⁴

On the other hand, the interaction of MX_5 (M = Nb, Ta; X = Cl, Br) with α -amino acids leads to dinuclear complexes containing bridging α -amino acidato ligands *via* HX release.¹⁵ Subsequent activation of the coordinated α -amino acidato moiety has been observed in mild conditions in some specific cases, leading to iminium salts.

Herein, we will present an extension of our study on the reactivity between α -amino acids and HVTMH, including the synthesis of TiCl₄ derivatives, the unprecedented metal mediated C–C dimerization of a α -amino acid (L-proline) and the stabilization of otherwise reactive α -ammonium acylchloride cations. We will also describe some reactivity of α -amino acid ester with niobium and tantalum pentahalides.^{16,17} All the reactions were carried out in a weakly coordinating solvent (CH₂Cl₂) by using enantiopure L-organic reactants in case. The metal products were characterized by elemental analysis, IR and NMR spectroscopy, while DFT calculations assisted the structural characterization. A DFT study was performed also in order to shed some light on the NbCl₅-directed unusual L-proline pseudo dimerization.

Results and discussion

Reactivity of MCl_n with α -amino acids

Titanium tetrachloride. Titanium tetrachloride smoothly reacted with a series of α-amino acids in dichloromethane affording moisture sensitive, yellow to orange solid materials 1a–d, in 70 to 78% yields (Scheme 1). Compounds 1 are coordination adducts: in this respect, the reactivity of TiCl₄ with α-amino acids differs from that of MCl₅ (M = Nb, Ta), featured by HCl release,¹⁵ and from those of MoCl₅ and WCl₆, leading to Cl/O interchange products (see Introduction).¹⁴

Compounds **1a–d** were characterized by analytical and spectroscopic methods. The IR spectra (solid state) contain one medium and one strong intensity absorption in the range 1600–1400 cm⁻¹. These two absorptions are due, respectively, to the asymmetric (v_a) and the symmetric (v_s) stretching vibrations of





the carboxylato group. In general, the wavenumber difference $(\Delta \nu_{\rm a-s} = \nu_{\rm a} - \nu_{\rm s})$ is considered as a useful parameter to discriminate between monodentate, chelating, and bridging bidentate carboxylato ligands. $\Delta \nu_{\rm a-s}$ values within the range 100 to 150 cm⁻¹ are typical of either chelating or bridging bidentate carboxylates.¹⁸ In view of the IR data available for **1a-d**, *i.e.* $\Delta \nu_{\rm a-s}$ varies between 103 (**1a**) and 135 (**1d**) cm⁻¹, and the DFT results (*vide infra*), we propose a bridging bidentate-coordination fashion. This implies that the amino acid ligand should be coordinated to titanium as a zwitterion. Accordingly, a broad IR absorption is observed at 3091 cm⁻¹ in the IR spectrum of **1d**, assigned to ammonium N–H stretching vibration.

The geometry proposed on the basis of spectroscopic data was supported by DFT calculations on the possible isomers of **1b**. The dinuclear structure $[TiCl_4(\mu-\kappa^2O-O_2CCH(CH_2Ph)NH_3)]_2$, depicted in Fig. 1, resulted meaningfully more stable than mononuclear structures (see Fig. S1 given as ESI[†]).

Compounds **1a–d** display low solubility in common organic solvents. The NMR spectra were recorded in CD₃CN, displaying single sets of resonances. The ¹H NMR spectra exhibit broad



Fig. 1 DFT-optimized geometry of the most stable isomer of **1b** (C-PCM/ M06 calculations). Selected computed bond lengths (Å): Ti1–O 2.020, 2.081; Ti2–O 1.990, 2.080; Ti1–Cl (*trans* O) 2.295, 2.232; Ti2–Cl (*trans* O) 2.232, 2.304; Ti1–Cl (*cis* O) 2.266, 2.392; Ti2–Cl (*cis* O) 2.291, 2.394; C–O 1.244, 1.261, 1.251, 1.252; N–H 1.025, 1.028, 1.038, 1.027, 1.028, 1.037. Selected computed angles (°): O–Ti1–O 86.1; O–Ti2–O 84.6; Ti1–O–C 149.8, 151.4; Ti2–O–C 139.2, 151.3; O–C–O 124.9, 126.4.



resonances in the 7.7–7.0 ppm range, related to the uncoordinated ammonium group. The ¹³C-NMR spectra of the more soluble **1a–b** show the resonance of the carboxylate carbon at 176.1 and 170.0 ppm, respectively. These values are similar to those reported for *O*,*O*-coordinated α -amino acids in NbCl₅ derivatives.¹⁵

The coordination of organic species to high valent transition metal chlorides represents, in a number of cases, the preliminary step of some activation process.^{13c,19} The activation is favoured by the strong Lewis acidity of the metal centre, and may be triggered by the addition of a Brönsted base. For instance, Peryshkov and coworkers recently described a C–H bond activation reaction of nitriles by means of NEt₃ upon coordination to TaCl₅.²⁰

Thus, the reaction of **1b** with NEt₃ proceeded with selective deprotonation of the ammonium group; analogous result was achieved by treatment of a $\text{TiCl}_4/\text{L-}N,N$ -dimethylphenylalanine mixture with NEt₃ (Scheme 2).²¹ The reactions of **1a,c,d** with NEt₃ were not straightforward, leading to non identified compounds; the solid isolated from **1a**/NEt₃ revealed to be paramagnetic.

The CH_2Cl_2 soluble compounds **2a-b** (Scheme 2) were isolated by addition of hexane to the respective reaction mixtures.²²

The ¹H NMR spectra of **2a–b** display a low field resonance accounting for the triethylammonium proton (*e.g.* at 9.09 ppm in the case of **2a**); the resonances of the N-bound protons within the anion undergo significant upfield shift on going from the amino acid unit in **1b** to the amino acidate one in **2b** ($\Delta \delta > 3$ ppm). The IR spectra of **2a–b** exhibit a strong absorption around 1700 cm⁻¹: this evidence suggests O,N-coordination of the aminoacidate moiety, leaving a uncoordinated C=O bond. The geometries of the **2a,b** anions were DFT optimized, on considering either mononuclear and dinuclear structures as starting points (Fig. S2[†]). Thus, mononuclear compounds bearing N,Ochelating α -aminoacidates (Fig. 2) resulted much more stable than dinuclear homologues (see ESI[†] for more details).

Niobium pentachloride. We reported that the 2 : 1 reactions of NbX₅ (X = Cl, Br) with a variety of α -amino acids afforded dinuclear α -aminoacidate complexes *via* HCl release. The addition of a further equivalent of organic reactant resulted in the decarboxylation of one amino acidate moiety, with consequent formation of iminium salts and Nb-formate species (see Scheme 3, showing the specific case of *N*,*N*-dimethylphenylalanine).¹⁵

With the aim of exploring the possibility of further activation pathways, we investigated the reactions of $Nb_2Cl_9(\alpha$ -amino



Fig. 2 DFT-optimized geometries of the most stable anions of **2a** and **2b** (C-PCM/M06 calculations). Selected computed bond lengths for **2a** (Å): Ti–O 1.909; Ti–N 2.240; Ti–Cl (*trans* O) 2.321; Ti–Cl (*trans* N) 2.263; Ti–Cl (*trans* Cl) 2.340, 2.353; C–O 1.316; C=O 1.212; N–H 1.018, 1.021. Selected computed angles for **2a** (°): O–Ti–N 76.0; O–Ti–Cl 87.8, 88.8, 97.6, 160.9; C–O–Ti 127.5. Selected computed bond lengths for **2b** (Å): Ti–O 1.893; Ti–N 2.434; Ti–Cl (*trans* O) 2.309; Ti–Cl (*trans* N) 2.271; Ti–Cl (*trans* Cl) 2.333, 2.354; C–O 1.316; C=O 1.212. Selected computed angles for **2b** (°): O–Ti–N 73.2; O–Ti–Cl 88.6, 89.7, 98.5, 160.3; C–O–Ti 130.2.



Scheme 3 Reaction of NbCl₅ with *N*,*N*-dimethylphenylalanine.



Scheme 4 Unusual activation of L-proline by NbCl₅/amine.

acidate) complexes with NEt₃. In general, the amino acidate moiety did not undergo activation under these conditions, with an exception provided by the Nb₂Cl₉(*t*-prolinate)/NEt₃ system. This latter evolved into a complicated mixture of products, including minor amounts of Nb₂Cl₈{ μ - κ^2 O, κ^2 N - [CH₂CH₂CH₂C(N)C(O)]₂, **3**. The use of NHⁱPr₂ in the place of NEt₃ allowed to isolate red crystals of 3 (12% yield), and also yellow crystals of [NH₂ⁱPr₂] [NbCl₆], 4 (40% yield), Scheme 4.



Fig. 3 ORTEP drawing of 3. Displacement ellipsoids are at the 50% probability level.

Table 1 Selected bond lengths (Å) and angles (°) for 3^a

Nb(1)-Cl(1)	2.3319(13)	Nb(1)-Cl(2)	2.2833(11)	
Nb(1)-Cl(3)	2.3454(12)	Nb(1)-Cl(4)	2.3453(12)	
Nb(1)-O(1)	1.927(3)	Nb(1)-N(1)	2.235(4)	
C(1)-O(1)	1.357(6)	$C(1)-C(1_1)$	1.337(10)	
C(1)-C(2)	1.471(7)	C(2)-C(3)	1.488(7)	
C(3)-C(4)	1.530(6)	C(4)-C(5)	1.528(6)	
N(1)-C(2)	1.276(6)	N(1)-C(5)	1.477(6)	
Cl(1)-Nb(1)-O(1)	158.18(10)	Cl(2)-Nb(1)-N(1)	167.62(11)	
Cl(3)-Nb(1)-Cl(4)	170.32(5)	O(1)-Nb(1)-N(1)	73.03(14)	
Nb(1)-O(1)-C(1)	122.9(3)	Nb(1)-N(1)-C(2)	117.6(3)	
Nb(1)-N(1)-C(5)	132.0(3)	C(2)-N(1)-C(5)	110.4(4)	
N(1)-C(2)-C(3)	115.8(4)	N(1)-C(2)-C(1)	110.7(4)	
C(1)-C(2)-C(3)	133.5(4)	C(2)-C(3)-C(4)	102.3(4)	
C(3)-C(4)-C(5)	105.8(4)	C(4)-C(5)-N(1)	105.7(4)	
O(1)-C(1)-C(2)	114.6(4)	O(1)-C(1)-C(1_1)	120.2(6)	
$C(2)-C(1)-C(1_1)$	125.2(6)			
^{<i>a</i>} Symmetry transfor	rmation used to	generate C(1_1): $-x +$	1, -y, -z + 1.	

The X-ray structure of **3** is shown in Fig. 3, with relevant bonding parameters listed in Table 1; the X-ray structure of **4** is given as ESI (Fig. S3; Tables S1A and S1B[†]).

Complex 3 displays crystallographic $\overline{1}$ (C_i) symmetry with the inversion centre located on the middle of the C(1)–C(1_1) bond. The complex is composed of an unprecedented anionic $[CH_2CH_2CH(N)CH(O)]_2^{2*}$ ligand (*E*-1,2-bis(3,4-dihydro-2*H*-pyrrol-5-yl)ethene-1,2-diolate) that is μ - κ^2 O, κ^2 N-coordinated over two [NbCl₄]⁺ cationic fragments. Such anionic ligand is almost perfectly planar (mean deviation from the least squares plane 0.0387 Å), being the two Nb atoms respectively 0.1879 Å above and below this plane. C(1), C(2) and N(1) [sum angles 360.0(9), 360.0(7) and 360.0(6)°, respectively] show a perfect sp² hybridization, and the C(1)–C(1_1) [1.357(6) Å] and C(2)–N(1) [1.276(6) Å] distances are typical for double bonds.²³

The {C₂O₂} core of the (*E*)-1,2-bis(3,4-dihydro-2*H*-pyrrol-5-yl) ethene-1,2-diolate ligand in **3** is a fully deprotonated 1,2-enediol. In general, 1,2-enediols are quite unstable species,²⁴ whose stabilization may be supplied by hydrogen-bonded protons,²⁵ or by chelating (N,O) coordination to transition metals.²⁶



Scheme 5 NbCl₅-promoted C–C pseudo-dimerization of L-proline (A); peptide bond formation from two L-proline units (B).

Basically, the process leading to **3** is a C–C bond forming condensation of two proline units (Scheme 5A). Differently, typical α -amino acid condensation generates a peptide bond (Scheme 5B).²⁷

In order to gain some insight into the mechanism of the low yield formation of **3**, a DFT study was undertaken (see Scheme S1 in the ESI[†]). It seems plausible that the C–C bond forming step consists in the coupling of two acylchloride units, accompanied by the release of HCl and assisted by the amine (see Scheme S1,[†] E \rightarrow F \rightarrow G). Indeed the side reactions NHⁱPr₂ + HCl \rightarrow [NH₂ⁱPr₂]Cl and [NH₂ⁱPr₂]Cl + NbCl₅ \rightarrow **4** should contribute to decrease the ΔG variation of the process leading to **3**.

The crystals of **3** exhibited insufficient solubility in suitable deuterated solvents, thus preventing the NMR characterization.

 $MCl_n/phosphorous$ pentachloride ($MCl_n = NbCl_5, WCl_6$). The carboxylic acid to acyl chloride conversion is an important preliminary step for the subsequent functionalization of α amino acids.²⁸ PCl₅ has been traditionally employed as Clsource in order to obtain the relevant a-ammonium acylchloride salts (*Fischer procedure*); the counterion is Cl^{-} or $[PCl_6]^-$ depending on the employed PCl_5/α -aminoacid molar ratio.²⁹ α-Ammonium acylchloride species stable at room temperature have been obtained only with primary ammonium groups and in the absence of donor atoms within the side chain. On the other hand, in the case of the L-proline derivative (secondary N), both Cl⁻ and [PCl₆]⁻ salts undergo quick degradation at room temperature, due to HCl release and subsequent condensation reactions.29 Furthermore, the reactions of PCl₅ with L-N,N-dimethylphenylalanine (tertiary N), sarcosine (secondary N) and L-methionine (thioether group), in CH₂Cl₂, proceed with the formation of complicated mixtures of products (¹H and ³¹P NMR spectroscopy). Thus, the L-N,Ndimethylphenylalanine and L-methionine acylchloride derivatives have not been known heretofore, while the highly moisture sensitive [NH(Me)CH2COCl][WOCl5] has been recently obtained by ourselves from sarcosine/WCl6.14a

Some of us recently reported^{13b,29} a straightforward and clean route to unusually stable salts of the acylchloride derivative of Lproline, by combination of the *traditional* PCl₅-chlorinating reaction with the considerable stability imparted by the $[MCl_6]^-$ (M = Nb, Ta) anions, Scheme 6.^{13b-c,30}

We reckoned that the easily available $[NbCl_6]^-$ anion could provide stability also to other unstable/unknown α -ammonium acylchloride cations (see above). Therefore, we tried to optimize

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The iminium salt $[PhCH_2=NMe_2][NbCl_6]$ (see Scheme 3) and the adduct $NbCl_5(O=PCl_3)$, 6, identified by comparison of the



Scheme 6 Stable pyrrolidinium-2-carbonylchloride salts from L-proline and MCl_5 (M = Nb, Ta).



Scheme 7 Formation of otherwise unstable α -ammonium acylchloride cations from α -amino acids and niobium and tungsten chlorides.

crystal cell data with those reported in the literature,³⁶ were obtained as minor products from NbCl₅/L-*N*,*N*-dimethylpheny-lalanine and NbCl₅/sarcosine, respectively.

The synthetic approach leading to 5a-b exploits the M–Cl (M = P, Nb) bond energy scale,^{13b,37} making PCl₅ a preferential chlorinating agent respect to NbCl₅, and the stability of the [NbCl₆]⁻ anion. Similar considerations led us to test the PCl₅/WCl₆ mixture; it should be noted that anionic simple derivatives of WCl₆ (*i.e.*, WCl₆, WOCl₅⁻) have recently proposed as effective partners for the stabilization of otherwise reactive cations.^{14a,38}

Hence, the reactions of PCl_5/WCl_6 (1:1 mixture) with L-proline, L-*N*,*N*-dimethylphenylalanine, sarcosine and L-methionine proceeded with PCl_5 to $POCl_3$ conversion (³¹P NMR), and straightforwardly afforded the respective α -ammonium acylchloride cations (Scheme 7). According to elemental analyses and magnetic measurements,³⁸ the cations were isolated in good yields as $[WCl_6]^-$ salts, **7a–b**, respectively from $PCl_5/WCl_6/$ L-proline and $PCl_5/WCl_6/L-N,N$ -dimethylphenylalanine. Otherwise, different anions were presumably associated with sarcosine and methionine derivatives, including $[WOCl_5]^-$ (few crystals of $[MeNH_2CH_2C(=O)Cl][WOCl_5]$ were isolated and X-ray characterized) and W(n) species.

The characterization of the 1:1 mixture WCl₆/PCl₅ suggested that both chlorides remained intact when mixed together (see Experimental for details). This implies that the WCl₆ to WCl₆⁻ reduction, as clearly observed in 7**a**-**b**, is promoted by the α -amino acid. Analogous WCl₆ reduction has



Fig. 4 ORTEP drawing of 5a. Displacement ellipsoids are at the 50% probability level.

Table 2 Selected bond lengths (Å) and angles (°) for 5a						
Nb(1)-Cl(1)	2.3305(11)	Nb(1)-Cl(2)	2.3344(11)			
Nb(1)-Cl(3)	2.2921(11)	Nb(1)-Cl(4)	2.3475(11)			
Nb(1)-Cl(5)	2.3844(11)	Nb(1)-Cl(6)	2.4206(11)			
C(1)-Cl(7)	1.758(5)	C(1)-O(1)	1.178(5)			
C(1)-C(2)	1.516(7)	C(2) - C(3)	1.553(6)			
C(2)-N(1)	1.500(6)	N(1)-C(4)	1.499(5)			
N(1)-C(5)	1.501(6)	C(3) - C(6)	1.516(6)			
Cl(1)-Nb(1)-Cl(4)	176.12(5)	Cl(2)-Nb(1)-Cl(5)	174.25(5)			
Cl(3)-Nb(1)-Cl(6)	179.14(4)	O(1)-C(1)-Cl(7)	120.6(4)			
O(1)-C(1)-C(2)	125.4(4)	Cl(7)-C(1)-C(2)	113.9(3)			
C(1)-C(2)-C(3)	115.1(4)	C(1)-C(2)-N(1)	107.3(4)			
C(2) - C(3) - C(6)	115.1(4)	C(2)-N(1)-C(4)	111.4(3)			
C(2)-N(1)-C(5)	114.9(3)	C(4)-N(1)-C(5)	111.4(4)			

been previously observed in a number of cases by interaction with organic compounds.^{38,39}

All the α -ammonium acylchloride cations produced from PCl₅/NbCl₅ and PCl₅/WCl₆ were fully characterized by IR and NMR spectroscopy, and those cations derived from L-*N*,*N*-dimethylphenylalanine and L-methionine are reported here for the first time. The chloro-acyl moiety manifests itself by a strong IR absorption in the region 1765–1783 cm⁻¹, other than the ¹³C NMR resonance in the range 169.0–171.7 ppm.

Reactivity of MCl_n with α -amino acid esters

Preparation of α-amino acid ester hydrochlorides and α-amino acid esters. The α-amino acid ester derivatives, 8, were prepared from the corresponding hydrochlorides, 8 · HCl, which were in general isolated (Scheme 8). Although most of the compounds 8 and 8 · HCl have been already appeared in the literature,⁴⁰ we decided to collect their preparations and IR and NMR data in this paper, in view of possible modifications to the reported procedures or additional spectroscopic data.

Reactions with niobium and tantalum pentahalides. The reactions of α -amino acid esters with NbF₅ are often non selective, affording in most cases mixtures of products where the only recognizable compounds are the scarcely soluble ammonium ester salts [RCH(NH₃)COOR'][NbF₆]. These might be formed as a consequence of some activation reaction or the

adventitious presence of water.⁴¹ We were able to isolate satisfactory yields of well defined coordination compounds only in two cases (Scheme 9).

Compound **9** can be viewed as a coordination compound resulting from the unsymmetrical rupture of the structure of NbF₅ (a tetramer in the solid state^{17*a*}).^{13*c*,42} The IR spectrum shows a strong absorption at 1648 cm⁻¹, attributed to the stretching vibration of the C=O bond belonging to the ester function. The *ca.* 100 cm⁻¹ shift to lower wavenumbers is in agreement with the coordination of the carbonyl moiety to niobium. The shift of the absorptions due to the stretching of the amino group from 3380 cm⁻¹ (in **8k**) to 3232 cm⁻¹ (in **9**) suggests that also the nitrogen atom is involved in the coordination to the metal centre. Accordingly, two low field ¹H NMR resonances have been found for the NH₂ group in **9** (δ = 8.6 and 7.0 ppm, CDCl₃ solution). On the other hand, the same group



Scheme 9 Coordination compounds from the reactions of NbF $_5$ with L-proline ethylester and L-leucine methylester.



Scheme 8 Synthesis of α-amino acid esters.

gives raise to a singlet at 1.65 ppm in the ¹H NMR spectrum

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of 8k.

In addition, the ¹⁹F and ⁹³Nb NMR spectra (decet at 103 ppm and septet at -1553 ppm, respectively) are unequivocal fingerprints for the presence of the $[NbF_6]^-$ anion in solution.^{13c,42,43}

In conclusion, on considering the tendency of NbF₅ to the unsymmetrical breaking of the Nb–F bridges, with formation of $[NbF_4]^+$ cations and $[NbF_6]^-$ anions,⁴² analytical and spectroscopic data suggest that **9** is a salt containing an octacoordinate $[NbF_4(Me_2CHCH_2CHNH_2CO_2Me)_2]^+$ cation, comprising two O,N-ligated α -amino acid esters, and a $[NbF_6]^-$ anion. The coordination number of the cation was confirmed by DFT calculations, being six-coordinate geometries less stable by more than 30 kcal mol⁻¹. The optimized geometry is shown in Fig. 5. DFT calculations with dichloromethane as implicit solvent also indicated that the $[NbF_4(Me_2CHCH_2CHNH_2CO_2-Me)_2][NbF_6]$ salt is slightly more stable compared to its neutral isomer NbF₅(Me_2CHCH_2CHNH_2CO_2Me). The metal centre in this last species should be eight-coordinated, the α -amino acid ester behaving as N,O-donor chelating ligand (Fig. S4[†]).

It worth noting that the majority of coordination complexes containing α -amino acid ester ligands are based on late transition metals (Ru, Os, Co, Rh, Pt, Zn).⁴⁴ Only few examples are known with group 6 elements⁴⁵ and also derivatives of group 4 and group 5 metals are very rare.

The reaction of NbF₅ with L-proline ethyl ester, **8b**, revealed a different outcome, and **10** was obtained under the same conditions employed for NbF₅/**8k**. The use of Nb/**8b** molar ratio = 2 afforded **10** with the best yield (Scheme 9). Ethyl fluoride was NMR identified as a co-product of the reaction performed in CD_2Cl_2 in a closed tube, while L-proline was recovered after hydrolysis of the reaction mixture. These experimental facts support the presence in **10** of a carboxylato moiety originated from the cleavage of the ester function.

Compound **10** is a colourless solid whose salient spectroscopic features are two IR bands at 3381 cm⁻¹ (N–H) and 1636 cm⁻¹ (C=O), and ¹H and ¹⁹F NMR resonances at 11.77 ppm (NH) and 100.9 ppm ([NbF₆]⁻), respectively. These data suggest a bidentate N,O-coordination of the α -amino carboxylate ligand. Dinuclear geometries with the α -amino acidate as bridging



Fig. 5 DFT-optimized geometry of the cation of **9** (C-PCM/M06 calculations). Selected computed bond lengths (Å): Nb-O 2.273, 2.273; Nb-N 2.364, 2.364; Nb-F 1.900, 1.900, 1.900, 1.900; C-O(Nb) 1.234, 1.234; C-O(Me) 1.303, 1.303; N-H 1.019, 1.019, 1.019, 1.020. Selected computed angles (°): O-Nb-N 68.3, 68.3; O-Nb-O 135.0; N-Nb-N 132.3; C-O-Nb 123.3, 123.3.

ligand were ruled out by DFT calculations. The optimized geometry of the cation of **10** is depicted in Fig. 6 (see also Fig. S5 given as ESI[†]).

We extended the present study to the interaction of α -amino acid esters with the heavier niobium pentahalides. These reactions led to complicated mixtures of metal products, with presumable activation of the organic substrates. Only in a few cases, all involving the metal pentachlorides, a clean reaction pathway was observed (Scheme 10).

All the identified products, **11a–d** and **12**, are colourless to pale yellow solids, being scarcely soluble in organic solvents. Spectroscopic considerations discussed for **10** are valid also for **11a–d**, thus suggesting the bidentate N,O coordination of two α amino acid ester ligands to the same metal centre within a cation. The presence of the [NbCl₆][–] anion in **11a–c** is the consequence of unsymmetrical cleavage of the dinuclear NbCl₅ structure, ^{13c,33,46} and was unambiguously evidenced by a sharp ⁹³Nb NMR resonance occurring in the interval 4–13 ppm.^{13c,33}

DFT calculations were carried out on the cation of **11a**, considering either one or two α -amino acid esters in the niobium sphere. The coordination of another equivalent of the α -amino acid ester to $[NbCl_4(Me_2CHCH_2CHNH_2CO_2Me)]^+$ resulted a favourable process, being the associated ΔG variation



Fig. 6 DFT-optimized geometry of the cation of 10 (C-PCM/M06 calculations). Selected computed bond lengths (Å): Nb-O 1.902; Nb-N 2.243; Nb-F 1.833, 1.835, 1.857; C-O 1.365; C=O 1.192; N-H 1.022. Selected computed angles (°): O-Nb-N 73.5; O-Nb-F 97.7, 97.7, 147.3; C-O-Nb 131.0.



Scheme 10 Synthesis of niobium pentachloride derivatives of α -amino acid esters.

around -25 kcal mol⁻¹. The DFT-optimized geometry of $[NbCl_4(Me_2CHCH_2CHNH_2CO_2Me)_2]^+$ is represented in Fig. 7.

A crop of X-ray quality crystals of **12** was obtained directly from the reaction mixture after separation from **11a**. Compound **12** differs from the previous compounds as far as the solubility is concerned. Once isolated in the solid state, it does not dissolve again in common organic solvents.

Compound **12** consists of an ionic packing of $[NbCl_5(-CH_3)_2CHCH_2CHNH_3CO_2Me)]^+$ cations and $[NbCl_6]^-$ anions. The cation is represented in Fig. 8, and the related bonding parameters are reported in Table 3. A view of the structure of the anion is given in Fig. S6,† the relevant bonding parameters being collected in Table S2A.† H-bonds between the NH₃-group of the cation and the chlorides of $[NbCl_6]^-$ are present within the crystals (see Table S2B† for details). Compound **12** crystallizes in the chiral space group $P2_1$ and the C(2) atom of the α -amino acid ester ligand displays *S* absolute configuration.

The source of protonation leading to **12** is not clear, being possibly the result of some activation of the organic reactant



Fig. 7 DF1-optimized geometry of the cation of IIa (C-PCM/M06 calculations). Selected computed bond lengths (Å): Nb–O 2.261, 2.266; Nb–N 2.377, 2.381; Nb–Cl 2.380, 2.382, 2.413, 2.414; C–O(Nb) 1.236, 1.237; C–O(Me) 1.294, 1.294; N–H 1.020, 1.021, 1.020, 1.021.
Selected computed angles (°): O–Nb–N 68.4, 68.4; O–Nb–O 136.2; N–Nb–N 132.9; C–O–Nb 123.7, 123.8.



Fig. 8 ORTEP drawing of the $[NbCl_5(CH_3)_2CHCH_2CHNH_3CO_2Me)]^+$ cation in **12**. The $[NbCl_6]^-$ anion is reported in Fig. S6.† Displacement ellipsoids are at the 50% probability level.

promoted by the strongly acidic niobium chloride. Nevertheless, the occurrence of fortuitous hydrolysis might play some role and should not be ruled out.

12 represents the second crystallographically characterized example where a cationic α -amino acid ester is coordinated to any metal centre, and the first one where the coordination occurs *via* oxygen. In fact, previous to this work, only the structure of a Ru(II) complex containing a η^6 -bonded L-phenyl-alaninium methyl ester was reported.⁴⁷ More commonly, α -amino acid esters act as ligands in the neutral form RCHNH₂-CO₂R', *via* the N-atom or both N and O.^{44,45}

We moved to study the reaction of NbCl₅ with L-serine isopropylester, **8h**, *i.e.* a α -amino acid ester bearing a peripheral OH group and potentially acting as a pincer ligand. The 1 : 1 reaction of NbCl₅ and **8h** in refluxing chloroform led to the formation of NbCl₃(OCHCH₂NHCOOⁱPr), **13**, as a colourless precipitate. The ν (C=O) stretching band in **13** (1732 cm⁻¹) is only slightly shifted respect to **8h** (1728 cm⁻¹), thus indicating that the ester group is not involved in coordination.

Table 3 Selected bond distances (Å) and angles (°) for the [NbCl₅(- CH₃)₂CHCH₂CHNH₃CO₂Me)]⁺ cation in **12**. The data relative to the [NbCl₆]⁻ anion are reported in Tables S2A and S2B

Nb(2)-Cl(7)	2.3555(9)	Nb(2)-Cl(8)	2.2641(10)
Nb(2)-Cl(9)	2.3318(10)	Nb(2)-Cl(10)	2.3250(10)
Nb(2)-Cl(11)	2.2918(9)	Nb(2)-O(1)	2.209(2)
C(1)-O(1)	1.231(4)	C(1)-O(2)	1.291(4)
C(7)-O(2)	1.473(3)	C(1)-C(2)	1.499(4)
C(2)-N(1)	1.506(4)	C(2)-C(3)	1.534(4)
Cl(7)-Nb(2)-Cl(10)	174.26(3)	Cl(9)-Nb(2)-Cl(11)	163.09(3)
Cl(8)-Nb(2)-O(1)	177.18(6)	Nb(2)-O(1)-C(1)	145.2(2)
O(1)-C(1)-O(2)	125.5(3)	O(1)-C(1)-C(2)	121.0(3)
O(2)-C(1)-C(2)	113.5(3)	C(1)-C(2)-C(3)	112.3(3)
C(1)-C(2)-N(1)	109.4(2)	N(1)-C(2)-C(3)	108.4(3)



Fig. 9 DFT-optimized geometry of **13** (C-PCM/M06 calculations). Selected computed bond lengths (Å): Nb–O 1.873; Nb–N 1.943; Nb– Cl 2.359, 2.368, 2.389; N–H 1.019. Selected computed angles (°): O– Nb–N 76.0; O–Nb–Cl 93.3, 96.7, 157.8; N–Nb–Cl 82.0, 109.3, 110.0.

Compound **13** showed a single set of signals in ¹H and ¹³C NMR spectra (in CD₃CN); ¹H resonances are shifted to higher ppm values with respect to **8h** [most notably δ (NH) from 2.6 ppm in **8h** to 7.0 ppm in **13**], while ¹³C resonances are shifted to lower ppm values [*e.g.*, δ (CO) from 175 ppm in **8h** to 167 ppm in **13**].

At variance to the other α -amino acid ester complexes of niobium in this work, compound **13** is a neutral species in solution with a single ⁹³Nb resonance at -493 ppm. DFT calculations suggest a mononuclear structure (Fig. 9) as the most probable geometry. A positive Gibbs energy variation (about 3.5 kcal mol⁻¹) is accompanied to the dimerization of this species to the dinuclear form (see Fig. S7 and S8⁺ for more details).

We could not cleanly isolate metal products from MBr_5/α amino acid ester (M = Nb, Ta). However, NMR investigations outlined the release of ethyl bromide from L-proline ethylester, in the presence of MBr_5 (see Experimental for details).

Conclusions

The reactions of TiCl₄ with a series of α-amino acids do not proceed with HCl release, in spite of the Lewis acidic character of the metal centre, and afford dinuclear coordination compounds containing zwitterionic ligands. Deprotonation of the ammonium function may be easily promoted by the addition of triethylamine, resulting in a modification of the coordination fashion of the α -amino acidic frame. On the other hand, the interaction of L-proline with NbCl₅/NHEt₂ has provided the first example of C-C bond forming selfcondensation of a α -amino acid, although in modest yield. The overall transformation may be regarded as a Lewis acid induced intramolecular redox reaction, in which the carboxylate group is reduced and the pyrrolidine ring is oxidized. The combination of NbCl₅ or WCl₆ with the chlorinating power of PCl₅ has been exploited to develop a simple synthetic method affording isolable salts of otherwise unstable *a*-ammonium acylchloride cations. a-Amino acid esters usually behave as bidentate O,N-ligands towards niobium and tantalum pentahalides, however activation of the ester function with release of alkyl halides has been observed in some cases.

Experimental

General

Warning: all the metal products reported in this paper are highly moisture-sensitive, thus rigorously anhydrous conditions were required for the reaction and crystallization procedures. The reaction vessels were oven dried at 140 °C prior to use, evacuated (10^{-2} mmHg) and then filled with argon. TiCl₄, NbX₅ (X = F, Cl), PCl₅ and WCl₆ were purchased from Strem (>98% purity) and stored in sealed tubes under argon atmosphere. NbBr₅ and TaBr₅ were prepared according to literature procedures and stored under argon atmosphere.⁴⁸ Once isolated, the metal products were conserved in sealed glass tubes under argon. The organic reactants were commercial products (Sigma-Aldrich) stored under argon atmosphere as received.

Solvents (Sigma-Aldrich) were distilled before use from appropriate drying agents. Chromatographic purification of organic products was carried out on columns of deactivated alumina (4% w/w water). Infrared spectra were recorded at 298 K on a FT IR-Perkin Elmer Spectrometer, equipped with a UATR sampling accessory. NMR spectra were recorded at 293 K on a Bruker Avance II DRX400 instrument equipped with a BBFO broadband probe. The chemical shifts for ¹H and ¹³C were referenced to the non-deuterated aliquot of the solvent; the chemical shifts for ⁹³Nb were referenced to external [NEt₄][NbCl₆]; the chemical shifts for ¹⁹F were referenced to external CFCl₃. Conductivity measurement was carried out using an Eutech Con 700 instrument (cell constant = 1.0 cm^{-1}).⁴⁹ Magnetic susceptibilities (reported per W atom) were measured on solid samples at 298 K with a Magway MSB Mk1 magnetic susceptibility balance (Sherwood Scientific Ltd). Diamagnetic corrections were introduced according to König.50 Carbon, hydrogen and nitrogen analyses were performed on a Carlo Erba mod. 1106 instrument. The chloride/bromide content was determined by the Mohr method⁵¹ on solutions prepared by dissolution of the solids in aqueous KOH and heated at boiling temperature for 72 hours, followed by cooling to room temperature and addition of HNO₃ up to neutralization. Titanium, niobium and tantalum were analyzed, respectively, as TiO_2 and M_2O_5 (M = Nb, Ta), obtained by hydrolysis of the samples followed by calcination in a platinum crucible.

Reactions of TiCl₄ with α -amino acids: synthesis of TiCl₄(aa) (aa = 1-proline, 1a; 1-phenylalanine, 1b; sarcosine, 1c; *N*,*N*-dimethylglycine, 1d)

General procedure. A suspension of the appropriate α -amino acid (1.50 mmol) in CH₂Cl₂ (*ca.* 15 mL) was treated with a solution (100 mg mL⁻¹) of TiCl₄ (1.50 mmol) in heptane. The mixture was stirred at room temperature overnight, then hexane (*ca.* 30 mL) was added. The precipitate was separated and dried *in vacuo*.

*TiCl*₄(*L*-*proline*), *1a*. Yellow solid, yield 321 mg (70%). Anal. calcd for C₅H₉Cl₄NO₂Ti: C, 19.70; H, 2.98; N, 4.60; Cl, 46.52; Ti, 15.70. Found: C, 19.39; H, 3.09; N, 4.52; Cl, 45.88; Ti, 15.89. IR (solid state): $\nu = 3219$ mw, 2962 w, 1570 m, 1544 vs, 1441 vs, 1367 m, 1331 ms, 1260 m, 1081 m, 1031 ms, 798 s cm⁻¹. ¹H NMR (CD₃CN): $\delta = 7.46$, 7.15 (br, 2H, NH₂); 4.53 (br, 1H, NCH); 3.54, 3.45, 2.42, 2.20, 2.06 (br, 6H, *CH*₂) ppm. ¹³C NMR (CD₃CN): $\delta = 176.1$ (OCO); 61.6 (CH); 47.6, 28.6, 23.7 (CH₂) ppm.

TiCl₄(L-phenylalanine), **1b**. Light orange solid, yield 388 mg (73%). Anal. calcd for C₉H₁₁Cl₄NO₂Ti: C, 30.46; H, 3.12; N, 3.95; Cl, 39.96; Ti, 13.49. Found: C, 30.60; H, 3.02; N, 4.13; Cl, 39.40; Ti, 13.28. IR (solid state): $\nu = 3030$ m-br, 1600 m, 1558 vs, 1445 vs-br, 1336 m, 1047 w, 744 m, 698 ms cm⁻¹. ¹H NMR (CD₃CN): $\delta = 7.39$ -7.30, 6.98 (8H, Ph + NH₃); 4.46 (m, 1H, CH); 3.28 (m, 2H, CH₂) ppm. ¹³C NMR (CD₃CN): $\delta = 170.0$ (OCO); 134.1 (*ipso*-Ph); 129.8, 129.2, 127.9 (C₆H₅); 61.8 (CH); 35.3 (CH₂) ppm.

TiCl₄(sarcosine), 1c. Yellow solid, yield 448 mg (78%). Anal. calcd for C₃H₇Cl₄NO₂Ti: C, 12.93; H, 2.53; N, 5.02; Cl, 50.87; Ti, 17.17. Found: C, 13.02; H, 2.43; N, 4.98; Cl, 36.23; Ti, 12.80. IR (solid state): $\nu = 3185$ m, 2930 vw, 2810 vw, 1575 ms, 1561 vs,

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1454 s, 1429 w, 1409 ms, 937 w, 792 s, 689 m-w cm⁻¹. ¹H NMR (CD₃CN): δ = 7.26, 7.00 (br, 2H, NH₂); 4.03 (br, 2H, CH₂); 2.81 (br, 3H, CH₃) ppm.

TiCl₄(N,N-dimethylglycine), **1d**. Yellow solid, yield 312 mg (71%). Anal. calcd for C₄H₉Cl₄NO₂Ti: C, 16.41; H, 3.10; N, 4.78; Cl, 48.43; Ti, 16.35. Found: C, 16.29; H, 3.17; N, 4.91; Cl, 48.26; Ti, 16.57. IR (solid state): $\nu = 3091$ m-w, 2983 w, 1573 vs, 1438 s, 1401 m, 1364 s, 1324 m, 1142 m-w, 932 m-w, 857 m, 716 m cm⁻¹. ¹H NMR (CD₃CN): $\delta = 7.7$ (br, 1H, NH); 4.09 (br, 2H, CH₂); 3.01 (br, 6H, CH₃) ppm.

Reactions of TiCl₄ with α -amino acid/NEt₃: synthesis of [NHEt₃][TiCl₄(aa)] (aa = 1-phenylalanine, 2a; *N*,*N*-dimethylphenylalanine, 2b)

General procedure. A suspension of the appropriate α -amino acid (1.00 mmol) in CH₂Cl₂ (*ca.* 10 mL) was treated with a solution (100 mg mL⁻¹) of TiCl₄ (1.00 mmol) in heptane. The mixture was stirred at room temperature overnight, then hexane (*ca.* 30 mL) was added. The liquors were eliminated with a syringe, then CH₂Cl₂ (20 mL) and NEt₃ (1.00 mmol) were added in the order given. The mixture was allowed to stir for 5 h, then hexane (30 mL) was added. The resulting precipitate was separated and dried *in vacuo*.

[*NHEt*₃][*TiCl*₄(*ι*-*phenylalanine*)], **2a**. Light brown solid, yield 297 mg (65%). Anal. calcd for C₁₅H₂₆Cl₄N₂O₂Ti: C, 39.50; H, 5.75; N, 6.14; Cl, 39.50; Ti, 10.50. Found: C, 39.33; H, 5.87; N, 6.16; Cl, 39.19; Ti, 10.61. IR (solid state): $\nu = 3306$ w, 3240 w-br, 2984 w-br, 2675 w-br, 2488 w-br, 1691 vs, 1652 vs, 1568 s, 1454 s, 1228 m, 1099 m-s, 1070 m-s, 749 vs, 702 s cm⁻¹. ¹H NMR (CD₂Cl₂): $\delta = 9.09$ (s, 1H, NH); 7.39–7.30 (5H, Ph); 4.38 (m, 1H, CH); 4.13, 3.62 (br, 2H, NH₂); 3.26 (m, 6H, NCH₂); 3.40, 3.17 (dd, 2H, *CH*₂Ph); 1.40 (t, ³*J*_{HH} = 7.34 Hz, 9H, NCH₂*CH*₃) ppm. ¹³C NMR (CD₂Cl₂): $\delta = 179.5$ (OCO); 135.9 (*ipso*-Ph); 129.4, 129.2, 127.5 (Ph); 61.7 (CH); 47.1 (NCH₂); 38.6 (*C*H₂Ph); 9.0 (NCH₂*C*H₃) ppm.

[*NHEt*₃][*TiCl*₄(*t*-*N*,*N*-*dimethylphenylalanine*)], **2b**. Yellow solid, yield 290 mg (60%). Anal. calcd for C₁₇H₃₀Cl₄N₂O₂Ti: C, 42.18; H, 6.25; N, 5.79; Cl, 29.29; Ti, 9.89. Found: C, 42.36; H, 6.08; N, 5.65; Cl, 29.41; Ti, 9.72. IR (solid state): $\nu = 3260$ w-br, 2963 w-br, 2679 w-br, 1702 s, 1660 vs, 1581 vs, 1454 vs, 1260 m, 1012 s, 741 s, 699 s cm⁻¹. ¹H NMR (CD₂Cl₂): $\delta = 8.0$ (br, 1H, NH); 7.36-7.26 (5H, Ph); 4.47 (m, 1H, CH); 3.32 (m, 6H, NCH₂); 3.19, 3.07 (m, 2H, CH₂Ph); 2.95, 2.73 (s, 6H, NMe₂); 1.42 (t, ³*J*_{HH} = 7.34 Hz, 9H, NCH₂CH₃) ppm. ¹³C NMR (CD₂Cl₂): $\delta = 178.8$ (OCO); 138.9 (*ipso*-Ph); 129.9, 129.2, 128.6, 126.6 (Ph); 75.6 (CH); 50.9, 47.6 (NMe₂); 47.5 (NCH₂); 30.8 (CH₂Ph); 8.9 (NCH₂CH₃) ppm.

Reaction of NbCl₅ with L-proline/NHⁱPr₂: synthesis and isolation of Nb₂Cl₈{ μ - κ^{2} O, κ^{2} N-[CH₂CH₂CH₂C(N)C(O)]₂, 3, and [NH₂ⁱPr₂] [NbCl₆], 4

NbCl₅ (0.385 g, 1.42 mmol) and L-proline (0.163 g, 1.42 mmol) were allowed to react in CH_2Cl_2 (20 mL). The solution was repetitiously purged with nitrogen gas in order to remove released HCl. After six hours, the yellowish mixture was treated with NH^iPr_2 (0.203 mL, 1.45 mmol), then the stirring was prolonged for additional 20 min. The final dark-red mixture was

filtered off in order to remove a minor amount of solid, layered with hexane and settled aside at -30 °C. Red crystals of 3 were recovered after 48 h. Yield 56 mg, 12%. Anal. calcd for C₁₀H₁₂-Cl₈N₂Nb₂O₂: C, 18.15; H, 1.83; N, 4.23; Cl, 42.86. Found: C, 18.43; H, 1.63; N, 4.51; Cl, 43.12. IR (solid state): $\nu = 1580$ m-s ($\nu_{C=N}$), 1183 m-s ($\nu_{C=O}$) cm⁻¹.

The mother liquors were dried *in vacuo*, hence the residue was dissolved into CHCl_3 (10 mL). The solution was layered with pentane and settled aside at -30 °C, thus 4 was isolated as a yellow-orange crystals after 48 h. Yield 238 mg, 40%. Anal. calcd for $C_7H_{16}Cl_6NNb$: C, 20.03; H, 3.84; N, 3.34; Cl, 50.67. Found: C, 19.84; H, 3.75; N, 3.20; Cl, 49.95. IR (solid state): $\nu = 3130$ s-br, 3080 s-br, 2987 w-m, 1606 m, 1565 m, 1424 m cm⁻¹. ¹H NMR (CD₃CN): $\delta = 6.30$ (br, 1H, NH); 3.48 (m, 2H, CH); 1.31 (m, 12H, Me) ppm. ¹³C NMR (CD₃CN): $\delta = 47.7$ (CH); 18.2 (Me) ppm. ⁹³Nb NMR (CD₃CN): $\delta = -0.2$ ($\Delta v_2^1 = 95$ Hz) ppm.

Reactions of α -amino acids with PCl₅/NbCl₅: synthesis of [(R)(Me)NHCH(R')C(O)Cl][NbCl₆] (R = Me, R' = CH₂Ph, 5a; R = R' = H, 5b)

General procedure. A suspension of PCl_5 (169 mg, 0.81 mmol) and $NbCl_5$ (220 mg, 0.81 mmol) in CH_2Cl_2 (10 mL) was stirred at room temperature for 2–3 h. Then the appropriate α -amino acid (0.81 mmol) was added. The resulting mixture was stirred at room temperature for 2 h. The solution was concentrated to 3–5 mL, then it was layered with pentane and stored in the freezer (–30 °C) for one week. A crop of crystalline material was collected and then stored at –30 °C. By slow evaporation of the crystallization solutions under inert atmosphere, few crystals of [PhCH₂ = NMe₂][NbCl₆] and NbCl₅(O=PCl₃), **6**, were obtained from PCl₅/NbCl₅/L-*N*,*N*-dimethylphenylalanine and PCl₅/NbCl₅/sarcosine, respectively.

[*Me*₂*NHCH*(*CH*₂*Ph*)*C*(*O*)*Cl*][*NbCl*₆], *5a.* Orange solid, yield 214 mg (47%) from PCl₅/NbCl₅/L-*N*,*N*-dimethylphenylalanine. Anal. calcd for C₁₁H₁₅Cl₇NbO: C, 25.49; H, 2.92; N, 2.70; Cl, 47.88; Nb, 17.92. Found: C, 25.23; H, 2.80; N, 2.67; Cl, 47.60; Nb, 18.10. IR (solid state): $\nu = 3087$ w-m, 3029 w, 2939 w, 1767 vs ($\nu_{C=O}$), 1460 s, 1440 m, 1411 w, 1374 w, 1340 w, 1287 w, 1198 w, 1172 w, 1138 w, 1078 w, 1033 m, 1016 w, 991 w-m, 936 m, 917 w, 896 vs, 839 m, 820 w, 745 m, 726 vs, 693 vs cm⁻¹. ¹H NMR (CD₂Cl₂): $\delta = 7.68$ (br, 1H, NH); 7.47–7.35 (m, 5H, Ph); 4.83 (s, 1H, CH); 3.62 (m, 2H, CH₂); 3.34, 3.22 (s, 6H, NMe₂) ppm. ¹³C NMR (CD₂Cl₂): $\delta = 170.6$ (C==O); 130.7, 130.0, 129.2 (Ph); 127.7 (*ipso*-Ph); 76.5 (CH); 44.4, 43.4 (NMe₂); 34.6 (CH₂) ppm. ⁹³Nb NMR (CD₂Cl₂): $\delta = 8.1$ ($\Delta \nu_{1}^{1} = 3\cdot10^{2}$ Hz) ppm.

[*MeNH*₂*CH*₂*C*(*O*)*Cl*][*NbCl*₆], *5b*. Yellow solid, yield 144 mg (43%) from PCl₅/NbCl₅/sarcosine. Anal. calcd for C₃H₇Cl₇-NNbO: C, 8.70; H, 1.70; N, 3.38; Cl, 59.92; Nb, 22.43. Found: C, 8.80; H, 1.59; N, 3.31; Cl, 59.60; Nb, 22.55. IR (solid state): $\nu = 3138 \text{ m-s} (\nu_{\text{N-H}})$, 2983 w, 2943 w, 1766 vs ($\nu_{\text{C=O}}$), 1546 w-m, 1454 m, 1430 m, 1414 m, 1397 vs, 1341 m, 1160 m, 1129 w, 1036 m, 999 vs, 926 vs, 789 vs, 767 vs cm⁻¹. ¹H NMR (CD₃CN): $\delta = 7.42$ (br, 2H, NH₂); 4.67 (t, 2H, ⁴*J*_{HH} = 5.38 Hz, CH₂); 3.21 (q, 3H, ⁴*J*_{HH} = 5.38 Hz, Me) ppm. ¹³C NMR (CD₃CN): $\delta = 0.1$ ($\Delta \nu_{2}^{1} = 31$ Hz) ppm.

Reactions of α -amino acids with PCl₅/WCl₆: synthesis of [NH₂(CH₂)₃CHC(O)Cl][WCl₆], 7a, [Me₂NHCH(CH₂Ph)C(O)Cl] [WCl₆], 7b, and [(R)NH₂CH(R')C(O)Cl]⁺ (R = Me, R' = H; R = H, R' = CH₂CH₂SMe)

General procedure. A suspension of PCl_5 (163 mg, 0.78 mmol) and WCl_6 (310 mg, 0.78 mmol) in CD_2Cl_2 (4 mL) was stirred at room temperature overnight. Then the appropriate α -amino acid (0.78 mmol) was added. The resulting mixture was stirred at room temperature for 3 h. Thus ³¹P NMR analyses revealed the presence of POCl₃ as unique phosphorous species. In addition, ¹H and ¹³C NMR analyses on $PCl_5/WCl_6/L-N,N$ -dimethylphenylalanine and $PCl_5/WCl_6/sarcosine$ solutions pointed out the clean formation of 7a and [MeNH₂CH₂C(O)Cl]⁺. These solutions were layered with hexane and stored at -30 °C for a few days, thus resulting in the isolation of 7a and a mixture of [MeNH₂CH₂C(O)Cl]⁺ salts. In the cases of $PCl_5/WCl_6/L$ -methionine and $PCl_5/WCl_6/L$ -proline, dark solid materials precipitated, which were isolated from the respective yellow solutions and dried *in vacuo*.

In a different experiment, a 1 : 1 PCl₅/WCl₆ mixture obtained in CD₂Cl₂ (3 mL) was analyzed. $\Lambda_{\rm M}$ (CD₂Cl₂) = 0.3 S cm² mol⁻¹. ³¹P NMR (CD₂Cl₂): $\delta = -81.1$ (PCl₅) ppm. Cl analysis was carried out on the solid residue obtained by removal of the volatiles *in vacuo*. Anal. calcd for Cl₁₁PW: Cl, 64.48. Found: Cl, 63.91. Magnetic measurement: diamagnetic.

[$NH_2(CH_2)_3CHC(0)Cl$][WCl_6], 7*a*. Green solid, yield 302 mg (73%) from PCl₅/WCl₆/L-proline. Anal. calcd for C₅H₉Cl₇NOW: C, 11.31; H, 1.71; N, 2.64; Cl, 46.72. Found: C, 11.20; H, 1.78; N, 2.50; Cl, 46.33. IR (solid state): $\nu = 3133 \text{ m} (\nu_{\text{N-H}})$, 3073 m, 2955 w, 1768 vs ($\nu_{\text{C=O}}$), 1563 m, 1455 w, 1375 w-m, 1343 w-m, 1079 w, 1043 m, 996 vs, 942 m, 906 w, 864 m cm⁻¹. ¹H NMR (CD₃CN): $\delta = 7.36$ (t, 2H, ³ $J_{\text{HH}} = 50$ Hz, NH₂); 4.80 (m, 1H, CH); 3.45 (m, 2H, NCH₂); 2.60, 2.36 (m, 2H, CH₂); 2.14–2.06 ppm (m, 2H, CH₂) ppm. ¹³C NMR (CD₃CN): $\delta = 171.7$ (C=O); 68.3 (CH); 48.4 (NCH₂); 29.1, 23.7 ppm (CH₂) ppm. Magnetic measurement: $\chi_{\text{MM}}^{\text{corr}} = 3.60 \times 10^{-4}$ cgsu, $\mu_{\text{eff}} = 0.93$ BM.

[*Me*₂*NHCH*(*CH*₂*Ph*)*C*(*O*)*Cl*][*WCl*₆], **7b**. Dark yellow – brown solid, yield 285 mg (60%) from PCl₅/WCl₆/L-*N*,*N*-dimethylphenylalanine. Anal. calcd for C₁₁H₁₅Cl₇NOW: C, 21.69; H, 2.48; N, 2.30; Cl, 40.73. Found: C, 21.78; H, 2.35; N, 2.20; Cl, 40.32. IR (solid state): $\nu = 3122$ m-br, 3073 m, 2987 w-m, 2946 w, 2789 w, 1783 vs ($\nu_{C=O}$), 1564 w, 1455 m, 1433 w-m, 1400 m, 1347 w, 1260 w, 1164 w-m, 1041 w, 1001 s, 952 m, 930 m, 857 s, 802 s, 774 s cm⁻¹. ¹H NMR (CD₂Cl₂): $\delta = 7.74$ (br, 1H, NH); 7.49–7.39 (m, 5H, Ph); 4.66 (br, 1H, CH); 3.62 (m-br, 2H, CH₂); 3.31, 3.19 (s, 6H, NMe₂) ppm. ¹³C NMR (CD₂Cl₂): $\delta = 170.8$ (C=O); 131.3, 130.0, 129.9, 129.2 (Ph); 78.3 (CH); 50.2, 48.1 (NMe₂); 36.7 (CH₂) ppm. Magnetic measurement: $\chi_{M}^{corr} = 4.02 \times 10^{-4}$ cgsu, $\mu_{eff} = 0.98$ BM.

[*MeNH*₂*CH*₂*C*(*O*)*Cl*]⁺. Green solid, from PCl₅/WCl₆/sarcosine. IR (solid state): $\nu = 3104$ m-br ($\nu_{\rm N-H}$), 2980 w, 2938 w, 2796 w, 1765 vs ($\nu_{\rm C=O}$), 1562 w-m, 1455 m, 1431 m, 1406 m, 1345 w-m, 1165 w-m, 1136 w, 1037 w, 1009 s, 991 vs, 951 m, 929 vs, 854 s, 806 vs, 773 vs, 728 s cm⁻¹. ¹H NMR (CD₃CN): $\delta = 7.29$ (br, 2H, NH₂); 4.58 (s, 2H, CH₂); 3.21 (s, 3H, Me) ppm. ¹³C NMR (CD₃CN): $\delta = 169.0$ (C=O); 63.2 (CH₂); 40.4 (Me) ppm. Magnetic measurement: $\chi_{\rm M}^{\rm corr} = 1.13 \times 10^{-3}$ cgsu. $[NH_3CH(CH_2CH_2SMe)C(O)Cl]^t$. Brown solid, from PCl₅/WCl₆/ L-methionine. IR (solid state): $\nu = 3350$ w-m (ν_{N-H}), 3008 w, 2921 w, 1779 s ($\nu_{C=O}$), 1569 w, 1480 w, 1449 w-m, 1415 m, 1367 w-m, 1309 w, 1263 w, 1176 w, 1144 w, 1094 w, 1025 m, 964 vs, 897 s, 759 s, 733 m, 699 w, 661 w cm⁻¹. Magnetic measurement: $\chi_M^{corr} = 7.08 \times 10^{-4}$ cgsu.

Synthesis of α-amino acid ester hydrochlorides

These compounds were obtained by a slight modification of the literature procedures.

Procedure A (compounds 8a–d·HCl). A 250 mL flask was charged with the appropriate alcohol (120 mL)/ α -amino acid (*ca.* 35 mmol) combination. SOCl₂ (12 mL, 170 mmol) was slowly added (3 h) to the suspension under vigorous stirring at room temperature. After 24 h stirring, volatiles were removed *in vacuo* at room temperature. The residue was suspended in Et₂O (50 mL) for 4 h. The suspension was filtered and the resulting solid was dried *in vacuo* at 40 °C.

Procedure B (compounds 8e-i·HCl). SOCl₂ (10 mL, 138 mmol) was slowly added (30 minutes) at 0 °C to the alcohol (80 mL) in a 500 mL Schlenk tube. The solution was then allowed to reach room temperature and the α -amino acid (24 mmol) was introduced. The mixture was refluxed for 8 h and a pale yellow solution was obtained. Afterwards, the volatiles were removed *in vacuo* and the residue was suspended in Et₂O (50 mL) for 2 h. The suspension was filtered and the resulting solid was dried *in vacuo* at 40 °C.

L-Proline methylester hydrochloride, **8a**·HCl.^{40a,b} Colourless solid, yield 97%. IR (liquid film): $\nu = 3115 \text{ w} (\nu_{\text{N-H}})$, 3021 w ($\nu_{\text{N-H}}$), 2805 m, 1755 vs ($\nu_{\text{C=O}}$), 1634 m, 1442 m, 1391 m-s, 1089 m, 1015 m cm⁻¹. ¹H NMR (CDCl₃): $\delta = 10.68$, 9.04 (br, 2H, NH₂); 4.51 (m, 1H, CH); 3.86 (s, 3H, OMe); 3.54 (m, 2H, NCH₂); 2.48, 2.20, 2.10 (m, 4H, CH₂) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 169.3$ (C=O); 59.2 (CH); 53.6 (OMe); 46.0 (NCH₂); 28.7, 23.6 (CH₂) ppm.

L-Proline ethylester hydrochloride, **8b**·HCl.^{40a} Colourless solid, yield 79%. ¹H NMR (CDCl₃): $\delta = 10.57$, 8.92 (br, 2H, NH₂); 4.42 (m, 1H, CH); 4.24 (q, ³J_{HH} = 5.87 Hz, 2H, OCH₂); 3.50 (m, 2H, NCH₂); 2.40, 2.10 (m, 4H, CH₂); 1.27 (t, ³J_{HH} = 5.87 Hz, 3H, OCH₂CH₃) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 168.7$ (C=O); 62.9 (OCH₂); 59.2 (CH); 45.9 (NCH₂); 28.7, 23.6 (CH₂); 14.0 (OCH₂CH₃) ppm.

L-Proline isopropylester hydrochloride, **8***c*·HCl.^{40a} Colourless solid, yield 97%. ¹H NMR (CDCl₃): δ = 10.70, 8.81 (br, 2H, NH₂); 5.11 (sept, ³J_{HH} = 6.2 Hz, 1H, OCH); 4.44 (m, 1H, CH); 3.57 (m, 2H, NCH₂); 2.43, 2.12, 2.03 (m, 4H, CH₂); 1.29 (pseudo-t, ³J_{HH} = 6.2 Hz, 6H, OCHMe₂) ppm. ¹³C{¹H} NMR (CDCl₃): δ = 168.3 (C= O); 71.2 (OCH); 59.3 (CH); 46.1 (NCH₂); 29.0, 23.6 (CH₂); 21.6 (OCHMe₂) ppm.

L-Phenylalanine methylester hydrochloride, **8**d·HCl.^{40b} Colourless crystalline solid, yield 83%. IR (solid state): $\nu = 3091$ m-br ($\nu_{\text{N-H}}$), 2944 w-sh, 2906 w-sh, 2838 w, 2625 w, 1743 vs ($\nu_{\text{C=O}}$), 1583 w-m, 1495 m, 1447 w-m, 1436 w-m, 1400 w, 1358 w, 1327 w, 1291 w-m, 1238 vs, 1214 vs, 1146 m, 1119 m, 1084 s, 1061 m, 1033 w, 990 m, 934 m, 865 w, 760 n, 741 vs, 701 vs cm⁻¹. ¹H NMR (CDCl₃): $\delta = 8.7$ (br, 3H, NH₃); 7.31, 7.28 (m, 5H, Ph); 4.38 (br, 1H, CH); 3.72 (s, 3H, OMe); 3.42 (m-br, 2H, CH₂) ppm. ¹³C {¹H} NMR (CDCl₃): δ = 169.2 (C=O); 133.9 (*ipso*-Ph); 129.6, 129.0, 127.7 (Ph); 54.4 (CH); 53.0 (OMe); 36.3 (CH₂) ppm.

L-Leucine ethylester hydrochloride, **8**e·HCl.^{40d} Colourless solid, yield 80%. ¹H NMR (CDCl₃): $\delta = 8.5$ (br, 3H, NH₃); 4.26 (m, 2H, OCH₂); 4.07 (m, 1H, CH); 1.92 (m, 1H, CHMe₂); 1.83 (m, 2H, CH₂); 1.31 (m, 6H, CHMe₂); 1.23 (t, ³J_{HH} = 6.85 Hz, 3H, OCH₂-CH₃) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 169.9$ (C=O); 62.6 (OCH₂); 51.9 (CH); 39.5 (CH₂); 24.5 (CHMe₂); 22.2, 22.1 (CHMe₂); 14.0 (OCH₂CH₃) ppm.

L-Leucine isopropylester hydrochloride, **8fH**Cl.^{40e} Colourless solid, yield 81%. ¹H NMR (CDCl₃): $\delta = 8.21$, 6.64 (br, 3H, NH₃); 5.11 (m, 1H, OCH); 4.01 (m, 1H, CH); 1.89 (m, 1H, CHMe₂); 1.81 (m, 2H, CH₂); 1.28 (m, 6H, CHMe₂); 0.98 (m, 6H, OCHMe₂) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 169.3$ (C=O); 70.8 (OCH); 52.0 (CH); 39.5 (CH₂); 24.8 (CHMe₂); 22.2 (CHMe₂); 21.6 (OCHMe₂) ppm.

Glycine isopropylester hydrochloride, **8g**·HCl.^{49f} Colourless solid, 97% yield. ¹H NMR (CDCl₃): δ = 8.50 (br, 3H, NH₃); 5.10 (sept, ³J_{HH} = 6.1 Hz, 1H, OCH); 3.96 (q, ³J_{HH} = 4.2 Hz, 2H, CH₂); 1.26 (d, ³J_{HH} = 6.1 Hz, 6H, CHMe₂) ppm.

L-Serine isopropylester hydrochloride, **8h**·HCl.^{40g} Colourless solid, 98% yield. ¹H NMR (DMSO-d₆): $\delta = 8.51$ (br, 3H, NH₃); 5.71–5.52 (m, 1H, OH); 5.05–4.94 (m, 1H, CHMe₂); 4.04–3.98 (m, 1H, CHH'); 3.83–3.78 (s, 2H, CHH' + CHN); 1.26–1.21 (m, 6H, CHMe₂) ppm. ¹³C{¹H} NMR (DMSO-d₆): $\delta = 167.6$ (CO); 69.6 (CHMe₂); 59.5 (CH₂); 54.4 (CHN); 21.5 and 21.4 (CHMe₂) ppm.

L-Tyrosine isopropylester hydrochloride, **8i**⁺HCl.^{40h} Colourless solid, 71% yield. ¹H NMR (DMSO-d₆): δ = 9.49 (s, 1H, OH); 8.49 (br, 3H, NH₃); 7.01 (d, ³*J*_{HH} = 8.2 Hz, 2H, Ar); 6.71 (d, ³*J*_{HH} = 8.2 Hz, 2H, Ar); 4.88 (sept, ³*J*_{HH} = 5.9 Hz, 1H, OCH); 4.07 (t, ³*J*_{HH} = 6.6 Hz, 1H, NCH); 3.17–2.83 (m, 2H, CH₂); 1.16 (d, ³*J*_{HH} = 6.2 Hz, 3H) and 1.06 (d, ³*J*_{HH} = 6.2 Hz, 3H, CH*Me*₂) ppm.

Synthesis of α -amino acid esters. Three different procedures were adopted. Compounds **8a–g** were prepared by treating the appropriate α -amino acid ester hydrochloride with NH_{3(aq)} as described in detail for **8a**. Compounds **8h–j** were prepared by treating the appropriate α -amino acid ester hydrochloride with NaOH_(aq) as described in detail for **8h**. Compounds **8k–l** were obtained directly from the appropriate alcohol/ α -amino acid (*ca.* 50 mmol) combination, followed by treatment with NH_{3(aq)}; attempts to isolate (**8k–l**)·HCl led to mixtures of products.

I-Proline methylester, 8a.^{40α,c} Compound **8a**·**HCl** (10.2 g, 60.4 mmol) was dissolved into CH₂Cl₂ (100 mL) and the solution was treated with a 28% w/w NH₃ aqueous solution until neutrality. The mixture was left stirring at room temperature for 24 h. The phases were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 50 mL). The organic phase was filtered through an alumina pad and solvent was removed by distillation at 40 °C and *p* = 700 mbar. The product was obtained as a pale orange liquid, which was stored under argon. Yield 4.71 g (60%). ¹H NMR (CDCl₃): δ = 3.54 (s, 3H, CH₃); 2.92 (m, 1H, CH); 2.48 (m, 2H, CH₂N); 1.99, 1.64 (m, 4H, CH₂) ppm.

L-Proline ethylester, **8b**.^{40α} Pale yellow liquid, yield 65%. IR (liquid film): $\nu = 1730$ vs ($\nu_{C=O}$) cm⁻¹. ¹H NMR (CDCl₃): $\delta = 4.12$ (q, ³*J*_{HH} = 6.85 Hz, 2H, OCH₂); 3.73 (m, 1H, CH); 3.03, 2.90 (m, 2H, NCH₂); 2.07, 1.81, 1.73 (m, 4H, CH₂); 1.22 (t, ³*J*_{HH} = 6.85 Hz, 3H, OCH₂CH₃) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 173.9$ (C=O);

61.0 (OCH₂); 59.7 (CH); 46.9 (NCH₂); 30.2, 25.4 (CH₂); 14.2 (OCH₂CH₃) ppm.

L-Proline isopropylester, 8c.^{40*i*} Pale orange liquid, yield 60%. ¹H NMR (CDCl₃): $\delta = 5.03$ (sept, ³*J*_{HH} = 6.3 Hz, 1H, OCH); 3.71 (dd, ³*J*_{HH} = 8.3, 5.6 Hz, 1H, CH); 3.12–3.04 (m, 1H, NCHH'); 2.94–2.85 (m, 1H, NCHH'); 2.28 (br, 2H, NH₂); 2.18–2.06 (m, 1H) and 1.86–1.67 (m, 3H, CH₂CH₂); 1.25 (d, ³*J*_{HH} = 3.5 Hz, 3H, CH*MeMe'*); 1.23 (d, ³*J*_{HH} = 3.6 Hz, 3H, CHMe*Me'*) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 174.6$ (C=O); 68.0 (OCH); 59.6 (CH); 46.8 (NCH₂); 30.1, 25.2 (CH₂); 21.5 (OCH*Me*₂) ppm.

L-Phenylalanine methylester, 8d.^{40c} Pale orange liquid, yield 63%. IR (liquid film): $\nu = 3381$ w (ν_{N-H}), 3062 w, 3028 w, 2951 w, 1732 vs ($\nu_{C=O}$), 1603 w-m, 1496 m, 1454 m, 1436 m, 1266 m, 1195 s, 1172 s, 1111 m, 1076 m, 1009 m, 826 m, 812 m, 744 m-s, 699 vs cm⁻¹. ¹H NMR (CDCl₃): $\delta = 7.24$ -7.12 (5H, Ph); 3.66 (m, 1H, CH); 3.63 (s, 3H, OMe); 3.01, 2.80 (m, 2H, CH₂) ppm. ¹³C {¹H} NMR (CDCl₃): $\delta = 175.3$ (C=O); 137.3 (*ipso*-Ph); 129.3, 128.5, 126.7 (Ph); 55.8 (OMe); 51.8 (CH); 41.0 (CH₂) ppm.

L-Leucine ethylester, 8e.^{40d} Pale yellow liquid, yield 74%. ¹H NMR (CDCl₃): δ = 4.16 (m, 2H, OCH₂); 3.44 (m, 1H, CH); 1.77 (m, 1H, CHMe₂); 1.65 (s, 2H, NH₂); 1.55, 1.43 (m, 2H, CH₂); 1.27 (t, ³*J*_{HH} = 6.85 Hz, 3H, OCH₂CH₃); 0.93 (m, 6H, CHMe₂) ppm. ¹³C{¹H} NMR (CDCl₃): δ = 176.5 (C=O); 60.7 (OCH₂); 52.9 (CH); 44.0 (CH₂); 24.7 (CHMe₂); 22.9, 21.8 (CHMe₂); 14.2 (OCH₂CH₃) ppm.

L-Leucine isopropylester, **8f**.^{40e} Pale yellow liquid, yield 55%. ¹H NMR (CDCl₃): δ = 5.05 (sept, 1H, OCH); 3.49 (m, 1H, CH); 2.90 (s, 2H, NH₂); 1.80 (m, 1H, CHMe₂); 1.57, 1.50 (m, 2H, CH₂); 1.26 (m, 6H, OCHMe₂); 0.94 (m, 6H, CHMe₂) ppm. ¹³C{¹H} NMR (CDCl₃): δ = 175.1 (C=O); 68.5 (OCH); 52.8 (CH); 43.5 (CH₂); 24.7 (CHMe₂); 22.9, 21.9 (CHMe₂); 21.7 (OCHMe₂) ppm.

Glycine isopropylester, **8**g.^{40f} Pale yellow viscous liquid, yield 60%. ¹H NMR (CDCl₃): δ = 5.05 (sept, ³*J*_{HH} = 6.3 Hz, 1H, OCH); 3.40 (s, 2H, CH₂); 1.89 (br, 2H, NH₂); 1.24 (d, ³*J*_{HH} = 6.2 Hz, 6H, CH*Me*₂) ppm.

L-Serine isopropylester, 8h.40f Compound 8h·HCl (1.61 g, 8.75 mmol) was dissolved in NaOH 1.0 M (8.75 mL, 8.75 mmol) in a 25 mL round bottom flask. The solution was stirred at room temperature for 10 minutes then extracted with CH_2Cl_2 (3 × 20 mL). The organic phase was dried with Na₂SO₄, filtered and the solvent was removed by distillation at 40 °C and p = 700 mbar. The product was obtained as a colourless viscous liquid, 1.11 g, yield 87%. IR(liquid film): $\nu = 3500-3100$ br, 3362 w, 3308 w, 2981 w, 2938 w, 2879 w, 1728 s ($\nu_{C=O}$), 1595 w, 1468 w, 1455 w, 1384 w-sh, 1375 m, 1326 w, 1209 m, 1179 m, 1145 m, 1105 s, 1039 m, 933 m, 903 w, 849 w, 822 m cm⁻¹. ¹H NMR (CDCl₃): $\delta =$ 5.07 (sept, ${}^{3}J_{HH} = 6.2$ Hz, 1H, OCH); 3.85 (dd, ${}^{2}J_{HH} = 10.2$ Hz, ${}^{3}J_{\rm HH} = 2.5$ Hz, 1H, CHH'); 3.71 (dd, ${}^{2}J_{\rm HH} = 10.7$ Hz, ${}^{3}J_{\rm HH} =$ 5.8 Hz, 1H, CHH'); 3.66-3.60 (m, 1H, CHN); 2.59 (br, 3H, NH₂ + OH); 1.29–1.25 (m, 6H, CHMe₂) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta =$ 173.3 (CO); 68.8 (OCH); 63.9 (CH₂); 56.1 (CHN); 21.7 (Me₂) ppm. ¹H NMR (CD₃CN): $\delta = 4.97$ (sept, ³ $J_{HH} = 6.3$ Hz, 1H, OCH); 3.62– $3.58 \text{ (m, 2H, CH}_2\text{)}; 3.38 \text{ (t, }^{3}J_{HH} = 4.8 \text{ Hz}, 1\text{H}, \text{CHN}\text{)}; 2.19 \text{ (br, 3H},$ $NH_2 + OH$; 1.23–1.20 (m, 6H, Me₂) ppm. ¹³C{¹H} NMR (CD₃CN): $\delta = 174.6$ (CO); 68.9 (OCH); 65.1 (CH₂); 57.3 (CHN); 22.0 and 21.9 (Me₂) ppm. The compound was dissolved in CDCl₃ and stored in a graduated Schlenk tube under nitrogen. The

concentration of the solution was determined by 1 H NMR, using CH₂Br₂ as internal standard.

L-Tyrosine isopropylester, 8i.^{49/} Colourless solid, yield 83%. ¹H NMR (CDCl₃): δ = 7.02 (d, ³*J*_{HH} = 8.3 Hz, 2H, Ph); 6.68 (d, ³*J*_{HH} = 8.3 Hz, 2H, Ph); 5.05 (sept, ³*J*_{HH} = 6.5 Hz, 1H, OCH); 3.73–3.63 (m, 1H, NCH); 3.23 (br, 3H, NH₂ + OH); 3.04 (dd, ²*J*_{HH} = 13.8 Hz, ³*J*_{HH} = 5.0 Hz, 1H, CHH'); 2.81 (dd, ²*J*_{HH} = 13.7 Hz, ³*J*_{HH} = 7.7 Hz, 1H, CHH'); 1.29–1.22 (m, 6H, CHMe₂) ppm.

L-Alanine ethylester, 8j.^{40k} Prepared from commercial Lalanine ethylester hydrochloride (Fluka). Pale yellow viscous liquid, yield 60%. ¹H NMR (CDCl₃): $\delta = 4.01$ (q, ³ $J_{\rm HH} = 6.6$ Hz, 2H, OCH₂); 3.37 (q, ³ $J_{\rm HH} = 6.6$ Hz, 1H, NCH); 1.54 (br, 2H, NH₂); 1.17 (d, ³ $J_{\rm HH} = 6.9$ Hz, 3H, CH₃CH); 1.13 (t, ³ $J_{\rm HH} = 7.1$ Hz, 3H, CH₃CH₂) ppm.

L-Leucine methylester, **8k**.⁴⁰ Pale yellow liquid, yield 42%. IR (liquid film): $\nu = 3380$ w-m ($\nu_{\rm N-H}$), 2956 m, 2932 w, 2869 w, 2851 w, 1740 vs ($\nu_{\rm C=O}$), 1610 w, 1468 m, 1436 m, 1384 w, 1368 w, 1316 w, 1270 m, 1198 s, 1145 m, 1010 m, 971 w, 921 w, 869 w, 837 w, 822 w cm⁻¹. ¹H NMR (CDCl₃): $\delta = 3.64$ (s, 3H, OMe); 3.41 (m, 1H, CH); 1.70 (m, 1H, CHMe₂); 1.65 (s, 2H, NH₂); 1.4 (m-br, 2H, CH₂); 0.86 (m, 6H, CHMe₂) ppm. ¹³C{¹H} NMR (CDCl₃): $\delta = 177.1$ (C=O); 53.0 (OMe); 52.0 (CH); 44.2 (CH₂); 24.8 (CHMe₂); 23.2, 22.0 (CHMe₂) ppm.

L-Phenylalanine isopropylester, 8.^{40m} Colourless solid, yield 36%. ¹H NMR (CDCl₃): δ: M7.32–7.22 (5H, Ph); 5.03 (sept, 1H, OCH); 3.71 (m, 1H, CH); 3.08, 2.90 (m, 2H, CH₂); 2.10 (s, 2H, NH₂); 1.24, 1.20 (m, 6H, OCH*M*e₂) ppm. ¹³C{¹H} NMR (CDCl₃): R (174.2 (C=O); 137.2 (*ipso*-Ph); 129.4, 128.5, 126.8 (Ph); 68.6 (OCH); 55.8 (CH); 40.9 (CH₂); 21.7 (OCH*M*e₂) ppm.

Reactions of NbF_5 with α -amino acid esters

Synthesis of [NbF₄(Me₂CHCH₂CHNH₂CO₂Me)₂][NbF₆], 9. A suspension of NbF₅ (167 mg, 1.15 mmol) in CH₂Cl₂ (15 mL) was treated with 8k (167 mg, 1.15 mmol). After 24 h stirring at room temperature, a colourless solution was obtained. By addition of hexane (10 mL), 9 was obtained as a colourless solid, which was recovered by filtration and dried in vacuo at room temperature. Yield 185 mg (48%). Anal. calcd for C₁₄H₃₀F₁₀N₂Nb₂O₄: C, 25.24; H, 4.54; N, 4.20; Nb, 27.89. Found: C, 25.01; H, 4.1; N, 4.01; Nb, 25.71. IR (solid state): $\nu = 3232$ w (ν_{N-H}), 3064 m, 2966 m, 2877 m, 1648 vs ($\nu_{C=O}$), 1499 m, 1456 m, 1374 w, 1328 w, 1289 w, 1262 w, 1170 w, 1129 w, 1067 w, 834 br-s cm⁻¹. ¹H NMR (CDCl₃): $\delta =$ 8.6, 7.0 (br, 2H, NH₂); 4.66 (s, 3H, OMe); 4.19 (m, 1H, CH); 2.02-1.52 (m, 3H, CHMe₂ + CH₂); 1.03 (m, 6H, CHMe₂) ppm. ¹⁹F NMR (CDCl₃): $\delta = 140 \ (\Delta \nu_{1/2} = 2.5 \times 10^3 \text{ Hz}, \text{NbF}_4^+); 103 \ (\text{decet}, {}^1J_{\text{NbF}})$ = 345 Hz, NbF₆⁻) ppm. ⁹³Nb NMR (CDCl₃): δ = -1553 (septet, ${}^{1}J_{\rm NbF} = 366 \text{ Hz}, \text{ NbF}_{6}^{-}) \text{ ppm}.$

Synthesis of [NbF₃(HNCH₂CH₂CH₂CH₂CH₂C₀)][NbF₆], **10**. Compound **10** was prepared by a procedure analogous to that described for the synthesis of **9**, from NbF₅ (222 mg, 1.18 mmol) and **8b** (85 mg, 0.59 mmol). Yield 167 mg (60%). Anal. calcd for C₅H₈-F₉NNb₂O₂: C, 12.75; H, 1.71; N, 2.97; Nb, 36.46. Found: C, 12.70; H, 1.61; N, 3.03; Nb, 36.27. IR (solid state): ν = 3381 vw, 2965 w, 1636 vs (ν _{C=O}), 1481 m, 1346 w, 1296 w, 1281 w, 1234 w, 1138 w, 1073 w, 1010 w, 983 m, 925 w, 910 w, 873 m, 765 m, 690 vs cm⁻¹. ¹H NMR (CD₃CN): δ = 11.77 (br, 1H, NH); 4.83 (m, 1H, CH); 3.84, 3.70 (m, 2H, NCH₂); 2.60, 2.28, 2.20 (m, 4H, CH₂) ppm. ¹⁹F NMR (CD₃CN): δ = 100.9 (decet, ¹*J*_{NbF} = 343 Hz, NbF₆⁻) ppm.

The reaction of NbF₅ (0.50 mmol) with **8b** (0.50 mmol) was performed also inside a sealed NMR tube (CD₂Cl₂, 0.70 mL). A complicated mixture of compounds were detected by NMR, including ethyl fluoride [δ (¹H) = 4.38 (m), 1.35 (t) ppm;⁵² δ (¹³C) = 77.6, 13.7 ppm;⁵² δ (¹⁹F) = -209.9 (br) ppm]. Addition of D₂O (*ca.* 0.1 mL) to a solution of **10** in CD₃CN (0.6 mL)⁵³ resulted in the formation of an abundant precipitate; a solution was separated whose ¹³C NMR analysis evidenced the presence of L-proline only.

Reactions of NbCl₅ with α-amino acid esters

Synthesis and isolation of [NbCl4(Me2CHCH2CHNH2CO2-Me)₂[[NbCl₆], 11a, and [NbCl₅(Me₂CHCH₂CHNH₃CO₂Me)] [NbCl₆], 12. A suspension of NbCl₅ (199 mg, 0.736 mmol) in CH_2Cl_2 (15 mL) was treated with 8k (97 mg, 0.74 mmol). After 24 h stirring at room temperature, a pale orange solution was obtained. By addition of hexane, 11a was obtained as a colourless solid. Yield 130 mg (44% based on Nb). Anal. calcd for C₁₄H₃₀Cl₁₀N₂Nb₂O₄: C, 20.24; H, 3.64; N, 3.37; Cl, 42.68; Nb, 22.37. Found: C, 20.05; H, 3.18; N, 3.27; Cl, 42.98; Nb, 23.13. IR (solid state): $\nu = 3292 \text{ w} (\nu_{\text{N-H}})$, 3243 w, 2959 w-m, 1633 m ($\nu_{\text{C}}=$ o), 1569 m, 1467 m, 1387 m, 1327 m, 1290 w-m, 1152 w-m, 1123 m, 1015 w-m, 934 w-m, 856 vs, 746 m-s cm⁻¹. ¹H NMR (CDCl₃): $\delta = 7.17$ (br, 2H, NH₂); 4.44 (s, 3H, OMe); 4.40 (m, 1H, CH); 1.96 (m, 1H, CHMe₂); 1.51 (m, 2H, CH₂); 1.10 (m, 6H, CHMe₂) ppm. ⁹³Nb NMR (CDCl₃): $\delta = 7.8 (\Delta v_{1/2} = 4 \times 10^2 \text{ Hz, NbCl}_6^-) \text{ ppm.}^{42a}$ Hydrolysis⁵³ of a CD₃CN solution (0.6 mL) of **11a** resulted in the formation of an abundant precipitate; a solution was separated whose ¹³C NMR analysis evidenced the presence of 8k only.

In a different experiment, the reaction solution was set aside at *ca.* -30 °C for two weeks. Thus pale yellow crystals of **12** were collected. Yield 65 mg (25% based on Nb). Anal. calcd for C₇-H₁₆Cl₁₁NNb₂O₂: C, 11.64; H, 2.23; N, 1.94; Cl, 54.01; Nb, 25.74. Found: C, 11.75; H, 2.16; N, 1.98; Cl, 53.70; Nb, 25.55. IR (solid state): $\nu = 3282$ w-br (ν_{N-H}), 3241 w, 2959 w-m, 1641 m ($\nu_{C=O}$), 1559 m, 1457 m-w, 1377 m, 1327 m, 1281 w-m, 1158 w, 1128 m, 1011 w-m, 922 w-m, 860 vs, 744 m-s cm⁻¹.

Synthesis of $[NbCl_4(Me_2CHCH_2CHNH_2CO_2Et)_2][NbCl_6]$, 11b, $[NbCl_4(HNCH_2CH_2CH_2CHCO_2Et)_2][NbCl_6]$, 11c. These products were prepared by a procedure analogous to that described for 11a, from the appropriate niobium pentahalide (*ca.* 0.70 mmol)/ α -amino acid ester combination.

11b. Colourless viscous solid, yield 73%. Anal. calcd for C₁₆-H₃₄Cl₁₀N₂Nb₂O₄: C, 22.38; H, 3.99; N, 3.26; Cl, 41.28; Nb, 21.64. Found: C, 22.25; H, 4.06; N, 3.16; Cl, 41.70; Nb, 21.29. ¹H NMR (CDCl₃): $\delta = 7.00$ (br, 2H, NH₂); 4.50 (m, 2H, OCH₂); 4.31 (br, 1H, CH); 1.96 (m, 1H, CHMe₂); 1.47 (m, 2H, CH₂); 1.42 (m, 3H, OCH₂CH₃); 1.07 (m, 6H, CHMe₂) ppm. ¹³C NMR (CDCl₃): $\delta =$ 170.2 (C=O); 65.5 (OCH₂); 54.1 (CH); 39.4 (CH₂); 25.0 (CHMe₂); 22.3, 21.9 (CHMe₂); 14.1 (OCH₂CH₃) ppm. ⁹³Nb NMR (CDCl₃): $\delta = 13 (\Delta v_{1/2} = 4 \times 10^2 \text{ Hz}, \text{NbCl}_6^{-}); -540 (\Delta v_{1/2} = 8 \times 10^3 \text{ Hz}, \text{NbCl}_4^{+})$ ppm.

11c. Colourless solid, yield 70%. Anal. calcd for $C_{14}H_{26}Cl_{10}$ - $N_2Nb_2O_4$: C, 20.34; H, 3.17; N, 3.39; Cl, 42.88; Nb, 22.48. Found:

Table 4 Crystal data and experimental details for 3, 4, 5a and 12

	3	4	5a	12
Formula	$\mathrm{C_{10}H_{12}Cl_8N_2Nb_2O_2}$	C ₆ H ₁₆ Cl ₆ Nb	$\mathrm{C_{10}H_{12}Cl_8N_2Nb_2O_2}$	C ₇ H ₁₆ Cl ₁₁ NNb ₂ O ₂
Fw	661.64	407.81	518.30	721.98
Т, К	100(2)	100(2)	100(2)	100(2)
Crystal system	Monoclinic	Monoclinic	Orthorhombic	Monoclinic
Space group	$P2_1/c$	$P2_1/c$	P21212	$P2_1$
a, Å	6.7734(9)	8.065(3)	6.9864(4)	10.045(2)
b, Å	13.4440(18)	18.492(7)	12.7821(7)	6.9628(17)
<i>c</i> , Å	11.9844(16)	10.184(4)	20.9934(11)	16.994(4)
<i>β</i> , °	103.854(2)	94.036(5)	90	106.428(3)
Cell volume, Å ³	1059.6(2)	1515.1(10)	1874.73(18)	1140.2(5)
Ζ	2	4	4	2
$D_{\rm c}$, g cm ⁻³	2.074	1.788	1.836	2.103
μ , mm ⁻¹	2.097	1.821	1.635	2.297
F(000)	640	808		700
Independent reflections	$2315 [R_{int} = 0.0261]$	$3507 [R_{int} = 0.0656]$	$3338 \left[R_{\rm int} = 0.0312 \right]$	$4492 \left[R_{\text{int}} = 0.0312 \right]$
Data/restraints/parameters	2315/0/109	3507/2/137	3338/0/195	4492/4/217
Goodness on fit on F^2	1.065	1.073	1.214	1.023
$R_1 \left(I > 2\sigma(I) \right)$	0.0406	0.0442	0.0249	0.0217
wR_2 (all data)	0.0913	0.0870	0.0575	0.0428
Largest diff. peak and hole, e $Å^{-3}$	2.092/-1.392	0.769 / -0.891	0.372/-0.692	0.286 / -0.360

C, 19.98; H, 3.54; N, 3.47; Cl, 42.09; Nb, 21.79. IR (solid state): $\nu = 2961$ w, 2880 w, 1609 vs ($\nu_{C=O}$), 1456 s, 1342 w, 1271 m, 1233 w-m, 1052 w, 974 m, 922 w-m, 869 m, 802 m, 756 m, 656 w-m cm⁻¹. ¹H NMR (CD₃CN): 7.2 (br, 2H, NH₂); 4.73 (m, 1H, CH); 4.31 (m, 2H, OCH₂); 3.79 (br, 2H, NCH₂); 2.58, 2.18 (m, 4H, CH₂); 1.30 (m, 3H, OCH₂CH₃) ppm. ⁹³Nb NMR (CD₃CN): $\delta = 4$ ($\Delta \nu_{1/2} = 3 \times 10^2$ Hz, NbCl₆⁻); -569 ($\Delta \nu_{1/2} = 2 \times 10^3$ Hz, NbCl₄⁺) ppm.

Synthesis of NbCl₃(OCH₂CHNHCO₂ⁱPr), 13. A suspension of NbCl₅ (258 mg, 0.954 mmol) and 8h (1.32 mL of a 0.72 M solution in CDCl₃, 0.95 mmol) in CHCl₃ (8 mL) was refluxed for 31 h. Then, the mixture was allowed to cool to room temperature and the colourless solution was separated from the colourless precipitate. The solid was washed with CH_2Cl_2 (2 \times 5 mL) and dried in vacuo. Yield 239 mg (66% based on Nb). Anal. calcd for C₆H₁₁Cl₃NNbO₃: Cl, 30.9. Found: Cl, 30.8.⁵⁴ IR (solid state): $\nu = 3580-3200$ br, 3120 m-br, 2985 m, 2965 m, 2940 m, $1732 \text{ s} (\nu_{C=0}), 1675 \text{ w}, 1576 \text{ m}, 1486 \text{ m}, 1471 \text{ m}, 1387 \text{ m}, 1377 \text{ m},$ 1304 w, 1258 s, 1184 w, 1146 m, 1098 s, 1080 s, 1033 s, 937 m, 895 m, 860 w, 800 s, 757 m cm⁻¹. ¹H NMR (CD₃CN): $\delta = 6.98$ (br, 1H, NH); 5.09 (sept, ${}^{3}J_{HH} = 6.2$ Hz, 1H, CHMe₂); 4.18-4.07 (m,1H, NCH), 4.01–3.92 (m, 2H, CH₂); 1.28 (d, ${}^{3}J_{HH} = 6.1$ Hz, 6H, CHMe₂) ppm. ¹³C{¹H} NMR (CD₃CN): $\delta = 167.3$ (C=O); 72.2 (CHMe₂); 63.4 (CH₂); 59.9 (br, NCH); 21.7, 21.6 (CHMe₂) ppm. ⁹³Nb{¹H} NMR (CD₃CN): $\delta = -493.7 (\Delta \nu_{1/2} = 2.3 \times 10^3 \text{ Hz}) \text{ ppm}.$

A sample of **13** was suspended in D_2O (1 mL) with vigorous stirring for 14 h.⁵³ NMR analysis of the solution revealed the presence of **8h** only (pH = 1.8).

MBr_5 (M = Nb, Ta) mediated formation of EtBr from L-proline ethylester

The reactions of MBr_5 (*ca.* 0.50 mmol) with **8b** (0.50 mmol) were carried out in CD_2Cl_2 (*ca.* 1 mL). Subsequent NMR analysis pointed out the formation of a complicated mixture of compounds, including ethyl bromide.⁵² In a different

experiment, the mixture obtained from NbBr₅/**8b** was eliminated of the volatile materials; the residue was dissolved into CD₃CN (0.6 mL) and then treated with H₂O (*ca.* 0.2 mL).⁵³ A yellow solution was separated from an abundant precipitate, and ¹³C NMR analysis of the solution evidenced the presence of L-proline only.

X-ray crystallographic studies

Crystal data and collection details for 3, 4, 5a and 12 are reported in Table 4. The diffraction experiments were carried out on a Bruker APEX II diffractometer equipped with a CCD detector and using Mo-K α radiation ($\lambda = 0.71073$ Å). Data were corrected for Lorentz polarization and absorption effects (empirical absorption correction SADABS).⁵⁵ Structures were solved by direct methods and refined by full-matrix least-squares based on all data using $F^{2,56}$ All non-hydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atoms were fixed at calculated positions and refined by a riding model, except hydrogens bonded to N(1) in 4, 5a and 12 which were located in the Fourier map and refined isotropically using the 1.2 fold (for 4 and 5a) and 1.5 fold (for 12) $U_{\rm iso}$ value of the parent N-atom. The N(1)–H distances were restrained to 0.93 Å (s.u. 0.02).

Computational studies

The computational geometry optimizations were carried out without symmetry constrains using the hybrid-GGA EDF2 functional,⁵⁷ in combination with the 6-31G** basis set (ECP-based LANL2DZ basis set for elements beyond Kr).⁵⁸ The "restricted" formalism was applied in all cases. The software used was Spartan 08.⁵⁹ Further computational geometry optimizations were carried out without symmetry constrains, using the hyper-GGA functional M06,⁶⁰ in combination with a polarized basis set composed by the 6-31G(d,p) set on the light atoms

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and the ECP-based LANL2TZ(f) on the metal centre.⁶¹ The C-PCM implicit solvation model ($\varepsilon = 9.08$) was added to M06 calculations.⁶² Gaussian '09 was used as software.⁶³ All the stationary points were characterized by IR simulations (harmonic approximation), from which zero-point vibrational energies and thermal corrections (T = 298.15 K) were obtained.⁶⁴ Vibrational simulation supported the interpretation of experimental IR data. Cartesian coordinates of the optimized geometries are collected in a separated. *xyz* file.

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Notes and references

- (a) K. Drauz, I. Grayson, A. Kleemann, H.-P. Krimmer, W. Leuchtenberger and C. Weckbecker, Amino Acids, Ullmann's Encyclopedia of Industrial Chemistry, Wiley-VCH, Weinheim, Germany, 2012, vol. 3, pp. 1–58; (b) K. Araki and T. Ozeki, Amino Acids, Kirk-Othmer Encyclopedia of Chemical Technology, John Wiley & Sons, Hoboken, NJ, USA, 5th edn, 2004, vol. 2, pp. 554–618.
- 2 (a) S. Kaneko, Y. Kumatabara and S. Shirakawa, Org. Biomol. Chem., 2016, 14, 5367–5376; (b) M. A. Ashley, J. S. Hieschi, J. A. Izzo and M. J. Vetticatt, J. Am. Chem. Soc., 2016, 138, 1756–1759; (c) X. Zhao, B. Zhou and Z. Jiang, Synlett, 2016, 26, 2216–2230; (d) M. Hibi and J. Ogawa, Appl. Microbiol. Biotechnol., 2014, 98, 3869–3876; (e) P. Singh, K. Samanta, S. K. Das and G. Panda, Org. Biomol. Chem., 2014, 12, 6297–6339; (f) V. Srivastava, Asymmetric organocatalysis, 2013, 1, 8–23; (g) J. G. Hernández and E. Juaristi, Chem. Commun., 2012, 48, 5396–5409; (h) J. Paradowska, M. Stodulski and J. Mlynarski, Angew. Chem., Int. Ed., 2009, 48, 4288–4297.
- 3 (a) K. M. Engle, Pure Appl. Chem., 2016, 88, 119–138; (b)
 M. A. Telpoukhovskaia and C. Orvig, Chem. Soc. Rev., 2013, 42, 1836–1846; (c) T. Kiss, J. Inorg. Biochem., 2013, 128, 156–163; (d) J. S. Casas, M. D. Couce and J. Sordo, Coord. Chem. Rev., 2012, 256, 3036–3062; (e) W. Maret, J. Inorg. Biochem., 2012, 111, 110–116; W. Beck, Z. Anorg. Allg. Chem., 2011, 637, 1647–1672. (f) R. H. Fish, Aust. J. Chem., 2010, 63, 1505–1513; (g) K. Severin, R. Bergs and W. Beck, Angew. Chem., Int. Ed., 1998, 37, 1634–1654.
- 4 (a) R. M. Palou, A. A. Vallejo Cardona, R. J. Reyes Avila, J. Clavel Lopez, C. Bernal Huicochea, M. Ramirez de Santiago and J. A. Aburto Anell, US 2016/0068471 A1, Istituto Mexicano del Petroleo, 2016; (b) G. Lahouar, R. Touati, S. Khamri, J. Marrot and B. Ben Hassine, J. Chem. Pharm. Res., 2015, 7, 283–288; (c) F.-S. He, J.-H. Jin, Z.-T. Yang, X. Yu, J. S. Fossey and W.-P. Deng, ACS Catal., 2016, 6, 652–656; (d) A. K. Padala, R. R. Kumar, S. Athimoolam and Q. N. Ahmed, Org. Lett., 2016, 18, 96–99; (e) Y. Kohsaka, Y. Matsumoto and T. Kitayama, Polym. Chem., 2015, 6, 5026–5029; (f) H. Waldmann, Liebigs Ann. Chem., 1990, 671–680.

- 5 B. Schreiner, B. Wagner-Schuh and W. Beck, *Z. Naturforschg.*, 2010, 65, 679–686; R. Krämer, M. Maurus, K. Polborn, K. Sünkel, C. Robl and W. Beck, *Chem.-Eur. J.*, 1996, 23, 1518–1526; K. Haas and W. Beck, *Eur. J. Inorg. Chem.*, 2001, 2485–2488.
- 6 G. C. Barrett, Amino Acids, Peptides and Proteins, R.S.C. Publications, London1996, vol. 27, p. 1.
- 7 R. H. Fish and G. Jaouen, *Organometallics*, 2003, 22, 2166–2177.
- 8 (a) L. Kiss, M. Kardos, E. Forró and F. Fülöp, Eur. J. Org. Chem., 2015, 1283-1289; (b) L. Konnert, B. Reneaud, R. Marcia de Figueiredo, J.-M. Campagne, F. Lamaty, J. Martinez and E. Colacino, J. Org. Chem., 2014, 79, 10132-10142; (c) E. A. Egbewande, L. E. H. Paul, B. Therrien and J. Furrer, Eur. J. Inorg. Chem., 2014, 1174-1184; (d) M. Periasamy, R. Gurubrahaman, N. Sanjeevakumar, Dalai, L. Anakonda, P. O. Reddy, S. Suresh, M. S. Satishkumar, M. Padmaja, M. N. Reddy, S. Suresh, S. Anwar, G. P. Muthukumaragopal, P. Vairaprakash and Seenivasaperumal, Chimia, 2013, 67, 23-29; (e) М V. K. Tandon, D. B. Yadav, R. V. Singh, A. K. Chaturvedi and P. K. Shukla, Bioorg. Med. Chem. Lett., 2005, 15, 5324-5328; (f) S. L. Manjinder, K. R. Yeeman, N. G. J. Michael and C. V. John, J. Org. Chem., 2002, 67, 1536-1547.
- 9 S. Jugé, J. Bayardon, E. Remond and M.-J. Ondel-Eymin, WO2013030193A1, CNRS and University of Bourgogne, France, 2013; J. Kim, J. Kim, S. Song, O. Jung and H. Suh, J. Inclusion Phenom. Macrocyclic Chem., 2007, 58, 187–192;
 G. Pollini, N. Baricordi, S. Benetti, C. De Risi and V. Zanirato, Tetrahedron Lett., 2005, 46, 3699–3701;
 C. Somlai, A. Peter, P. Forgo and B. Penke, Synth. Commun., 2003, 33, 1815–1820; J.-A. Ma, Angew. Chem., Int. Ed., 2003, 42, 4290–4299; T. Zieliński, M. Achmatowicz and J. Jurczak, Tetrahedron: Asymmetry, 2002, 13, 2053–2059.
- 10 A. C. Fonseca, M. H. Gil and N. P. Simoes, *Prog. Polym. Sci.*, 2014, 39, 1291–1311; A. M. Perez-Lopez, D. Gonzalez-Calderon, A. Occorso, J. Galindo-Angel, J. F. Dominguez-Seglar, J. A. Tamayo, M. Diaz-Gavilan and J. A. Gomez-Vidal, *Synlett*, 2014, 25, 2319–2322; N. H. Powell, G. J. Clarkson, R. Notman, P. Raubo, N. G. Martin and M. Shipman, *Chem. Commun.*, 2014, 50, 8797–8800; E. Aresu, S. Fioravanti, S. Gasbarri, L. Pellacani and F. Ramadori, *Amino Acids*, 2013, 44, 977–982; R. Katsarava and Z. Gomurashvili, *Handbook of Biodegradable Polymers*, ed. A. Lendlein and A. Sisson, 2011, pp. 107–131; A. Nagai, T. Miyagawa, H. Kudo and T. Endo, *Macromolecules*, 2003, 36, 9335–9339; H. Waldmann, *Synlett*, 1995, 133–141.
- 11 G. E. Sapper and U. Wannag, Z. Anorg. Allg. Chem., 1975, 417, 41–43.
- 12 D. S. Aspandiyarova and S. S. Uskova, *Izv. Akad. Nauk Kazakhskoi SSR, Ser. Khim.*, 1983, 76–78; *Chem. Abstr.*, 1983, 98, 209055.
- 13 (a) C. Evangelisti, M. Hayatifar, F. Marchetti, M. Marelli,
 G. Pampaloni and F. Piccinelli, *Inorg. Chem.*, 2016, 55, 1816–1820; (b) F. Marchetti, G. Pampaloni and S. Zacchini, *RSC Adv.*, 2014, 4, 60878–60882; (c) F. Marchetti and

G. Pampaloni, *Chem. Commun.*, 2012, **48**, 635–653, and references therein.

- 14 (a) M. Bortoluzzi, F. Marchetti, G. Pampaloni, M. G. Murrali and S. Zacchini, *Dalton Trans.*, 2014, 44, 8729–8738; (b)
 M. Bortoluzzi, G. Bresciani, F. Marchetti, G. Pampaloni and S. Zacchini, *Dalton Trans.*, 2014, 44, 10030–10037.
- 15 M. Bortoluzzi, M. Hayatifar, F. Marchetti, G. Pampaloni and S. Zacchini, *Inorg. Chem.*, 2015, **54**, 4047–4055.
- 16 The pentafluorides of niobium and tantalum have tetranuclear structure in the solid state,^{17a} while the heavier pentahalides^{17b,c} are dinuclear in the solid state. All of these compounds will be mentioned by the empirical formulas MX_5 throughout this paper.
- 17 (a) A. J. Edwards, J. Chem. Soc., 1964, 3714-3718; (b)
 A. F. Wells, Structural Inorganic Chemistry, Clarendon Press, Oxford, 5th edn, 1993; (c) K. Habermehl, I. Pantenburg and G. Meyer, Acta Crystallogr., 2010, 66, i67.
- 18 (a) G. B. Deacon and R. J. Phillips, *Coord. Chem. Rev.*, 1980, 33, 227–250; (b) S. D. Robinson and M. F. Uttley, *J. Chem. Soc., Dalton Trans.*, 1973, 1912–1920; (c) K. Nakamoto, J. Fujita, S. Tanaka and S. Kobayashi, *J. Am. Chem. Soc.*, 1957, 79, 4904–4908.
- 19 V. Lacerda Jr, D. Araujo dos Santos, L. C. Da Silva-Filho, S. J. Greco and R. Bezerra dos Santos, *Aldrichimica Acta*, 2012, 45, 19–27; Y. Satoh and Y. Obora, *Eur. J. Org. Chem.*, 2015, 5041–5054.
- 20 M. M. Rahman, M. D. Smith and D. V. Peryshkov, *Inorg. Chem.*, 2016, **55**, 5101–5103.
- 21 The combination of TiCl₄ with *N*,*N*-dimethylphenylalanine, in the absence of NEt₃, afforded a mixture of two products, that could not be unambiguously characterized.
- 22 The reaction of the $TiCl_4$ L-proline adduct **1a** with NEt₃ affords is a deep green, paramagnetic compound probably formed *via* activation reactions involving the amino acid ligand.
- 23 F. H. Allen, O. Kennard, D. G. Watson, L. Brammer, A. G. Orpen and R. Taylor, *J. Chem. Soc., Perkin Trans.* 2, 1987, S1–S19.
- 24 (a) M.-C. Lasne and J.-L. Ripoll, *Tetrahedron Lett.*, 1982, 23, 1587–1588; (b) F. Tureček and Z. Havlas, *J. Org. Chem.*, 1986, 51, 4066–4067.
- 25 (a) C. A. Buehler and J. O. Harris, *J. Am. Chem. Soc.*, 1950, 72, 5015–5017; (b) E. Lippmann and V. Becker, *Z. Chem.*, 1974, 14, 357–358; (c) M. Weissenfels and J. Punkt, *Tetrahedron*, 1978, 34, 311–316.
- 26 C. M. Mayhan, T. J. Szabo, J. E. Adams and C. A. Deakyne, *Struct. Chem.*, 2013, 24, 2089–2099; J. Szklarzewicz and K. Stadnicka, *Inorg. Chim. Acta*, 2012, 392, 131–136; F. Dong, S. Heinbuch, Y. Xie, J. J. Rocca, E. R. Bernstein, Z.-C. Wang, K. Deng and S.-G. He, *J. Am. Chem. Soc.*, 2008, 130, 1932–1943; A. Kovacs and G. Frenking, *Organometallics*, 2001, 20, 2510–2524.
- 27 (a) Y. Tsuda and Y. Okada, Solution-Phase Peptide Synthesis in Amino Acids, Peptides and Proteins in Organic Chemistry, ed. A.
 B. Hughes, Wiley-VCH, Weinheim, 2011, vol. 3; (b)
 A. D. Borthwick, Chem. Rev., 2012, 112, 3641–3716.

- 28 (a) Aminoacids, peptides and proteins in organic chemistry, ed.
 A. B. Hughes, Wiley, New York, 2011; (b) Á. L. Fuentes de Arriba, L. Simón, C. Raposo, V. Alcázar and J. R. Morán, *Tetrahedron*, 2009, 65, 4841–4845.
- 29 M. Bortoluzzi, F. Marchetti, M. G. Murrali and G. Pampaloni, *Inorg. Chim. Acta*, 2015, **427**, 150–154, and references therein.
- 30 F. Marchetti, G. Pampaloni and C. Pinzino, *Chem.-Eur. J.*, 2013, **19**, 13962–13969.
- 31 The analogous reaction PCl₅/NbCl₅/L-methionine proceeded with clean formation of POCl₃, and led to the isolation of a brown, solid product containing [NbCl₆]⁻, which could not be clearly identified.
- 32 H. Chen, X. Xu, L. Liu, G. Tang and Y. Zhaoa, *RSC Adv.*, 2013, 3, 16247–16250.
- 33 (a) M. Bortoluzzi, E. Ferretti, F. Marchetti, G. Pampaloni and S. Zacchini, *Dalton Trans.*, 2016, 45, 6939–6948; (b)
 W. Levason, M. E. Light, G. Reid and W. Zhang, *Dalton Trans.*, 2014, 43, 9557–9566; (c) R. Bondi, F. Marchetti, G. Pampaloni and S. Zacchini, *Polyhedron*, 2015, 100, 192–198.
- 34 F. H. Allen, O. Kennard, D. G. Watson, L. Brammer, A. G. Orpen and R. Taylor, *J. Chem. Soc., Perkin Trans.* 2, 1987, S1–S19.
- 35 S. Parsons, H. D. Flack and T. Wagner, *Acta Crystallogr.*, 2013, **69**, 249–259.
- 36 C.-I. Bränden and I. Lindqvist, *Acta Chem. Scand.*, 1963, **17**, 353–361.
- 37 D. V. Drobot and E. A. Pisarev, Russ. J. Coord. Chem., 1981, 26, 1–8.
- 38 M. Bortoluzzi, F. Marchetti, G. Pampaloni and S. Zacchini, *Chem. Commun.*, 2015, **55**, 1323–1325.
- 39 (a) M. Bortoluzzi, F. Marchetti, G. Pampaloni, C. Pinzino and S. Zacchini, *Inorg. Chem.*, 2016, 55, 887–893; (b) M. Bortoluzzi, F. Marchetti, G. Pampaloni and S. Zacchini, *New J. Chem.*, 2016, 40, 8271–8281; (c) M. Bortoluzzi, F. Marchetti, G. Pampaloni and S. Zacchini, *Inorg. Chem.*, 2014, 53, 3832–3838; (d) S. Dolci, F. Marchetti, G. Pampaloni and S. Zacchini, *Dalton Trans.*, 2013, 42, 5635–5648.
- 40 (a) A. Sferrazza, A. Triolo, L. M. Migneco and R. Caminiti, Curr. Org. Chem., 2015, 19, 99-104; (b) J. Li and Y. Sha, Molecules, 2008, 13, 1111-1119; (c) T. Yamada, P. J. Lukac, T. Yu and R. G. Weiss, Chem. Mater., 2007, 19, 4761-4768; (d) R. H. Mazur, J. A. Reuter, K. A. Swiatek and J. M. Schlatter, J. Med. Chem., 1973, 16, 1284-1287; (e) Q.-Y. Wei, H. Jiang, J.-X. Zhang, C. Zhang and P.-F. Guo, Asian J. Chem., 2012, 24, 2383–2388; (f) R. Quevedo, L. Pabon and Y. Quevedo-Acosta, J. Mol. Struct., 2013, 1041, 68-72; (g) M. van Dijk, T. M. Postma, D. T. S. Rijkers, R. M. J. Liskamp, C. F. van Nostrum and W. E. Hennink, Polymer, 2010, 51, 2479-2485; (h) V. M. Zakharova, M. Serpi, I. S. Krylov, L. W. Peterson, J. M. Breitenbach, K. Z. Borysko, J. C. Drach, M. Collins, J. M. Hilfinger, B. A. Kashemirov and C. E. McKenna, J. Med. Chem., 2011, 54, 5680-5693; (i) T. J. Trivedi, K. S. Rao, T. Singh, S. K. Mandal, N. Sutradhar, A. B. Panda and A. Kumar,

ChemSusChem, 2011, 4, 604–608; (j) N. Nuñez-Dallos, A. Reyes and R. Quevedo, *Tetrahedron Lett.*, 2012, 53, 530– 534; (k) I. Aviva and Z. Gross, *Chem. Commun.*, 2006, 4477– 4479; (l) H. Chen, M. He, Y. Wang, L. Zhai, Y. Cui, Y. Li, H. Zhou, X. Hong and Z. Deng, *Green Chem.*, 2011, 13, 2723–2726; (m) L. Fabian, M. Gómes, J. A. Caturelli Kuran, G. Moltrasio and A. Moglioni, *Synth. Commun.*, 2014, 44, 2386–2392.

- 41 (a) M. Hayatifar, F. Marchetti, G. Pampaloni and S. Zacchini, *Polyhedron*, 2014, **70**, 6–10; (b) F. Marchetti, G. Pampaloni, C. Pinzino and S. Zacchini, *Eur. J. Inorg. Chem.*, 2013, 5755–5761; (c) M. Jura, W. Levason, G. Reid and M. Webster, *Dalton Trans.*, 2009, 7610–7612.
- 42 (a) W. Levason, G. Reid and W. Zhang, J. Fluorine Chem., 2015, 172, 62–67; (b) W. Levason, G. Reid, J. Trayer and W. Zhang, Dalton Trans., 2014, 43, 3649–3659; (c) R. Haiges, P. Deokar and K. O. Christe, Z. Anorg. Allg. Chem., 2014, 640, 1568–1575; (d) M. Jura, W. Levason, R. Ratnani, G. Reid and M. Webster, Dalton Trans., 2010, 39, 883–891.
- 43 S. L. Benjamin, W. Levason and G. Reid, *Chem. Soc. Rev.*, 2013, **42**, 1460–1499.
- 44 (a) Ru(II): P. Pelagatti, A. Bacchi, F. Calbiani, M. Carcelli, L. Elviri, C. Pelizzi and D. Ragolino, J. Organomet. Chem., 2005, 690, 4602-4610; (b) Ru(II): H. Dialer, P. Mayer, K. Polborn and W. Beck, Eur. J. Inorg. Chem., 2011, 1051-1055; (c) Ru(II): T. Reiner, D. Jantke, X. Miao, A. N. Marziale, F. Kiefer and J. Eppinger, Dalton Trans., 2013, 42, 8692-8703; (d) Ru(IV): K. Severin, S. Mihan and W. Beck, Chem. Ber., 1995, 128, 1117-1125; (e) Os(II): R. A. Sarfraz, T. G. Kazi, S. Iqbal, H. I. Afridi, M. K. Jamali, N. Jalbani and M. B. Arain, Adv. Organomet. Chem., 2008, 22, 187–192; (f) Co(II): Z. P. Zubreichuk, T. O. Denisova, V. A. Knizhnikov and S. E. Nefedov, Russ. Chem. Bull., 2004, 53, 1770-1772; (g) Co(III): Y. Ohgo, Y. Ohashi, M. Hagiwara, M. Shida, Y. Arai, S. Takeuchi and Y. Ohgo, Acta Crystallogr., 2000, C56, 1062–1064; (h) Rh(I): J. Rauzaud, M. D. Jones, R. Raja, B. F. G. Johnson, J. M. Thomas and M. J. Duer, Helv. Chim. Acta, 2003, 86, 1753–1759; (i) Ru(III), Ir(III), Pd(II): W. Hoffmüller, K. Polborn, J. Knizek, H. Noth and W. Beck, Z. Anorg. Allg. Chem., 1997, 623, 1903-1911; (j) Pt(II): E. Chardon, G. Dahm, G. Guichard and S. Bellemin-Lapollaz, Chem.-Asian J., 2013, 8, 1232-1242; (k) Pd(II), Pt(II): E. Schumann, C. Robl and W. Beck, Z. Naturforsch., 1994, 49B, 1569-1579.
- 45 (a) W.-Q. Zhang, A. C. Whitwood, I. J. S. Fairlamb and J. M. Lynam, *Inorg. Chem.*, 2010, 49, 8941–8952; (b) R. Bergs, R. Krämer, M. Maurus, B. Schreiner, R. Urban, C. Missling, K. Polburn, K. Sunkel and W. Beck, *Z. Naturforsch.*, 1996, 51B, 187–200.
- 46 P. J. McKarns, M. J. Heeg and C. H. Winter, *Inorg. Chem.*, 1998, **37**, 4743–4747.
- 47 J. M. Wolff and W. S. Sheldrick, *J. Organomet. Chem.*, 1997, 531, 141–149.

- 48 F. Calderazzo, P. Pallavicini, G. Pampaloni and P. F. Zanazzi, J. Chem. Soc., Dalton Trans., 1990, 2743–2746.
- 49 (a) A. Jutand, Eur. J. Inorg. Chem., 2003, 2017–2040; (b)
 W. J. Geary, Coord. Chem. Rev., 1971, 7, 81–122.
- 50 E. König, Magnetische Eigenschaften der Koordinations und Metallorganischen Verbindungen der Übergangselemente, in Landolt-Börnstein, Zahlenwerte und Funktionen aus Naturwissenschaften und Technik, Springer-Verlag, Berlin, Göttingen, Heidelberg, 6th edn, 1966, vol. 2, p. 16.
- 51 D. A. Skoog, D. M. West, F. J. Holler and S. R. Crouch, *Fundamentals of Analytical Chemistry*, Thomson Learning Inc, Belmont, CA, 8th edn, 2004.
- 52 (a) N. J. R. van Eikema Hommes and T. Clark, *J. Mol. Model.*,
 2005, 11, 175–185; (b) Spectral Database for Organic Compounds, SDBS at: http://www.sdbs.db.aist.go.jp/sdbs/cgi-bin/direct_frame_top.cgi.
- 53 The addition of water to the reaction mixtures facilitates the release of the organic material from the highly oxophilic metal species, and allows the spectroscopic identification of the former. This strategy has been successfully adopted by ourselves in previous works, having proved that H₂O is generally inert towards ligand activation reactions.³⁸
- 54 Determined by potentiometric titration at pH = 2.0.
- 55 G. M. Sheldrick, *SADABS, Program for empirical absorption correction*, University of Göttingen, Göttingen, Germany, 1996.
- 56 G. M. Sheldrick, *SHELX97*, University of Göttingen, Göttingen, Germany, 1997.
- 57 C. Y. Lin, M. W. George and P. M. W. Gill, *Aust. J. Chem.*, 2004, 57, 365–370.
- 58 (a) M. Dolg, Modern Methods and Algorithms of Quantum Chemistry, ed. J. Grotendorst, John Neumann Institute for Computing, NIC series, Jülich, 2000, vol. 1, p. 479; (b) P. J. Hay and W. R. Wadt, J. Chem. Phys., 1985, 82, 270–283; (c) P. J. Hay and W. R. Wadt, J. Chem. Phys., 1985, 82, 299–310; (d) W. J. Henre, R. Ditchfield and J. A. Pople, J. Chem. Phys., 1972, 56, 2257–2261.
- 59 Spartan '08, version 1.1.1, Wavefunction, Inc., Irvine CA, 2009Except for molecular mechanics and semi-empirical models, the calculation methods used in Spartan have been documented in: Y. Shao, *et al.*, *Phys. Chem. Chem. Phys.*, 2006, **8**, 3172–3191.
- 60 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, **120**, 215–241.
- 61 (a) L. E. Roy, P. J. Hay and R. L. Martin, J. Chem. Theory Comput., 2008, 4, 1029–1031; (b) M. M. Francl, W. J. Pietro, W. J. Hehre, J. S. Binkley, D. J. DeFrees, J. A. Pople and M. S. Gordon, J. Chem. Phys., 1982, 77, 3654–3665.
- 62 (*a*) M. Cossi, N. Rega, G. Scalmani and V. Barone, *J. Comput. Chem.*, 2003, **24**, 669–681; (*b*) V. Barone and M. Cossi, *J. Phys. Chem. A*, 1998, **102**, 1995–2001.
- 63 M. J. Frisch, *et al.*, *Gaussian 09, Revision C.01*, Gaussian, Inc., Wallingford CT, 2010.
- 64 C. J. Cramer, *Essentials of Computational Chemistry*, Wiley, Chichester, 2nd edn, 2004.