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Diazepinium perchlorate: a neutral catalyst for mild, solvent-free acetylation of carbohydrates and other substances†

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Diazepinium perchlorate, an essentially neutral organic salt possessing excellent stability, has been found to be well suited for the acetylation of free as well as partially protected sugars, phenols, thiophenols, thiols and other alcohols as well as amines. The diazepinium perchlorate-catalyzed acetylation is mild, organic and solvent-free and leaves acid sensitive protecting groups such as TBDMS/TBDPS/Tr ethers and isopropylidene/benzylidene acetals present on a substrate unaffected. Regioselective hydroxyl protection in partially protected carbohydrate derivatives/polyhydroxylic compounds was possible and was proved to be a convenient time-saving alternative to the conventional synthesis of such compounds. Easy preparation of the catalyst, mild reaction conditions and an environmentally benign protocol are some of the notable features of this reaction. The results obtained on the acetylation of phenols and thiophenols could be rationalized through their local nucleophilicity index obtained from DFT calculations.

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1. Introduction

Acyl transfer reaction is one of the most commonly used reactions in synthetic organic chemistry, in particular, synthetic carbohydrate and natural products chemistry, for the protection of hydroxyl groups as esters and therefore application of new catalysts for this category of reactions is in continuous demand. It is usually carried out by reacting the substrate with the

desired acyl donor in the presence of a catalyst. Both acidic as well as basic catalysts are equally effective and have used for this purpose. Examples of substances that are neutral in nature and used as acyl transfer agents can also be seen in the literature. An exhaustive list of the various classes of catalysts used in the peracetylation of carbohydrates can be seen from a recently published review.¹ Additionally, for non-carbohydrate alcohols a variety of other catalysts such as Bu_3P ,² CoCl_2 ,³ TaCl_2 ,⁴ TMSOTf ,⁵ iminophosphorane bases (with enol esters)⁶ and distannoxane catalysts, *etc.* have also been used in the presence of excess amounts of acetic anhydride in various solvents.⁷ The relatively strong basicity ($\text{p}K_{\text{a}} = 13.4$)⁸ of the 2,3-dihydro-5,7-dimethyl-1,4-diazepine (**1a**) and the excellent stability of its neutral perchlorate salt **2** (ref. 9) (Fig. 1) attracted our attention in this context and prompted us to explore the suitability of the latter as a possible acylation catalyst with potential benefits over many of the hitherto known acylation catalysts. In particular, it was felt that the mild nature of **2** may offer good possibilities for the selective formation/hydrolysis of isopropylidene acetals besides possible selective acetolysis of one or more benzyl ether protecting groups in sugars carrying multiple benzyl ether protecting groups. Thus, herein we describe the use of **2** as an efficient acyl transfer catalyst for the acetylation of various classes of organic compounds containing $-\text{OH}$, $-\text{SH}$ and NH_2 groups in stoichiometric acetic anhydride (see Scheme 1 to follow).

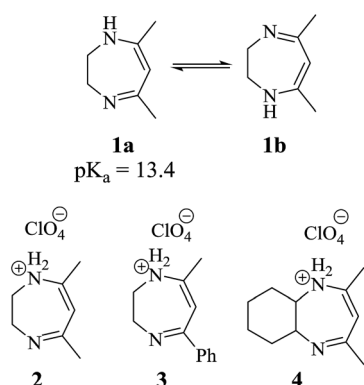


Fig. 1 Diazepinium perchlorate catalysts.

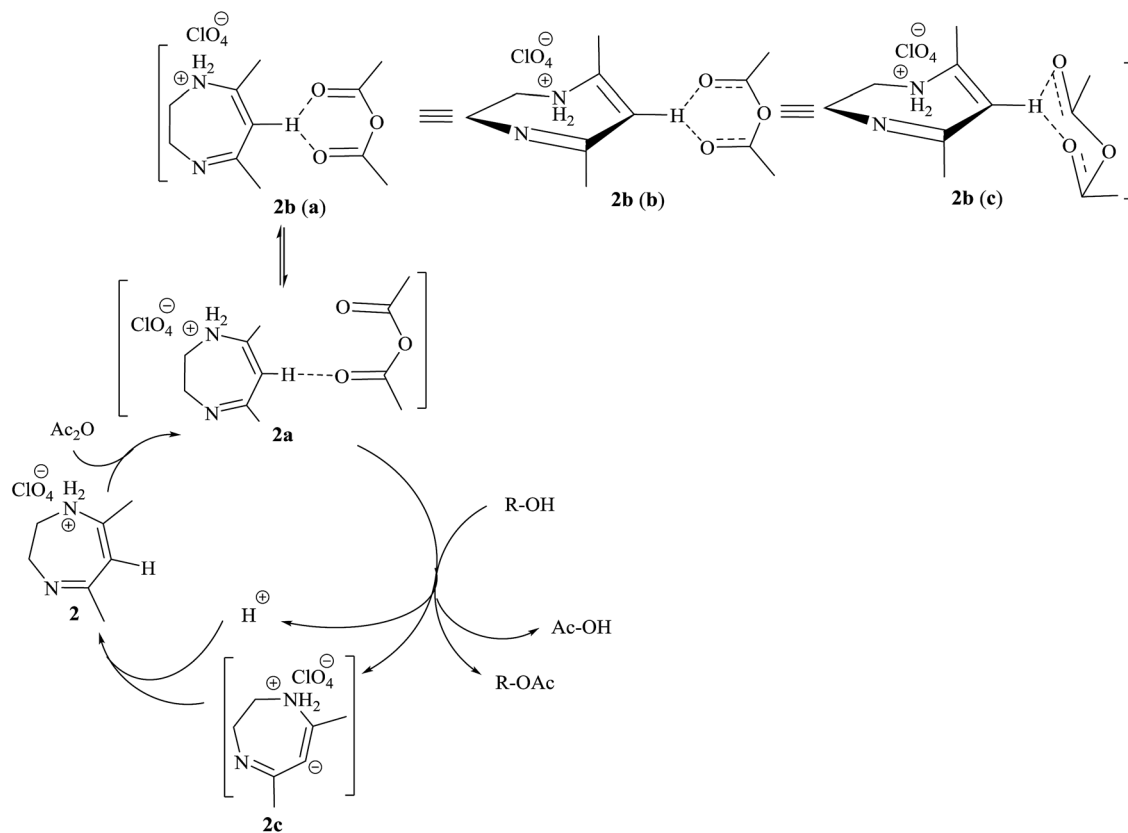
2. Results and discussion

The diazepinium salt **2** (ref. 10) was prepared by the method reported by Lloyd *et al.*⁹ and was used for the acetylation reaction as described below. Considering the more challenging

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Scheme 1 Diazepinium perchlorate-catalyzed acyl transfer reaction using Ac_2O as the acyl donor.

Table 1 Optimization of the reaction condition for the acetylation of D-glucose (5) in presence of 2 as the promoter

Entry	R-OH ^a	Acyl donor ^b	Catalyst (mol%)	Temp. (°C)	Time	R-OAc	Yield ^c (%)
1	D-Glucose (5)	Ac_2O	10	25	24 h	D-Glucose pentaacetate (6)	—
2	5	Ac_2O	20	25	24 h	6	—
3	5	Ac_2O	30	25	24 h	6	40
4	5	Ac_2O	30	35–40	10 min	6	99 ^d
5	5	Ac_2O	25	35–40	20 min	6	99
6	5	AcOH	30	35–40	24 h	—	—
7	<i>n</i> -BuOH	AcOH	30	35–40	30 min	<i>n</i> -BuOAc	75 ^e
8	5	AcCl	30	35–40	24 h	6	20

^a 1 mmol substrate was taken for the reaction. ^b The acyl donor was taken in excess. ^c Isolated yields are reported. ^d Anomeric ratio of $\alpha : \beta = 98 : 2$ was obtained. ^e Yield (conversion) was determined by gas chromatography (GC).

demand of a carbohydrate molecule structurally, as compared to many of the other simple hydroxylic compounds, a common monosaccharide, D-glucose (5), was chosen as the substrate for the initial experiments on acetylation using perchlorate 2 as the promoter. Thus when 5 was treated with acetic anhydride in the presence of 2 (10 and 20 mol% in two separate parallel

experiments; see Table 1, entries 1 and 2), no acetylated product was found to be formed during a reaction time of up to 24 h at 25 °C. Compound 5 remained insoluble. This was not very surprising from the neutral nature of 2. Under the same conditions at 30 mol% of 2, however, 5 reacted to a notable degree to give a 40% yield of the desired 6 (entry 3, Table 1). A



partial dissolution of **5** had been observed in this case. And, when the reaction was performed at a slightly higher temperature of nearly 40 °C (entry 4, Table 1) the sugar started to go into solution in a few minutes and a complete dissolution of **5** was observed in 10 min (entry 4, Table 1). A clear colourless solution had been resulted which had also become hot, typical of an exothermic reaction. Analysis of the reaction mixture by TLC (eluent, EtOAc : *n*-heptane, 2 : 3) showed the presence of only one product (spot), that corresponded to an authentic sample of the expected penta-*O*-acetate **6** (ref. 11) in *R_f* value; and importantly, no partially acetylated glucose derivative was present in the mixture (as evidenced by the absence of any slower moving products on TLC). Compound **6** isolated in near-quantitative yield (2.15 g, 99%) following a simple aqueous workup and crystallization was practically pure and in a form suitable for employing in reactions further. In a subsequent reaction wherein **2** was used at 25 mol% concentration (entry 5, Table 1), the same reaction was found to be complete in about 20 min of reaction time with **6** again being obtained in near-quantitative yield. Acetylation of **5** using Ac₂O as the acyl donor agent in the presence of **2** (25–30 mol%) carried out in common solvents such as MeCN, CH₂Cl₂, etc. for comparative purposes was found to be inferior indeed in achieving the objective (results not shown in the table); and was therefore not pursued further. Thus, solvent-free reaction with a slight excess over the

stoichiometric requirement of Ac₂O in the presence of 25–30 mol% **2** at 35–40 °C was found to be optimum for the complete acetylation of a compound such as **5**.

Further, although a reaction of **5** carried out with AcOH as the acetyl donor agent in the presence of **2** (30 mol%) was proved incapable of serving the purpose (entry 6, Table 1), was found somewhat effective for the acetylation of simple alcohols such as *n*-BuOH (entry 7, Table 1). Thus, under these conditions (entry 7, Table 1) conversion of the alcohol to an extent of 75% (by GC) had been observed. On the other hand, when AcCl was used as the acyl donor in the reaction of **5**, the acetylated product **6** (entry 8, Table 1) was obtained in 20% yield. It may be noted that reaction of polyhydroxylic compounds such as **5** with AcCl (conducted in the absence of any acid scavenger) is usually accompanied by release of stoichiometric quantities of HCl (gas). Taking into account the solubility of hydrogen halides in Ac₂O/AcOH, this opportunity can be utilized in effectively trapping the HCl generated *in situ* and utilizing the same for the conversion of the glycosyl acetate formed in the reaction to the corresponding glycosyl chloride in the same pot.¹³ This possibility in the instance above shall be investigated in the future.

The diazepinium perchlorate (**2**)-catalysed acetylation can be depicted as shown in Scheme 1. When a solution of perchlorate **2** in Ac₂O (0.28 M) was prepared and the mass spectrum (LCMS-LTQ ESI) was recorded after 30 min of standing, peaks

Table 2 Results of DFT study on the complexes formed by Ac₂O with diazepinium perchlorate (**2**) and MeSO₃H, calculated at B3LYP/6-31+G(d) level of theory

Entry	Structure of the complex	Atom	NBO charge
1		H	0.282
		O-1	-0.563
		O-2	-0.555
2		H	0.541
		O-1	-0.611
		O-2	-0.548



corresponding to the following m/z values were obtained: (i) 125.02 of $[2 - \text{ClO}_4]^+$ and (ii) 349.12 of $[2 + \text{Ac}_2\text{O} + \text{Na}]^+$, the latter clearly suggesting the possible 2-Ac₂O adduct in the solution. On the other hand under the same conditions *n*-Bu₄N(ClO₄) gave only one peak, of m/z 242.19 that corresponded to $[\text{Bu}_4\text{N}]^+$ as also CsClO₄ but at an m/z value of 125.01 that corresponded to $[\text{Ac}_2\text{O} + \text{Na}]^+$. LiClO₄, however, did not give any notable mass spectral signals at all under these conditions. It may be emphasized that among the four perchlorates mentioned above (that is, 2, *n*-Bu₄N(ClO₄), CsClO₄, and LiClO₄), LiClO₄ was also insoluble in Ac₂O.¹⁴ Again, in reactions in Ac₂O using these catalysts (25 mol% with respect to the sugar, D-glucose) for acetylation at 35–40 °C for 30 min, only the case of 2 led to the formation of the acetylated product 6. However, in the case of LiClO₄, the reaction employing the catalyst at 50 mol% level for 12 h led to a successful acetylation as described by Huang and coworkers.¹⁵ Further, in the case of acetylation of the monosaccharide 5 promoted by 2, mass spectroscopic analysis of the reaction mixture revealed that the intensity of the peak corresponding to $m/z = 349.12$ ($[2 + \text{Ac}_2\text{O} + \text{Na}]^+$) was significantly weaker towards the completion of the reaction as compared to the initial stages of the reaction.

In view of the above observations made in the course of the mass spectral analysis of the acetylation mixture, a DFT study on the system was also carried out and the results of this have been shown in Tables 2 and 3 (respectively, the energy-minimized 2-Ac₂O complex 2b showing the possible H-bond distances and listing of the energy values observed). The energy-minimized structure clearly shows that the H-6 of the diazepinium moiety is very much within the range of hydrogen bond distances considered reasonable for C–H hydrogens.^{16,17} A value of +0.282 NBO charge for the H-6 as against the values of –0.563 and –0.555 for the acetic anhydride–oxygen atoms is also consistent with H-6 acting as a Lewis acid as depicted in Scheme 1. It was then thought worthwhile to compare the complex 2b with another formed between a conventional acid/Lewis acid catalyst and Ac₂O. Thus a recently reported MeSO₃H–Ac₂O that has been found to be highly efficient in acetylating various hydroxylic substrates was chosen for this purpose.¹⁸ The minimized structure of the complex 2d along with the hydrogen bond distances and the energy of stabilization are also shown in Tables 2 and 3. Thus, while the stronger hydrogen bond in complex 2d along with a higher energy of stabilization (compared to those of 2b) are consistent with this (latter) powerful acetylation system,¹⁸ it also indeed lends

support to the mechanism of Lewis acid-like catalysis as proposed in Scheme 1 for the activation of Ac₂O in the acetylation reaction promoted by 2.

The bond angle α in C–H···O interaction is in between 90° and 180° in complex 2b which are necessary for the H-bonding (entry 1, Table 2) to be facilitated with ease. The data summarized in Table 3 on the calculated energies for 2, Ac₂O and their complex 2b, for which the net energy of stabilization was found to be lower than the value for MeSO₃H–Ac₂O complex (2d) as expected, also is in agreement with the activation of Ac₂O proposed for acetylation in Scheme 1.

Arguably, analogous to the situation in metal triflate-catalyzed acetylations where an *in situ*-formed TFOH could be attributed to partially account for the catalysis of the acylation reaction,¹⁹ an *in situ*-formation of HClO₄ in 2-Ac₂O system can also be considered reasonable and to similarly account for the observed catalysis. In view of this possibility, we tried to examine the acidity of In(OTf)₃–Ac₂O system,¹² a typical metal triflate–Ac₂O system, as well as 2-Ac₂O system using a standard pH meter. Thus, while a “pH” value of 2.73–3.2 was obtained for neat Ac₂O, the corresponding reading obtained for a solution of In(OTf)₃ (catalytic) in Ac₂O was well below zero (and outside the limits of the pH meter). However, the corresponding reading for the 2-Ac₂O system was 3.44. Similarly, for the 2-Ac₂O–glucose system (after completion of the acetylation of the sugar) the “pH” reading obtained was 2.71. Thus, in contrast to Ac₂O turning highly acidic upon dissolution of a catalytic amount of the metal triflate in it, the dissolution of 2 only rendered the resultant solution turn less acidic. This is in accord with the reaction sequence outlined in Scheme 1. In order to verify the possibility for any base-catalyzed activation of the alcohol residue by the perchlorate anion, work is underway for synthesizing diazepinium salts with other counter anions such as those of triflic acid, hexafluorophosphoric acid, tetrafluoroboric acid, *etc.* This will also enable us to conclude on the possibilities for finding low melting diazepinium salts that we hope could also possibly serve as ionic liquids.

In the acetylation of D-glucose described above, it may also be pointed out that the by-product (AcOH) of the reaction served as an excellent solvent for the per-*O*-acetylated product but could also be removed easily in the aqueous alkali workup involved in the acetylation. Needless to mention, in small scale reactions it can also be conveniently removed by co-distillation with toluene under a reduced pressure.

Table 3 Energies for the complexes formed by Ac₂O with diazepinium perchlorate (2) and MeSO₃H, respectively calculated at B3LYP/6-31+G(d) level of theory^a

Entry	Catalyst/complexing agent	Calculated absolute energy (kcal mol ^{–1})			
		E_1	E_2	E_3	$E_3 - E_2$
1	Diazepinium perchlorate (2)	–718239.306	–957727.049	–957731.374	–4.325
2	MeSO ₃ H	–416830.609	–656318.352	–656327.777	–9.424
3	Ac ₂ O	–239487.742	—	—	—

^a E_1 = absolute energy of substrate; E_2 = sum of (catalyst + Ac₂O) energy; E_3 = energy of complex with catalyst and Ac₂O; $E_3 - E_2$ = energy difference.



Table 4 Diazepinium perchlorate-catalyzed peracetylation of sugars, alcohols, phenols, thiols, thiophenols and amines^a

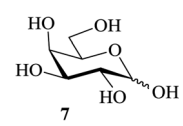
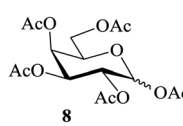
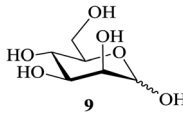
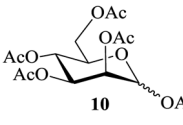
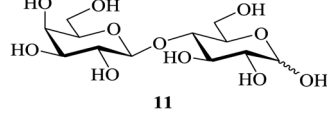
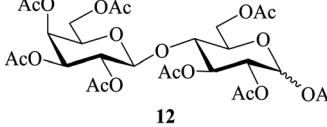
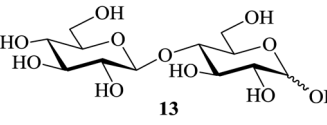
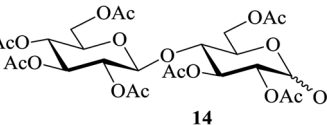
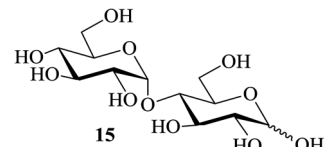
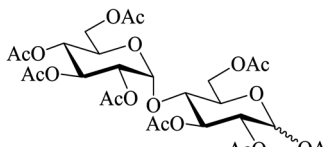
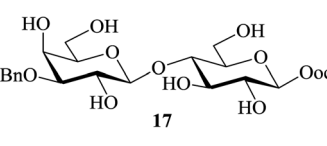
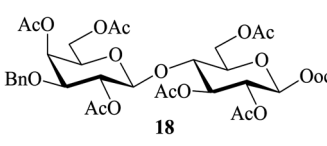
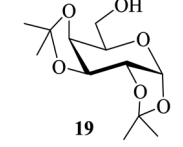
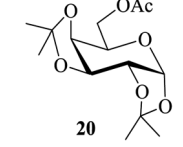
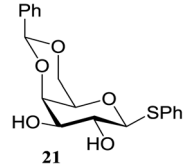
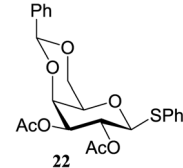
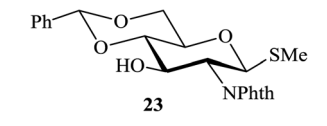
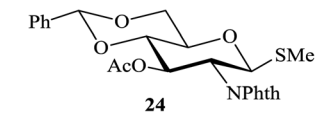
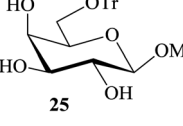
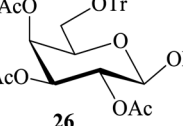
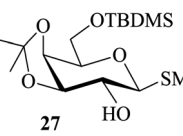
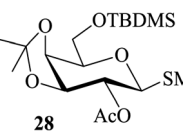
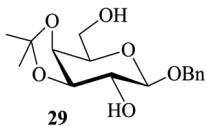
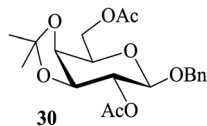
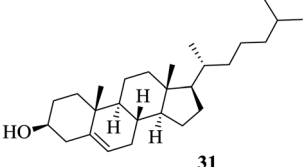
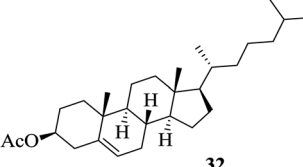
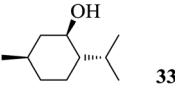
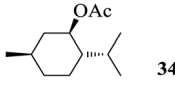
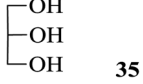
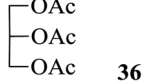
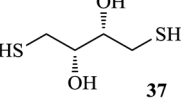
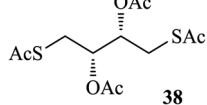
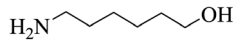
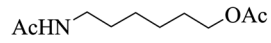
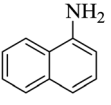
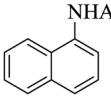
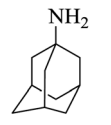
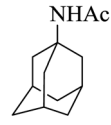
Entry	Substrate	Product	Reaction time (min)	% yield ^b (α : β)	Ref.
1			30	98 (2 : 3)	11
2			20	96 (3 : 1)	11
3			45	96 (2 : 1)	11
4			180	99 (1 : 2)	20
5			45	96 (1 : 5)	21
6 ^c			24 h	95	—
7			8 h	98 ^{d,e}	18
8			60	97	22
9			15	95	—
10			120	96	23
11			5 h	96 ^d	24



Table 4 (Contd.)

Entry	Substrate	Product	Reaction time (min)	% yield ^b (α : β)	Ref.
12			12 h	98 ^d	25
13			120	96 ^d	26
14			30	99	26
15			30	97	27
16			30	98	18
17	39 R ¹ = CH ₂ OH, R ² = H	40 R ¹ = CH ₂ OAc, R ² = H	30	99	26
18	41 R ¹ = OH, R ² = H	42 R ¹ = OAc, R ² = H	3 h	96	28
19	43 R ¹ = OH, R ² = CH ₃	44 R ¹ = OAc, R ² = CH ₃	2.5 h	95	29a
20	45 R ¹ = OH, R ² = NO ₂	46 R ¹ = OAc, R ² = NO ₂	24 h	No reaction	—
21	47 R ¹ = SH, R ² = CH ₃	48 R ¹ = SAc, R ² = CH ₃	15	96	29a
22	49 R ¹ = SH, R ² = OH	50 R ¹ = SAc, R ² = OAc	2 h	96	29a
23	41 + 45	42 + unchanged 45	3 h	95	28
24	51 R ¹ = OH, R ² = NH ₂	52 R ¹ = OAc, R ² = NHAc	1 h	99 ^d	30
25			10	96 ^d	31
26			15	97	30
27			60	95	32

^a The substrate was treated with Ac₂O (1.2 equiv. per OH/SH group except for entry 7, 11, 12, 13) in the presence of diazepinium perchlorate (2, 25 mol%) under neat reaction conditions at room temperature (40 °C). ^b Isolated yield has been reported. ^c 3.0 equiv. of 2 was used at room temperature (40 °C). ^d 2.5 equiv. of Ac₂O was used at room temperature (40 °C). ^e 50 mol% of 2 needed for the reaction.

When the reaction under identical conditions was carried out using D-galactose (7, entry 1, Table 4) as the starting material, the corresponding pentaacetate 8 was also obtained in 98%

yield. Likewise another common reducing sugar (D-mannose, 9) was also converted at ease to its per-O-acetylated derivative 10 in 96% yield (entry 2, Table 4). In the same manner, disaccharides



[lactose (11), cellobiose (13) and maltose (15)] could also be converted to their corresponding octaacetates (12, 14 and 16, respectively) with equal efficiency (entries 3–5, Table 4).

In view of the above successful results obtained with 2, its analogues 3 and 4 were also investigated for its possible application in the above class of acylation reactions using 5 as the substrate. It was found that under the reaction conditions described above, the perchlorates 3 and 4 were in fact ineffective as a promoter for the acetylation of 5. These salts were therefore not investigated further.

In most of the acid-catalyzed acetylation reactions of sugars difficulties arise when acetylation of their derivatives bearing acid-labile substituents such as acetals and trityl/TBDMS/TBDPS ethers are to be carried out. One of the approaches towards overcoming such problems has been to perform the reaction under milder and controlled conditions (for example, at lower temperatures, reduced catalyst-concentration, *etc.*). Therefore care-free methods requiring no such special conditions for the success of this reaction would be greatly beneficial. It was in this context that the following sets of partially protected sugar derivatives were examined as substrates for acetylation in the presence of 2. Thus, the partially benzylated lactoside 17, the isopropylidene acetal of galactopyranose 19, the hexopyranosides-derived benzylidene acetals 21 and 23, the trityl ether 25, the silyl ether 27 and the benzyl glycoside 29 also gave their respective acetylated derivatives 18, 20, 22, 24, 26, 28 and 30 respectively in excellent (near-quantitative) yields in reactions carried out at 35–40 °C (entries 6–12, Table 4). The NMR spectra of these compounds obtained as crude products after the aqueous workup of the reaction mixture (please see the ESI†) stand as testimony to their excellent purity.

In order to reconfirm the stability of the acid-sensitive protecting groups on the substrates in the acetylating solution, the reaction mixture from a parallel reaction carried out on the acetal-protected 29 was left for four weeks after the formation of 30 was complete.

The product isolated subsequently by aqueous work up was indeed found to be the unchanged 30. The importance of this observation lies in the fact that all these compounds are extremely vulnerable to acetolytic conditions (Ac₂O-acidic catalyst systems) usually employed for the reaction. Further, alcohols such as β-cholesterol (31), (–)-menthol (33), glycerol (35), dithiothreitol (37) and benzyl alcohol (39) also gave their acetylated derivatives 32, 34, 36, 38 and 40 (entries 13–17, Table 4), respectively, in excellent yields on treatment with Ac₂O in the presence of 2. Likewise, examples of phenols, thiophenols and their derivatives were also tested as substrates for acetylation. It was found that phenol (41), 4-methylphenol (43), 4-methylthiophenol (47) and 4-hydroxythiophenol (49) gave their respective acetylated derivatives 42, 44, 48 and 50 (entries 18, 19, 21 and 22, Table 4) in excellent yields. Thus, no chemoselectivity towards acetylation under the above condition was observed in the case of the phenolic hydroxyl and sulfhydryl groups. On the other hand, 4-nitrophenol (45) in the presence of 2 in Ac₂O did not give the acetylated product 46 even after 24 h of treatment under conditions identical for phenol (entry 20, Table 4). As theoretical data of 4-methylphenol (43) and 4-nitrophenol (45)

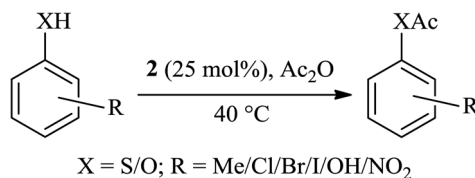
like pK_a and σ (Hammett constants)^{29b} are 10.174, –0.17 and 7.161, 0.78 indicated that latter case is more acidic and rather electrophilic (entries 19 and 20, Table 4). Thus, the deactivating effect of the *para*-substituted electron withdrawing nitro group was clearly evident. Moreover, the reported values^{29b} for the pK_a and the σ (Hammett constants) for these compounds are also supportive of this observation. As a result thus, when a mixture of 41 and 45 was subjected to treatment with Ac₂O in the presence of 2, only the former underwent the reaction enabling the isolation of 42 by aqueous workup in excellent yield along with the near quantitative recovery of 45 (entry 23, Table 4). Therefore this could serve as a simple protocol for the effective separation of such compounds in a mixture. In continuation of this study, a set of typical examples of amino compounds were also subjected to acetylation under the conditions noted above. Thus, aminophenols/alcohols such as 51 and 53 as well as amines such as 55 and 57 when treated with Ac₂O in the presence of 2 also underwent the reaction smoothly to give the corresponding acetylated derivatives 52, 54, 56 and 58, respectively (Table 4, entries 24–27) in excellent yields. Whereas in case of 51 and 53 (Table 4, entries 24–25), no chemoselectivity in the *N*-/*O*-acetylation was observed.

2.1 Rationalization of the reactivity differences observed in the phenolic/thiophenolic compounds

In the acylation reactions such as those described above, the reactivity differences observed in the case of substituted phenols/thiophenols, *etc.* are generally justified (and as described herein) by the activating/deactivating effect of the substituent groups present on the aromatic ring. However, no quantitative estimates in respect of the above, in terms of the calculated nucleophilicity of phenol/thiophenol substrates have been available. In view of this an attempt was made to perform a computational study by using Gaussian09 package employing a density functional B3LYP/6-31+G(d) method to calculate the nucleophilicity parameters as local nucleophilicity index (N_k^-), global nucleophilicity index (N), and Fukui function for electrophilic attack (f_k^-). The Fukui function allows to predict the most electrophilic and nucleophilic sites in a molecule which can be further used to determine the reactivity of the molecules under study. When 39, 41, 43, 47, 49, 59, 61, 63, 67, 69, 71 and 73 were treated with acetic anhydride in the presence of 2 at 40 °C yielded respectively 40, 42, 44, 48, 50, 60, 62, 64, 68, 70, 72 and 74 in excellent yields (Table 5, entries 1–16). On the other hand, the fluoro- (65) and the phenol, thiophenol and the substituted phenols/thiols used in this study have been listed in Table 5 in the decreasing order of their nucleophilicity, based on the calculated local nucleophilicity index, N_k^- (see in ESI†). It can be seen from the table that for the substrates used in this study, the nucleophilicity index in general followed the order, thiophenols > substituted thiophenols > phenols > substituted phenols, with the exception of *p*-nitrothiophenol occupying a place among that of the substituted phenols, not surprisingly though considering the strong electron withdrawing (inductively as well as, significantly more strongly, by resonance) nature of this substituent group. The experimental results



Table 5 Calculated nucleophilicity of phenols/thiophenols and experimental support for the order of nucleophilicity of phenols/thiophenols and their derivatives in the acetylation catalyzed by **2**^{a,b}



Entry	Phenol/thiophenol		Calculated values		Experimental observations/results	
	Substrate	Product	N_k^-	f_k^-	Reaction time (h)	Yield ^c (%)
1	Ph-SH (59)	Ph-SAc (60)	-0.35	-0.36	0.5	99
2	<i>p</i> -Me-Ph-SH (47)	<i>p</i> -Me-Ph-SAc (48)	-0.32	-0.32	0.25	96
3	<i>o</i> -Br-Ph-SH (61)	<i>o</i> -Br-Ph-SAc (62)	-0.29	-0.35	1.5	99
4	<i>p</i> -Cl-Ph-SH (63)	<i>p</i> -Cl-Ph-SAc (64)	-0.25	-0.31	1.5	99
5	<i>p</i> -HO-Ph-SH (49)	<i>p</i> -AcO-Ph-SAc (50)	-0.17	-0.20	2	96
6	<i>p</i> -Me-Ph-OH (43)	<i>p</i> -Me-Ph-OAc (44)	-0.16	-0.15	2.5	95
7	Ph-OH (41)	Ph-OAc (42)	-0.16	-0.16	3	96
8	<i>p</i> -F-Ph-OH (65)	<i>p</i> -F-Ph-OAc (66)	-0.14	-0.16	24	Traces ^d
9	<i>o</i> -Cl-Ph-OH (67)	<i>o</i> -Cl-Ph-OAc (68)	-0.14	-0.16	3.5	98
10	<i>p</i> -O ₂ N-Ph-SH (69)	<i>p</i> -O ₂ N-Ph-SAc (70)	-0.12	-0.35	4	98
11	<i>p</i> -Cl-Ph-OH (71)	<i>p</i> -Cl-Ph-OAc (72)	-0.12	-0.14	3.5	98
12	<i>p</i> -Br-Ph-OH (73)	<i>p</i> -Br-Ph-OAc (74)	-0.11	-0.13	4	90
13	Bn-OH (39)	Bn-OAc (40)	-0.10	-0.12	0.5	99
14	<i>m</i> -O ₂ N-Ph-OH (75)	<i>m</i> -O ₂ N-Ph-OAc (76)	-0.06	-0.17	24	73
15	<i>p</i> -O ₂ N-Ph-OH (45)	<i>p</i> -O ₂ N-Ph-OAc (46)	-0.06	-0.16	24	No reaction
16	<i>o</i> -O ₂ N-Ph-OH (77)	<i>o</i> -O ₂ N-Ph-OAc (78)	-0.05	-0.18	24	No reaction

^a N_k^- = local nucleophilicity, f_k^- = Fukui function. ^b All reactions were carried out using 1 mmol substrate in 1 mL of acetic anhydride in the presence of 25 mol% **2**. ^c Isolated yields are reported. ^d Not isolated.

obtained for these compounds are also in general in accordance with the above, as can be seen from the reaction time noted in Table 5 for the respective compounds.

However, the experimental results obtained in the case of *p*-fluorophenol (entry 8, Table 5) showing it to be virtually unreactive under the conditions, has been an exception in this case. The reactivity differences between 4-fluorophenol (**65**) and 2-chlorophenol (**67**) have also been noted previously by others.^{46a} This certainly points to the important additional factors, such as the intermolecular hydrogen bonding, in operation in the case of this halogenophenol. Likewise, although *p*-thiocresol (**47**) was proved experimentally to be completing the reaction faster as expected from the activating effect of the *p*-Me substituent in it, was shown to be marginally lower in the order of nucleophilicity index (entries 1 and 2, Table 5). Similarly, *p*-cresol (**43**) though was more reactive (reacting completely in 2.5 h) than phenol (**41**, that required 3 h in comparison), had identical nucleophilicity indices (entries 6 and 7, Table 5). In the case of the three (*o*-, *m*- and *p*-) nitrophenols (entries 14–16, Table 5), the *m*-nitrophenol (**75**) was proved more reactive than the other two (**77** and **45**) as expected. The strong electron withdrawing nature of the nitro group is still evident (in spite of the absence of any resonance effect) from the relatively low yielding reaction observed in it (entry 14 *versus* 15 and 16, as well as other entries, Table 5).

Additional information emerged from the experiments carried out on various sugars and their partially protected analogues was that, in general, the partially protected sugars required longer time for the completion of the reaction. Although it is consistent with the exothermic nature of the reaction, it also served to suggest that selective acetylation leading to partially protected sugars could perhaps be achievable in the case of partially protected sugar derivatives with more than one hydroxyl functionality, perhaps placed strategically, on them.^{33,34}

Thus, the 6-*O*-silylated mannosides **79** and **81** were employed as substrates for acetylation with Ac₂O in the presence of **2** (entries 1 and 2, Table 6). Indeed, the respective di-*O*-acetylated products **80** and **82**, with the OH-4 remaining free, were obtained as the sole respective products in excellent yields (entries 1 and 2, Table 6). The partially protected compounds **80** and **82** are extremely valuable glycosyl acceptor substrates in oligosaccharide synthesis and their preparation from their respective precursor-glycosides would have involved significantly more steps. In contrast, the outcome of the acetylation of the *p*-nitrophenyl galactoside **83** carried out subsequently was comparatively less dramatic in its effectiveness (entry 3, Table 6) in that the selectively protected **84** could be obtained in only 60% yield, the remaining being the tetra-*O*-acetate **85** although no difficulty in the separation of the two (**84** and **85**) by column chromatography on silica gel was experienced. Thus, under the



Table 6 Diazepinium perchlorate-catalyzed regioselective acetylation^a

Entry	Substrate	Catalyst (mol%)	Time (h)	Ratio of products ^b	Yield ^c (%)
1		30	2		96
2		30	4		95
3		100	3		95
4		100	3		96
5		100	3		92 ^d
6		100	4		90
7		100	4		96
8		50	5		97
9		30	4		96

^a The substrate was treated with Ac₂O (2 equiv.) in the presence of diazepinium perchlorate 2 at room temperature (35–40 °C). ^b On the basis of isolated yield and ¹H NMR, ratios of the products were reported. ^c Isolated yield of the product was reported. ^d 90 and 91 are inseparable in the ratio of 1 : 0.6 and 92 and 93 are inseparable in the ratio of 1 : 0.7 where overall ratio of 90 : 91 : 92 : 93 = 1 : 0.6 : 2 : 1.4.

conditions of acetylation studied the HO-2 of the galactoside 83 was found less reactive than the other secondary hydroxyls present in the molecule. Among the secondary hydroxyl groups

of galactopyranosides such as 83 the HO-3 is known for its relative high reactivity compared to HO-2 and HO-4. However, the lower reactivity of HO-2 compared to that of HO-4 observed

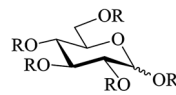
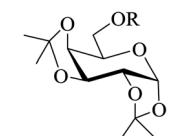


in the above case was somewhat surprising and could have been due to the deactivating nature of the *p*-nitrophenyl group present on O-1 of the sugar moiety leading to a possibly reduced, despite perhaps only marginally, nucleophilicity of its O-2. In an attempt to rationalize this observation further the acetylation of *m*-nitrophenyl β -D-galactoside **86** was carried out under the same conditions as for the analogous galactoside **83**. Here again, the selectively 2,3,6-tri-*O*-acetylated product **87** was obtained along with the tetra-*O*-acetylated galactoside derivative **88**. However, compounds **87** and **88** were obtained in 80% and 20% yields (entry 4, Table 6), respectively, after separation by chromatography on a column of silica gel. Thus, although the *m*-nitrophenyl group at the anomeric centre of **86** is expected to influence the nucleophilicity of its O-4 to a lesser extent, as compared to the *p*-nitrophenyl substituent present in **83**, a significantly enhanced yield of **87** (bearing a free hydroxyl on C-4) was indeed obtained in the reaction of **86**.

It therefore appeared that the influence of the *m*-nitrophenyl group present at the anomeric centre on **86** may have been more steric (than electronic) in nature. It may be noted that as compounds **87** and **88** had significantly different R_f values their separation was achieved efficiently and therefore the extremely valuable partially protected galactoside **87** could be obtained in pure form without any practical difficulty. Its traditional synthesis would have indeed been multi-step in nature. At this point a study on the acetylation of the *o*-nitrophenyl β -D-galactoside **89** was thought beneficial.

Thus, when **89** was treated with Ac_2O in the presence of **2** and the product emerged was analyzed by TLC, it was revealed that the outcome was of a more complex nature. A set of three spots with R_f values 0.45 (A), 0.43 (B) and 0.23 (C) was obtained on TLC (eluent, EtOAc : *n*-heptane, 2 : 1) of the product. Attempts to separate them by chromatography on a column of silica, resulted in the elution of the first two (A and B) closely-moving components as a mixture. Analysis of this mixture by NMR spectroscopy, however, enabled the identification of these products as the 2,4,6-tri-*O*-acetylated compound **90** and the 3,6-di-*O*-acetylated compound **91** (A and B, respectively) in the ratio 1 : 0.6 (entry 5, Table 6). Following the above, component C was eluted and was identified by NMR spectroscopy as a mixture of two regioisomeric diacetates, namely, the 4,6-di-*O*-acetylated compound **92** and the 2,6-di-*O*-acetylated compound **93** in a ratio of 1 : 0.7. The proportion in which these eluates (A + B and C) were present in the mixture was 1 : 2 [(**90** + **91**) : (**92** + **93**)], thus the overall ratio of **90** : **91** : **92** : **93** formed being 1 : 0.6 : 2 : 1.4. A surprising observation on the outcome of the foregoing reaction was the unusually low proportions of the product-fraction **91** bearing a 3-*O*-acetyl moiety, in spite of the known relatively high reactivity of the HO-3 of unprotected D-galactopyranosides. Moreover, compound **91** was the sole 3-*O*-acetyl group-containing product formed in the acetylation of **89**. In contrast, the 2-*O*-acetyl group-containing products formed in the acetylation of **89** were significantly more, indicating a relatively more facile acetylation at HO-2. This was suggestive of

Table 7 Comparative study of acetylation using different reported catalysts

Substrate Product	Catalyst						
	Name	Mol%	Ac_2O (equiv.)	Solvent	Time (min)	Reported yield (%)	Ratio (α/β)
 5 R = H 6 R = Ac	$\text{In}(\text{OTf})_3$ (ref. 12)	0.16	8.8	None	3	100	14/1
	$\text{Ce}(\text{OTf})_3$ (ref. 35)	1	7.5	Ac_2O	150	>98	—
	$\text{Sc}(\text{OTf})_3$ (ref. 36)	0.5	6	None	90	96	—
	CoCl_2 (ref. 3)	5	10	CH_3CN	6 h	75 ^a	—
	InCl_3 (ref. 37)	0.1	7.5	CH_3CN	1.5	94 ^b	4/1
	I_2 (ref. 11)	3.5	9.5	None	30	90	25/1
	$[\text{bmim}][\text{HSO}_4]^{38}$	100	5.5	Ac_2O	No reaction	—	—
	$[\text{bmim}][\text{OAc}]^{38}$	100	5.5	Ac_2O	15	90	—
	LiClO_4 (ref. 15a)	50	5.8	None	12 h	94 ^c	—
	CuClO_4 (ref. 39)	1	5.05	None	30	97	1/0
	Montmorillonite- K^{40}	1 g	13.8	None	6 h	94	—
	Imidazole ⁴¹	60	6	CH_3CN	5 h	98	4/1
	Pyridine ^{42a}	Excess	7.5	CH_3CN	18 h	33	3.7/1
	—do— ^{42b}	Excess	Excess	Ac_2O	18 h	95	—
 19 R = H 20 R = Ac	Diazepinium perchlorate (2)	25	6.6	None	30	99 ^c	49/1
	I_2 (ref. 11)	2.5	13	None	5	98 ^d	—
	LiClO_4 (ref. 15b)	10	1.1	None	48 h	96 ^c	—
	Al_2O_3 (ref. 43)	200	1.5	None	12 h	30 ^e	—
	InCl_3 (ref. 37)	0.1	7.5	CH_3CN	2	35 ^{b,f}	—
	Pyridine ⁴²	Excess	2.5	Pyridine	45	95	—
	Imidazole ⁴¹	60	1.2	CH_3CN	2 h	98	—
	DABCO ⁴⁴	100	1.2	None	20	98	—
	Diazepinium perchlorate (2)	50	2.5	None	8 h	98 ^c	—

^a The reaction was performed in CH_3CN at 80 °C. ^b The reaction was performed under microwave irradiation. ^c The reaction was performed at 40 °C.

^d The reaction was carried out at ice-bath temperature. ^e Reaction did not go to completion. ^f Degraded product was obtained.



a possible steric interaction (1,3-*cis*-) exerted by the anomeric *O*-nitrophenyl substituent in **89** on its HO-3.

Moreover, in the case of the 4,6-*O*-benzylidene-protected diol **94** the mono- and di-*O*-acetates **95** and **96**, obtained in 2 : 1 ratio, were the only products of acetylation, with no selectively 2-*O*-acetylated product being obtained in the reaction. Our attempts to gather theoretical support for the observations noted in the regioselective acetylations of galactosides **83**, **86**, **89** and **90** described above, by subjecting these structures to studies by Gaussian09 package unfortunately were turned out inconclusive (and therefore are not discussed here).

p-Acetyl-*o*-methoxyphenyl β -D-glucoside **97** when treated with acetic anhydride in the presence of **2** the selectively 2,3,6-tri-*O*-acetylated product **98** was obtained in 80% yield along with the tetra-*O*-acetylated glucoside derivative **99** obtained in 20% yield (entry 7, Table 6) in consistence with the known order of reactivity of a glucopyranoside-hydroxyl groups. Likewise, in the case of the reaction of methyl 6-*O*-trityl α -D-glucoside **100** with Ac₂O in the presence of **2** also, the selectively 2,3-di-*O*-acetylated product **101** was indeed the major partially acetylated product formed (65% yield along with tri-*O*-acetylated product **102** in 32% yield, entry 8, Table 6). Moreover, preference for the acetylation of a primary hydroxyl group over a secondary was also evident from the results obtained from the reaction of the partially protected glycerol derivative **103** yielding selectively mono-*O*-acetylated product **104** as the major product (along with di-*O*-acetate **105** in a ratio of 9 : 1 in 96% combined yield, entry 9, Table 6).

Further, a comparison of different reported catalysts with the diazepinium salt **2** for acetylation reaction of **5** and **19** has been illustrated in Table 7. Thus, it becomes clear that while the current method is comparable to the best literature methods in terms of the reaction yield, it is solvent-free and requires only close-to-stoichiometric quantities of the acyl donor agent (like some of the known entries in the table) and, unlike many of the others, is greener (organic, non-corrosive, non-hazardous and not sensitive to air/moisture).

3. Conclusions

Diazepinium perchlorate (**2**) has been found to be an attractive alternative neutral catalyst for acyl transfer reactions of mono- and disaccharides and their various partially protected derivatives as well as various alcohols, phenols, thiols, thiophenols and amines using acetic anhydride as the acyl donor agent. Regioselective acetylations of various partially protected monosaccharides were also successful and were proved effective time-saving alternatives to their conventional multi-step synthesis. The results obtained on the acetylation of phenols and thiophenols were rationalized through their local nucleophilicity index from DFT calculations.

4. Experimental

4.1 General

All reagents (Aldrich India Ltd) used were of analytical grade and were used as purchased without further purification.

Reactions were monitored by TLC, which was performed using 0.2 mm pre-coated silica gel 60 F₂₅₄ aluminium sheets. Compounds were detected by dipping the TLC plates in an ethanolic solution of sulphuric acid (5%, v/v) and heating or using a reagent prepared by dissolving a mixture of ammonium molybdate (4.2 g), cerium sulfate tetrahydrate (0.2 g) and sulphuric acid (6.2 mL) in water (94 mL). Column chromatography was performed using silica gel (100–200 mesh, ASTM). ¹H/¹³C NMR spectra were recorded in CDCl₃ at 400/100 MHz on a Bruker Avance^{III} spectrometer. The chemical shifts have been reported in ppm using TMS as the internal standard. The multiplicity of the signals have been shown as, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet) with the coupling constant (*J* value) expressed in Hz. Dept 135 and 2D NMR like COSY, HSQC and HMBC were also used for characterizing synthesized molecules. The anomeric ratios of products reported in Table 1 were determined from their NMR spectra. Mass spectra were obtained on Ultraflex TOF/TOF MALDI or HRMS equipment. Specific rotations were determined on an AUTOPOL IV polarimeter at 20–24 °C. IR spectra were recorded on a Nicolet FT-IR Impact 410 instrument either as neat or as KBr pellets. Melting points were determined on a melting point apparatus.

4.2 Typical procedure for the preparation of **2**

A mixture of acetylacetone (10 mL, 97.8 mmol) and ethylenediamine (10 g, 166.3 mmol) was heated at 120 °C for 2 min. Acetic acid (20 mL) was then added to it and the reaction mixture was stirred for 10 min at 120 °C. It was then brought to room temperature, and after adding water (200 mL) followed by aqueous HClO₄ (20%, 60 mL) and stirring, the mixture was allowed to stand overnight to form crystals. The crystals were then separated by filtration through a fritted glass funnel and were washed with cold water followed by *n*-hexane/*n*-heptane on the funnel. They were then dried under reduced pressure (48 h) to yield **2** as a colourless crystalline product (36 g, 96%). The product was characterized by comparison of its melting point and spectral data (NMR and MS) with literature values which were in agreement with each other. Colourless needle shaped crystal; mp 140–141 °C; ¹H NMR (CD₃OD): δ (ppm) 5.03 (s, 1H, CH), 4.88 (bs, 4H, 2 \times CH₂), 3.65 (bs, 2H, NH₂), 2.22 (s, 6H, 2 \times CH₃); ¹³C NMR: (CD₃OD): δ (ppm) 167.3, 90.9 (CH), 47.9 (CH₂), 47.5 (CH₂), 22.4 (2 \times CH₃). IR: (KBr) ν_{\max} 3335, 2477, 1620, 1552, 1328, 1100, 819, 703, 619 cm⁻¹; MALDI-TOF/TOF: *m/z* calcd for C₇H₁₃NaClN₂O₄ 247.04. Found: 247.69 [M + Na]⁺.

4.3 General procedure for the acetylation reaction in the presence of **2**

The substance to be acetylated was added portionwise to a mixture of **2** (25–30 mol%) and acetic anhydride (1.2–2 mol equiv./OH/SH/NH₂ group) and the mixture was stirred at 35–40 °C. On completion of the reaction (as confirmed by TLC), the mixture was directly poured onto crushed ice, neutralized with NaHCO₃ and was extracted with EtOAc in a separating funnel. The organic extract was then washed with brine and was dried over anhydrous Na₂SO₄. After separation by filtration it was



concentrated under reduced pressure and vacuum-dried to yield the respective peracetylated product in near-quantitative yields. The crude product thus obtained was practically pure (see the NMR spectra) and were suitable for further use directly. Analytically pure samples were obtained by either recrystallization or by chromatography on a column of silica. Compounds **6**,¹¹ **8**,¹¹ **10**,¹¹ **12**,¹¹ **14**,²⁰ **16**,²¹ **20**,¹⁸ **22**,²² **26**,²³ **28**,²⁴ **30**,²⁵ **32**,²⁶ **34**,²⁶ **36**,²⁷ **38**,¹⁸ **40**,²⁶ **42**,²⁸ **44**,^{29a} **46**,^{29a} **48**,^{29a} **50**,^{29a} **52**,³⁰ **54**,³¹ **56**,³⁰ **58**,³² **60**,²⁸ **62**,⁴⁵ **64**,²⁸ **66**,²⁸ **68**,⁴⁶ **70**,¹⁸ **72**,²⁸ **74**,²⁹ **76**,¹⁸ **78**,²⁸ **85**,⁴⁷ **88**,⁴⁷ **96**,⁴⁸ **99**,⁴⁹ **101**,⁵⁰ **102** (ref. 18) and **105** (ref. 51) (Tables 1–6) have been reported in the literature. Compounds **18**, **24**, **80**, **82**, **84**, **87**, **90**, **91**, **92**, **93**, **95**, **98** and **104** are new and their characterization data have been described below. For the NMR spectra of these latter compounds, please see the ESI† provided.

4.4 Typical procedure for a practical-scale preparation of **6**

D-Glucose (20 g, 111 mmol) was added portionwise (over a period of 15 min) to a solution of **2**, (6 g, 26.7 mmol) in acetic anhydride (80 mL) during stirring at room temperature (35–40 °C) for 15 min by which time TLC (EtOAc : *n*-heptane, 2 : 3, v/v) showed complete consumption of the sugar. The solution had become hot due to the exothermic nature of the reaction. It was then poured, while being stirred, onto crushed ice in a beaker containing sodium bicarbonate and was left in a refrigerator for the solids to separate. The solids were then collected over a glass filter by filtration at the pump and were washed with cold water followed by cold *n*-hexane/*n*-heptane. Drying under reduced pressure yielded the desired product, 1,2,3,4,6-penta-*O*-acetyl-D-glucopyranose (**6**, 42.8 g, 99%) as a colourless solid homogeneous in all respects with an authentic sample of **6** and as confirmed by physical constants and spectral data.

4.5 Typical procedure for the regioselective acetylation of the triol **79**: preparation of **80**

Octyl 6-*O*-*tert*-butyldimethylsilyl- α -D-mannopyranoside (**79**, 1 g, 2.31 mmol) was added to a solution of diazepinium perchlorate (**2**, 165 mg, 30 mol%) in acetic anhydride (5 mL) and was stirred for 2 h at 35–40 °C when TLC (EtOAc : *n*-heptane, 2 : 3, v/v) showed complete consumption of the triol **79**. The reaction mixture was diluted with EtOAc (20 mL) and was washed successively with cold aqueous NaHCO₃ solution and water (to neutrality) and dried (Na₂SO₄). Removal of the solvent under reduced pressure and purification of the residue on a short column of silica (eluent, EtOAc : Hex, 30 : 70) afforded octyl 2,3-di-*O*-acetyl-6-*O*-*tert*-butyldimethylsilyl α -D-mannopyranoside (**80**, 1.16 g, 96%) as a colourless liquid as was confirmed by spectral analysis (see below).

4.6 Characterization of compounds

4.6.1 Octyl 2,3,6,2',4',6'-hexa-*O*-acetyl-3'-*O*-benzyl- β -D-lactoside (18**).** Colourless viscous liquid (0.14 g, 95%), [α]_D²⁰ +15.8° (*c* = 1, CH₂Cl₂); ¹H NMR (CDCl₃): δ (ppm) 7.34–7.30 (m, 3H, ArH), 7.25–7.23 (m, 2H, ArH), 5.48 (d, *J* = 2.6 Hz, 1H, H-4'), 5.19 (t, 1H, *J* = 9.3 Hz, H-3), 5.04 (dd, *J*₁ = 8.0 Hz, *J*₂ = 9.9 Hz, 1H, H-2), 4.88 (dd, 1H, *J*₁ = 8.0 Hz, *J*₂ = 9.9 Hz, 1H, H-2'), 4.70 (d, *J* = 12.1 Hz, 1H, CH of PhCH), 4.46 (d, *J* = 7.9 Hz, 1H, H-1'), 4.44–

4.43 (m, 1H, H-4), 4.40 (d, *J* = 8.0 Hz, 1H, H-1), 4.37 (d, *J* = 11.9 Hz, 1H, CH of PhCH), 4.13–4.12 (m, 3H, H-6, CH₂ & CH of octyl), 3.86–3.72 (m, 3H, H-5 & CH₂ of H-6'), 3.62 (m, 1H, H-5'), 3.50 (dd, *J*₁ = 3.4 Hz, *J*₂ = 9.9 Hz, 1H, H-3'), 3.48–3.47 (m, 1H, CH of octyl), 2.17 (s, 3H, COCH₃), 2.11 (s, 6H, 2 × COCH₃), 2.08 (s, 6H, 2 × COCH₃), 2.04 (s, 3H, COCH₃), 1.27 {m, 12H, (CH₂)₆ of octyl}, 0.89 (m, 3H, CH₃ of octyl); ¹³C NMR: (CDCl₃): δ (ppm) 170.4, 170.3, 169.8, 169.6, 169.1 (C=O), 137.2, 128.4, 127.9, 127.7, 101.2, 100.5, 76.3, 72.9, 72.6, 71.7, 71.4, 70.8, 70.6, 70.2, 65.3, 62.3, 61.4, 31.9, 31.7, 29.7, 29.3, 29.2, 25.7, 22.6, 20.8, 20.7, 20.6 (3 × CH₃ of COCH₃), 14.2, 14.1. IR: (neat) ν _{max} 3475, 2926, 2855, 2255, 1751, 1458, 1369, 1226, 1170, 1054, 913, 743, 648 cm⁻¹; ESI-HRMS: *m/z* calcd for C₃₉H₆₀NO₁₇ 814.3861. Found: 814.3861 [M + NH₄]⁺.

4.6.2 Methyl 3-*O*-acetyl-4,6-benzylidene-2-deoxy-2-*N*-phthalimido- β -D-1-thio-glucopyranoside (24**).** Colourless solid (0.23 g, 95%), mp = 220–222 °C; [α]_D²⁰ = -0.6° (*c* = 1, CHCl₃); ¹H NMR (CDCl₃): δ (ppm) = 7.89 (m, 2H, ArH, Phth), 7.77–7.74 (m, 2H, ArH), 7.49–7.49 (m, 2H, ArH), 7.40–7.36 (m, 3H, ArH), 5.97 (t, *J* = 9.2 Hz, 1H, H-3), 5.55 (s, 1H, CH of PhCH), 5.49 (d, *J* = 10.5 Hz, 1H, H-1), 4.47 (dd, *J*₁ = 9.7 Hz, *J*₂ = 5.9 Hz, 1H, H-4), 4.42 (t, *J* = 10.3 Hz, 1H, H-2), 3.85–3.77 (m, 3H, H-5 & H-6, CH₂), 2.17 (s, 3H, COCH₃), 1.87 (s, 3H, SMe); ¹³C NMR: (CDCl₃): δ (ppm) = 170.7 (CO of COCH₃), 167.7 (CO of Phth), 167.5 (CO of Phth), 136.86, 134.4, 131.7, 129.2, 128.2, 126.2, 123.7, 101.6, 81.3, 79.3, 70.6, 68.6, 53.6 (C-6), 20.6 (CH₃ of COCH₃), 11.7 (CH₃ of SMe). IR (KBr) ν _{max}: 3453, 2883, 1719, 1387, 1226, 1098, 1049, 761, 720 cm⁻¹. ESI-HRMS: *m/z* calcd for C₂₄H₂₃NO₇SK 508.0832. Found: 508.0832 [M + K]⁺.

4.6.3 Octyl 2,3-di-*O*-acetyl-6-*O*-*tert*-butyldimethylsilyl- α -D-mannopyranoside (80**).** Colourless syrup (0.11 g, 96%); [α]_D²⁰ = +30.7° (*c* = 1.5, CH₂Cl₂); ¹H NMR (CDCl₃): δ (ppm) 5.21 (dd, *J*₁ = 3.4 Hz, *J*₂ = 12.6 Hz, 1H, H-3), 5.17 (s, 1H, H-2), 4.70 (s, 1H, H-1), 3.98 (t, *J* = 9.4 Hz, 1H, H-4), 3.93 (dd, *J*₁ = 4.6 Hz, *J*₂ = 10.6 Hz, 1H, OCH₂), 3.85 (dd, *J*₁ = 5.3 Hz, *J*₂ = 10.5 Hz, 1H, OCH₂), 3.68–3.67 (m, 1H, H-5), 3.65–3.63 (m, 1H, H-6a), 3.39–3.37 (m, 1H, H-6b), 3.06 (s, 1H, OH), 2.08 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 1.58 (m, 2H, CH₂), 1.33–1.23 (m, 10H, 5 × CH₂), 0.89 (s, 9H, *tert*-butyl), 0.86 (t, *J* = 7.0 Hz, 3H, CH₃ of octyl), 0.09 (s, 6H, 2 × CH₃ of TBDMS). ¹³C NMR: (CDCl₃): δ (ppm) 170.7, 170.1 (C=O), 97.4 (C-1), 72.8, 71.8, 71.3, 69.8, 68.1, 64.2 (C-6), 31.8, 29.3, 26.0, 25.8, 22.6, 20.9, 18.2, 14.0, -5.4 {2 × CH₃ of Si(CH₃)₂}; IR: (neat) ν _{max} 3436, 2925, 2854, 1751 (C=O), 1634, 1372, 1249, 1084, 836 cm⁻¹; ESI-HRMS: *m/z* calcd for C₂₄H₄₆NaO₈Si 513.2864. Found: 513.2854 [M + Na]⁺.

4.6.4 Methyl *N*-(benzoyloxycarbonyl)-*O*-(2,3-di-*O*-acetyl-6-*O*-*tert*-butyldimethylsilyl- α -D-mannopyranosyl)-*L*-serinate (82**).** Solid (0.22 g, 95%); mp 78 °C; [α]_D²⁰ +27.9° (*c* = 1, CH₂Cl₂); ¹H NMR (CD₃OD): δ (ppm) 7.37–7.31 (m, 5H, Ar-H), 5.88 (d, *J* = 8.5 Hz, 1H, NH), 5.16 (dd, *J*₁ = 3.6 Hz, *J*₂ = 1.7 Hz, 1H, H-2), 5.13 (s, 2H, CH₂Ph), 5.10 (dd, *J*₁ = 10.0 Hz, *J*₂ = 3.4 Hz, 1H, H-3), 4.80 (s, 1H, H-1), 4.54 (t, *J* = 4.0 Hz, 1H, CHNH), 3.99 (dd, *J*₁ = 10.4 Hz, *J*₂ = 4.0 Hz, 1H, CH of CH₂O of serine), 3.92 (t, *J* = 4.2 Hz, 1H, H-4), 3.89 (dd, *J*₁ = 7.2 Hz, *J*₂ = 3.0 Hz, 3H, H-6 & CH of CH₂O of serine), 3.77 (s, 3H, OCH₃), 3.68–3.63 (m, 1H, H-5), 2.09 (s, 3H, COCH₃), 2.03 (s, 3H, COCH₃), 0.90 (s, 9H, C(CH₃)₃), 0.08 (s, 6H, Si(CH₃)₂); ¹³C NMR: (CDCl₃): δ (ppm)



170.7, 170.2, 170.1, 170.0 (C=O), 156, 136, 128.5, 128.3, 98.2 (C-1), 72.2, 71.5, 69.3, 68.9, 67.1, 63.5, 54.2, 52.8, 29.6 ($2 \times \text{CH}_3$ of COCH_3), 25.8 (CH_3 of $\text{C}(\text{CH}_3)_3$), 20.9 ($2 \times \text{CH}_3$ of COCH_3), 18.2 (CH_3 of COCH_3), -5.4 (CH_3 of $\text{Si}(\text{CH}_3)_2$); IR: (neat) ν_{max} 3361, 3034, 2927, 2855, 1748 (CO), 1530, 1462, 1438, 1371, 1248, 1141, 1048, 984, 836 cm^{-1} ; ESI-HRMS: m/z calcd for $\text{C}_{28}\text{H}_{44}\text{NO}_{12}\text{Si}$ 614.2633. Found: 614.2632 $[\text{M} + \text{H}]^+$.

4.6.5 *p*-Nitrophenyl 3,4,6-tri-*O*-acetyl- β -D-galactopyranoside (84). Viscous oil (0.8 g, 57%); $[\alpha]_{\text{D}}^{20} -15.2^\circ$ ($c = 1$, CH_2Cl_2); ^1H NMR (CDCl_3): δ (ppm) 8.22 (dd, $J_1 = 9.2$ Hz, $J_2 = 3.7$ Hz, 2H, H-3', H-5', ArNO_2), 7.16 (dd, $J_1 = 9.2$ Hz, $J_2 = 3.2$ Hz, 2H, H-2', H-6', ArNO_2), 5.46 (d, $J = 3.1$ Hz, 1H, H-4), 5.14 (d, $J = 7.6$ Hz, H-1), 5.07 (dd, $J_1 = 3.4$ Hz, $J_2 = 10.2$ Hz, 1H, H-3), 4.23–4.11 (m, 4H, H-2, H-5, H-6a & H-6b), 2.16 (s, 3H, COCH_3), 2.04 (s, 6H, $2 \times \text{COCH}_3$), 2.02 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 170.4, 170.3, 170.1 (C=O), 161, 143.0, 125.7, 116.6, 100.4 (C-1), 72.5, 71.3, 68.5, 66.9, 61.9, 20.7 (CH_3 of COCH_3), 20.5 ($2 \times \text{CH}_3$ of COCH_3); IR: (neat) ν_{max} 3439, 2924, 2927, 1747 (CO), 1611, 1593, 1518, 1372, 1345, 1240, 1074, 862, 751 cm^{-1} ; ESI-HRMS: m/z calcd for $\text{C}_{18}\text{H}_{21}\text{NaNO}_{11}$ 450.1007. Found: 450.1003 $[\text{M} + \text{Na}]^+$.

4.6.6 *m*-Nitrophenyl 2,3,6-tri-*O*-acetyl- β -D-galactopyranoside (87). Crystalline solid (1.0 g, 78%), mp 149–160 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{20} -10.6^\circ$ ($c = 0.5$, CH_2Cl_2); ^1H NMR (CDCl_3): δ (ppm) 7.97 (d, $J = 8.0$ Hz, 1H, H-4', ArNO_2), 7.90 (d, $J = 2.0$ Hz, 1H, H-2', ArNO_2), 7.49 (t, $J = 8.2$ Hz, 1H, H-5', ArNO_2), 7.72 (d, $J = 8.0$ Hz, 1H, H-6', ArNO_2), 5.59 (dd, $J_1 = 7.9$ Hz, $J_2 = 10.2$ Hz, 1H, H-2), 5.14 (d, $J = 7.9$ Hz, 1H, H-1), 5.09 (dd, $J_1 = 10.3$ Hz, $J_2 = 3.12$ Hz, 1H, H-3), 4.45 (dd, $J_1 = 4.8$ Hz, $J_2 = 11.8$ Hz, 1H, H-6a), 4.37 (dd, $J_1 = 7.4$ Hz, $J_2 = 11.8$ Hz, 1H, H-6b), 4.19–4.16 and 4.04–4.00 (2m, 3H, H-4, H-5 & OH), 2.19 (s, 3H, COCH_3), 2.09 (s, 6H, $2 \times \text{COCH}_3$), 2.02 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 170.5, 170.1, 169.9, 169.2 (C=O), 157.0, 149.1, 130.17, 123.7, 118.1, 111.3, 99.2 (C-1), 71.6, 70.6, 68.3, 66.9, 61.7, 20.6 (CH_3 of COCH_3), 20.5 ($3 \times \text{CH}_3$ of COCH_3); IR: (neat) ν_{max} 3459, 2933, 1746 (CO), 1634, 1532, 1352, 1230, 1080, 1141, 1048, 749 cm^{-1} ; ESI-HRMS: m/z calcd for $\text{C}_{21}\text{H}_{26}\text{O}_{11}\text{Na}$ 477.1373. Found: 477.1370 $[\text{M} + \text{Na}]^+$.

4.6.7 *o*-Nitrophenyl 2,4,6-tri-*O*-acetyl- β -D-galactopyranoside (90). ^1H NMR (CDCl_3): δ (ppm) 7.88 (dd, $J_1 = 8.1$ Hz, $J_2 = 1.6$ Hz, 1H, H-3', ArNO_2), 7.58–7.56 (m, 1H, H-4', ArNO_2), 7.42–7.40 (m, 1H, H-5', ArNO_2), 7.23–7.19 (m, 1H, H-6', ArNO_2), 5.43–5.42 (m, 1H, H-4), 5.33 (dd, $J_1 = 10.0$ Hz, $J_2 = 7.8$ Hz, 1H, H-2), 5.07 (d, $J = 7.9$ Hz, 1H, H-1), 4.41 (d, $J = 6.4$ Hz, 2H, H-6), 3.97 (dd, $J_1 = 9.7$ Hz, $J_2 = 3.2$ Hz, 1H, H-3), 3.96 (t, 1H, $J = 6.4$ Hz, H-5), 2.23 (s, 3H, COCH_3), 2.22 (s, 3H, COCH_3), 2.11 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 171.3, 170.7, 170.3 (C=O), 149.4, 141.4, 134.5, 125.6, 123.5, 119.4, 103.6 (C-1), 74.0, 72.8, 71.5, 69.2, 66.7, 61.9, 20.7 ($3 \times \text{CH}_3$ of COCH_3); IR: (Neat) ν_{max} 3445, 2924, 2854, 1743 (CO), 1607, 1529, 1487, 1370, 1237, 1073, 748 cm^{-1} . ESI-HRMS: m/z calcd for $\text{C}_{18}\text{H}_{21}\text{NNaO}_{11}$ 450.1007. Found: 450.1006 $[\text{M} + \text{Na}]^+$.

4.6.8 *o*-Nitrophenyl 3,6-di-*O*-acetyl- β -D-galactopyranoside (91). ^1H NMR (CDCl_3): δ (ppm) 7.82 (dd, $J_1 = 8.0$ Hz, $J_2 = 1.6$ Hz, 1H, H-3', ArNO_2), 7.58–7.55 (m, 1H, H-4', ArNO_2), 7.38–7.28 (m, 1H, H-5', ArNO_2), 7.24–7.20 (m, 1H, H-6', ArNO_2), 5.01 (dd, $J_1 = 10.2$ Hz, $J_2 = 3.2$ Hz, 1H, H-3), 4.97 (d, $J = 7.6$ Hz, 1H, H-1), 4.23 (dd, 1H, $J_1 = 7.8$ Hz, $J_2 = 10.2$ Hz, H-2), 4.22 (d, $J = 6.3$ Hz, 2H, H-

6), 4.14 (d, 1H, $J = 2.9$ Hz H-4), 4.03 (dt, $J_1 = 7.4$ Hz, $J_2 = 0.9$ Hz, 1H, H-5), 2.13 (s, 3H, COCH_3), 2.10 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 170.7, 170.3 (C=O), 150.3, 141.4, 133.6, 125.1, 123.4, 118.9, 100.4 (C-1), 74.0, 71.1, 68.7, 66.7, 62.2, 20.7 ($2 \times \text{CH}_3$ of COCH_3); IR: (neat) ν_{max} 3445, 2924, 2854, 1743 (CO), 1607, 1529, 1487, 1370, 1237, 1073, 748 cm^{-1} . ESI-HRMS: m/z calcd for $\text{C}_{16}\text{H}_{19}\text{NNaO}_{10}$ 408.0901. Found: 408.0900 $[\text{M} + \text{Na}]^+$.

4.6.9 *o*-Nitrophenyl 4,6-di-*O*-acetyl- β -D-galactopyranoside (92). ^1H NMR (CDCl_3): δ (ppm) 7.88 (dd, $J_1 = 8.1$ Hz, $J_2 = 1.4$ Hz, 1H, H-3', ArNO_2), 7.58–7.56 (m, 1H, H-4', ArNO_2), 7.39–7.36 (m, 1H, H-5', ArNO_2), 7.21–7.17 (m, 1H, H-6', ArNO_2), 5.44 (d, $J = 2.8$ Hz, 1H, H-4), 4.97 (d, $J = 7.4$ Hz, 1H, H-1), 4.22–4.20 (m, 2H, H-6), 4.09 (t, $J = 6.3$ Hz, 1H, H-5), 4.02 (t, $J = 4.0$ Hz, 1H, H-2), 3.96 (dd, $J_1 = 9.7$ Hz, $J_2 = 3.3$ Hz, 1H, H-3), 2.21 (s, 3H, COCH_3), 2.12 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 170.9, 170.5 (C=O), 150.3, 141.3, 134.4, 125.6, 123.3, 119.4, 102.8 (C-1), 72.9, 71.8, 71.3, 68.7, 62.7, 20.8 ($2 \times \text{CH}_3$ of COCH_3); IR: (Neat) ν_{max} 3419, 2924, 1742 (CO), 1607, 1528, 1370, 1238, 1070, 749 cm^{-1} . ESI-HRMS: m/z calcd for $\text{C}_{16}\text{H}_{19}\text{NNaO}_{10}$ 408.0907. Found: 408.0903 $[\text{M} + \text{Na}]^+$.

4.6.10 *o*-Nitrophenyl 2,6-di-*O*-acetyl- β -D-galactopyranoside (93). ^1H NMR (CDCl_3): δ (ppm) 7.79 (dd, $J_1 = 8.1$ Hz, $J_2 = 1.4$ Hz, 1H, H-3', ArNO_2), 7.56–7.52 (m, 1H, H-4', ArNO_2), 7.39–7.37 (m, 1H, H-5', ArNO_2), 7.21–7.19 (m, 1H, H-6', ArNO_2), 5.33 (t, $J = 9.0$ Hz, 1H, H-2), 5.00 (d, $J = 7.9$ Hz, 1H, H-1), 4.42–4.41 (m, 2H, H-6), 4.03–4.00 (m, 1H, H-4), 3.90 (t, $J = 6.2$ Hz, 1H, H-5), 3.80 (dd, $J_1 = 9.5$ Hz, $J_2 = 3.1$ Hz, 1H, H-3), 2.21 (s, 3H, COCH_3), 2.11 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 171.7, 170.9 (C=O), 149.5, 140.5, 133.6, 125.0, 123.2, 118.6, 100.4 (C-1), 72.2, 71.8, 71.5, 68.4, 61.9, 20.7 ($2 \times \text{CH}_3$ of COCH_3); IR: (neat) ν_{max} 3419, 2924, 1742 (CO), 1607, 1528, 1370, 1238, 1070, 749 cm^{-1} . ESI-HRMS: m/z calcd for $\text{C}_{16}\text{H}_{19}\text{NNaO}_{10}$ 408.0901. Found: 408.0902 $[\text{M} + \text{Na}]^+$.

4.6.11 *p*-Nitrophenyl 3-*O*-acetyl-4,6-*O*-benzylidene- β -D-galactopyranoside (95). Colourless solid (0.7 g, 60%), mp 201–208 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{20} -18.3^\circ$ ($c = 1$, CHCl_3); ^1H NMR (CDCl_3): δ (ppm) 8.22 (dt, $J_1 = 9.2$ Hz, $J_2 = 3.3$ Hz, 2H, H-3', ArNO_2), 7.51–7.49 (m, 2H, ArH), 7.39–7.35 (m, 3H, ArH), 7.16 (dt, $J_1 = 9.2$ Hz, $J_2 = 3.3$ Hz, 2H, H-2', ArNO_2), 5.54 (s, 1H, CH of PhCH), 5.14 (d, $J = 7.7$ Hz, 1H, H-1), 4.99 (dd, $J_1 = 10.2$ Hz, $J_2 = 3.5$ Hz, 1H, H-3), 4.49 (d, $J = 2.9$ Hz, 1H, H-4), 4.38–4.34 (m, 1H, H-2), 4.35 (dd, $J_1 = 12.6$ Hz, $J_2 = 1.5$ Hz, 1H, H-6a, CH_2), 4.15 (dd, $J_1 = 12.6$ Hz, $J_2 = 1.5$ Hz, 1H, H-6b, CH_2), 3.73 (s, 1H, H-5), 2.18 (s, 3H, COCH_3); ^{13}C NMR: (CDCl_3): δ (ppm) 171.0 (C=O), 161.6, 142.9, 137.3, 129.1, 128.2, 126.1, 125.7, 116.7, 101.0, 100.5, 73.4, 73.0, 68.7, 68.0, 66.8, 21.0 (CH_3 of COCH_3); IR: (neat) ν_{max} 3482, 2992, 2936, 2864, 1755 (C=O), 1593, 1519, 1347, 1230, 1089, 994, 910, 850, 750 cm^{-1} ; anal. calcd for $\text{C}_{21}\text{H}_{21}\text{NO}_9$: C, 58.47; H, 4.91; N, 3.25. Found: C, 58.12; H, 5.02; N, 2.87. ESI-MS: m/z calcd for $\text{C}_{21}\text{H}_{21}\text{NO}_9$ 431.12. Found: 431.21 $[\text{M}]^+$.

4.6.12 3-Methoxy-4-(2,3,6-tri-*O*-acetyl- β -D-glucopyranosyloxy)acetophenone (98). Viscous oil (1.05 g, 77%); $[\alpha]_{\text{D}}^{20} -39.5^\circ$ ($c = 1.5$, CH_2Cl_2); ^1H NMR (CDCl_3): δ (ppm) 7.52 (dd, $J = 2$ Hz, 1H, H-2, ArNO_2), 7.51 (dd, $J_1 = 8.2$ Hz, $J_2 = 1.9$ Hz, 1H, H-6, ArNO_2), 7.14 (d, $J = 8.2$ Hz, 1H, H-5, ArNO_2), 5.28 (dd, $J_1 = 7.7$ Hz, $J_2 = 9.5$ Hz, 1H, H-2'), 5.14 (dd, $J_1 = 8.8$ Hz, $J_2 = 9.4$ Hz, 1H, H-3'), 5.06 (d, $J = 7.7$ Hz, 1H, H-1'), 4.49 (dd, $J_1 =$



12.3 Hz, $J_2 = 5.0$ Hz, 1H, H-6a'), 4.39 (dd, $J_1 = 12.3$ Hz, $J_2 = 2.0$ Hz, 1H, H-6b), 3.87 (s, 3H, OCH₃), 3.70 (t, $J = 8.6$ Hz, 1H, H-4'), 3.67–3.65 (m, 1H, H-5'), 2.57 (s, 3H, ArCOCH₃), 2.12 (s, 3H, COCH₃), 2.10 (s, 3H, COCH₃), 2.07 (s, 3H, COCH₃); ¹³C NMR: (CDCl₃): δ (ppm) 196.9 (C=O), 171.5, 171.4, 169.5 (C=O), 150.4, 150.2, 133.3, 122.4, 117.1, 111.6, 99.9 (C-1), 75.1, 74.4, 70.9, 68.7, 62.6, 56.1, 26.4 (CH₃ of ArCOCH₃), 20.8 (3 \times CH₃ of COCH₃), 14.1; IR: (neat) ν_{\max} 3459, 2924, 2853, 2855, 1747 (C=O), 1674, 1593, 1416, 1367, 1233, 1080, 1035, 908, 805, 744 cm⁻¹; ESI-HRMS: m/z calcd for C₂₁H₂₆O₁₁Na 477.1373. Found: 477.1370 [M + Na]⁺.

4.6.13 1-O-Acetyl-3-O-(tert-butylidiphenylsilyl)glycerol

(104). ¹H NMR (CDCl₃): δ (ppm) 7.68–7.65 (m, 4H, ArH), 7.48–7.39 (m, 6H, ArH), 4.19–4.16 (m, 2H, CH₂), 3.95–3.93 (m, 1H, CH), 3.72–3.69 (m, 2H, CH₂), 2.07 (s, 3H, COCH₃), 1.09 {s, 9H, CH₃, C(CH₃)₃}; ¹³C NMR: (CDCl₃): δ (ppm) 171.1 (C=O), 135.5, 133.0, 129.9, 127.8, 71.7, 65.2, 64.4, 26.7 {C(CH₃)₃}, 21.0 (CH₃ of COCH₃); IR: (neat) ν_{\max} 3435, 2963, 1770 (CO), 1428, 1371, 1260, 1136, 1012, 799 cm⁻¹. ESI-HRMS: m/z calcd for C₂₁H₂₈NaO₄Si 395.1649. Found: 395.1647 [M + Na]⁺.

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