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Arylidene indanone scaffold: medicinal chemistry and structure–activity relationship view

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Arylidene indanone (AI) scaffolds are considered as the rigid cousins of chalcones, incorporating the α,β -unsaturated ketone system of chalcones forming a cyclic 5 membered ring. They are generally synthesized from 1-indanone and benzaldehydes *via* an aldol reaction. The furnished molecules have been explored as inhibitors of cholinesterases towards the treatment of Alzheimer's disease, as tubulin depolymerizing agents, as inhibitors of breast cancer and leukemia, inhibitors of dual specificity phosphatase (DUSP), as antimalarials, and for many other uses. This review is an effort to highlight the biochemical effects of arylidene indanones designed from natural or known drug compounds, discuss their structure–activity relationships (SAR), and correlate them with related chalcones providing insights for further development of this scaffold.

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1. Introduction

Natural indanones like pterosin P **1** from fern (*Pteridium*) species,^{1–3} 4-hydroxy-7-methylindanone **2** and 7-formyl-3-methoxy-5-methylindanone **3** from marine cyanobacterium^{4,5} have shown antibacterial,⁴ antispasmodic,⁶ inhibition of hypoxia-induced activation of vascular endothelial growth factor (VEGF)⁵ and cytotoxic activities.³ The 4,5,6-

trimethoxyindanone-estradiol hybrid **4** was more potent against MCF-7 breast cancer cells in comparison to the chalcone-estradiol precursor **5** (see Fig. 1 for structures).⁷

Arylidene indanones (AI) are mostly synthetic molecules, consisting of the natural/synthetic indanone core (see the part of the structure in green lines) and an aryl group linked through an exocyclic double bond at the 2-position of this indanone core (for the numbering system see Fig. 1). This AI scaffold often connoted as cyclic chalcone can be considered as rigid cousins or homologs of chalcones, incorporating the α,β -unsaturated ketone (red dashed circles) into a cyclic 5 member ring which is attached to the 2-position of chalcone ring (red arrow) and to the α -position of the unsaturated bond (blue arrow), making it

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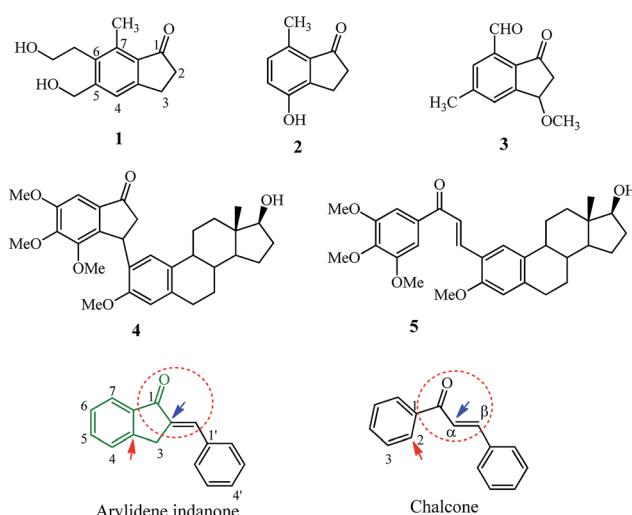


Fig. 1 Natural and synthetic indanones and the basic similarities between arylidene indanone and its cousin, the chalcone.



a planar/rigid exocyclic α,β -unsaturated ketone system. Chalcones, belong to the class of flavonoids and have been explored as anti-infective, anti-inflammatory and anti-cancer agents.^{8,9} The interaction of natural or synthetic chalcones with molecular targets like tubulin, kinases, oxidoreductases, hydrolases, esterases,¹⁰ and cell cycle blockers have been reviewed.¹¹ Recent developments of the synthetic chalcone scaffold with different heterocyclic rings or hybrids with other natural compounds for various therapeutic roles have been compiled.¹²

The position of the arylidene ring with respect to the indanone core results in a planar structure and allows transmission of the electron donor effects of the substituents *via* the double bond onto the carbonyl group of indanone, thus lowering the wavenumber-frequency more effectively compared to chalcones.¹³ Higher values were observed in the ^{13}C NMR of the α,β -unsaturated ketone carbons of AIs compared with chalcones which had similar electron donating substituents in the aryl ring.^{13,14} The position of absorption maxima (UV-vis) for AI is higher in comparison with chalcones having similar *para*-substituents in the aryl ring indicating the strongest conjugation due to its planar structure.¹⁵ The lipophilicity of AI can be increased by geometric inversion of the double bond from the *E* to *Z*-isomer which occurs due to UV-light.¹⁶ In the case of chalcones, the double bond can have either the *s-cis* or *s-trans* conformation while in the case of AI the molecule is forced to adopt the *s-cis* conformation due to the cyclic ring. The introduction of an α -substituent on the double bond makes the chalcone adopt the *s-trans* conformation and this was explored to design potent tubulin inhibitors.^{17,18}

Retrosynthesis of AI leads to 1-indanones (natural or synthetic) which are assembled from phenyl/aryl propionic acid *via* cyclization reactions using polyphosphoric acid,¹⁹ methane sulphonic acid,²⁰ Friedel-Crafts acylation under microwave heating and ultrasonication,²¹ and Nazarov cyclization of chalcones.⁷ Further reaction of suitably substituted 1-indanone with benzaldehyde derivatives under aldol conditions or potassium carbonate and microwave irradiation leads to the synthesis of AI.²²⁻²⁴ Substitution in the indanone ring as well as in the benzaldehyde moiety has furnished AI scaffolds having therapeutic effects against breast cancer,²⁵ leukemia,²⁶ and Alzheimer's disease,²⁷ AI scaffolds causing inhibition of DUSP^{28,29} and AI scaffolds acting as potential antimalarial agents.^{30,31} This review discusses the various therapeutic roles of AIs, their structure-activity relationships and correlates them wherever possible with structurally similar or related chalcones. The web-based Molinspiration software and the related molecular properties (using the Lipinski rule) were utilized to predict theoretical milog *P* (octanol-water partition coefficient) of selected groups of molecules.^{32,33}

2. Effect of AI in Alzheimer's disease

Alzheimer's disease (AD) is a neurodegenerative disorder which is the cause of dementia (loss of memory) in the later stages of adult life and its prevalence is estimated to rise up to 106 million by 2050.³⁴ The neuropathological features of AD are extracellular deposits of amyloid beta peptide (A β) called

amyloid plaques and hyperphosphorylated neurofibrillary tangles (NFTs).³⁵ Cleavage of the amyloid protein precursor (APP) by secretases (α -, β -, and γ -secretase) results in the production of various species of A β which due to non-clearance from the brain form A β 42 oligomers which hydrophobically self-aggregate and give rise to diffuse senile plaques (SPs).³⁵

The human brain has two major cholinesterases; acetylcholinesterase (AChE) and butyrylcholinesterase (BuChE) both differing in their genetics, structure, and kinetics. In patients with AD, BuChE is found in neurons and glial cells as well as in neuritic plaques and tangles.³⁶ Acetylcholine, a neurotransmitter molecule linked with intellectual abilities decreases in the brain due to enhanced acetylcholinesterase activity resulting from AD or old age. The acetylcholinesterase (AChE) enzyme structure determined for *Torpedo californica* (Tc) consists of a catalytic triad (having residues Ser200, His440, and Glu327) and a peripheral anionic site (PAS).³⁷ The active site also contains an anionic subsite consisting of the key residue Trp84 located near the bottom of the cavity. This site interacts with the quaternary ammonium group of acetylcholine substrates and other ligands *via* cation- π interaction.³⁷ While the PAS of the acetylcholinesterase enzyme is located at the entrance of the narrow catalytic gorge which serves as a transient binding site for substrates entering the active site. The A β peptide interacts with the PAS resulting in the folding of these proteins and the formation of plaques.³⁷ Donepezil hydrochloride **6** (see Fig. 2) interacts with these dual binding sites of the AChE enzyme³⁸ and is the presently used drug (Aricept) for all stages of Alzheimer's disease.³⁹ Several other drugs like rivastigmine **7**, galantamine **8**, memantine **9** and the combination of donepezil and memantine have been approved.³⁹ Hybrid compounds for the treatment of AD have been derived by combining fragments or drug molecules like **6** and tacrine **10**;^{40,41} **8** and **9**;⁴² and natural molecules like curcumin, berberine, ferulic and caffeic acid,^{42,43} chalcones and coumarin derivatives,⁴³ and resveratrol and clioquinol.⁴⁴

Most of the compounds discussed in this section have been derived from the indanone core of **6** with modification in the 5,6-position or substitution of arylidene with fragments from

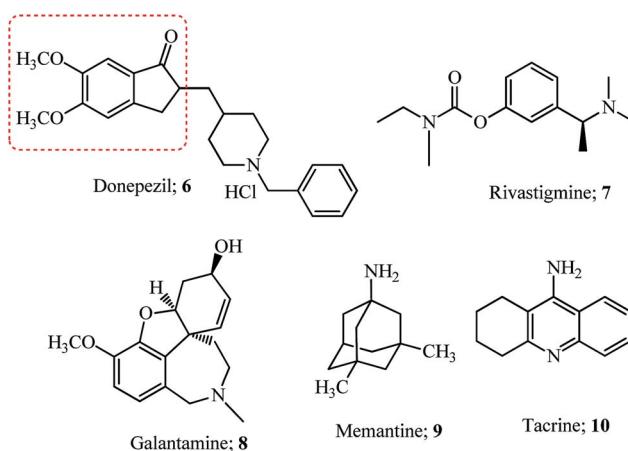


Fig. 2 Drugs approved for the treatment of Alzheimer's disease.



other drugs. Hybrid molecules designed by combining the phenyl-*N*-methylbenzylamine fragment of AP2238 (see the structure and fragment in the blue ellipsoid; Fig. 3), with the indanone core of donepezil **6** (see the red dashed box in Fig. 2) resulted in several new indanone derivatives with potencies against AChE (5 examples).⁴⁵

Although the 5,6-dimethoxy groups of donepezil have efficient interaction with the PAS of the cholinesterase enzyme, the methoxy at 6-position of indanone **11a** gave a hydrophobic interaction with the side chain of the Leu289 residue and was more potent for AChE inhibition.⁴⁵ Further modification of the 6-position by introducing alkyl chains of varying lengths having different terminal amine groups was studied for efficient interaction with the PAS of AChE (12 examples). The optimum length of the alkyl spacer was 5 carbons, with the terminal *N,N*-diethyl substituent (**11b**). The ability of **11b** to prevent AChE-induced and self A β aggregation was noted.⁴⁵

APP is cleaved by a β -secretase known as the β -site APP cleaving enzyme, BACE1, and this is the rate-limiting step in the generation of A β peptide fragments.³⁵ Inhibition of this enzyme was attempted by retaining the basic structure of **11b** and introducing piperazine on the pentyl side chain with a di-substituent on the second nitrogen (8 examples). Substitution with bis(4-fluorophenyl)methyl resulted in the molecule **11c** which showed potent activity against BACE1 ($IC_{50} = 2.49 \mu M$).⁴⁶ Docking studies of **11c** with the BACE1 enzyme showed interaction with amino acid residues outside the binding pocket but it was unable to enter the narrow gorge of hAChE due to the bulky nature of the bis(4-fluorophenyl)methyl group and it had low neurotoxicity in human neuronal SH-SY5Y cells.⁴⁶

Meng and co-workers evaluated the modification of the 6-position of indanone by derivatives of the type **12** (13 examples).⁴⁹ The introduction of the piperidinyl ethoxy substituent at the 6-position of indanone **12a** and pyridinyl methylene at the 2-position made indanone **12b** more potent than donepezil **6** and the starting indanone **12a** (see Fig. 3 and Table 1 for IC_{50} values). The introduction of the piperidinyl ethoxy group in the

6-position of the starting indanone **12a** had no improvement on the inhibition efficiency indicating the crucial nature of the double bond for the activity. Chelation of biometals like Cu^{2+} , Fe^{3+} and Zn^{2+} is an important strategy for treatment of AD since these metals accumulate on the formation of amyloidal plaques and SPs.^{53,54} Metal chelation studies using **12b** and a solution of Cu^{2+} , Fe^{3+} and Zn^{2+} indicated a blue shift using UV-vis spectrophotometry (a decrease in wavelength or a move towards the blue end of the visible spectrum) from 360 nm to 352 nm, 346 nm, and 357 nm respectively.⁴⁹ The piperidinyl ethoxy arm of **12b** (2 carbon spacer) functions/mimics as the biometal chelator group is also present in clioquinol (see the dashed oval shape for the clioquinol structure in Fig. 3) and is used as a reference for these studies. This fact was fortified by using a compound with a carbon spacer of 5 (C = 5) which did not show any metal chelation.⁴⁹ In the case of clioquinol, the donor groups N and O may form a bidentate chelate which can form a five-membered ring/complex with bivalent metals and the five-membered ring complex formed during chelation is more effective, compared to any other ring size.⁵⁵ The benefits/challenges of using chelating therapy for AD have been given an overview, which includes clioquinol among other molecules.^{53,54}

The 6-piperidinylethoxy-5-methoxy indanone moiety from **12b** was retained while the 4-position on the arylidene ring was varied to design “multi-target one ligand” molecules (11 examples). Derivatives **13a**–**13c** (Fig. 3) indicated a higher potency for inhibition of AChE with nanomolar IC_{50} values compared with **10** and similar ones to **6** (see Table 1 for IC_{50} values).⁵¹ The substituent at the 4-position of the phenyl ring showed no relevant role as the potencies were similar for compounds **13a**–**13c**.⁵¹ Inhibition of A β aggregation by **13a** and **13b** was noted. Antioxidant activity was highly diminished for **13c** highlighting the importance of the amino group at the 4-position on the phenyl ring.⁵¹ The chalcone derivative **13d** (Fig. 3) having 2-pyrrolidinyl ethoxy at the 4'-position and a fluoro group at the 4-position showed potency for AChE

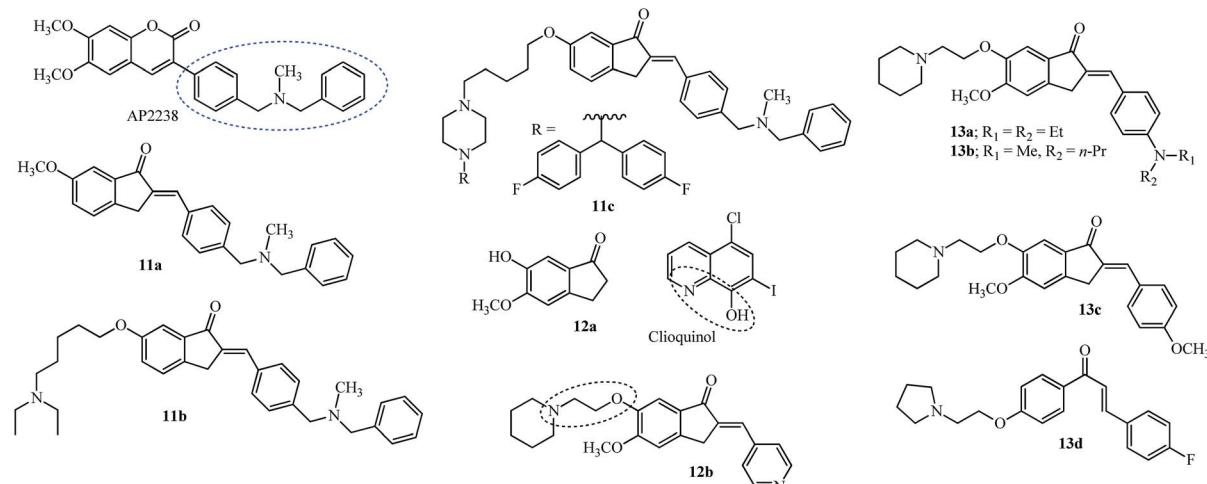


Fig. 3 Cholinesterase inhibitor hybrids derived from donepezil and AP2238.



Table 1 IC_{50} of AIs used for therapy of Alzheimer's disease

Compound	IC_{50} hAChE (μ M)	IC_{50} hBuChE (μ M)	Method/substrate and Ref	Compound	IC_{50} hAChE (μ M)	IC_{50} hBuChE (μ M)	Method/substrate and Ref
11a	0.63	7.93	Ellman ⁴⁵	7	10.54	0.26	Ellman ⁴⁷
11b	0.18	10.0	Ellman	14a	0.045	3.015	eeAChE/eqBuChe ⁴⁸
6	0.02	7.42	Ellman ⁴⁵	14b	0.025	2.434	eeAChE/eqBuChe
AP2238	0.044	48.9	Ellman ⁴⁵	14c	0.034	1.969	eeAChE/eqBuChe
11c^a	2.49 ^a	ND	hBACE1	6	0.039	3.36	eeAChE/eqBuChe ⁴⁸
12a	91.96	258.03	Ellman ⁴⁹	14d	4.895	—	eeAChE/Ellman ⁵⁰
12b	0.0018	9.5	Ellman	6	0.2197	—	eeAChE/Ellman ⁵⁰
6	0.026	4.66	Ellman ⁴⁹	10	0.3715	—	eeAChE/Ellman ⁵⁰
13a	14.8 nM	1384.9	eeAChE/eqBuChe ⁵¹	15a	0.42	315	Ellman ⁵²
13b	18.6 nM	1417.1	eeAChE/eqBuChe	15b	0.035	82	Ellman
13c	14.8 nM	3213.6	eeAChE/eqBuChe	15c	0.045	26.6	Ellman
6	38.2 nM	3365.4	eeAChE/eqBuChe ⁵¹	15d	0.10	84.6	Ellman
10	109 nM	15.2	eeAChE/eqBuChe ⁵¹	7	2.07	1.41	Ellman ⁵²
13d	0.21	13.65	Ellman ⁴⁷	—	—	—	—

^a Human recombinant BACE1 (hBACE1) using FRET assay; ND-not determined.

compared to BuChE and this was correlated by docking analysis which showed a dual interaction with the catalytic site and the PAS of the AChE enzyme.⁴⁷

The bis(4-fluorophenyl)methyl piperazine group (see R of **11c** in Fig. 3) was introduced on the 4-position of the arylidene group (**14a**; Fig. 4) providing the most potent acetylcholinesterase inhibition amongst the 15 tested compounds.⁴⁸ The pyrimidine **14b** and 3-furoyl groups **14c** also gave potent acetylcholinesterase and butyrylcholinesterase inhibition compared with **6** (see Table 1).⁴⁸ The substitution on the piperazine ring by *N,N*-dimethyl aminopropyl group gave the most potent molecule **14d** having AChE inhibition activity (among 24 examples) and was compared with standard reference donepezil **6** and tacrine **10**.⁵⁰

Incorporating the *N,N*-dimethyl amino side chain similar to that found in rivastigmine **7** (Fig. 2), at the *para* position of the benzene ring and varying the substituents to diethyl, pyrrolidine and piperidine resulted in 20 compounds of the type **15a**–

d (see Fig. 4). The *para* isomers were more effective than their *meta* isomers. The diethyl **15b** and pyrrolidine **15c** AIs were more potent compared with dimethyl amino **15a**, piperidine **15d**, and rivastigmine **7** (see IC_{50} values in Table 1). In general, the AIs were more potent than their reduced counterparts and selective for AChE over BuChE.⁵² The active indanone derivative **15b** improved the acquisition and memory loss induced by scopolamine^{52,56} sodium nitrite, and ethanol⁵⁶ and showed neuroprotective functions in PC12 cells.⁵⁶

Interaction of the diethylamine nitrogen of **15b** with the Trp84 residue situated at the bottom of the AChE gorge and of the indanone ring with the indole ring of Trp279 at the top of the gorge was shown by docking studies based on the X-ray crystal structure of the TcAChE-E2020 (donepezil) complex (see Fig. 5 reproduced with permission from Elsevier Masson SAS).⁵² These docking interactions were verified by independent QSAR and docking studies of **15b** which revealed similar findings.⁵⁷ Another recent docking study also verified these finding

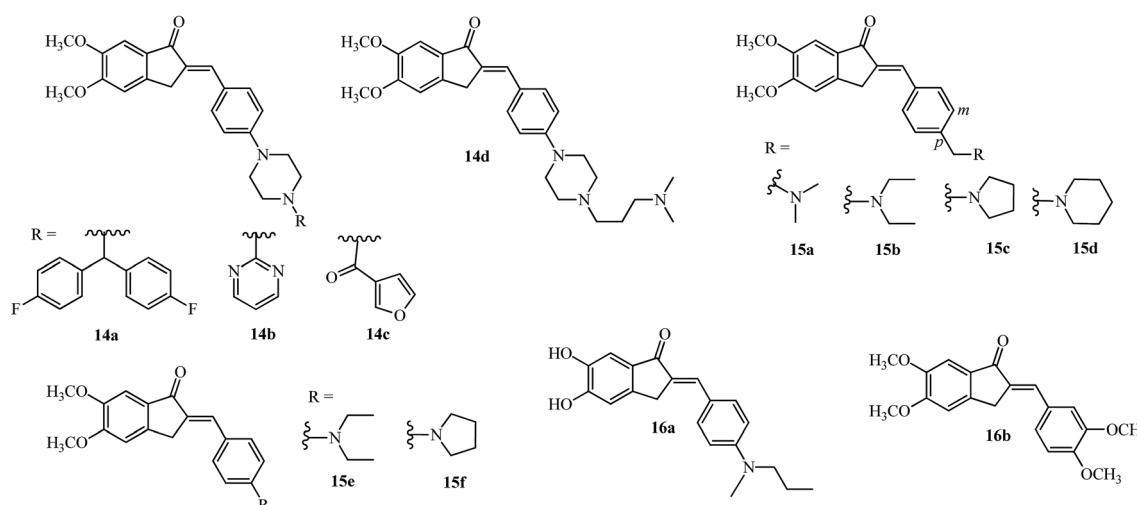


Fig. 4 Inhibitors of acetylcholinesterase and $\text{A}\beta$ aggregation and metal chelation agents.



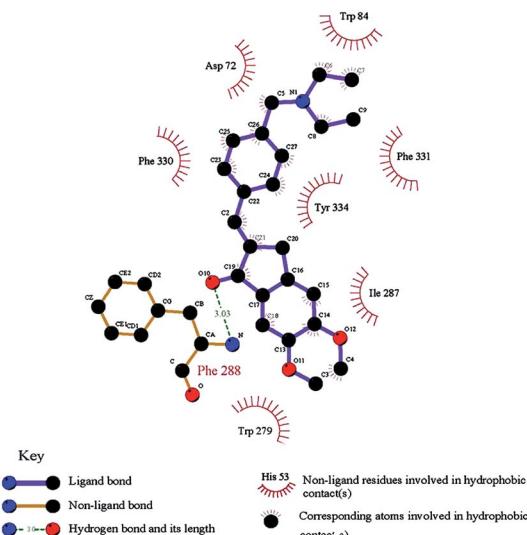


Fig. 5 Docking of **15b** (carbon atoms in black and bonds in purple), into the gorge of the TcAChE enzyme. This figure has been reproduced from ref. 52 with permission from Elsevier Masson SAS.

and found the pyrrolidinyl derivative **15c** which is a non-classical bioisostere of **15b** to have a similar potency.⁵⁸ A recent study with the AI derivatives **15e** and **15f** having the *N,N*-diethylamino and pyrrolidine groups directly attached to the 4-position of aryl ring didn't show promising AChE or BuChE inhibition activities,⁵⁹ pointing out the importance of methylene substitution at the 4-position of aryl ring as seen in **15b** and **15c**.⁵²

Self-mediated $\text{A}\beta_{1-42}$ aggregation which is implicated in the progression of AD, was targeted by **16a** (Fig. 4; highest potency of 80.1% at a concentration of 20 μM) which was among 19 derivatives having the 5,6-position of the indanone core substituted by methoxy and hydroxy groups while the arylidene ring had methoxy and alkylamino or dialkylamino substitution at the 3',4'-position.²⁷ Stoichiometric interaction of **16a** (ratio of 1 : 1) with Cu^{2+} solutions showed a red shift (437 nm to 451 nm) by UV-vis spectrophotometry, while no significant change for Fe^{3+} and Zn^{2+} metals were observed.²⁷ The *ortho*-dihydroxy group is also present in natural flavonoids like quercetin and myricetin and has shown chelation of copper ions at physiological pH.⁵⁹

Monoamine oxidases (MAO) A and B are enzymes which belong to the family of flavin adenine dinucleotides (FADs) responsible for catalyzing the oxidative deamination of neurotransmitters like serotonin, dopamine and adrenaline in the CNS due to the progression of AD.⁶⁰ Inhibitors of MAO-A/B may increase the levels of these neurotransmitters improving the learning and memory deficit of patients having AD. Selective inhibition of MAO-B by **16a** was observed (7.50 μM); while 5-methoxy-6-hydroxy substitution diminished this activity (40.5 μM), indicating the importance of the dihydroxyl substitution.²⁷ Various scaffolds like coumarins and indole have been synthesized and tested as dual MAO-B and AChE inhibitors,⁶⁰ indicating the unexplored potential of the arylidene indanone scaffold.

Although the dihydroxy-substituted **16a** showed MAO-B inhibition, several new compounds like **16c–16h** (Fig. 6) similar to this scaffold have shown inhibition with mono hydroxy/methoxyl substitution at the 5-position.^{61,62} The 5-monohydroxyl compound **16c** showed better inhibition of MAO-B compared to the 6-hydroxy isomer **16d** (see IC_{50} values in Table 2). In a previous study, the appropriate benzyloxy substituent on the 5 or 6-position of the indanone ring was shown to have influence in the inhibition of MAO-B over MAO-A.⁶³ Moreover, a comparison of the MAO-B inhibition data of the 5 and 6-hydroxyindanones **16c'** and **16d'** (ref. 63) with the arylidene indanones **16c** and **16d** indicates that the inhibition potency improved with the introduction of the benzylidene moiety.⁶¹ The calculated milog *P* values of **16c** and **16d** (3.24) are higher than **16c'** and **16d'** (1.22) indicating an increase of lipophilicity due to the benzylidene moiety.³²

Substitution on the arylidene ring at the 4'-position with bromo (**16e**) and methyl (**16f**) gave potent results among 22 tested compounds. These compounds were more potent for MAO-B than as MAO-A inhibitors. The lipophilic 5-methoxyindanone with the benzylidene moiety **16g** improved the MAO-B inhibition value compared to that observed for **16c**. The modification of the arylidene group using hetero arylidene moieties like pyridine, furan and thiophene gave the equipotent compound **16h** which has a 5-bromo-2-furan substituent.⁶² Chalcones **16i** and **16j** having some structural similarities with the AIs discussed above were more potent for MAO-B compared to their use as MAO-A inhibitors (see Table 2).^{64,65}

Detection of senile plaques (SPs) which are composed of $\text{A}\beta$ protein aggregates and NFTs may help in the diagnosis of AD.⁶⁶ The importance of single photon emission computed tomography (SPECT) radiotracers for imaging amyloid plaques in the brain of AD patients has been an active area of research using different types of molecules.⁶⁶ Amyloid imaging probes *e.g.*, the radioactive labeled Pittsburgh compound B (PIB), and its ¹⁸F analog are routinely used for detection (Fig. 7). AIs **17a–17d** (Fig. 7) were evaluated as amyloid imaging agents *in vitro* and *in vivo* since they qualify as a neutral molecule possessing at least two phenyls (or heterocyclic) rings and a linker and are in conjugation with each other, and the molecule as a whole has a hydrophobic, planarized π system.⁶⁷

A binding affinity assay with human AD brain homogenates showed that **17a** has the highest binding constant ($K_i = 16 \pm 1$ nM) while the 5-methoxy **17b** showed an increase in the binding constant ($K_i = 21 \pm 1$ nM). The iodinated derivatives **17c–17d** were bound to the SPs in less than 20 minutes but were unusable for *in vivo* experiments due to their high binding constants (>20 nM) while **17d** had excellent fluorescence staining in SPs. Radioiodination derivatives **17c** [¹²⁵I] and **17d** [¹²⁵I] showed good brain uptake/penetration (2.74% and 4.58% ID g⁻¹ (injected dose per gram) respectively; 2 minutes after intravenous injection), clearance rates (0.40% and 0.45% ID g⁻¹ respectively; 60 minutes) and labeling of SPs with as intense signals as those of 4-(6-iodoimidazo[1,2-*a*]pyridin-2-yl)-*N,N*-dimethylaniline (IMPA). They were also more stable in human brain homogenates (**17c** [¹²⁵I]; 98% for 60 min) compared to those in mice (**17c** [¹²⁵I]; 13% for 30 min).⁶⁷

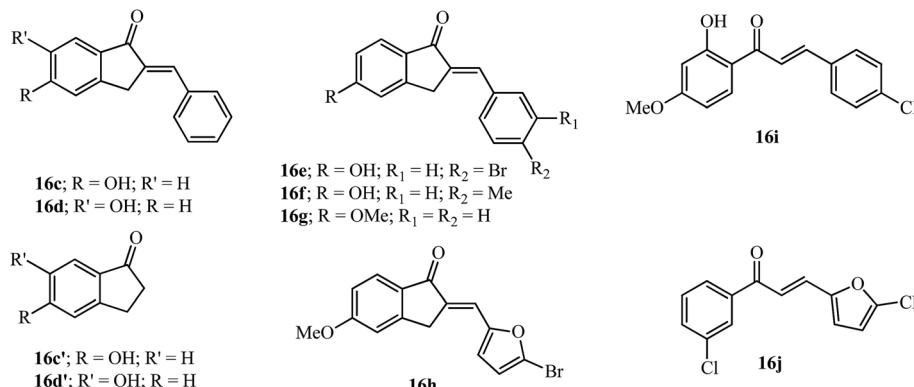


Fig. 6 AIs and chalcones tested as inhibitors of MAO-A/B.

Table 2 IC₅₀ of AIs evaluated for MAO-A/B inhibition

Compound	milog P	IC ₅₀ MAO-A (μM)	IC ₅₀ MAO-B (μM)
16c	3.24	1.94	0.376
16d	3.24	0.440	2.42
16c'	1.22	61.7	>100
16d'	1.22	64.7	85.4
16e	4.05	1.05	0.0053
16f	3.69	3.22	0.0052
16g	3.77	NI ^a	0.0092
16h	3.79	0.183	0.0044
16i	4.46	—	0.0044
16j	4.34	28.6	0.174

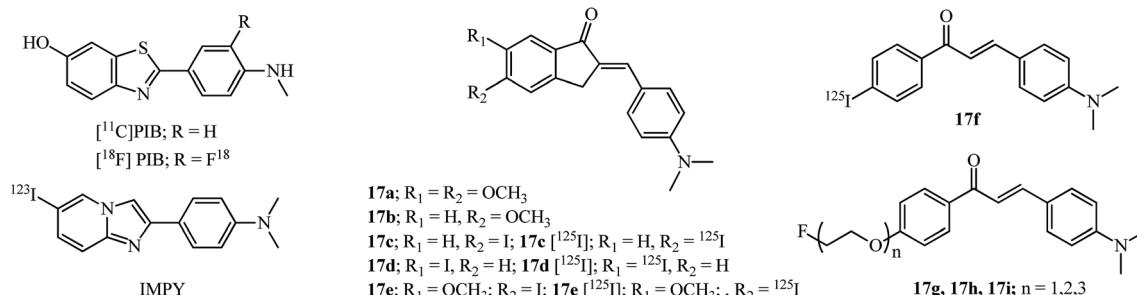
^a NI-no inhibition at maximum tested concentration of 100 μM.

Modification of the same scaffold of **17a–17d**, with the incorporation of a methoxy group at the 6-position and iodine at the 5-position, improved the binding affinity ($K_i = 28.63$ nM) of **17e**, in comparison to **17c** and **17d** (see discussion in the previous paragraph and Fig. 7). Furthermore, **17e** showed good brain uptake/penetration (5.29% ID g⁻¹; 2 minutes after intravenous injection) which can be explained by the increase of lipophilicity due to the methoxy group and correlated with the partition coefficient value of log P 3.45. The stability of radio-iodinated **17e** in human AD brain homogenates was higher (**17e** [¹²⁵I]; 98% for 60 min) compared to those in mice (**17e** [¹²⁵I]; 9.9% for 30 min).⁶⁸

The radio labelled chalcone **17f** having similar substitution of *N,N*-dimethyl and an iodine group at the 4 and 4'-positions also showed excellent binding affinities ($K_d = 4.2 \pm 1.1$ nM), rapid uptake (2.04% ID g⁻¹; 2 minutes after intravenous injection) and clearance (0.40% ID g⁻¹; 30 min) from the brain.⁶⁹ Based on the same substitution pattern fluoropolyethyleneglycol (PEG) chalcones **17g**, **17h** and **17i** were evaluated for detection of plaques and showed binding affinities (inhibition constant; $K_i = 20\text{--}50$ nM), [¹¹C] radio labelled **17i** had rapid uptake (4.31% ID g⁻¹; 2 minutes after intravenous injection) and clearance (0.35% ID g⁻¹; 60 min) from mice brains and [¹⁸F] radio labelled chalcone **17i** intensely labeled existing amyloid plaques in the brain tissues from AD patient thus having potential as positron emission computed tomography (PET) probe.⁷⁰

3. Arylidene indanones as inhibitors of tubulin assembly

Microtubules are dynamic polymers, stiff hollow cylindrical structures with an outer diameter of about 25 nm and a wall approximately 5 nm thick, formed from globular protein subunits of α -tubulin and β -tubulin of about 55 kDa each that are tightly bound together by noncovalent bonds.⁷¹ They assemble/disassemble continuously in the cell filling the cytoplasm and are equipped to transmit signals throughout the cell and interact with signaling proteins and organelles. The

Fig. 7 Amyloid imaging probes and potential AIs and chalcones as imaging probes for β -amyloid plaques.

nucleation–elongation pathway for the polymerization of $\alpha\beta$ -tubulin dimers occurs by forming a short polymer nucleus to which subunits are added non-covalently. The association of tubulin subunits to the growing polymer is faster compared to the depolymerization and at the steady state; there is a balance between these two processes. This dynamic steady state, wherein the microtubules alternate between a period of formation and disassembly is termed dynamic instability. Chemical agents interact with microtubules promoting either disassembly or assembly, thereby blocking microtubule dynamics, modifying biological processes and signaling pathways and leading ultimately to apoptosis.⁷¹

Indanocine **18** was identified as a potent microtubule-depolymerizing agent with anti-proliferative activity by the National Cancer Institute's (NCI) Therapeutic Program (see Fig. 8).²⁵ Indanocine **18** blocked tubulin polymerization, induced apoptosis in stationary phase multidrug resistant cancer cells and didn't affect the viability of non-proliferating cells.²⁵ Assessment of the *in vivo* antitumor activity of **18** was hampered due to poor water solubility (soluble in DMSO up to 20 mM and up to 100 μ M in aqueous solutions). This solubility issue was overcome by designing the 4-phosphate analog **18a**, having similar *in vitro* activity but reduced comparatively to **18** which is maybe due to the size of a phosphate group, low penetration of the drug in cell membranes and dephosphorylation kinetics.⁷²

The importance and structure–activity relationships of the 4'-hydroxy and 3',5'-dimethyl substitution in the arylidene moiety and amino group in the 7-position were confirmed by designing appropriate derivatives (24 examples) and using them against Jurkat cells.⁷³ A docking based study of various agents that bind to the colchicine site on tubulin was evaluated to get a seven point structure-based pharmacophore which is required for effective binding. The red, pink and yellow spheres (hydrogen bond acceptors; see Fig. 8) and blue sphere (hydrophobic center) mark the similar type of binding sites on the colchicine molecule as well as on indanocine **18**.⁷⁴ The overall geometric and steric requirements of binding with the colchicine molecule were satisfied by the rigid portion of the indanocine scaffold which is caused by the hydrophobic ring A and planar ring C (**18** has six out of the seven-point pharmacophores).⁷⁴ These results fortified the SAR derived earlier for indanocine derivatives.⁷³ The theoretically calculated milog *P*

and TPSA values for **18** and its 4-phosphate derivative **18a** (see Table 3) corroborate well with the observed IC₅₀ values reinforcing the basic structural requirements. Based on these theoretical milog *P* calculations development of the mono-hydrochloride salt of **18** may have better cell penetration and bioavailability as observed for a milleepachine derivative (see **22d** in Fig. 11).⁷⁵ At a concentration of 5 nM, **18** inhibited and strongly suppressed microtubule dynamics and resulted in concentration-dependent slow wound healing of MDA-MB-231 cells in the scratch wounds healing assay indicating a reduction in the migration of cells.⁷⁶

The incorporation of indole motif as bioisostere for the dimethoxyaniline group of **18** provided with indole indanone derivative called indolocine **18b** (Fig. 8), among 8 other derivatives studied as tubulin interacting agents.⁷⁷ Indolocine **18b** tested positive against the Non-Small Cell Lung NCI-H522 cell line in a nanomolar concentration. Retention of the 4'-hydroxy and 3',5'-dimethyl substitution in the arylidene moiety of the new indolocine derivatives indicated that most derivatives were equipotent, and complemented the previous SAR⁷³ as well as a pharmacophoric model.⁷⁴ A striking resemblance to the 7-amine substitution from **18** is seen by incorporation of natural motifs like indole into the indanone structure like dilemmaones A–C (see red dashed box on structures) from South African sponge *Ectyonanchora flabellata*.⁷⁸

3.1 Potential of tri-methoxy substitution on the indanone core for inhibiting cancer cell growth *via* tubulin interaction

The tri-methoxy benzene substituted moiety is common to several important tubulin binding natural molecules like colchicine, combretastatins-A4 and A-1 and podophyllotoxin (see Fig. 8 and 9), forming the hydrophobic center and interacting with the major binding site on tubulin.⁷⁴ The red and pink spheres on combretastatin A-4 and pink and yellow spheres on podophyllotoxin are hydrogen bond acceptors (Fig. 9) while the blue sphere (hydrophobic center) marks the similar type of binding sites seen on the colchicine molecule in tubulin by pharmacophore modeling.⁷⁴ The incorporation of the 3,4,5-trimethoxyphenyl group found in ring A of colchicine into the ring A of indanone **19** and modification at the 3-position with another aryl ring having a tri-methoxy substitution pattern (see the E-ring in podophyllotoxin; Fig. 9) imparts anticancer properties to the indanone **19** scaffold against breast

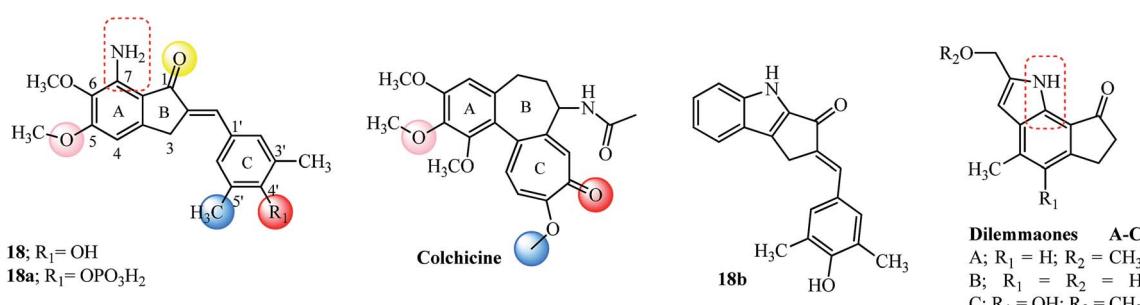


Fig. 8 Indanocine and its derivatives, colchicine as tubulin binding agents.



Table 3 IC_{50} and theoretical milog P calculation of AI as tubulin inhibitors

Compound	M. wt	milog P	n -ROTB	n -ON acceptors	n -OHNNH donors	n -Violations	Volume	TPSA	IC_{50} μ M	Cell lines
18	339.39	3.51	3	5	3	0	311.33	81.79	0.001	Jurkat
18a	419.37	2.50	5	8	4	0	356.68	128.32	30 nM	HUVEC
18b^a	303.36	4.71	1	3	2	0	277.93	53.09	0.093	NCI-H522
18.HCl^b	340.40	0.77	3	5	4	0	312.13	83.41	—	—
19	388.42	2.54	7	7	0	0	350.50	72.47	2.20 (1.88)	MCF-7 (tubulin)
19a	566.60	4.19	11	10	0	1	509.16	100.18	1.0 (3.04)	MCF-7 (tubulin)
19b	520.23	4.45	8	9	0	1	456.45	90.94	10 nM (0.63)	MCF-7 (tubulin)
19c	494.51	4.73	8	7	0	0	437.45	72.47	0.68 (0.73)	MCF-7 (tubulin)
19d	521.52	4.52	9	10	0	1	455.86	118.30	4.0 (1.54)	MCF-7 (tubulin)
PDT^c	412.44	2.31	4	7	1	0	362.25	83.47	3.5 (0.74)	MCF-7 (tubulin)

^a Non-Small Cell Lung NCI-H522. ^b Properties calculated with/without chloride counter ion mass gave same results. ^c PDT-podophyllotoxin.

cancer cells ($IC_{50} = 2.20 \mu$ M).^{79,80} The introduction of the arylidene moiety resulted in AIs of the type **19a–19c** among the 15 derivatives tested against human cancer cell lines.⁸¹ Compounds **19a**, **19b**, and **19c** were superior to the parent indanone, indicating an improvement in activity on the introduction of the arylidene moiety. Indanone **19b** inhibited tubulin polymerization in the same range when compared to podophyllotoxin (see Table 3 for comparison of IC_{50} values). Further *in vivo* acute oral toxicity tests in Swiss albino mice indicated 1000 mg kg^{-1} body weight as a single acute oral dose.⁸¹ Indanone **19b** showed antiproliferative activity against the ER + breast cancer cell line (MCF-7, $IC_{50} = 0.010 \mu$ M) and didn't allow the cells to form colonies in a concentration-dependent manner. The *in vivo* oral toxicity test using **19b** in Swiss albino mice indicated a 100 mg kg^{-1} body weight dose once in 28 days as a single acute dose and a 300 mg kg^{-1} body weight dose as a single dose.^{82,83}

Structurally the methylenedioxy group is an important moiety for cancer drug design which may interact favorably with the molecular targets in cancer cells and function as a prodrug by the action of cytochrome p450 enzymes.^{84–86} The 3,4-methylenedioxyphenyl moiety in polygamain (see Fig. 9), a natural

product isolated from *Amyris madrensis* shares the same space occupied by the tri-methoxy benzene moiety found in podophyllotoxin and shows a hydrophobic interaction within the colchicine-binding site on tubulin.⁸⁷ The presence of a fluorine atom in compound **19e** shows better experimental potency in both tests, and plays a critical role in the design of several anti-cancer agents.⁸⁸ Chalcone **19e** having a similar substitution pattern with respect to the positioning of the fluorine and tri-methoxy groups had the highest potency (30 nM) towards inhibition of NO production in RAW macrophages.⁸⁹

Further development of the indanone scaffold (7 examples) with the introduction of the nitro group in the *para*-position gave **19d** which has potent activity in several cancer cell lines and was tolerated up to 300 mg kg^{-1} doses in acute oral toxicity experiments.⁹⁰ In spite of **19d** having an almost similar molar mass and calculated theoretical volume as **19b**, it violates Lipinski rule but was found to effectively cross the blood–brain barrier due to its lipophilic nature (milog P -4.52; Table 3).⁹⁰

Recently AI incorporating methoxy groups in the indanone core similar to the colchicine A-ring were evaluated in several human cancer cell lines as antitumor agents (18 examples). The studies showed that AI with 4,5,6-trimethoxyl substitution

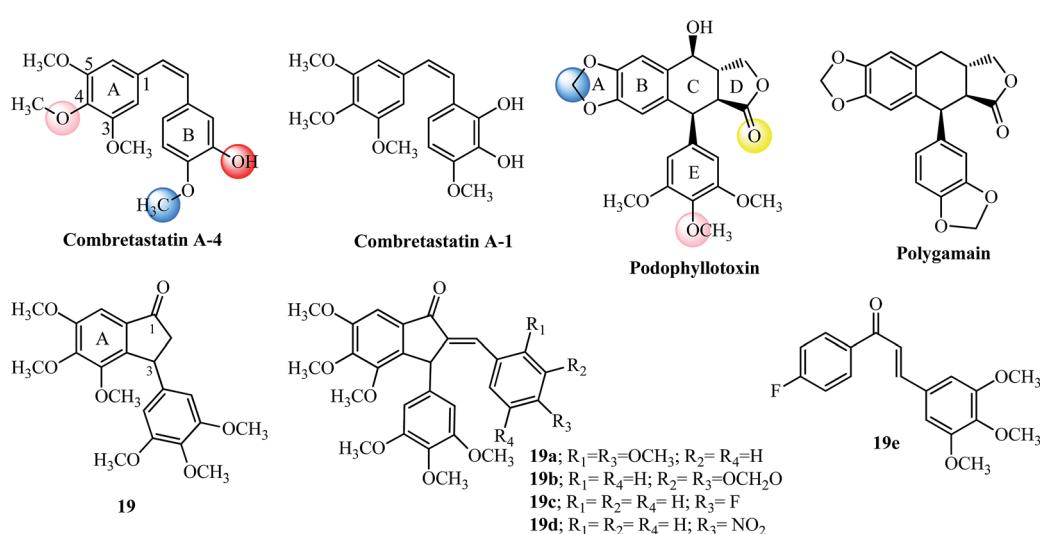


Fig. 9 Natural anti-tubulin agents and AI designed based on tri-methoxy substitution.



similar to the colchicine A-ring (**20a–20b**; Fig. 10) was more potent compared to the isomeric 5,6,7-trimethoxyl, 5-methoxy-6-hydroxy and 5,6-dimethoxy indanone moieties tested.⁹¹ The synthetic aurone derivative **20c** (an oxygen analog of active indanone **20a**; Fig. 10) gave similar results against the K562 human chronic myelogenous leukemia cell line.⁹² It is worthwhile pointing out that the most active indanone derivative **20a** shares the same tri-methoxy and isovanillin type substitution pattern as synthetic aurone and chalcone SD400 (ref. 92 and 93) (see Fig. 10 for structures and a comparison of the colored spheres with the binding sites seen on the colchicine molecule in tubulin by pharmacophore modelling⁷⁴ in Fig. 8). Several trimethoxy substituted chalcones among others with diverse substitution patterns and the ability to inhibit tubulin have been reviewed.¹⁰

Incorporating medicinally important scaffolds like indoles⁹⁴ with 4,5,6-trimethoxy indanones gave hybrid compounds with promising anti-cancer activity (14 examples). The linkage with the indole moiety was varied between the 3, 4 and 5 positions. The potent compounds (**21a–21b** and **22a–22b**; Fig. 11) gave promising results in several cancer cell lines and inhibited tubulin with no influence of the position of contact with the indole moiety.⁹⁵ Similarly, millepachine-indolyl chalcone **22c** with substitution at the 5-position of *N*-methyl indole was the most potent among others and showed cytotoxicity against all tested cancer cells.⁹⁶ The bioavailability of millepachine derivative **22d** was improved by 47% after forming its hydrochloride salt.⁷⁵ This rationale may also increase the bioavailability of derivative **20b** which is a potent anti-tubulin agent, equipotent with **20a** (see Fig. 10) and has structural similarity with the aryl ring in **22d**.

Several chalcone derivatives attached to the 3-position of the indole ring have shown induction of methuosis, a non-apoptotic cell death,⁹⁷ use as microtubule depolymerizing agents^{98,99} and have anti-inflammatory activities¹⁰⁰ while when connected to the 5-position of indole ring function as imaging agents by competitively binding to the A β fibers for detection of β -amyloid plaques in the brains of AD patients.¹⁰¹

4. Therapeutic potential of AI against breast cancer

Aromatase is a cytochrome P450 enzyme, which functions as a rate-limiting enzyme in the conversion of androgens to estrogens. Nonsteroidal aromatase inhibitors of the type; 2-(4-

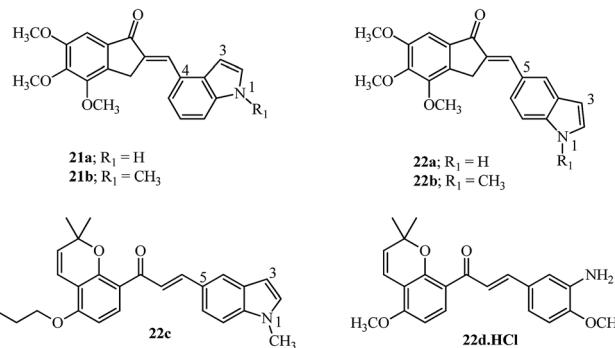


Fig. 11 Indolyl-indanones, indolyl-millepachine hybrid, and millepachine hydrochloride.

pyridylmethylene)-1-indanone **23** (Fig. 12) showed selectivity and increased potency toward aromatase as compared to aminoglutethimide.^{102,103} On similar lines, the relative potency of imidazolyl substituted derivative **24** against aromatase inhibition was 54 times more potent than aminoglutethimide (4 examples) and fared better in comparison with indanone **23**. This potency may be attributed to the orientation of the side chain which influences the binding at the active site of aromatase.¹⁰⁴ 2-Benzylidene-4,7-dimethyl indan-1-one **25** showed inhibition against the MCF-7 cell line and was non-toxic to the normal breast cell line HBL-100 up to 100 μ M.¹⁰⁵ The 2-arylidene-1-indanones **25** and **26** (Fig. 12) were evaluated in MCF-7 cells showing superior activity (7.3 μ M) in a dose-dependent manner.¹⁰⁶

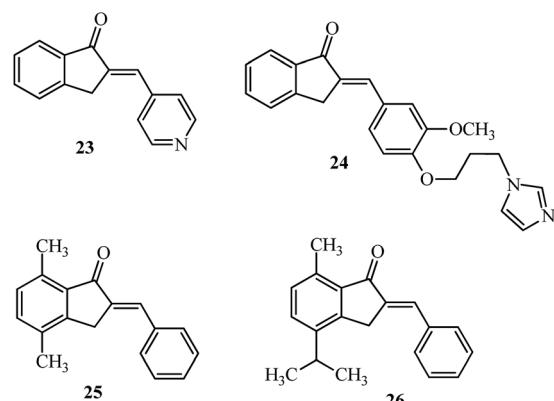


Fig. 12 AI and their derivatives inhibiting aromatase, MCF-7 cell growth and lung cancer.

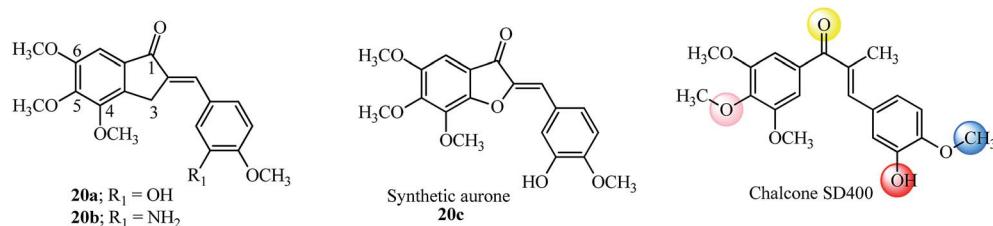


Fig. 10 Anti-proliferative AI, aurone and chalcone SD400 based on colchicine tri methoxyl substitution.



5. Arylidene indanones in treatment of lung cancer

Akt activation and phosphorylation is the key to the survival of cancer cells and has been linked to lung cancer resistant to chemotherapy. The over-expression of Akt provides resistance to chemotherapy and makes it an interesting therapeutic tool for developing anti-cancer agents. Indan-1-one **25** inhibited Akt enzyme activity by 0.20 μ M and did not alter the IC_{50} value on increasing the ATP concentration, indicating allosteric binding of the molecule to the enzyme. Indan-1-one **25** reduced cell proliferation of non-small cell lung cancer cell lines, NCI-H 460 and A549 cells with a GI_{50} of 0.28 μ M and 0.77 μ M *via* the caspase-activated apoptosis pathway.¹⁰⁷

6. Arylidene indanones in treatment of leukemia

The therapeutic effects of natural and synthetic flavonoids including chalcones with respect to structure activity relationships on various forms of leukemia has been recently reviewed.¹⁰⁸ AIs being the rigid cousins of chalcones also play important roles in the therapy of leukemia. Indanocine **18** (Fig. 8) a potent tubulin binding agent, induces apoptosis in malignant B cells from patients with chronic lymphocytic leukemia (CLL) but remains ineffective in resistant CEM human lymphoblastoid cells due to a single point mutation of the nucleotide close to the colchicine binding site of tubulin.²⁶ The combination of an equimolar concentration of **18**, an anti-tubulin agent and mizoribine, a prodrug, synergistically led to a decrease in the viability of chronic lymphocytic leukemia (CLL) cells.¹⁰⁹

7. Arylidene indanones as dual specificity phosphatase (DUSP) inhibitors

Dual specificity phosphatases (DUSP) are enzymes that dephosphorylate and inactivate mitogen-activated protein kinases (MAPKs). DUSP6, also known as mitogen-activated protein kinase phosphatase 3 (MKP3), functions as a feedback regulator of fibroblast growth factor (FGF) signaling to limit the activity of extracellular signal-regulated kinase (ERK) 1 and 2. The catalytic activity of members of the DUSP family like DUSP6 and DUSP1 is substrate specific which on interaction with ERK is enhanced, while DUSP5 is active and independent of substrate binding.²⁸ FGFs play crucial roles in development, proliferation, and cellular homeostasis.²⁸ These FGFs also bind to tyrosine kinase receptor proteins to activate the signaling of the rat sarcoma homolog RAS/MAPK cascade among others.¹¹⁰

BCI **27** was identified as a specific inhibitor of DUSP6 and DUSP1 but not of DUSP5 using a zebrafish chemical screen which is an animal model well suited for small molecule screening (Fig. 13).^{28,29} BCI **27** at a concentration of 10 μ M,

functions as an inhibitor of DUSP6 with increased expression of FGF in treated embryos. Furthermore, using analogs **27a** and **27b** it was shown that cyclohexylamine, as well as the benzylidene moiety, plays an important role in the inhibition (compare EC_{50} values in Table 4).²⁸

A total of 11 compounds (from 29 examples) effectively activated FGF signaling in a concentration-dependent manner in *Tg(dusp6:EGFP)^{P16}* embryos with most of them being as potent as the starting pharmacophore **27**.²⁹ Toxicity studies of the compounds indicated BCI analog **28a** showed no toxicity even at twice the EC_{50} (20 μ M) required for activation of FGF signaling indicating an attractive and clean molecule to investigate DUSPs.²⁹ The theoretical milog *P* calculations (Table 4) provide results which would disqualify the active molecules **27** and **28a** but substantiate the result for **28b**, which follows the Lipinski "rule of five" with no violations. However, it is not a necessity to qualify the "rule of five" to have drug-likeness.¹¹¹

The study revealed the more potent derivative **28b** due to the introduction of a heteroatom in the ring at the 3-position of indanone and the fluoro substituent in **28c** compared to **27**.²⁹ The importance of the carbonyl functionality was assayed by testing its reduced form as hydroxyl derivative **28d** (Fig. 12), which showed no activity.²⁹

The selective DUSP6 inhibitor, indanone **27**, induced cell death in patient-derived pre-B acute lymphoblastic leukemia (ALL) cells and also overcame mechanisms of drug-resistance. Treatment of patient-derived ALL cells led to hyperactivation of Erk and compromised the negative control of Erk signaling causing cell death.¹¹²

8. Quinolinylmethylidene indanones as antimalarial agents

Malaria is an infectious disease which causes deaths worldwide and is caused by transmission of *Plasmodium* strains by mosquito bites. Natural chalcones, like licochalcone A isolated from Chinese licorice roots protected mice infected with *P. yoelii* for 3 to 6 days.¹¹³ The quinoline moiety is important in drugs like chloroquine used for the treatment of malaria infections.¹¹⁴ Hybrids resulting from substituted indanones and quinolines led to indanones **29a** and **29b** (Fig. 14) as potent antimalarial agents (13 examples).³⁰ SAR studies indicated that monosubstitution in the quinoline ring at position 8 were effective inhibitors of haem polymerization (**29a** and **29b**). The *in vivo* tests using mice infected with *Plasmodium berghei* ANKA, showed survival up to 12 days after infection by treatment with indanone **29b** (at a concentration of 20 mg kg⁻¹ intraperitoneal once daily for 4 days post infection).³⁰

The SAR was carried out by substitution in the indanone ring at positions 5 and 7 by a methoxy group as well as positions 6, 7 and 8 of the quinoline moiety (10 examples). *In vivo* tests in mice infected with *Plasmodium berghei*, showed a survival rate of 11 days for indanones **30a** and **30b** (at concentration of 20 mg kg⁻¹ intraperitoneal once daily for 4 days post infection).³¹



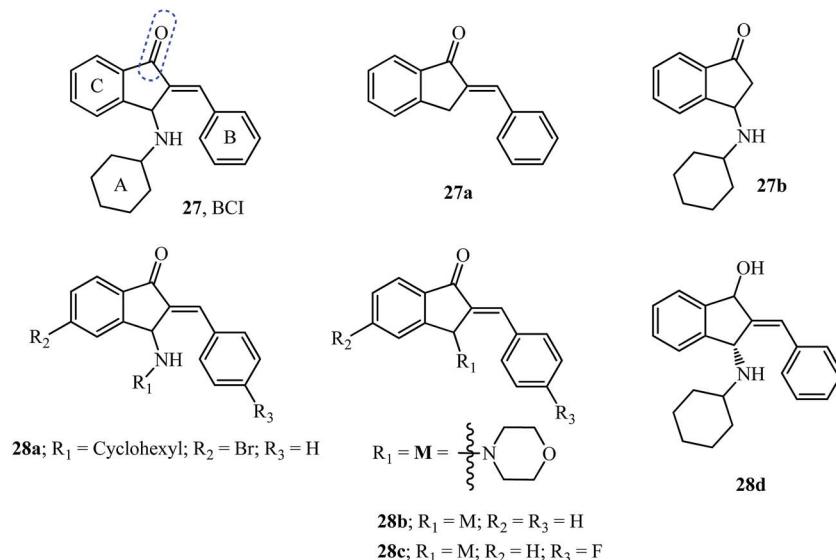


Fig. 13 BCI and its analogs tested as DUSP inhibitors.

9. Arylidene indanone as an aryl hydrocarbon receptor antagonist

Recently, the simple *E/Z*-2-benzylindene-5,6-dimethoxy-3,3-dimethylindan-1-one, **31** (Fig. 15) was shown to transiently act as an aryl hydrocarbon receptor (AhR) antagonist and selectively inhibit the UVB-induced expression of the CYP1A1 gene in normal human epidermal keratinocytes (NHEK). Results of the topical application of ointment containing 0.5% **31** protected human skin from UV-B radiation by reduced expression of CYP1A1, COX-2 and MMP-1 genes.¹¹⁵

10. Bioavailability and SAR considerations

Although no bioavailability studies are reported for the AIs mentioned in this review, a concise description of the analysis, degradation and metabolism of acetylcholinesterase inhibitors *i.e.*, donepezil, the most structurally related molecule to the AI scaffold has been reviewed.¹¹⁶ The current status of clinical trials using disease modifying drugs in AD has also been

reviewed.¹¹⁷ The increased lipophilicity of 6-substituted derivatives **12b**, **13a–13c** which were equipotent to **6** (Table 1) may improve their bioavailability by delaying metabolic demethylation.¹¹⁶

Bioavailability of chalcone and arylidene indanone was shown to be dependent on the formation of ionized species of molecules (pK_a) which have hydroxyl or *N,N*-dimethyl amino groups, which may inhibit their passive diffusion across cell membranes.¹¹⁸ The dimethylamino containing derivatives remain uncharged at pH 7.4; therefore, they may easily pass through the cellular membrane while the hydroxyl group may get ionized with the negative charge which may inhibit their diffusion. Inclusion complexes with cyclodextrins have shown great potential for drug delivery systems by increasing their bioavailability.^{119,120}

Although, the 4'-phosphate derivative of indanocine showed reduced cytotoxicity compared to **18**,⁷² the phosphate derivatives of combretastatin-like chalcones have functioned as pro-drugs and were equipotent to the parent molecules with respect to cell proliferation. While the same compounds were inactive for tubulin inhibition probably due to the *in vitro* assay not being cell-based and no hydrolysis by phosphatases can occur.⁹³

Table 4 EC₅₀ for FGF activation, IC₅₀ of DUSP6 and DUSP1 inhibition and theoretical milog P calculation of AI tested as DUSP inhibitors

Compound	Mol. weight	milog P	<i>n</i> -ROTB	<i>n</i> -ON acceptors	donors	<i>n</i> -Violations	Volume	TPSA	EC ₅₀ μM ^a	IC ₅₀ of DUSP6 inhibition	IC ₅₀ of DUSP1 inhibition
27	317.43	5.21	3	2	1	1	310.24	29.10	10.6 ± 0.8	13.3 ± 1.8	8.0 ± 0.6
27a	220.27	3.74	1	1	0	0	207.81	17.07	Inactive	ND	ND
27b	229.32	3.19	2	2	1	0	228.81	29.10	Inactive	ND	ND
28a	396.33	6.00	3	2	1	1	328.12	29.10	12.0 ± 3.0	55.9 ± 8.6	28.6 ± 3.8
28b	305.38	3.40	2	3	0	0	285.97	29.54	4.5 ± 0.6	50.8 ± 4.1	28.4 ± 1.8
28c	323.37	3.56	2	3	0	0	290.90	29.54	10.3	ND	ND
28d	319.45	4.88	3	2	2	0	316.10	32.26	>>25	ND	ND

^a EC₅₀-concentration required for FGF activation in zebrafish assay; ND-not determined.



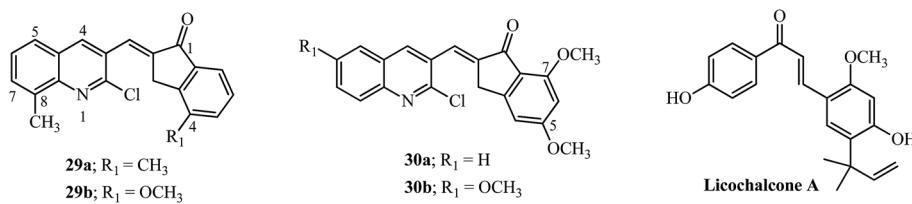


Fig. 14 Quinolinylmethylidene indanones and licochalcone A as antimarial agents.

Several combretastatin analogs like A-1 and A-4 phosphates are in various stages of clinical trials.¹²¹ The ethoxy derivative had a better half-life compared to its methoxy derivative and also improved the potency by delaying possible metabolic inactivation.¹²² The lipophilic acetate derivative of the natural flavonoid quercetin increased uptake in cells and delayed metabolism.¹²³

Inhibition of P-glycoprotein (P-gp), the drug efflux transporter in cancer cells can help increase the bioavailability of anti-cancer drugs. Natural flavonoids have been shown to be inhibitors of P-gp and have been co-administered with anti-cancer drugs.¹⁰⁸ Indanone derivatives like 32a and 32b have shown inhibition activity in the nanomolar range towards efflux of daunomycin using the CCRF vcr1000 cell line (Fig. 15).¹²⁴ The SAR provides insights into the higher activity of 32b based on the presence of one more rotatable bond than 32a while the remaining structure is identical.¹²⁴ Indanones 33a–33e have shown modulation of the ABCG2 transporter which is responsible for the efflux of a wide variety of drugs resulting in drug resistance.^{125,126} The resulting SAR indicated the presence of the dimethoxylated A-ring, adjacent to the carbonyl group and the exocyclic double bond to be key features for such activity. The activity was also dependent on the mono or disubstituted aryl ring having hydroxyl or methoxyl groups (33b–33e).¹²⁵

The majority of publications discussed in this review have a substitution at the *para* position of the arylidene ring, with *ortho* positions effectively not explored. The substitution of an *ortho* position, in comparison to the *para* isomer, can be differentiated by ESI-MS in AI as well as chalcones.^{23,127}

Moreover the *o*-substituted chalcone derivative has higher potency compared to *p*-substituted when the other structural substitutions are maintained.^{84,128} The ring B of flavopiridol and combretastatin A-1 are classical examples of *ortho* substituents influencing the potential of these scaffolds as therapeutic drugs.¹²¹ The potency of combretastatin derivatives having a 2-amino substitution in the B-ring was improved as inhibitors of tubulin assembly, cytotoxicity and *in vivo* reduction of blood flow to tumors.¹²⁹

Incorporation of the *N*-methyl-piperidinyl alkaloid moiety found in flavopiridol without the hydroxyl group in chalcones 34 has shown anti-cancer properties by inhibiting cell cycle progression^{11,130} and can be explored on the AI scaffold. Incorporation of indole or quinoline moieties with indanone may be beneficial in the design of new antimicrobial or antimarial agents,¹¹⁴ which may become more effective for resistant strains of bacteria on quaternization of nitrogen and improved solubility in the physiological medium.¹³¹

11. Conclusions

Arylidene indanones tested as acetylcholinesterase inhibitors have shown that the 6-position of the indanone core is important for interactions with the PAS of the enzyme⁴⁵ and subsequent modifications improve the efficiency of interaction.^{45,49,51} Chelation of metals like copper is critical for Alzheimer's therapy and chelating groups like piperidinyl ethoxy mimicking clioquinol in 12b or the catechol moiety in 16a serve as an

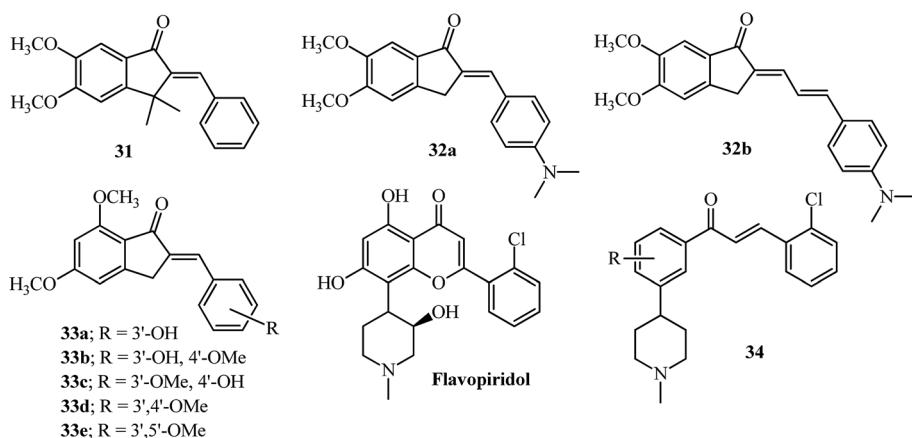


Fig. 15 Aryl hydrocarbon Receptor (AhR) antagonist, p-glycoprotein and ABCG2 inhibitors based on AI; flavopiridol and its chalcone derivative as inhibitors of cell cycle progression.



important aspect for new drug design.^{27,49} Incorporation of bulky groups like bis(4-fluorophenyl)methyl on piperazine either on the indanone side **11c** or arylidene ring **14a** can modulate either as a BACE1 inhibitor or acetylcholinesterase inhibitor.^{46,48} Tubulin depolymerization agents require the trimethoxy substitution on the indanone ring to mimic the colchicine type activity.^{74,80,81,91,95}

Tunability of the α,β -unsaturated ketone system as a Michael acceptor is paramount to the type of biological activity,¹³² since these systems are present in many natural products like chalcones as well as AI, where the molecule is more planar comparatively and the transmission of electronic effects of the aryl substituents are directed towards the carbonyl group.¹³ The importance of the double bond at the 2-position in arylidene indanone has been verified by SAR studies, indicating no DUSP inhibition activity if removed²⁸ or improved tubulin activity compared to unsubstituted compounds^{79,81} or diminished AChE activity if removed or hydrogenated.^{49,52} Similarly increased MAO-B inhibition was seen for benzylidene substituted 5-hydroxyindanones.^{61,63}

The chalcones have been merited with a lot of attention by structural modification of the position of the hydroxyl, methoxyl and other substitutions such as incorporation of heteroaryl moieties like coumarins, naphthyl, furan *etc.* displaying diverse activities.^{10–12} Similarly, in-depth studies of novel arylidene indanone scaffolds by pharmacophore modeling may give better insights into the unexplored potential of this scaffold.

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References

- 1 A. I. Syrchina and A. A. Semenov, *Chem. Nat. Compd.*, 1983, **18**, 1–11.
- 2 N. V. Kovganko, Z. N. Kashkan and S. N. Krivenok, *Chem. Nat. Compd.*, 2004, **40**, 227–229.
- 3 X. Ge, G. Ye, P. Li, W.-J. Tang, J.-L. Gao and W.-M. Zhao, *J. Nat. Prod.*, 2008, **71**, 227–231.
- 4 B. Jaki, J. Heilmann and O. Sticher, *J. Nat. Prod.*, 2000, **63**, 1283–1285.
- 5 D. G. Nagle, Y.-D. Zhou, P. U. Park, V. J. Paul, I. Rajbhandari, C. J. G. Duncan and D. S. Pasco, *J. Nat. Prod.*, 2000, **63**, 1431–1433.
- 6 H. Sheridan, N. Frankish and R. Farrell, *Eur. J. Med. Chem.*, 1999, **34**, 953–966.
- 7 H. O. Saxena, U. Faridi, J. K. Kumar, S. Luqman, M. P. Darokar, K. Shanker, C. S. Chanotiya, M. M. Gupta and A. S. Negi, *Steroids*, 2007, **72**, 892–900.
- 8 Z. Nowakowska, *Eur. J. Med. Chem.*, 2007, **42**, 125–137.
- 9 M. Das and K. Manna, *J. Toxicol.*, 2016, **2016**, 7651047.
- 10 B. Zhou and C. Xing, *Med. Chem.*, 2015, **5**, 388–404.
- 11 A. Boumendjel, X. Ronot and J. Boutonnat, *Curr. Drug Targets*, 2009, **10**, 363–371.
- 12 P. Singh, A. Anand and V. Kumar, *Eur. J. Med. Chem.*, 2014, **85**, 758–777.
- 13 P. Perjési, A. Perjéssy, E. Kolehmainen, E. Ősz, M. Samaliková, J. Linnanto and E. Virtanen, *J. Mol. Struct.*, 2004, **697**, 41–47.
- 14 P. Perjési, J. Linnanto, E. Kolehmainen, E. Ősz and E. Virtanen, *J. Mol. Struct.*, 2005, **740**, 81–89.
- 15 K. Fodor, V. Tomescova, T. Kőszegi, I. Kron and P. Perjési, *Monatsh. Chem.*, 2011, **142**, 463–468.
- 16 P. Perjési, *Monatsh. Chem.*, 2015, **146**, 1275–1281.
- 17 S. Ducki, *Anti-Cancer Agents Med. Chem.*, 2009, **9**, 336–347.
- 18 N. J. Lawrence, R. P. Patterson, L.-L. Ooi, D. Cook and S. Ducki, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 5844–5848.
- 19 S. P. Kamat, J. C. Menezes, B. M. Siddhaye and S. K. Paknikar, *Indian J. Chem.*, 2008, **47B**, 1597–1599.
- 20 V. Premasagar, V. A. Palaniswamy and E. J. Eisenbraun, *J. Org. Chem.*, 1981, **46**, 2974–2976.
- 21 M. Oliverio, M. Nardi, P. Costanzo, L. Cariati, G. Cravotto, S. V. Giofrè and A. Procopio, *Molecules*, 2014, **19**, 5599–5610.
- 22 A. Lévai, *ARKIVOC*, 2004, 15–33.
- 23 J. C. J. M. D. S. Menezes, J. A. S. Cavaleiro and M. R. M. Domingues, *Rapid Commun. Mass Spectrom.*, 2013, **27**, 2461–2471.
- 24 R. Mondal, T. K. Mandal and A. K. Mallik, *Org. Chem. Int.*, 2012, 456097.
- 25 L. M. Leoni, E. Hamel, D. Genini, H. Shih, C. J. Carrera, H. B. Cottam and D. A. Carson, *J. Natl. Cancer Inst.*, 2000, **92**, 217–224.
- 26 X. H. Hua, D. Genini, R. Gussio, R. Tawatao, H. Shih, T. J. Kipps, D. A. Carson and L. M. Leoni, *Cancer Res.*, 2001, **61**, 7248–7254.
- 27 L. Huang, C. Lu, Y. Sun, F. Mao, Z. Luo, T. Su, H. Jiang, W. Shan and X. Li, *J. Med. Chem.*, 2012, **55**, 8483–8492.
- 28 G. Molina, A. Vogt, A. Bakan, W. Dai, P. Q. D. Oliveira, W. Znosko, T. E. Smithgall, I. Bahar, J. S. Lazo, B. W. Day and M. Tsang, *Nat. Chem. Biol.*, 2009, **5**, 680–687 and references cited therein.
- 29 V. N. Korotchenko, M. Saydmohammed, L. L. Vollmer, A. Bakan, K. Sheetz, K. T. Debiec, K. A. Greene, C. S. Aglioni, I. Bahar, B. W. Day, A. Vogt and M. Tsang, *ChemBioChem*, 2014, **15**, 1436–1445.
- 30 J. E. Charris, J. N. Domínguez, N. Gamboa, J. R. Rodrigues and J. E. Angel, *Eur. J. Med. Chem.*, 2005, **40**, 875–881.
- 31 J. E. Charris, G. M. Lobo, J. Camacho, R. Ferrer, A. Barazarte, J. N. Domínguez, N. Gamboa, J. R. Rodrigues and J. E. Angel, *Lett. Drug Des. Discovery*, 2007, **4**, 49–54.
- 32 <http://www.molinspiration.com>.
- 33 C. A. Lipinski, F. Lombardo, B. W. Dominy and P. J. Feeney, *Adv. Drug Delivery Rev.*, 1997, **23**, 3–25.
- 34 H. W. Querfurth and F. M. LaFerla, *N. Engl. J. Med.*, 2010, **362**, 329–344.



35 A. Kumar, A. Singh and Ekavali, *Pharmacol. Rep.*, 2015, **67**, 195–203.

36 E. Giacobini, *Pharmacol. Res.*, 2004, **50**, 433–440.

37 M. Singh, M. Kaur, H. Kukreja, R. Chugh, O. Silakari and D. Singh, *Eur. J. Med. Chem.*, 2013, **70**, 165–188 and references cited therein.

38 Y. Kawakami, A. Inoue, T. Kawai, M. Wakita, H. Sugimoto and A. J. Hopfinger, *Bioorg. Med. Chem.*, 1996, **4**, 1429–1446.

39 Current Alzheimer's Treatments, http://www.alz.org/research/science/alzheimers_disease_treatments.asp, accessed 13 October, 2015.

40 P. Camps, X. Formosa, C. Galdeano, T. Gómez, D. Muñoz-Torrero, M. Scarpellini, E. Viayna, A. Badia, M. V. Clos, A. Camins, M. Pallas, M. Bartolini, F. Mancini, V. Andrisano, J. Estelrich, M. Lizondo, A. Bidon-Chanal and F. J. Luque, *J. Med. Chem.*, 2008, **51**, 3588–3598.

41 P. Camps, X. Formosa, C. Galdeano, T. Gómez, D. Muñoz-Torrero, L. Ramírez, E. Viayna, E. Gómez, N. Isambert, R. Lavilla, A. Badia, M. V. Clos, M. Bartolini, F. Mancini, V. Andrisano, A. Bidon-Chanal, Ó. Huertas, T. Dafni and F. J. Luque, *Chem.-Biol. Interact.*, 2010, **187**, 411–415.

42 K. S. T. Dias and C. Viegas-Junior, *Curr. Neuropharmacol.*, 2014, **12**, 239–255.

43 N. Guzior, A. Więckowska, D. Panek and B. Malawska, *Curr. Med. Chem.*, 2015, **22**, 373–404.

44 F. Mao, J. Yan, J. Li, X. Jia, H. Miao, Y. Sun, L. Huang and X. Li, *Org. Biomol. Chem.*, 2014, **12**, 5936–5944.

45 S. Rizzo, M. Bartolini, L. Ceccarini, L. Piazz, S. Gobbi, A. Cavalli, M. Recanatini, V. Andrisano and A. Rampa, *Bioorg. Med. Chem.*, 2010, **18**, 1749–1760.

46 A. Rampa, F. Mancini, A. D. Simone, F. Falchi, F. Belluti, R. M. C. D. Martino, S. Gobbi, V. Andrisano, A. Tarozzi, M. Bartolini, A. Cavalli and A. Bisi, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 2804–2808.

47 H.-R. Liu, C. Zhou, H.-Q. Fan, J.-J. Tang, L.-B. Liu, X.-H. Gao, Q.-A. Wang and W.-K. Liu, *Chem. Biol. Drug Des.*, 2015, **86**, 517–522.

48 C. B. Mishra, S. Kumari, A. Manral, A. Prakash, V. Saini, A. M. Lynn and M. Tiwari, *Eur. J. Med. Chem.*, 2017, **125**, 736–750.

49 F.-C. Meng, F. Mao, W.-J. Shan, F. Qin, L. Huang and X.-S. Li, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 4462–4466.

50 B. N. Sağlık, S. İlgin and Y. Özkar, *Eur. J. Med. Chem.*, 2016, **124**, 1026–1040.

51 L. Huang, H. Miao, Y. Sun, F. Meng and X. Li, *Eur. J. Med. Chem.*, 2014, **87**, 429–439.

52 R. Sheng, Y. Xu, C. Hu, J. Zhang, X. Lin, J. Li, B. Yang, Q. He and Y. Hu, *Eur. J. Med. Chem.*, 2009, **44**, 07–17.

53 M. L. Hegde, P. Bharathi, A. Suram, C. Venugopal, R. Jagannathan, P. Poddar, P. Srinivas, K. Sambamurti, K. J. Rao, J. Scancar, L. Messor, L. Zecca and P. Zatta, *J. Alzheimer's Dis.*, 2009, **17**, 457–468.

54 M. A. Santos, K. Chand and S. Chaves, *Coord. Chem. Rev.*, 2016, **327–328**, 287–303.

55 S. J. S. Flora and V. Pachauri, *Int. J. Environ. Res. Public Health*, 2010, **7**, 2745–2788.

56 J. Zhang, D. Zhu, R. Sheng, H. Wu, Y. Hu, F. Wang, T. Cai, B. Yang and Q. He, *Eur. J. Pharmacol.*, 2009, **613**, 1–9 and references cited therein.

57 L.-l. Shen, G.-x. Liu and Y. Tang, *Acta Pharmacol. Sin.*, 2007, **28**, 2053–2063.

58 N. Chitranshi, S. Gupta, P. K. Tripathi and P. K. Seth, *Med. Chem. Res.*, 2013, **22**, 2328–2345.

59 L. Mira, M. T. Fernandez, M. Santos, R. Rocha, M. H. Florencio and K. R. Jennings, *Free Radical Res.*, 2002, **36**, 1199–1208.

60 J. Joubert, J. P. Petzer, L. H. A. Prins, B. P. Repsold and S. F. Malan, *Drug Design and Discovery in Alzheimer's Disease*, 2014, ch. 5, pp. 291–365.

61 M. S. Nel, A. Petzer, J. P. Petzer and L. J. Legoabe, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 4599–4605.

62 M. S. Nel, A. Petzer, J. P. Petzer and L. J. Legoabe, *Bioorg. Chem.*, 2016, **69**, 20–28.

63 S. Mostert, A. Petzer and J. P. Petzer, *ChemMedChem*, 2015, **10**, 862–873.

64 F. Chimenti, R. Fioravanti, A. Bolasco, P. Chimenti, D. Secci, F. Rossi, M. Yáñez, F. Orallo, F. Ortuso and S. Alcaro, *J. Med. Chem.*, 2009, **52**, 2818–2824.

65 S. J. Robinson, J. P. Petzer, A. Petzer, J. J. Bergh and A. C. U. Lourens, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 4985–4989.

66 M. Ono and H. Saji, *Int. J. Mol. Imaging*, 2011, **2011**, 543267.

67 J.-P. Qiao, C.-S. Gan, C.-W. Wang, J.-F. Ge, D.-D. Nan, J. Pan and J.-N. Zhou, *ChemBioChem*, 2012, **13**, 1652–1662.

68 D.-D. Nan, C.-S. Gan, C.-W. Wang, J.-P. Qiao, X.-M. Wang and J.-N. Zhou, *Eur. J. Med. Chem.*, 2016, **124**, 117–128.

69 M. Ono, M. Haratake, H. Mori and M. Nakayama, *Bioorg. Med. Chem.*, 2007, **15**, 6802–6809.

70 M. Ono, R. Watanabe, H. Kawashima, Y. Cheng, H. Kimura, H. Watanabe, M. Haratake, H. Saji and M. Nakayama, *J. Med. Chem.*, 2009, **52**, 6394–6401.

71 F. Mollinedo and C. Gajate, *Apoptosis*, 2003, **8**, 413–450 and references cited therein.

72 L. M. Leoni, 2001, <http://www.dtic.mil/cgi/tr/fulltext/u2/a424665.pdf>.

73 H. Shih, L. Deng, C. J. Carrera, S. Adachi, H. B. Cottam and D. A. Carson, *Bioorg. Med. Chem. Lett.*, 2000, **10**, 487–490.

74 T. L. Nguyen, C. McGrath, A. R. Hermone, J. C. Burnett, D. W. Zaharevitz, B. W. Day, P. Wipf, E. Hamel and R. Gussio, *J. Med. Chem.*, 2005, **48**, 6107–6116.

75 Z. Yang, W. Wu, J. Wang, L. Liu, L. Li, J. Yang, G. Wang, D. Cao, R. Zhang, M. Tang, J. Wen, J. Zhu, W. Xiang, F. Wang, L. Ma, M. Xiang, J. You and L. Chen, *J. Med. Chem.*, 2014, **57**, 7977–7989.

76 S. Kapoor and D. Panda, *Biochem. Pharmacol.*, 2012, **83**, 1495–1506 and references cited therein.

77 G. A. Tunbridge, J. Oram and L. Caggiano, *MedChemComm*, 2013, **4**, 1452–1456.

78 D. R. Beukes, M. T. Davies-Coleman, M. Kelly-Borges, M. K. Harper and D. J. Faulkner, *J. Nat. Prod.*, 1998, **61**, 699–701.

79 H. O. Saxena, U. Faridi, S. Srivastava, J. K. Kumar, M. P. Darokar, S. Luqman, C. S. Chanotiya, V. Krishna,



A. S. Negi and S. P. S. Khanuja, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 3914–3918.

80 D. Chanda, S. Bhushan, S. K. Guru, K. Shanker, Z. A. Wani, B. A. Rah, S. Luqman, D. M. Mondhe, A. Pal and A. S. Negi, *Eur. J. Pharm. Sci.*, 2012, **47**, 988–995.

81 A. P. Prakasham, A. K. Saxena, S. Luqman, D. Chanda, T. Kaur, A. Gupta, D. K. Yadav, C. S. Chanotiya, K. Shanker, F. Khan and A. S. Negi, *Bioorg. Med. Chem.*, 2012, **20**, 3049–3057.

82 A. Singh, K. Fatima, A. Singh, A. Behl, M. J. Mintoo, M. Hasanain, R. Ashraf, S. Luqman, K. Shanker, D. M. Mondhe, J. Sarkar, D. Chanda and A. S. Negi, *Eur. J. Pharm. Sci.*, 2015, **76**, 57–67.

83 A. S. Negi, A. P. Prakasham, A. K. Saxena, S. Luqman, D. Chanda, T. Kaur and A. Gupta, *US Pat.*, US 8,633,242 B2, 2014.

84 B. Orlikova, J. C. J. M. D. S. Menezes, S. Ji, S. P. Kamat, J. A. S. Cavaleiro and M. Diederich, *Eur. J. Med. Chem.*, 2014, **84**, 173–180 and references cited therein.

85 S. Sale, R. G. Tunstall, K. C. Ruparelia, P. C. Butler, G. A. Potter, W. P. Steward and A. J. Gescher, *Invest. New Drugs*, 2006, **24**, 459–464.

86 R. D. Bruno and V. C. O. Njar, *Bioorg. Med. Chem.*, 2007, **15**, 5047–5060.

87 R. M. Hartley, J. Peng, G. A. Fest, S. Dakshanamurthy, D. E. Frantz, M. L. Brown and S. L. Mooberry, *Mol. Pharmacol.*, 2012, **81**, 431–439.

88 C. Isanbor and D. O'Hagan, *J. Fluorine Chem.*, 2006, **127**, 303–319.

89 J. Rojas, M. Payá, J. N. Dominguez and M. L. Ferrández, *Bioorg. Med. Chem. Lett.*, 2002, **12**, 1951–1954.

90 A. Singh, K. Fatima, A. Srivastava, S. Khwaja, D. Priya, A. Singh, G. Mahajan, S. Alam, A. K. Saxena, D. M. Mondhe, S. Luqman, D. Chanda, F. Khan and A. S. Negi, *Chem. Biol. Drug Des.*, 2016, **88**, 625–634.

91 J. Hu, J. Yan, J. Chen, Y. Pang, L. Huang and X. Li, *MedChemComm*, 2015, **6**, 1318–1327.

92 N. J. Lawrence, D. Rennison, A. T. McGown and J. A. Hadfield, *Bioorg. Med. Chem. Lett.*, 2003, **13**, 3759–3763.

93 S. Ducki, D. Rennison, M. Woo, A. Kendall, J. F. D. Chabert, A. T. McGown and N. J. Lawrence, *Bioorg. Med. Chem.*, 2009, **17**, 7698–7710.

94 N. K. Kaushik, N. Kaushik, P. Attri, N. Kumar, C. H. Kim, A. K. Verma and E. H. Choi, *Molecules*, 2013, **18**, 6620–6662.

95 J. Chen, J. Yan, J. Hu, Y. Pang, L. Huang and X. Li, *RSC Adv.*, 2015, **5**, 68128–68135.

96 G. Wang, C. Li, L. He, K. Lei, F. Wang, Y. Pu, Z. Yang, D. Cao, L. Ma, J. Chen, Y. Sang, X. Liang, M. Xiang, A. Peng, Y. Wei and L. Chen, *Bioorg. Med. Chem.*, 2014, **22**, 2060–2079.

97 M. W. Robinson, J. H. Overmeyer, A. M. Young, P. W. Erhardt and W. A. Maltese, *J. Med. Chem.*, 2012, **55**, 1940–1956.

98 A. Boumendjel, A. McLeer-Florin, P. Champelovier, D. Allegro, D. Muhammad, F. Souard, M. Derouazi, V. Peyrot, B. Toussaint and J. Boutonnat, *BMC Cancer*, 2009, **9**, 242.

99 V. Martel-Frachet, M. Keramidas, A. Nurisso, S. DeBonis, C. Rome, J.-L. Coll, A. Boumendjel, D. A. Skoufias and X. Ronot, *Oncotarget*, 2015, **6**, 14669–14686.

100 A. Özdemir, M. D. Altintop, G. Turan-Zitouni, G. A. Çiftçi, I. Ertorun, Ö. Alataş and Z. A. Kaplancıklı, *Eur. J. Med. Chem.*, 2015, **89**, 304–309.

101 M. Cui, M. Ono, H. Kimura, B. L. Liu and H. Saji, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 980–982.

102 R. W. Hartmann, H. Bayer and G. Grün, *J. Med. Chem.*, 1994, **37**, 1275–1281.

103 R. Gupta, D. P. Jindal, B. Jit, G. Narang, A. Paluszak and R. W. Hartmann, *Arch. Pharm.*, 2004, **337**, 398–401.

104 R. Bansal, G. Narang, C. Zimmer and R. W. Hartmann, *Med. Chem. Res.*, 2011, **20**, 661–669.

105 R. Prasanna and C. C. Harish, *Oncol. Res.*, 2010, **18**, 1–8.

106 R. Suresha, C. C. Kanakam, K. Dineshkumar and W. Hopper, *Am. J. Biomed. Sci. Eng.*, 2015, **1**, 32–38.

107 P. Rajagopalan, K. A. Alahmari, A. A. Elbessoumy, M. Balasubramaniam, R. Suresh, M. E. A. Shariff and H. C. Chandramoorthy, *Cancer Chemother. Pharmacol.*, 2016, **77**, 393–404.

108 J. C. J. M. D. S. Menezes, B. Orlikova, F. Morceau and M. Diederich, *Crit. Rev. Food Sci. Nutr.*, 2015, **56**, S4–S28 and references cited therein.

109 D. A. Carson, M. D. Rosenbach, C. J. Carrera and L. M. Leoni, *US Pat.*, US 2004/0096436 A1, 2004.

110 B. W. Day, M. W. Tsang, V. N. Korotchenko, *WO 2010/108058 A2*, 2010.

111 G. Vistoli, A. Pedretti and B. Testa, *Drug Discovery Today*, 2008, **13**, 285–294.

112 S. Shojaee, R. Caeser, M. Buchner, E. Park, S. Swaminathan, C. Hurtz, H. Geng, L. N. Chan, L. Klemm, W.-K. Hofmann, Y. H. Qiu, N. Zhang, K. R. Coombes, E. Paietta, J. Molkentin, H. P. Koeffler, C. L. Willman, S. P. Hunger, A. Melnick, S. M. Kornblau and M. Müschen, *Cancer Cell*, 2015, **28**, 114–128.

113 M. Chen, T. G. Theander, S. B. Christensen, L. Hviid, L. Zhai and A. Kharazmi, *Antimicrob. Agents Chemother.*, 1994, **38**, 1470–1475.

114 K. Kaur, M. Jain, R. P. Reddy and R. Jain, *Eur. J. Med. Chem.*, 2010, **45**, 3245–3264.

115 J. Tigges, T. Haarmann-Stemmann, C. F. A. Vogel, A. Grindel, U. Hübenthal, H. Brenden, S. Grether-Beck, G. Vielhaber, W. Johncock, J. Krutmann and E. Fritsche, *J. Invest. Dermatol.*, 2014, **134**, 556–559.

116 L. A. Marques, M. Giera, H. Lingeman and W. M. Niessen, *Biomed. Chromatogr.*, 2011, **25**, 278–299.

117 S. Salomone, F. Caraci, G. M. Leggio, J. Fedotova and F. Drago, *Br. J. Clin. Pharmacol.*, 2011, **73**, 504–517.

118 I. Kron, Z. Pudychová-Chovanová, B. Veliká, J. Guzy and P. Perjési, *Monatsh. Chem.*, 2012, **143**, 13–17.

119 R. L. Carrier, L. A. Miller and I. Ahmed, *J. Controlled Release*, 2007, **123**, 78–99.

120 G. Tiwari, R. Tiwari and A. K. Rai, *J. Pharm. BioAllied Sci.*, 2010, **2**, 72–79.

121 G. C. Tron, T. Pirali, G. Sorba, F. Pagliai, S. Busacca and A. A. Genazzani, *J. Med. Chem.*, 2006, **49**, 3033–3044.



122 Y.-S. Wu, M. S. Coumar, J.-Y. Chang, H.-Y. Sun, F.-M. Kuo, C.-C. Kuo, Y.-J. Chen, C.-Y. Chang, C.-L. Hsiao, J.-P. Liou, C.-P. Chen, H.-T. Yao, Y.-K. Chiang, U.-K. Tan, C.-T. Chen, C.-Y. Chu, S.-Y. Wu, T.-K. Yeh, C.-Y. Lin and H.-P. Hsieh, *J. Med. Chem.*, 2009, **52**, 4941–4945.

123 K. Sakao, M. Fujii and D.-X. Hou, *BioFactors*, 2009, **35**, 399–405.

124 Z. Parveen, G. Brunhofer, I. Jabeen, T. Erker, P. Chiba and G. F. Ecker, *Bioorg. Med. Chem.*, 2014, **22**, 2311–2319.

125 H. M. Sim, K. Y. Loh, W. K. Yeo, C. Y. Lee and M. L. Go, *ChemMedChem*, 2011, **6**, 713–724.

126 H.-M. Sim, C.-P. Wu, S. V. Ambudkar and M.-L. Go, *Biochem. Pharmacol.*, 2011, **82**, 1562–1571.

127 J. C. J. M. D. S. Menezes, J. A. S. Cavaleiro, S. P. Kamat, C. M. R. F. Barros and M. R. M. Domingues, *Rapid Commun. Mass Spectrom.*, 2013, **27**, 1303–1310.

128 P. Boeck, P. C. Leal, R. A. Yunes, V. C. Filho, S. López, M. Sortino, A. Escalante, R. L. E. Furlán and S. Zacchino, *Arch. Pharm.*, 2005, **338**, 87–95.

129 K. A. Monk, R. Siles, M. B. Hadimani, B. E. Mugabe, J. F. Ackley, S. W. Studerus, K. Edvardsen, M. L. Trawick, C. M. Garner, M. R. Rhodes, G. R. Pettit and K. G. Pinney, *Bioorg. Med. Chem.*, 2006, **14**, 3231–3244.

130 X. Liu and M.-L. Go, *Bioorg. Med. Chem.*, 2006, **14**, 153–163.

131 M. C. Jennings, K. P. C. Minbile and W. M. Wuest, *ACS Infect. Dis.*, 2015, **1**, 288–303.

132 S. Amslinger, *ChemMedChem*, 2010, **5**, 351–356.

