Near-infrared probes based on fluorinated Si-rhodamine for live cell imaging†

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The syntheses and biological applications of three Si-rhodamine probes with substituent groups on the pendant phenyl ring are reported. In solution, these Si-rhodamine probes (AZSiR) show slight aggregation. By introducing a methyl group at the 2-position of the pendant phenyl ring, the AZSiR-2 probe shows almost unchanged absorption and emission peaks, and a three times higher fluorescence quantum yield than that of AZSiR-1. However, the photostability of the AZSiR-2 probe becomes poor. By changing the substituent groups from methyl to trifluoromethyl, the AZSiR-3 probe displays slightly red-shifted absorption and emission peaks, and good photostability. Furthermore, the bulky groups on the phenyl ring of Si-rhodamine prevent nucleophilic attack through steric hindrance, and endow Si-rhodamine probes good chemical stability in nucleophilic systems. These Si-rhodamine probes have excellent live cell permeability and low cytotoxicity. Importantly, the Si-rhodamine probe with trifluoromethyl at the 2-position of the pendant phenyl ring retains high brightness and excellent stability even in a harsh physiological environment.

Introduction

Rhodamines are well-known fluorescent dyes that display water solubility, high fluorescence quantum yields and high molar extinction coefficients,1,2 and they are widely utilized in various fluorescent probes for bioimaging.1–5 However, these dyes basically have green to red fluorescence. Near-infrared (NIR) (650–900 nm) fluorescence dyes are attractive for biological applications because of minimum photo-damage to biological samples, deep tissue penetration, and minimum interference from background auto-fluorescence in the living systems.6,7 Recently, a new class of NIR silicon-substituted rhodamines, in which the O atom at the 10-position of the xanthene moiety is replaced with a SiMe2 group, have been developed for fluorescence bioimaging.5–10

Latestly, Lavis replaced these dimethyl amino groups of the classic tetramethylrhodamine with four-membered azetidines, and created so called Janelia Fluor (JF) dyes.11 This is a minor change in structure but causes a large increase in brightness and photostability. However, these fluorescent rhodamines are cationic conjugated compounds, and the nucleophilic attack on the 9th carbon atom of the xanthene ring of these rhodamine derivatives would occur easily, and induce drastic changes in optical properties.12,13 Thus, the Si-rhodamine probes with high stability are highly desired for practical images in vivo. Here, we report the NIR Si-rhodamine probes (AZSiR) with the bulky groups on benzene at 9-position of the xanthene moiety (Chart 1). The AZSiR probes possess high fluorescence quantum yields, improved stability in the nucleophilic biology system.

Results and discussion

The AZSiR probes were synthesized according to Scheme 1. 3,6-Dihydroxy-Si-xanthone (1) was obtained according to literature procedures.6 Triflation of 3,6-dihydroxy-Si-xanthone (1) afforded compound ditriflate-Si-xanthone (2). Then the important intermediate of diazetidine-Si-xanthone (3) was synthesized by compound 2 and azetidine under the catalysis of Pd2dba3 and XPhos. The azetidinyl-Si-rhodamines were synthesized by nucleophilic addition of the various kinds of freshly prepared appropriate benzene lithium solution to the diazetidine-Si-xanthone.

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Scheme 1 Reagents and conditions: (a) 3,6-dihydroxy-Si-xanthone, CH₂Cl₂, 0 °C, pyridine, trifluoromethanesulfonic anhydride, room temperature, 2 h. (b) Compound 1, azetidine, Pd₂dba₃, XPhOS, Cs₂CO₃, 1,4-dioxane, 100 °C, 4 h. (c) Appropriate bromobenzene, tert-butyllithium (1.3 M), 2-methyltetrahydrofuran –116 °C, –116 °C to rt, 2 N HCl aq.

These AZSiR probes were fully characterized by H-NMR spectroscopy, C-NMR spectroscopy, and mass spectrometry, which can be found in the ESI.† These AZSiR probes show water solubility, and high solubility in common organic solvents such as dichloromethane, tetrahydrofuran, dimethyl sulfoxide. The UV-Vis absorption spectroscopy and fluorescence spectroscopy of the AZSiR probes in dichloromethane are shown in Fig. 1, and the photophysical data are summarized in Table 1.

In dichloromethane solution, AZSiR-1 exhibited NIR absorption and emission with maxima at λₘ₃ = 652 nm and λₘ₄ = 663 nm respectively. Introducing 2-methyl group on the pendant phenyl ring, AZSiR-2 showed almost same absorption maxima (λₘ₃ = 653 nm) and emission maxima (λₘ₄ = 667 nm). Compared with those of AZSiR-2, AZSiR-3 with trifluoromethyl substituent displayed slight red-shifted absorption and emission maxima (λₘ₃ = 664 nm and λₘ₄ = 681 nm). The bathochromic electronic transitions observed for AZSiR-3 predominantly arises from a stabilization of the LUMO by the electron-withdrawing trifluoromethyl substituent. A ZSiR-1 showed relatively low fluorescence quantum yield (24%). In contrast, the fluorescence quantum yield of AZSiR-2 was significantly increased to 76% by introducing a stopper 2-methyl group on the phenyl ring, which can maintain an orthogonal relationship between the benzene moiety and the fluorophore, and reduce the energy loss by restricting rotation. Furthermore, AZSiR-3 with trifluoromethyl groups also maintained a high fluorescence quantum yield (74%).

The optical properties of AZSiR probes in dichloromethane solution were furthermore investigated via concentration-dependent steady state and transient state fluorescence spectroscopy (Fig. 1 and S2†). When the solution concentration increased from 5.0 x 10⁻⁶ to 5.0 x 10⁻⁴ mol L⁻¹, the absorption peaks and extinction coefficient of AZSiR probes did not change. However, within the same concentration range, the emission maximum peaks were continuously red-shifted by 21 nm (AZSiR-1: from 663 to 684 nm), 26 nm (AZSiR-2: from 670 to 696 nm), and 31 nm (AZSiR-3: from 681 to 712 nm). These results suggest that AZSiR probes have slight aggregation in dichloromethane solution. To further demonstrate the aggregation behavior of AZSiR probes, time resolved fluorescence measurements were performed. The curves of the fluorescence decays for three AZSiR probes were prolonged in the concentration range from 5.0 x 10⁻⁶ to 5.0 x 10⁻⁴ mol L⁻¹ (Fig. S2†). The prolonged lifetimes upon increasing concentration are attributed to the monomer transfer to aggregates. These AZSiR probes showed good solubility in PBS, which is important for probes used in biology system. Furthermore, the fluorescence quantum yields of AZSiR probes with bulky groups on 2-position of the pendant phenyl ring (AZSiR-1 and AZSiR-2) in PBS solution were lower than those in organic solution but still remained high (54–57%).

In order to compare the photostability of AZSiR probes reported here with fluorophores commonly used NIR probes, the
monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. As shown in Fig. 2B, only under nucleophilic condition, changes in absorbance were monitored upon addition of cysteine. 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position of the pendant phenyl ring are beneficial to improve fluorescence quantum yield, and prevent the nucleophilic attack for good chemical stability. Furthermore, the chemical inertness of trifluoromethyl groups endow Si-rhodamine probes better photostability. High fluorescence quantum yields, good photo-chemistry stability, live cell permeability, and low cytotoxicity make the fluorinated Si-rhodamine dyes excellent probes for live-cell imaging.

**Experimental**

**Materials and instrumentation**

All the chemicals used in synthesis are analytical pure and were used as received. UV/Vis spectra were recorded on a Shimadzu UV-2550 spectrophotometer. Fluorescence spectra were recorded on a Shimadzu RF-5301 fluorescence spectrophotometer. The H-NMR spectra and C-NMR spectra were recorded at 20 °C on 600 MHz or 150 MHz NMR spectrometer (Bruker). Mass spectra were carried out with Agilent LC/MSD XCT Trap. CLSM images were obtained using Olympus confocal laser scanning microscopy (Olympus Fluoview 1000). The fluorescence lifetimes were recorded on an Edinburgh Analytical Instruments FL980 spectrometer, equipped with a supercontinuum ultrafast fiber lasers (Fianium), using the time correlated single photon-counting (TCSPC) method. Typically, 10,000 counts were collected at the peak channel, and the decay curves were fitted by least-squares deconvolution with original Edinburgh instrument software; the quality of the parameters were judged by the reduced $\chi^2$ values and the randomness of the weighted residuals. The instrument used for HPLC analysis is Shimadzu LC-6AD high performance liquid chromatography, and the eluent were A (water : acetonitrile, v/v = 90 : 10) and B (acetonitrile : water, v/v = 90 : 10).

**Synthetic procedures**

3,6-Ditri fluoride-Si-xanthone (2). Compound 1 (300 mg, 1.11 mmol) was suspended in 10 mL of anhydrous CH$_2$Cl$_2$, and the solution was stirred at 0 °C. Pyridine (1.44 mL, 17.76 mmol, 16 eq.) was added and stirred uniformly. Triflic anhydride (1.50 mL, 8.88 mmol, 8 eq.) was added dropwise, and stirred at room temperature for 2 h. It was subsequently diluted with water and extracted with dichloromethane. The combined organic phase was washed with water and brine, dried over Na$_2$SO$_4$, filtered and evaporated. The resultant residue was quickly purified by silica gel chromatography (AcOEt/hexanes, 4 : 3, v/v) yielded 539 mg (1.01 mmol, 91%) of pure product 2 as a colorless solid.

1H-NMR (600 MHz, CDCl$_3$): $\delta$ 8.33 (d, $J$ = 8.4 Hz, 2H), 7.99 (d, $J$ = 2.4 Hz, 2H), 7.66–7.64 (dd, $J$ = 2.49 Hz, 2H), 0.46 (s, 6H); 13C-NMR (150 MHz, CDCl$_3$): $\delta$ 52.5, 30.4, 30.1, 30.0, 17.0, 1.8, 168.9, 153.5, 148.5, 148.7, 141.9, 141.2, 128.9, 128.5, 120.0, 112.4, 52.5, 30.4, 30.1, 17.0, 1.8, 0.2. MS (ESI): calcd for: C$_{28}$H$_{31}$N$_2$Si $m/z$ = 450.2, found: 450.2 [M$^+$].

AZSiR-1. In a nitrogen-flushed flask fitted with a septum cap, bromobenzene (224.5 mg, 1.43 mmol, 10 eq.) was dissolved in anhydrous 2-methyltetrahydrofuran (10 mL) and the solution was cooled to –116 °C, tert-BuLi (1.1 mL, 1.43 mmol, 10 eq.) was slowly added dropwise, and the mixture was stirred at the same for 20 minutes. Compound 3 (50 mg, 0.143 mmol, 1 eq.) in dry 2-methyltetrahydrofuran (5 mL) was slowly added dropwise via a syringe at –116 °C. Stirring was continued for 20 minutes, then the solution was warmed to room temperature, further stirred for 2 h, quenched with 2 N HCl aq., and basified with sat. NaHCO$_3$ aq. The aqueous solution was extracted with dichloromethane, and the combined organic phase was washed with water and brine, dried over Na$_2$SO$_4$, filtered and evaporated. The resultant residue was quickly purified by HPLC (eluent: A/B = 90/10 to 10/90, 40 min) to afford AZSiR-1 (20.5 mg, 35% yield) as blue solid. 1H-NMR (600 MHz, CDCl$_3$): $\delta$ 7.47–7.46 (m, 3H), 7.17–7.15 (m, 2H), 7.00 (d, $J$ = 9 Hz), 6.84 (d, $J$ = 2.4 Hz, 2H), 6.22 (q, $J$ = 2.4, 9 Hz, 2H), 4.40 (s, 8H), 2.60 (m, 4H), 0.57 (s, 6H); 13C-NMR (150 MHz, CDCl$_3$): $\delta$ 168.9, 153.5, 148.5, 148.7, 141.9, 129.9, 128.9, 128.5, 120.0, 112.4, 52.5, 30.4, 30.1, 17.0, 1.8 –0.2. MS (ESI): calcd for: C$_{27}$H$_{30}$SiN$_2$ $m/z$ = 409.2, found: 409.2 [M$^+$].

AZSiR-2. In a nitrogen-flushed flask fitted with a septum cap, 2-bromotoluene (244.6 mg, 1.43 mmol, 10 eq.) was dissolved in anhydrous 2-methyltetrahydrofuran (10 mL) and the solution was cooled to –116 °C, tert-BuLi (1.1 mL, 1.43 mmol, 10 eq.) was slowly added dropwise, and the mixture was stirred at the same for 20 minutes. Compound 3 (50 mg, 0.143 mmol, 1 eq.) in dry 2-methyltetrahydrofuran (5 mL) was slowly added dropwise via a syringe at –116 °C. Stirring was continued for 20 minutes, then the solution was warmed to room temperature, further stirred for 2 h, quenched with 2 N HCl aq., and basified with sat. NaHCO$_3$ aq. The aqueous solution was extracted with dichloromethane, and the combined organic phase was washed with water and brine, dried over Na$_2$SO$_4$, filtered and evaporated. The resultant residue was quickly purified by HPLC (eluent: A/B = 70/30 to 10/90, 50 min) to afford AZSiR-2 (22.9 mg, 38% yield) as blue solid. 1H-NMR (600 MHz, CDCl$_3$): $\delta$ 7.35 (t, $J$ = 7.2, 7.2 Hz, 1H), 7.26 (t, $J$ = 8.4, 7.8 Hz, 2H), 7.00 (d, $J$ = 7.8 Hz, 2H), 6.92 (d, $J$ = 9.6 Hz, 2H), 6.77 (d, $J$ = 2.4 Hz, 2H), 6.19 (q, $J$ = 2.4, 2.4 Hz, 2H), 4.35 (s, 8H), 2.58–2.52 (m, 4H), 1.94 (s, 3H), 0.54 (d, $J$ = 2.4, 2.4 Hz, 6H). 13C-NMR (150 MHz, CDCl$_3$): $\delta$ 168.1, 151.9, 147.0, 140.1, 129.2, 127.9, 127.8, 126.5, 124.6, 118.1, 110.9, 51.2, 28.7, 28.5, 28.3, 26.2, 21.7, 18.3, 15.1, 13.1, –1.8, –2.1. MS (ESI): calcd for: C$_{25}$H$_{31}$Si $m/z$ = 432.2, found: 432.3 [M$^+$].
AZSiR-3. In a nitrogen-flushed flask fitted with a septum cap, 2-bromobenzotrifluoride (321.8 mg, 1.43 mmol, 10 eq.) was dissolved in anhydrous 2-methyltetrahydrofuran (10 mL) and the solution was cooled to −116 °C, tert-BuLi (2.2 mL, 2.86 mmol, 20 eq.) was slowly added dropwise, and the mixture was stirred at the same for 20 minutes. Compound 3 (50 mg, 0.143 mmol, 1 eq.) in dry 2-methyltetrahydrofuran (5 mL) was slowly added dropwise via a syringe at −116 °C. Stirring was continued for 20 minutes, then the solution was warmed to room temperature, further stirred for 2 h, quenched with 2 N HCl aq., and basified with sat. NaHCO3 aq. The aqueous solution was extracted with dichloromethane, and the combined organic fraction was washed with water and brine, dried over Na2SO4, filtered and evaporated. The resultant residue was quickly purified by HPLC (elucent: A/B = 70/30 to 10/90, 50 min) to afford AZSiR-3 (19 mg, 28% yield) as blue solid. 1H-NMR (600 MHz, CDCl3): δ 7.78 (d, J = 7.8 Hz, 2H), 7.69–7.61 (m, 4H), 7.22 (t, J = 7.2, 7.2 Hz, 1H), 6.79 (q, J = 1.8, 9.6 Hz, 4H), 6.22 (q, J = 1.8, 2.4 Hz), 4.39 (s, 8H), 2.58–2.56 (m, 4H), 0.61 (s, 3H), 0.47 (s, 3H). 13C-NMR (150 MHz, CDCl3): δ 163.4, 151.7, 146.8, 140.1, 136.3, 130.8, 130.1, 128.2, 126.8, 125.6, 118.2, 110.8, 51.3, 30.9, 28.7, 21.7, 15.1, 13.1, −1.3, −2.9. MS [ESI]: calcd for: C28H28F3N2Si+ 411, found: 411. [116] This journal is © The Royal Society of Chemistry 2017

Cell culture and imaging

HeLa cells were cultured in Dulbecco’s modified Eagle’s medium (high glucose), supplemented with 10% (v/v) fetal bovine serum, penicillin (100 units per mL) and streptomycin (100 µg mL−1) at 37 °C in a 5% CO2/95% air incubator in a humidified atmosphere, and culture media were replaced with fresh media two to three days. For fluorescence imaging, HeLa cells were grown in DMEM on a 35 mm glass bottom culture dish for at least 24 h to enable adherence to the bottom.

The cells were loaded with AZSiR according to a following procedure. Briefly, the 50 µL of 100 µM AZSiR probes PBS solution was added to the dish (final concentration of AZSiR probes is 5 µM), and then the cells were incubated for 30 min at 37 °C. Afterward, the cells were washed several times with PBS for removing AZSiR. In CLSM imaging, the excitation wavelength was fixed at 635 nm and fluorescent signals were collected from 650 nm to 750 nm.

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Notes and references


