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## A general strategy for controllable synthesis of $\text{Ba}_3(\text{MO}_4)_2:\text{Mn}^{5+}$ ( $\text{M} = \text{V}, \text{P}$ ) nanoparticles

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The second near-infrared window (NIR-II, 1000–1400 nm) is ideal for fluorescence imaging owing to negligible tissue scattering and minimal autofluorescence. NIR-II luminescence nanophosphors are promising materials for fluorescence imaging.  $\text{Mn}^{5+}$  is emerging as a new luminescent indicator owing to its typical NIR-I (800 nm) to NIR-II (1190 nm) downconversion luminescence. Herein, we report a facile synthesis approach to prepare  $\text{Mn}^{5+}$  doped  $\text{Ba}_3(\text{MO}_4)_2$  ( $\text{M} = \text{V}, \text{P}$ ) nanoparticles with a remarkably sharp emission peak around 1190 nm in the NIR-II region for the first time to our knowledge. The developed two-step approach based on efficient anion exchange reaction is able to not only stabilize the valence but also keep the controlled morphology and uniform size. It is highly possible that the general approach will provide a new perspective on synthesizing special valence ion doped nanosized materials.

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### 1. Introduction

Recently, a large amount of attention has been devoted to the investigation of fluorescence imaging, which provides visible information about disease with high sensitivity and fast feedback.<sup>1,2</sup> Biological tissues such as skin and blood scatter and absorb less near-infrared (NIR) light than visible light so that the light in the NIR biological window can penetrate these tissues more efficiently.<sup>3</sup> In the past years, extensive investigation has been focused on fluorescent probes emitting in the conventional near-infrared biological window (NIR-I, wavelength of 700–1000 nm).<sup>4–6</sup> Recently, it has been demonstrated that the phosphors for emission in the second near-infrared biological window (NIR-II, wavelength of 1000–1400 nm) and excitation in the NIR-I are optimal for the fluorescence imaging of live animals owing to their lower background noise, greater tissue penetration depth, and reduced photon scattering and photo damage effects.<sup>7,8</sup> Therefore, they are expected to become more desirable in biological imaging. Up to now, numerous bioluminescence probes emitting in the NIR-II region have been developed, such as quantum dots,<sup>9</sup> carbon nanotubes,<sup>10</sup> and lanthanide doped nanomaterials.<sup>11</sup> However, most of those probes suffer from poor photostability, short luminescence lifetime, poor biocompatibility, high toxicity and even high cost.<sup>12</sup> Therefore, their further application is still limited and it is desirable to develop a novel NIR-II luminescence probe for fluorescence imaging.

$\text{Mn}^{5+}$ , a transition-metal ion with  $3d^2$  electron configuration, is stabilized in a tetrahedral coordination environment, where it

has the  $^1\text{E}$  state below the  $^3\text{T}_2$  state, giving rise to a longer-lived NIR emission in the 1000–1400 nm region.<sup>13</sup>  $\text{Mn}^{5+}$  doped phosphate and vanadate are considered promising candidates for solid-state lasers owing to their long fluorescence lifetime, the strong absorption and effective stimulated emission cross section in the near-infrared region.<sup>14–16</sup> These unique optical characteristics also enable  $\text{Mn}^{5+}$  doped vanadate and phosphate as potentially ideal bio-probes. So far, there still have been no reports about the application of these materials in fluorescence imaging because not only is  $\text{Mn}^{5+}$  rare but also  $\text{Mn}^{5+}$  doped nanosized materials are difficult to synthesize through traditional chemical routes.  $\text{Mn}^{5+}$  is an unstable ion and valence reduction often occurs during synthesis in aqueous solution, resulting in the formation of undesired  $\text{MnO}_2$  precipitate. Although doping with  $\text{Mn}^{5+}$  has been achieved through solid-state reaction under atmosphere control in previous studies,<sup>17</sup> it is still impractical to control the particle size and morphology *via* this route. In addition, high reaction temperature and long reaction time are required in the whole preparation process. To the best of our knowledge, nanosized materials with  $\text{Mn}^{5+}$  as the fluorescence center have never been reported. Therefore, it is highly desirable to develop a general but low-cost route to fabricate  $\text{Mn}^{5+}$  doped nanoparticles.

In this work, we demonstrate a strategy to fabricate for the first time, we believe,  $\text{Mn}^{5+}$  doped  $\text{Ba}_3(\text{MO}_4)_2$  ( $\text{M} = \text{V}, \text{P}$ ) nanoparticles in order to achieve fluorescence imaging.  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  nanoparticles with stable valence and controlled size were first successfully synthesized by a convenient two-step method, which not only diminishes the reaction time, but also high reaction temperature and oxygen atmosphere are not required. Stabilized  $\text{Mn}^{5+}$  ion was obtained through redox reaction in  $\text{H}_2\text{O}_2/\text{K}_2\text{MnO}_4$  solution. We also demonstrate that the method is general to the synthesis of other  $\text{Mn}^{5+}$  doped phosphors such

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as  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  and  $\text{BaSO}_4:\text{Mn}^{5+}$ . The luminescence properties, the crystalline structure and the element composition of  $\text{Mn}^{5+}$  ions in  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles have been investigated. The strategy based on anion exchange reaction is discussed in detail. Moreover, we made a simple demonstration of  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  for potential application in fluorescent imaging.

## 2. Experimental section

### 2.1 Synthesis of undoped $\text{Ba}_3(\text{VO}_4)_2$ nanoparticles

Undoped  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles were synthesized by the hydrothermal method. In a typical procedure, 0.8 mmol barium chloride ( $\text{BaCl}_2$ ) and 1.6 mmol sodium citrate were dissolved in 40 mL distilled water to form a clear liquid. Then, 1.2 mmol sodium orthovanadate ( $\text{Na}_3\text{VO}_4 \cdot 12\text{H}_2\text{O}$ ) was slowly added to the above reaction liquid and precipitated precursor of  $\text{Ba}_3(\text{VO}_4)_2$  appeared. The mixture was stirred for 10 minutes at room temperature. After that, the turbid liquid was transferred into a 50 mL Teflon-lined stainless steel autoclave and treated at 140 °C for 24 h. After the mixture fully cooled down to room temperature, the precipitate was collected by centrifugation at 8000 rpm for 5 min. The as-prepared powder was washed with distilled water and ethanol three times and dried at 60 °C for 6 h in an oven. All the above chemicals were purchased from the Aladdin Chemical Reagent Company, and used as received without any further purification.

### 2.2 Preparation of manganate(v) solution

Typically, potassium permanganate ( $\text{KMnO}_4$ ) was added to a highly alkaline solution (5–10 M KOH) at room temperature and stirred for 12–24 h to form a stable aqueous solution of manganate(vi). Manganate(v) solution was prepared by adding a theoretical amount of 0.1 wt% hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) to an ice-cold solution of manganate(vi) in 10 M KOH under vigorous stirring for 1 h and this was followed by filtering to remove  $\text{MnO}_2$  precipitate. The color of the solution gradually changed from dark green to sky blue, indicating  $\text{MnO}_4^{3-}$  was fully generated.

### 2.3 Synthesis of $\text{Mn}^{5+}$ doped $\text{Ba}_3(\text{VO}_4)_2$ nanoparticles

An anion exchange approach was employed for the synthesis of  $\text{Ba}_3(\text{VO}_4)_2:1 \text{ mol}\% \text{ Mn}^{5+}$  nanoparticles as an example. Typically, 0.10 g of  $\text{Ba}_3(\text{VO}_4)_2$  powder was first dissolved in 5 mL distilled water. Then, 0.16 mL of the 0.01 M  $\text{K}_3\text{MnO}_4$  solution was added to the obtained solution under vigorous stirring for 15 min. The solution gradually changed color from blue to colorless, indicating a decrease of  $\text{Mn}^{5+}$  concentration in the solution. The precipitate was collected by centrifugation at 8000 rpm for 5 min and washed with distilled water and ethanol three times. The final purified products were dried under vacuum.  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  was also prepared by the same method.

### 2.4 Characterization

Powder X-ray diffraction (XRD) was performed on an X'Pert PRO X-ray diffractometer (PANalytical, The Netherlands) using Cu K $\alpha$  ( $\lambda = 1.5418 \text{ \AA}$ ) radiation. The excitation and emission spectra decay curves were measured using a spectrofluorometer (iHR 320,

Jobin-Yvon, France) equipped with a 150 W xenon lamp, a monochromator and a liquid nitrogen-cooled DSS-IGA020L InGaAs detector (800–1600 nm). All spectra were corrected for the optical system responses. Thermogravimetric analysis (TG) and differential scanning calorimetry (DSC) were carried out using a thermal analyzer (NETZSCH, STA449C, Germany). The samples were heated at a rate of 10 °C  $\text{min}^{-1}$  in nitrogen atmosphere. The particle size and distribution of samples were determined by dynamic light scattering (DLS) using an electrophoretic light scattering spectrophotometer (Malvern Instruments Ltd, Zetasizer Nano-ZS, UK). The Raman spectrum was measured with a Raman spectrometer (Renishaw inVia, UK) using a 532 nm laser as an excitation source. The doping concentration of  $\text{Mn}^{5+}$  was measured by atomic absorption spectroscopy (AAS). A high-resolution transmission electron microscope (HR-TEM, 2100F, JEOL, Japan) was employed to measure the size and morphology of all the samples. Scanning electron microscopy (SEM) images were taken with a field-emission scanning electron microscope (Nova NanoSEM 430, FEI, The Netherlands). Luminescence tissue imaging was performed with a modified imaging system including a Germany pro camera as the signal collector. All luminescence images were taken in a dark room and analyzed with home-made software. An 808 nm laser (LEO, China) was selected as excitation source. Pork tissue samples were purchased from a local market and placed in a dark room kept at 25 °C. All the measurements were carried out at room temperature.

## 3. Results and discussion

Fig. 1 schematically shows an illustration of the chemical transform approach for synthesizing  $\text{Mn}^{5+}$  doped  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles. The main difficulty in the synthesis procedure is to obtain stabilized  $\text{Mn}^{5+}$  ions. In the first step, 0.1 wt%  $\text{H}_2\text{O}_2$  solution is employed to effectively reduce  $\text{Mn}^{6+}$  to  $\text{Mn}^{5+}$  in our system. The ice-cold  $\text{H}_2\text{O}_2/\text{KOH}$  solution offers an alkaline environment to prevent  $\text{Mn}^{5+}$  being reduced to  $\text{Mn}^{4+}$ . Subsequently, the undoped  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles are dissolved in a small volume of  $\text{KOH}/\text{K}_3\text{MnO}_4$  solution under vigorous stirring to form a muddy mixture, where  $\text{VO}_4^{3-}$  ions are partially replaced by  $\text{MnO}_4^{3-}$  ions. Finally, the product powder is isolated by centrifugation and washed with ethanol and distilled water. In our experiment, the expected doping concentration of  $\text{Mn}^{5+}$  is designed to be 0.1, 0.5, 1, 1.5 and 2 mol%. According to the compositional analyses by means of AAS, the evaluated

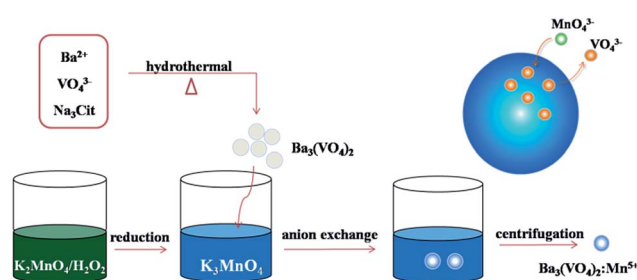


Fig. 1 Schematic illustration of anion exchange procedure for synthesizing  $\text{Mn}^{5+}$  doped  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles.



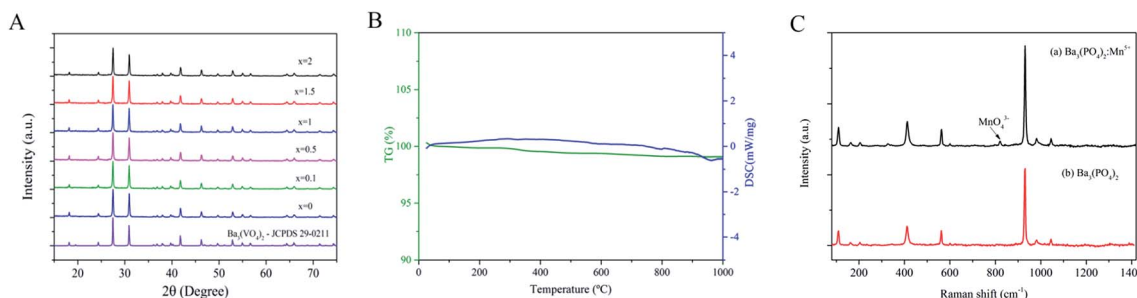


Fig. 2 (A) XRD spectra of  $\text{Ba}_3(\text{VO}_4)_2:x$  mol%  $\text{Mn}^{5+}$  ( $x = 0, 0.1, 0.5, 1, 1.5, 2$ ); (B) TG and DSC curves of  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$ ; (C) Raman spectra of (a)  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  and (b)  $\text{Ba}_3(\text{PO}_4)_2$ . a.u., arbitrary units.

doping concentrations of  $\text{Mn}^{5+}$  were 0.021, 0.072, 0.124, 0.361, 0.542 mol%. This reveals that the  $\text{Mn}^{5+}$  in the solution can only partially be doped into the  $\text{Ba}_3(\text{VO}_4)_2$  host.

Fig. 2A presents the XRD pattern of the  $\text{Ba}_3(\text{VO}_4)_2:x$  mol%  $\text{Mn}^{5+}$  nanoparticles ( $x = 0, 0.1, 0.5, 1, 1.5, 2$ ). All the diffraction peaks can be well indexed to the standard PDF card (JCPDS no. 29-0211) of rhombohedral phase ( $R\bar{3}m$ )  $\text{Ba}_3(\text{VO}_4)_2$ . The sharp diffraction peaks show that single-phase product with high crystallinity can be obtained by this method. With increasing doping amount of  $\text{Mn}^{5+}$ , no obvious changes or other detectable Mn impurity were observed, indicating that the occupation of  $\text{Mn}^{5+}$  did not change the crystal structure of  $\text{Ba}_3(\text{VO}_4)_2$ . Fig. 2B shows the TG and DSC curves of  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$ . The product exhibits good thermal stability and less mass loss is observed. The result indicates that the nanoparticles have few organic compounds outside. Fig. 2C presents the Raman spectra of undoped  $\text{Ba}_3(\text{PO}_4)_2$  and  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  excited at 532 nm. The Raman signal from  $\text{MnO}_4^{3-}$  is completely covered by the signal from  $\text{VO}_4^{3-}$  so that the vibration modes of  $\text{MnO}_4^{3-}$  are hardly detected in  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$ . As a consequence, we use  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  as example. As shown in Fig. 2C, we observe a group of Raman peaks at 112, 406, 562, 931, 983 and 1043  $\text{cm}^{-1}$  in undoped  $\text{Ba}_3(\text{PO}_4)_2$  and  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$ , which can be attributed to stretching and bending of  $\text{PO}_4^{3-}$ . The weak Raman peaks at 818  $\text{cm}^{-1}$  indicated by an arrow can be attributed to the vibration modes of  $\text{MnO}_4^{3-}$ .<sup>18</sup>

Fig. 3 shows the NIR emission spectra of  $\text{Ba}_3(\text{MO}_4)_2$  ( $M = \text{V}, \text{P}$ ) nanoparticles at room temperature. Under 808 nm excitation, both of the samples exhibit a sharp emission band at 1190 nm originating from  ${}^1\text{E} \rightarrow {}^3\text{A}_2$  transition of  $\text{Mn}^{5+}$  ions. In addition, the two vibronic emission bands centered at 1250 nm and 1300 nm are also detected, which can be assigned to the vibronic transitions of the  $\nu_1(\text{E})$  and  $\nu_2(\text{A})$  modes, respectively, of  $\text{MnO}_4^{3-}$ .<sup>19</sup> It appears that the line shape and luminescence center of  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  are quite similar to those of  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  because  $\text{Mn}^{5+}$  in tetrahedral coordination has the  ${}^1\text{E}$  state below the  ${}^3\text{T}_2$  state, so that the spin-forbidden emission is weakly dependent on crystal field strength.<sup>20</sup> Furthermore, the concentration dependence of emission intensity at 1190 nm for  $\text{Ba}_3(\text{M}_x\text{V}_{1-x}\text{O}_4)_2$  is given in the inset. The intensity increases with increasing  $\text{Mn}^{5+}$  doping concentration and reaches a saturation point at  $x = 0.1$ , then the intensity begins to decrease with further increasing  $\text{Mn}^{5+}$  doping concentration owing to the concentration quenching effect.

The photoluminescence excitation (PLE) and photoluminescence (PL) spectra of  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  and  $\text{BaSO}_4:\text{Mn}^{6+}$  phosphor are shown in Fig. 4A. By monitoring at  $\lambda_{\text{em}} = 1080$  nm,  $\text{BaSO}_4:\text{Mn}^{6+}$  in Fig. 4A(a) exhibits a relatively intense excitation band from 450 to 650 nm and a weak excitation band centered at 750 nm, which can be attributed to the charge transfer transition and  ${}^2\text{T} \rightarrow {}^2\text{E}$  transition, respectively, of  $\text{Mn}^{6+}$  ions. The PLE spectrum of  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  in Fig. 4A(b) obtained by monitoring at 1190 nm covers a broad spectral region from 500 to 850 nm and consists of three main excitation bands originating from the d-d transitions of  $\text{Mn}^{5+}$  ions, including the broad excitation band ranging from 650 to 750 nm assigned to  ${}^3\text{T}_1$  to  ${}^3\text{A}_2$  transition, the 770 nm band originating from  ${}^3\text{A}_2$  to  ${}^1\text{A}_1$  transition and the 850 nm band originating from  ${}^3\text{A}_2$  to  ${}^3\text{T}_2$  transition. In addition, the broad excitation before 350 nm is due to charge transfer transition between  $\text{O}^{2-}$  and  $\text{Mn}^{5+}$  ions. In our previous study,  $\text{BaSO}_4:\text{Mn}^{6+}$  was considered a promising fluorescent probe because it can offer a broad emission ranging from 900 to 1400 nm, which covers the entire NIR-II biological window. However, most excitation bands of  $\text{Mn}^{6+}$  ions are in the visible region. In contrast,  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  extends the excitation band into the NIR region, realizing an efficient NIR-I (800 nm) to NIR-II (1190 nm) downconversion luminescence, which has a better performance in biological imaging.

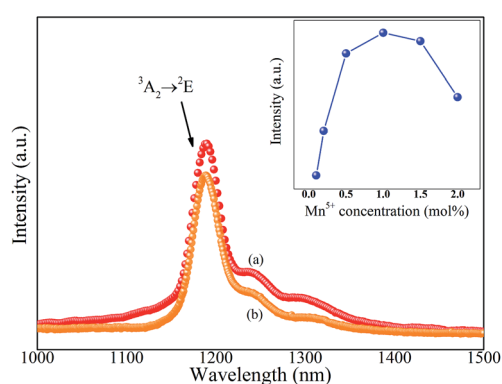


Fig. 3 NIR emission spectra of (a)  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  and (b)  $\text{Ba}_3(\text{PO}_4)_2:\text{Mn}^{5+}$  phosphors under 808 nm excitation. Inset: concentration dependence of NIR emission intensity of  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  phosphor. a.u., arbitrary units.



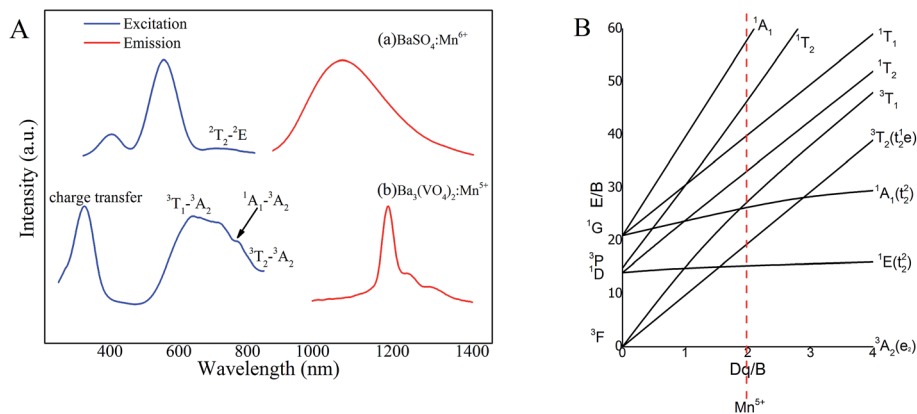


Fig. 4 (A) Photoluminescence excitation and photoluminescence spectra of (a) BaSO<sub>4</sub>:Mn<sup>6+</sup> and (b) Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:Mn<sup>5+</sup>; (B) Tanabe–Sugano energy diagram of a 3d<sup>2</sup> system in a tetrahedral crystal field of Mn<sup>5+</sup>. a.u., arbitrary units.

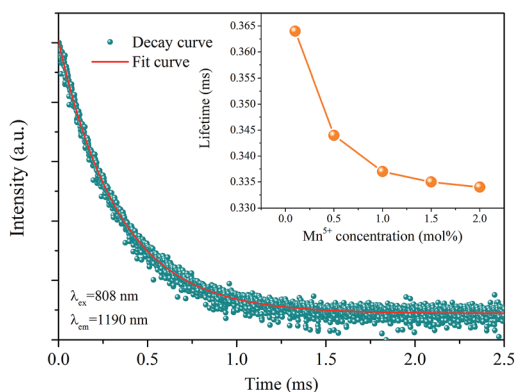


Fig. 5 Decay curve of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:Mn<sup>5+</sup> phosphor. The red curve is a fit of the experimental data to a single exponential decay equation. Inset: the concentration dependence of fluorescence lifetime of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:x mol% Mn<sup>5+</sup> phosphor (x = 0.1, 0.5, 1, 1.5, 2). a.u., arbitrary units.

The energy levels of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:Mn<sup>5+</sup> can be expressed by a Tanabe–Sugano energy level diagram of a 3d<sup>2</sup> system in tetrahedral crystal field, as shown in Fig. 4B. According to crystal field theory, the energy level sequence of Mn<sup>5+</sup> ion in tetrahedral environment should be <sup>3</sup>A<sub>2</sub> < <sup>1</sup>E < <sup>1</sup>A<sub>1</sub> < <sup>3</sup>T<sub>2</sub>.<sup>16,21</sup> The

<sup>1</sup>E → <sup>3</sup>A<sub>2</sub> transition of Mn<sup>5+</sup> ions is not sensitive to local coordination because in a strong field situation, the ground state is a spin triplet <sup>3</sup>A<sub>2</sub> level and the <sup>1</sup>E level is below the <sup>3</sup>T<sub>2</sub> level, becoming the first excited state. Based on the Tanabe–Sugano energy level diagram, the photoluminescence process can be described as follows. Firstly, the ground-state electrons of Mn<sup>5+</sup> ions are promoted to the <sup>3</sup>T<sub>1</sub>, <sup>1</sup>A<sub>1</sub> or <sup>3</sup>T<sub>2</sub> level after appropriate light excitation. Then, the excited electrons will relax to the lower <sup>1</sup>E level, giving rise to a longer-lived NIR emission due to <sup>1</sup>E → <sup>3</sup>A<sub>2</sub> transition.

Fig. 5 exhibits the luminescence decay curves of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:Mn<sup>5+</sup> excited at 808 nm and monitored at 1190 nm. The red curve is a fit of the experimental data to a single exponential decay equation. The concentration dependence of the fluorescence lifetime of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:Mn<sup>5+</sup> phosphors is given in the inset. It is found that the decay lifetime is well fitted by a first-order exponential equation:

$$I(t) = I_0 + A \exp(-t/\tau) \quad (1)$$

where  $I(t)$  and  $I_0$  represent the emission intensity at times  $t$  and 0, respectively.  $A$  is a constant and  $\tau$  is the decay time. The fitted decay times are calculated to be 0.364, 0.344, 0.337, 0.334 and 0.332 ms for  $x = 0.1, 0.5, 1, 1.5$  and 2, respectively. It is widely

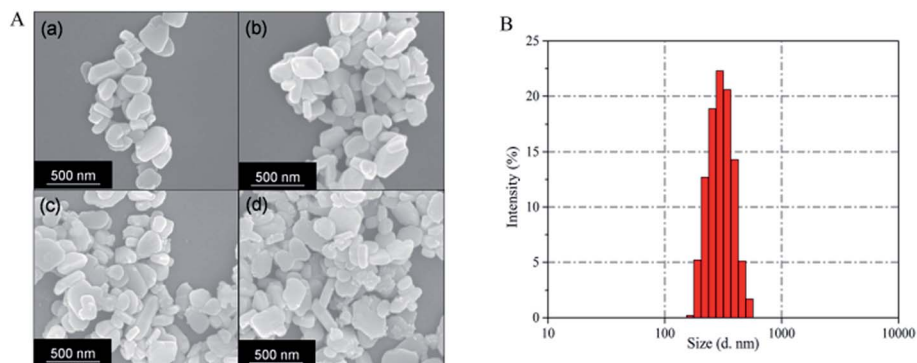


Fig. 6 (A) SEM image of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:x mol% Mn<sup>5+</sup> nanoparticles: (a) x = 0, (b) x = 0.5, (c) x = 1, (d) x = 2. (B) Particle size distribution of Ba<sub>3</sub>(VO<sub>4</sub>)<sub>2</sub>:1 mol% Mn<sup>5+</sup> nanoparticles, as measured by DLS.



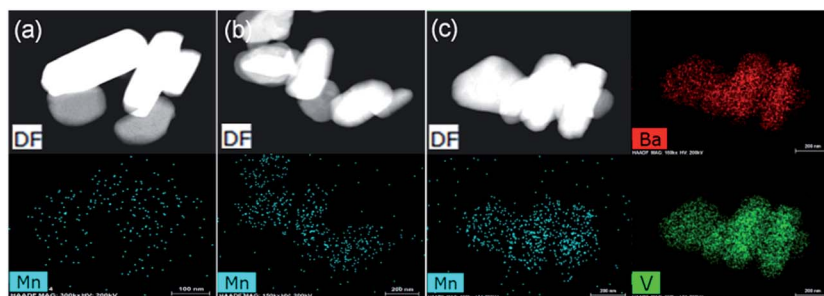


Fig. 7 EDX mapping of  $\text{Ba}_3(\text{VO}_4)_2 \cdot x \text{ mol\% Mn}^{5+}$  with different doping concentrations: (a)  $x = 0.5$ , (b)  $x = 1$ , (c)  $x = 1.5$ . DF, dark field image.

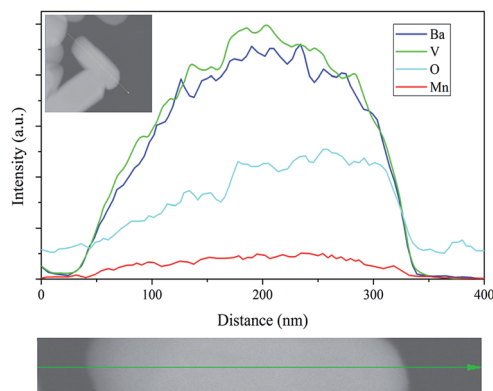


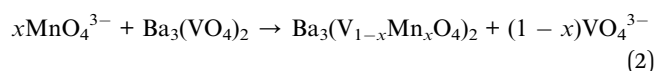
Fig. 8 EDX line scanning of  $\text{Ba}_3(\text{VO}_4)_2 \cdot \text{Mn}^{5+}$ , green, blue and red colors represent V, Ba and Mn elements, respectively. a.u., arbitrary units.

accepted that when  $\text{Mn}^{5+}$  doping concentration increases, the  $\text{Mn}^{5+}$ – $\text{Mn}^{5+}$  distance decreases, which increases the probability of energy transfer between  $\text{Mn}^{5+}$  ions and energy transfer to traps or quenching sites. As a consequence, the lifetime is shortened as  $\text{Mn}^{5+}$  concentration increases.

Fig. 6A presents SEM images of  $\text{Ba}_3(\text{VO}_4)_2 \cdot \text{Mn}^{5+}$  nanoparticles before and after anion exchange reaction. The  $\text{Ba}_3(\text{VO}_4)_2 \cdot \text{Mn}^{5+}$  nanoparticles exhibit flake-like micromorphology with a particle size in the range 150–450 nm. The shape and size of particles remain unchanged after anion exchange reaction at low doping concentration. Fig. 6B shows the particle size distribution of  $\text{Ba}_3(\text{VO}_4)_2 \cdot 1 \text{ mol\% Mn}^{5+}$  nanoparticles, measured by DLS. The mean particle diameter is 295 nm, which corresponds to that in the SEM images. In order to further confirm  $\text{Mn}^{5+}$  was successfully doped into the  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles, we carried out elemental mapping measurements and elemental line scanning.

Fig. 7a–c shows the EDX mapping of  $\text{Ba}_3(\text{VO}_4)_2 \cdot x \text{ mol\% Mn}^{5+}$  with different doping concentrations ( $x = 0.5, 1, 1.5$ ). The white color represents the digital TEM image of  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles. The green, blue and red colors represent V, Mn and Ba elements, respectively. As we can see,  $\text{Mn}^{5+}$  is distributed uniformly across the nanoparticles. With increasing  $\text{Mn}^{5+}$  doping concentration, the signal intensity of Mn element becomes higher. A similar result was also obtained in the EDX line scanning of  $\text{Ba}_3(\text{VO}_4)_2 \cdot \text{Mn}^{5+}$ , as shown in Fig. 8. The EDX analysis reveals that the elemental composition of the nanoparticles is Ba, V, O, and Mn. The signal intensity of Mn element distributed at the edges of particles is much lower than that in the center, indicating that the majority of  $\text{Mn}^{5+}$  ions are distributed on the surface of the nanoparticles. The results demonstrate that  $\text{Mn}^{5+}$  ions are successfully doped into the  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles by the chemical transform approach.

In consideration of the similar valence and effective ionic radius,  $\text{Mn}^{5+}$  ions prefer to occupy the  $\text{V}^{5+}$  sites in the host. The overall chemical reaction of  $\text{Ba}_3(\text{VO}_4)_2 \cdot \text{Mn}^{5+}$  can be represented by the following equation:



Generally, the diffusion rate in ion exchange reaction is an important factor to control the formation of the particles.<sup>22</sup> In bulk materials, the diffusion rates of exchanged ions are quite low owing to the larger particle size. When it comes to nano-sized materials, it has been reported that the diffusion rates are millions of times faster than in the bulk materials.<sup>23</sup> However, anions have larger ionic radius than cations, so that the anion exchange reaction usually has a lower diffusion rate, which always makes rigorous reaction conditions necessary. It is acknowledged that the faster diffusion of cations than

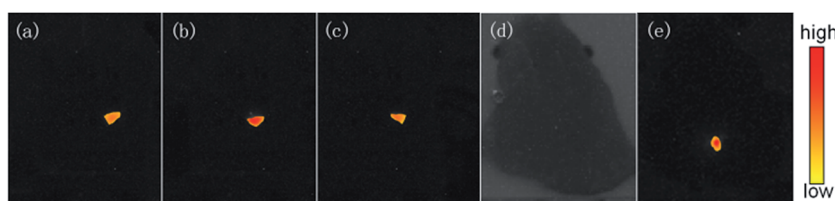


Fig. 9 NIR emission CCD pictures of the  $\text{Ba}_3(\text{VO}_4)_2 \cdot x \text{ mol\% Mn}^{5+}$  nanoparticles with doping concentration at (a)  $x = 0.5$ , (b)  $x = 1$ , (c)  $x = 1.5$ . (d) Optical and (e) fluorescence images of  $\text{Ba}_3(\text{VO}_4)_2 \cdot 1 \text{ mol\% Mn}^{5+}$  nanoparticles in pork tissue, excited by 808 nm laser.



incoming anions based on the nanoscale Kirkendall effect is the main mechanism to explain the reaction of anion exchange.<sup>24</sup> The nanoscale reduces the reaction barrier of nucleation so that  $\text{Ba}^{2+}$  are inclined to diffuse outward to the surface of particles and meet with the incoming  $\text{MnO}_4^{3-}$  ions, leading to the precipitation of  $\text{Mn}^{5+}$  doped  $\text{Ba}_3(\text{VO}_4)_2$  nanoparticles. The doping concentration of incoming  $\text{MnO}_4^{3-}$  ions is too low to change the rigid framework of  $\text{VO}_4^{3-}$  anionic groups. Hence, the initial shape and size of particles is unchanged after reaction.

NIR emission charge-coupled device (CCD) images of the  $\text{Ba}_3(\text{VO}_4)_2:x \text{ mol\% Mn}^{5+}$  nanoparticles with doping concentrations at  $x = 0.5, 1, 1.5$  are given in Fig. 9a–c; these images were obtained by using an AsGaIn camera in a dark room. An 808 nm continuous-wave (CW) laser was used as the excitation source, in combination with a long pass optical filter with a 1000 nm cut-off wavelength. It was clearly observed that the  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  emits intense NIR-I to NIR-II light, indicating the nanoparticles can be effectively excited by an 808 nm laser. To further demonstrate whether the intensity of emission from the  $\text{Ba}_3(\text{VO}_4)_2:\text{Mn}^{5+}$  is high enough for fluorescence imaging, we have obtained tissue images by injecting the nanoparticles into pork tissue at a depth of 5 mm. Fig. 9d and e show, respectively, optical and fluorescence images of  $\text{Ba}_3(\text{VO}_4)_2:1 \text{ mol\% Mn}^{5+}$  nanoparticles in pork tissue under 808 nm laser light. As shown in Fig. 9e, clear high-contrast NIR luminescence with no auto-fluorescence can be detected with an excitation power density of approximately  $500 \text{ mW cm}^{-2}$ , suggesting that the novel  $\text{Mn}^{5+}$  doped nanoparticles are potentially an ideal fluorescent probe in the NIR-II biological window.

## 4. Conclusions

In conclusion, we provide, for the first time, we believe, a facile two-step strategy based on efficient anion exchange reaction to fabricate  $\text{Mn}^{5+}$  doped nanoparticles. This novel approach not only can stabilize the valence, but also is able to control the uniform size. The  $\text{Mn}^{5+}$  doped  $\text{Ba}_3(\text{MO}_4)_2$  ( $\text{M} = \text{V}, \text{P}$ ) nanoparticles exhibit a remarkable NIR-II emission around 1190 nm with a broad excitation band in the visible and NIR region, which indicates  $\text{Mn}^{5+}$  doping has potential application in advanced NIR-II fluorescence imaging. By our approach, other candidate hosts for  $\text{Mn}^{5+}$  ions, consisting of  $\text{Ca}_3(\text{VO}_4)_2$ ,  $\text{Sr}_3(\text{VO}_4)_2$  and  $\text{Ba}_2\text{SiO}_4$ , can also be fabricated. To sum up, the general strategy illustrated in this paper will likely stimulate new ideas to prepare special valence ion doped nanosized materials.

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## References

- 1 R. Wang and F. Zhang, *J. Mater. Chem. B*, 2014, **2**, 2422–2443.
- 2 M. Chen and M. Yin, *Prog. Polym. Sci.*, 2013, **39**, 365–395.
- 3 A. M. Smith, M. C. Mancini and S. Nie, *Nat. Nanotechnol.*, 2009, **4**, 710–711.
- 4 V. J. Pansare, S. Hejazi, W. J. Faenza and R. K. Prud'Homme, *Chem. Mater.*, 2012, **24**, 812–827.
- 5 W. W. Yu, Y. A. W. And and X. Peng, *Chem. Mater.*, 2003, **15**, 4300–4308.
- 6 J. Chen, C. Guo, M. Wang, L. Huang, L. Wang, C. Mi, J. Li, X. Fang, C. Mao and S. Xu, *J. Mater. Chem.*, 2011, **21**, 2632–2638.
- 7 G. Hong, J. T. Robinson, Y. Zhang, S. Diao, A. L. Antaris, Q. Wang and H. Dai, *Angew. Chem.*, 2012, **51**, 9818–9821.
- 8 J. T. Robinson, G. Hong, Y. Liang, B. Zhang, O. K. Yaghi and H. Dai, *J. Am. Chem. Soc.*, 2012, **134**, 10664–10669.
- 9 Y. Zhang, G. Hong, Y. Zhang, G. Chen, F. Li, H. Dai and Q. Wang, *ACS Nano*, 2012, **6**, 3695–3702.
- 10 J. T. Robinson, K. Welsher, S. M. Tabakman, S. P. Sherlock, H. Wang, R. Luong and H. Dai, *Nano Res.*, 2010, **3**, 779–793.
- 11 U. Rocha, K. U. Kumar, C. Jacinto, I. Villa, F. Sanz-Rodríguez, A. Juarranz, E. Carrasco, F. C. J. M. V. Veggel and E. Bovero, *Small*, 2014, **10**, 1141–1154.
- 12 Y. T. Lim, S. Kim, A. Nakayama, N. E. Stott, M. G. Bawendi and J. V. Frangioni, *Mol. Imaging*, 2003, **2**, 50–64.
- 13 U. Oetliker, M. Herren, H. U. Güdel, U. Kesper, C. Albrecht and D. Reinen, *J. Chem. Phys.*, 1994, **100**, 8656–8665.
- 14 L. D. Merkle, A. Pinto, H. R. Verdún and B. Mcintosh, *Appl. Phys. Lett.*, 1992, **61**, 2386–2388.
- 15 M. F. Hazenkamp, H. U. Güdel, S. Kück, G. Huber, W. Rauw and D. Reinen, *Chem. Phys. Lett.*, 1997, **265**, 264–270.
- 16 R. Moncorgé, H. Manaa and G. Boulon, *Opt. Mater.*, 1994, **4**, 139–151.
- 17 T. C. Brunold, M. F. Hazenkamp and H. U. Guedel, *J. Am. Chem. Soc.*, 2002, **117**, 5598–5599.
- 18 R. Cao, J. Qiu, X. Yu and X. Sun, *ECS J. Solid State Sci. Technol.*, 2013, **2**, R237–R240.
- 19 L. C. Ferracin, M. R. Davolos and L. A. O. Nunes, *J. Lumin.*, 1997, **72–74**, 185–187.
- 20 M. Herren, T. Riedener, H. U. Güdel, C. Albrecht, U. Kaschuba and D. Reinen, *J. Lumin.*, 1992, **53**, 452–456.
- 21 J. A. Capobianco, G. Cormier, M. Bettinelli, R. Moncorgé and H. Manaa, *J. Lumin.*, 1992, **54**, 325.
- 22 R. D. Robinson, B. Sadtler, D. O. Demchenko, C. K. Erdonmez, L. W. Wang and A. P. Alivisatos, *Science*, 2007, **317**, 355–358.
- 23 C. Dong and F. C. J. M. V. Veggel, *ACS Nano*, 2009, **3**, 123–130.
- 24 J. Park, H. Zheng, Y. W. Jun and A. P. Alivisatos, *J. Am. Chem. Soc.*, 2009, **131**, 13943–13945.

