



Received 6th December 2016  
 Accepted 22nd February 2017

DOI: 10.1039/c6ra27830b  
[rsc.li/rsc-advances](http://rsc.li/rsc-advances)

## An overview of chemical constituents from *Alpinia* species in the last six decades†

Xiao-Ni Ma,<sup>ab</sup> Chun-Lan Xie,<sup>ab</sup> Zi Miao,<sup>a</sup> Quan Yang<sup>b</sup> and Xian-Wen Yang<sup>\*a</sup>

*Alpinia* species is one of the most important genera of the Zingiberaceae family. In Asia, they have been widely used as food and traditional medicines for centuries. This review focuses on their chemical constituents and their relevant biological activities with 252 references covering from 1955 to 2015. In total, 544 compounds were isolated from 35 *Alpinia* species. The major ones are terpenoids (207) and diarylheptanoids (143). The crude extracts and identified compounds exhibited a broad spectrum of bioactivities including antiemetic, antiulcer, antibacterial, anti-inflammatory, anti-amnesic, anticancer, etc.

### 1. Introduction

The genus *Alpinia* is an important member of the Zingiberaceae family. It includes *ca.* 230 species.<sup>1</sup> Most of them are distributed in tropical and subtropical Asia, including India, Malaysia, China, and Japan. A few are found in Australia and the Pacific Islands.<sup>1–3</sup> Plants of this genus have been extensively used for different purposes for centuries. For example, *A. vittata*, *A. purpurata* (Vieill.) K. Schum., *A. calcarata* Rosc., and *A. zerumbet* are cultivated as ornamental plants;<sup>3,4</sup> *A. blepharocalyx* K. Schum. is a natural dye;<sup>5</sup> *A. galanga* (L.) Willd is an important ingredient for curries and has been broadly utilized as a flavoring in the preparation of meats and soups in Southeast Asia<sup>6–8</sup> and in the preparation of beverages in Europe;<sup>9</sup> and *A. officinarum* Hance, listed as medicinal and edible food by the Chinese Ministry of Health, are used in medicinal diets,<sup>10–15</sup> wines,<sup>16</sup> sauces, and flavorings.<sup>17–19</sup> Moreover, *A. galanga* (L.) Willd is also applied to preserve food and fruits.<sup>8,20</sup> Most important of all, *Alpinia* plants are also broadly used as traditional medicines in India, China, and Japan to treat many diseases such as indigestion, gastralgia, vomiting, enterozoa etc.<sup>21–23</sup> Thus, a growing investigation on the chemical constituents and bioactivities of this genus has been carried out since 1955.<sup>24</sup> Consequently, *Alpinia* species were proved to have various biological activities including antiulcer,<sup>25</sup> antiemetic,<sup>26–28</sup> antibacterial,<sup>29–31</sup> antitumor,<sup>32–34</sup> hypoglycemic,<sup>35</sup> cardioprotection,<sup>36</sup> anti-fungi,<sup>37</sup> neuroprotection,<sup>38,39</sup> and antianxiety activities.<sup>40</sup>

Up to 2015, this genus contributed about 252 papers. However, only seven review articles were published, five of which were on chemical constituents and biological activities of single plant. And the rest two were on two major components of *Alpinia* species. The first review came out in 2010 regarding distributions, physiological activities and <sup>13</sup>C NMR spectroscopic data of 307 naturally occurring diarylheptanoids, which were mainly isolated from *Alpinia* species.<sup>41</sup> In 2011, the pharmacological and phytochemical studies of *A. galanga* (L.) Willd were summarized with 30 references. Although it was claimed to concern new phytoconstituents that have appeared in recent years for *A. galanga*, it actually collected all reported compounds including volatile oil.<sup>42</sup> In 2012, structural characterization and biological effects of constituents from the seeds of *A. katsumadai* was described. Sixty compounds were reported together with their structures and bioactivities with 18 references.<sup>43</sup> In 2013, chemical constituents in fruits of *A. oxyphylla* and their pharmacological activities were summarized. Eighty-five compounds were obtained from this species between 2001 and 2012, with the major component of sesquiterpenes (61.2%). It possessed a variety of pharmacological activities, including neuroprotection, learning and memory-improving function, anticancer, anti-aging, anti-inflammation, and anti-anaphylaxis.<sup>44</sup> In 2015, a comprehensive review on the ethnomedical uses, chemical constituents, and the pharmacological profile of *A. calcarata* Roscoe was published with particular attention given to the pharmacological effects of the essential oil.<sup>45</sup> In the same year, the phytochemistry of *A. purpurata* with pharmacological properties of antioxidant, antibacterial, larvicidal, cytotoxic, and vasodilator activities were reported together with another ornamental ginger, *Hedychium coronarium*. As a matter of fact, little research was performed on *A. purpurata*.<sup>46</sup> In addition, the isolation, synthesis, and characterization of dihydro-5,6-dehydrokavain, the major constituent of *A. zerumbet* were also reviewed.<sup>47</sup> However, so far there has been no comprehensive review for chemical constituents of this species. Herein, we describe all isolated compounds and their

<sup>a</sup>State Key Laboratory Breeding Base of Marine Genetic Resources, Key Laboratory of Marine Genetic Resources, Fujian Key Laboratory of Marine Genetic Resources, Third Institute of Oceanography, State Oceanic Administration, 184 Daxue Road, Xiamen 361005, PR China. E-mail: yangxianwen@tio.org.cn

<sup>b</sup>Department of Traditional Chinese Medicine, Guangdong Pharmaceutical University, Guangzhou 510006, China

† Electronic supplementary information (ESI) available: The name, source, plant part, and reference for each compound. A comparison of *Alpinia* species names from the references and the accepted name in The Plant List. See DOI: 10.1039/c6ra27830b



relevant bioactivities of *Alpinia* species reported in the last six decades from 1955 to 2015.

## 2. Terpenoids

### 2.1. Monoterpeneoids

A total number of 17 monoterpeneoids were obtained from *Alpinia* species (Fig. 1). Rubraine (1), isorubraine (2), and sumadain C (3) were three new monoterpene-chalcone conjugates obtained from *A. katsumadai*.<sup>48</sup> They were tested for cytotoxic activities against three tumor cell lines of HepG2, MCF-7, and MAD-MB-435. Sumadain C (3) exhibited very weak effect with IC<sub>50</sub> value of around 40.0  $\mu$ M.<sup>48</sup> *A. katsumadai* Hayata yielded a new monoterpene-kavalactone conjugate, katsumadain (4) and a new (E)-1-(1-terpinen-4-olyl)-3-methoxystilbene (5).<sup>49</sup> While *A. densibracteata* T. L. Wu and Senjen yielded two diastereoisomers of cinnamate esters, 2 $\alpha$ -cinnamoyl cineole (6) and 2 $\beta$ -cinnamoyl cineole (7).<sup>50</sup> From rhizomes of *A. tonkinensis* Gagnep., 2 $\alpha$ -(*p*-hydroxycinnamoyl) cineole (8) was isolated.<sup>50,51</sup> Two endoperoxides, (1S,4R,6R)-1,4-epidioxy-*p*-menth-2-ene (9) and (1R,4S,6R)-1,4-epidioxy-*p*-menth-2-ene (10), were isolated from aerial parts of *A. densibracteata* T. L. Wu and Senjen.<sup>50</sup> Whilst (3R,4R,6S)-3,6-dihydroxy-1-menthene (11) and 1-terpinen-4-ol (12) were obtained from *A. sichuanensis* Z. Y. Zhu (a synonym of *A. jianganfeng* T. L. Wu) and *A. katsumadai* Hayata, respectively.<sup>49,52</sup> Fruit of *A. oxyphylla* Miq. was the source of (1R,2R)-*p*-menth-3-ene-1,2-diol (13).<sup>53</sup> And aerial parts of *A. densibracteata* T. L. Wu and Senjen yielded 3,4-dihydroxy-*p*-menth-1-ene (14).<sup>50</sup> Compounds 15–17 were three hydroxyl-1,8-cineole glucopyranosides, which were mainly isolated from rhizomes of *A. galanga* (L.) Willd.<sup>54,55</sup>

### 2.2. Sesquiterpenoids

To date, 132 sesquiterpenoids were reported from *Alpinia* species (Fig. 2). They were divided into acyclic sesquiterpenoids (18 and 19), eremophilanes (20–40), eudesmanes (41–84), cadinanes (85–100), guaianes (101–117), caryophyllanes (118–120), bisabolanes (121–137), humulanes (138–140), drimane (141), elemene (142), carabane (143), oplopanane (144), and others (145–149).

Seeds of *A. katsumadai* Hayata produced an acyclic sesquiterpenoid, *trans,trans*-farnesol (18), which exerted weak neuraminidase inhibitory activity *in vitro* (IC<sub>50</sub> = 81.4  $\mu$ M).<sup>56</sup> Nerolidol (19), another acyclic sesquiterpene, was obtained from rhizomes of *A. japonica*.<sup>57</sup>

Investigations on fruits of *A. oxyphylla* Miq. afforded 16 eremophilanes (20–35). Epinootkatol (29) and nootkatone (30) displayed insecticidal activities against larvae and adults of *Drosophila melanogaster* with IC<sub>50</sub> values of 11.5  $\mu$ M and 96  $\mu$ g per adult, respectively.<sup>58</sup> While 9 $\beta$ -hydroxynootkatone (31), (11S)-12-chloronootkatone-11-ol (32), and (11R)-12-chloronootkatone-11-ol (33) displayed anti-acetylcholinesterase (AChE) activities by TLC-bioautographic assays.<sup>59,60</sup> 12-Nornootkatone-6-en-11-one (35) was a novel nor-eremophilane. It showed potent anti-AChE bioactivity at 10 nM using the same TLC-bioautographic assay.<sup>59</sup> The rest of five eremophilanes (36–40) were isolated from three different species. Eremophilene-10 $\beta$ -ol (36) and eremophilene-11-ol (37) were obtained from *A. intermedia* Gagnep. and *A. japonica* (Thunb.) Miq., respectively,<sup>61,62</sup> whilst

nootkatene (38), valencene (39), and dehydro-nootkatone (40) were all identified from *A. oxyphylla* Miq.<sup>59,63–65</sup>

Among 44 eudesmane sesquiterpenoids, oxyphyllones A and B (41 and 42) were isolated from *A. oxyphylla*. They were the first two examples of 4,5-secoeudesmanes in the Zingiberaceae family.<sup>66</sup> Oxyphyllone A displayed moderate anti-AChE activity.<sup>59</sup> Also obtained from *A. oxyphylla* Miq. were compounds 43–63.<sup>67,68</sup> *A. intermedia* Gagnep. was the source of intermedeol (64) and  $\beta$ -selinene (65).<sup>61</sup> Investigations of *A. japonica* (Thunb.) Miq. led to the identification of 66–75.<sup>21,57,69,70</sup> Two novel trinoreudesmanes, oxyphyllanenes A (76) and B (77) were obtained from *A. oxyphylla*, together with four known ones (78–81).<sup>71,72</sup> Investigation on *A. oxyphylla* Miq. provided three nor-eudesmane sesquiterpenoids, oxyphyllanene C (82), (5R,7S,10S)-5-hydroxy-13-noreudesma-3-en-2,11-dione (83), and 4-methoxy-oxyphyllene A (84).<sup>67,71,73</sup>

A new 1,10-seco-15-norcadinane sesquiterpene nominated oxyphenol A (85) was isolated from *A. oxyphylla*.<sup>65</sup> Fruits of *A. oxyphylla* Miq. also provided one tricyclic sesquiterpene, mustakone (86), nine nor-cadinanes, 87–94 and 2 $\beta$ -hydroxy- $\delta$ -cadinol (95).<sup>53,59,68,74</sup> *A. oxymitra* K. Schum. was the source of (–)-(1R,4S)-8-hydroxy-13-calamenenoic acid (96).<sup>75</sup> Alpinaterpene A (97) was provided by *A. officinarum* Hance,<sup>76</sup> while 4(15)-cadinene-6,10-diol (98) by *A. tonkinensis* Gagnep.<sup>51</sup> Two new compounds (99 and 100) were isolated from fruits of *A. oxyphylla* Miq. And 100 exhibited moderate hypoglycemic activity with inhibitory rate of 11.5%, compared to 41.9% of the positive control acarbose (41.9%) at 90  $\mu$ M.<sup>77</sup>

Rhizomes of *A. japonica* (Thunb.) Miq. produced alpinenone (101), an inhibitor of AChE.<sup>59,60</sup> Hanamyol (102), containing a cyclic ether linkage, was also isolated from *A. japonica* (Thunb.) Miq.<sup>78</sup> Rhizomes of *A. intermedia* Gagnep. provided hanalpinol peroxide (103), isohanalpinol (104), and aokumanol (105).<sup>61</sup> While *A. intermedia* Gagnep. and *A. japonica* (Thunb.) Miq. produced hanalpinol (106), hanalpinone (107), and isohanalpinone (108).<sup>61,79</sup> From *A. japonica* (Thunb.) Miq. and *A. intermedia* Gagnep., furopelargones A (109) and B (110) were obtained.<sup>61,78,80</sup> Later on, 110 was also found from *A. formossana*.<sup>81</sup> Compounds 111–114 were four secoguaiane-type sesquiterpenes with an  $\alpha$ , $\beta$ -unsaturated butenolide. *A. intermedia* Gagnep. produced epialpinolide (111), whilst *A. japonica* (Thunb.) Miq. yielded alpinolide peroxide (112), 6-hydroxy-alpinolide (113), and alpinolide (114).<sup>61,78,79</sup> A 1,10-seco-guaiane sesquiterpene, (+)-mandassidion (115), and two 1,10-seco-15-norguaiane sesquiterpenes, mandassions A (116) and B (117) were obtained from fruits of *A. oxyphylla* Miq.<sup>65</sup>

Caryophyllene oxide (118), caryophyllenol-I (119), and caryophyllenol-II (120) were caryophyllanes from *A. galanal*. In addition, caryophyllene oxide was also distributed in rhizomes of *A. conchigera* Griff.<sup>24,82</sup>

Investigation of the aerial parts of *A. densibracteata* T. L. Wu and Senjen led to the isolation of two bisabolane endoperoxides (121 and 122), three bisabolane hydroperoxides (123–125), and one 3,4-dihydroxy-bisabola-1,10-diene (126).<sup>50</sup> Compounds 127–137 were reported from rhizomes of *A. japonica* (Thunb.) Miq.<sup>83</sup>

*A. oxyphylla* Miq. was the source of 3(12),7(13),9(E)-humulatriene-2,6-diol (138).<sup>84</sup> While *A. formossana* and *A. japonica* produced humulene epoxideII (139).<sup>57,81</sup> (9E)-Humulene-2,3;6,7-diepoxyde (140) was reported from the fruits of *A. oxyphylla* Miq. However, its relative configuration remained undetermined. It



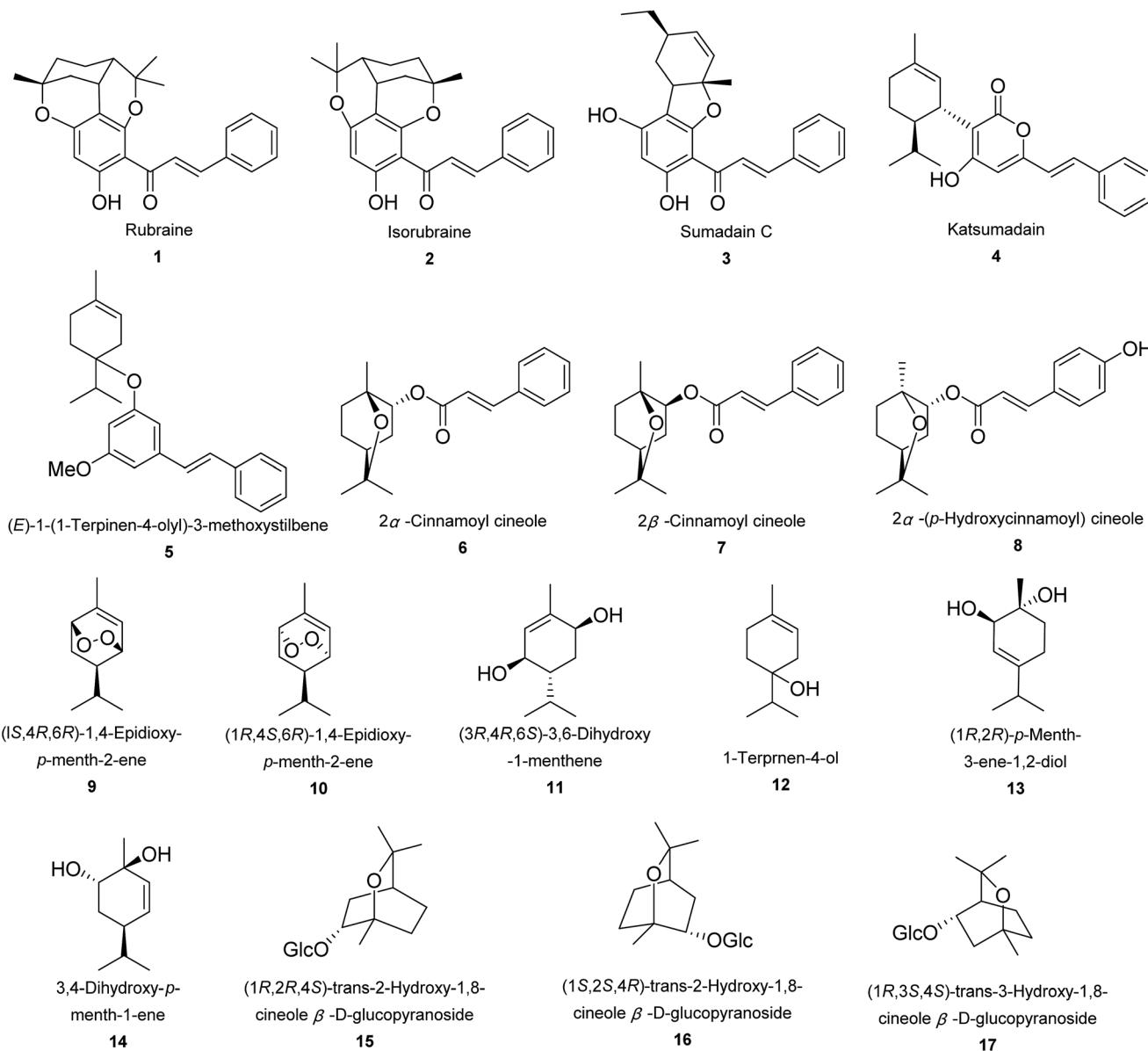


Fig. 1 Monoterpenoids from *Alpinia* species.

exhibited moderate anti-AChE activity in bioautographic assay at 10 nM.<sup>59,84</sup> Interestingly, the structure and molecular formula for **140** (CAS Registry Number: 21956-93-4) provided by SciFinder were not correct. It should be C<sub>15</sub>H<sub>24</sub>O<sub>2</sub> instead of C<sub>14</sub>H<sub>21</sub>O<sub>2</sub>.

Rhizomes of *A. calcarata* Rosc. afforded a drimane-type sesquiterpene ( $\gamma$ -bicyclohomofarnesal, **141**),<sup>85</sup> and an elemene one (shybunone, **142**).<sup>83</sup> Pubescone (**143**) was isolated from *A. oxyphylla* Miq. and showed weak anti-AChE activity at the concentration of 100  $\mu$ M.<sup>59</sup> (–)Oplopanone (**144**) and oxyphyllone F (**145**) were obtained from fruits of *A. oxyphylla* Miq.<sup>84</sup> (*Z*)-4-(2,6-Dimethylhepta-1,5-dien-1-yl)-1-methyl-cyclobut-1-ene (**146**) was a novel nor-sesquiterpene incorporating cyclobutene ring from *A. oxyphylla* Miq.<sup>74</sup> Seeds of *A. galanga* (L.) Willd produced caryolane-1,9 $\beta$ -diol (**147**), which suppressed the proliferation of four cancer cell lines of HeLa, A549, HepG2, and SMMC-7721 with IC<sub>50</sub> values ranged from 252 to 378  $\mu$ M.<sup>86</sup> *A. japonica* (Thunb.) Miq. yielded

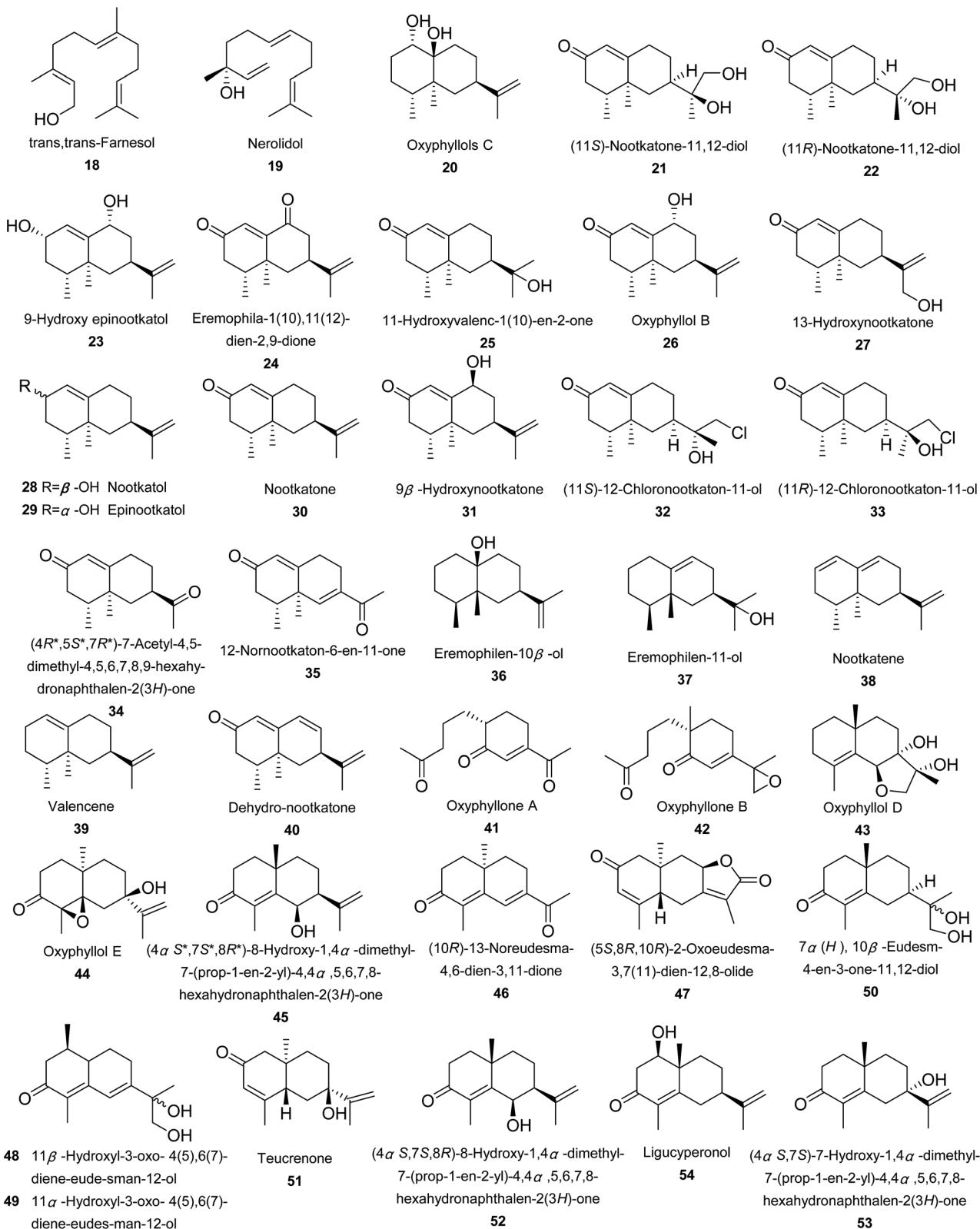
alpiniol (**148**).<sup>87</sup> Compound 2-ethyl-6-isopropyl-7-hydroxymethyl naphthalene (**149**) was a novel naphthalene from *A. oxyphylla*.<sup>77</sup> It showed bioactive activity with the inhibitory rates of 10.3%, compare to 41.9% of the positive control acarbose at 0.9 mM.<sup>77</sup>

Noteworthily, compounds **22–31**, **34**, **48–56**, **58–63**, **79–82**, **87–89**, **117**, and **129–136** exerted NO production inhibitory activities at different levels.<sup>58–60,65,67,71,73,83,88–90</sup> While (10*R*)-13-noreudesma-4,6-dien-3,11-dione (**46**), (5*S*,8*R*,10*R*)-2-oxo-13-noreudesma-3,7(11)-dien-12,8-olide (**47**), (5*R*,7*S*,10*S*)-5-hydroxy-13-noreudesma-3-en-2,11-dione (**83**), and (4*S*)-10-nor-calamenen-10-one (**90**) showed potent auxo-action of NO production at 10  $\mu$ M induced by lipopolysaccharide (LPS) in microglia.<sup>71</sup>

### 2.3. Diterpenoids

Labdane diterpenes is undoubtedly predominant in Zingiberaceae family, notably in *Alpinia* genus. Almost all diterpenes are

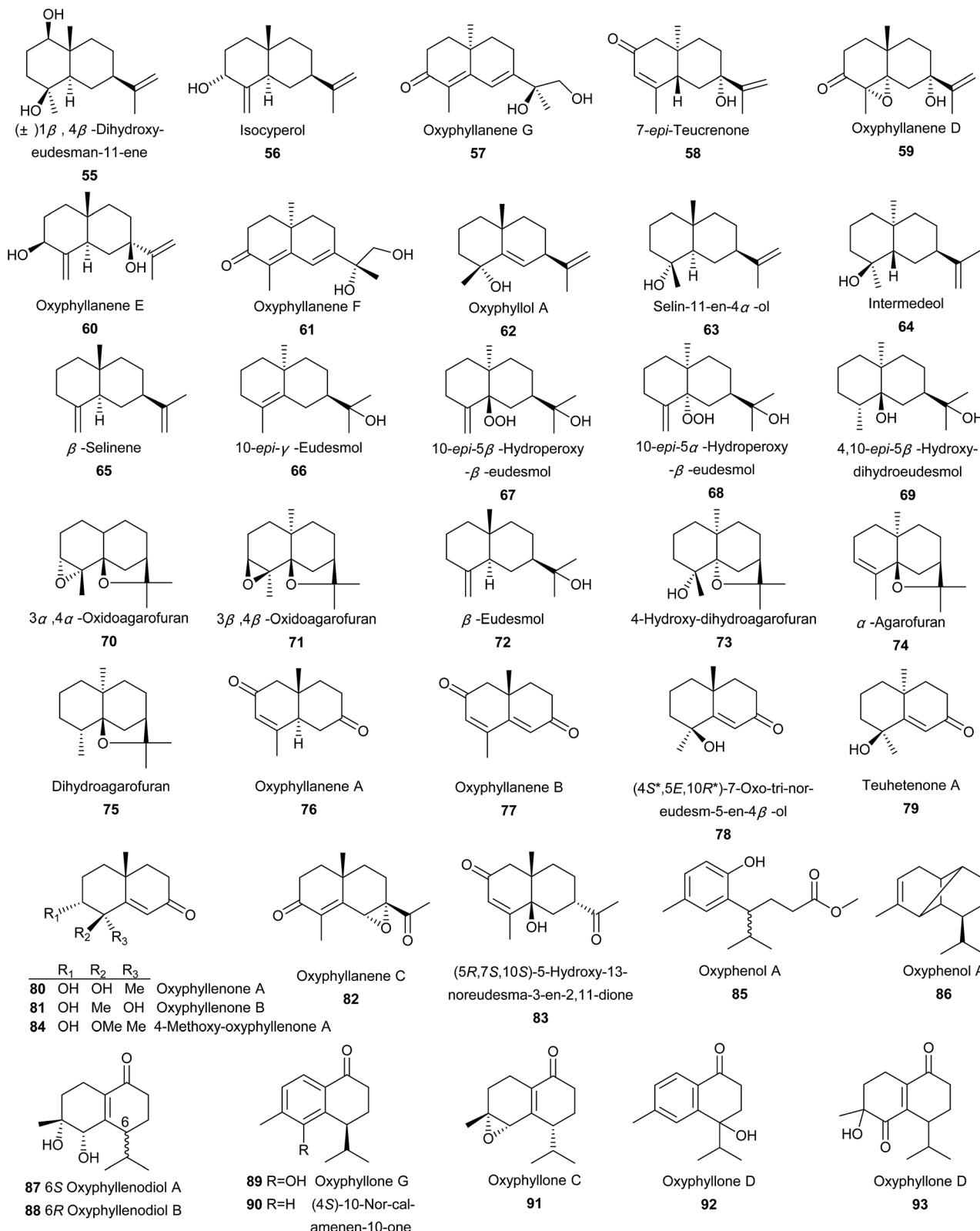


Fig. 2 Sesquiterpenoids from *Alpinia* species.

labdanes (150–205). Only one grayanane diterpene was found (206) (Fig. 3).

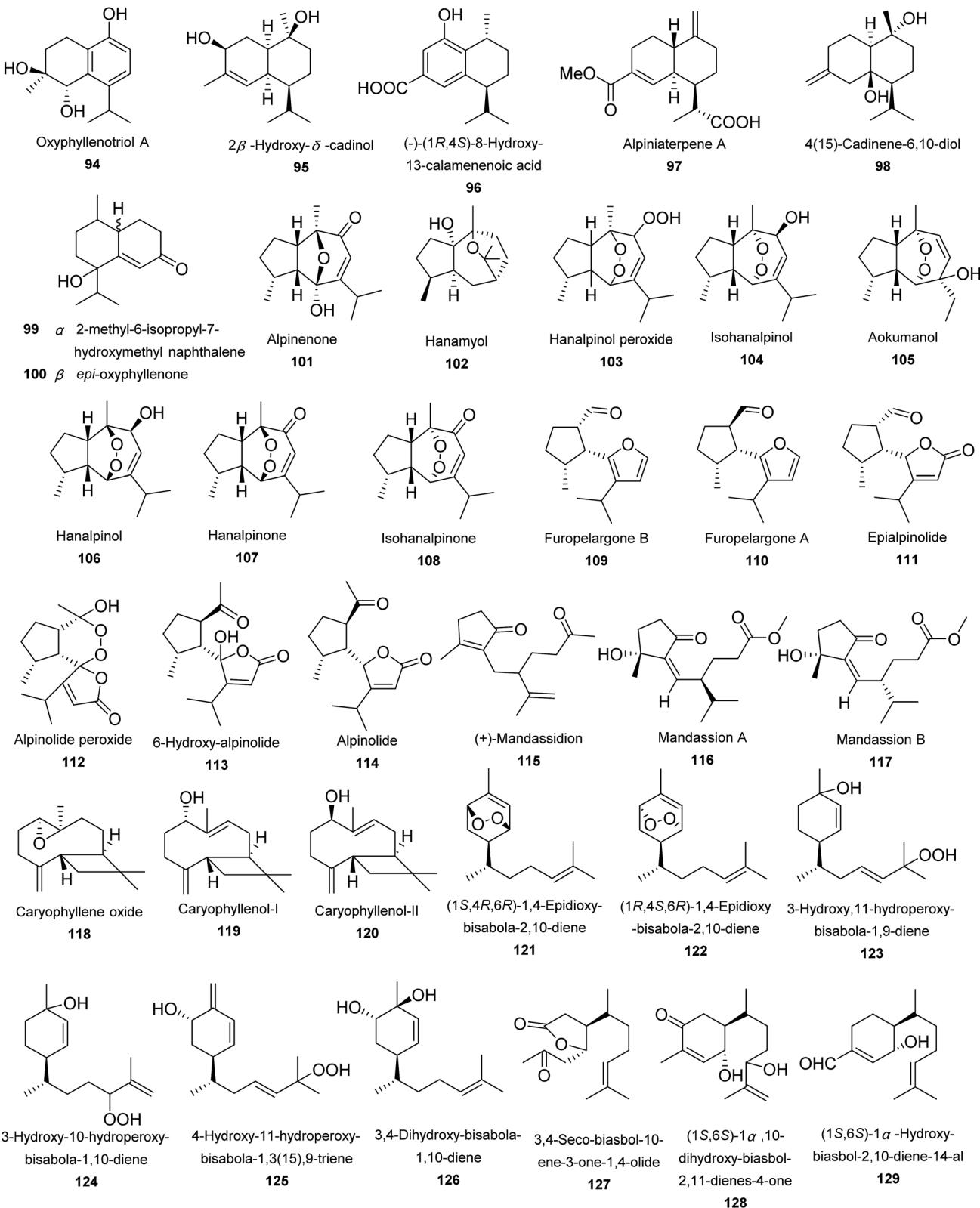
(E)-Labda-8(17),12-diene-15,16-dial (150) is widely distributed in *Alpinia*. It exhibited a number of bioactivities, such as

antibacterial,<sup>91</sup>  $\alpha$ -glucosidase inhibition,<sup>92</sup> NO production inhibition,<sup>88</sup> antifungal,<sup>93</sup> antiglycation,<sup>94</sup> HIV-1 integrase, and neuraminidase inhibitory activities.<sup>95</sup> *A. katsumadai* Hayata, *A. galanga* (L.) Willd, and *A. nigra* yielded (E)-8 $\beta$ ,17-epoxylabd-12-

Fig. 2 Sesquiterpenoids from *Alpinia* species (continued).

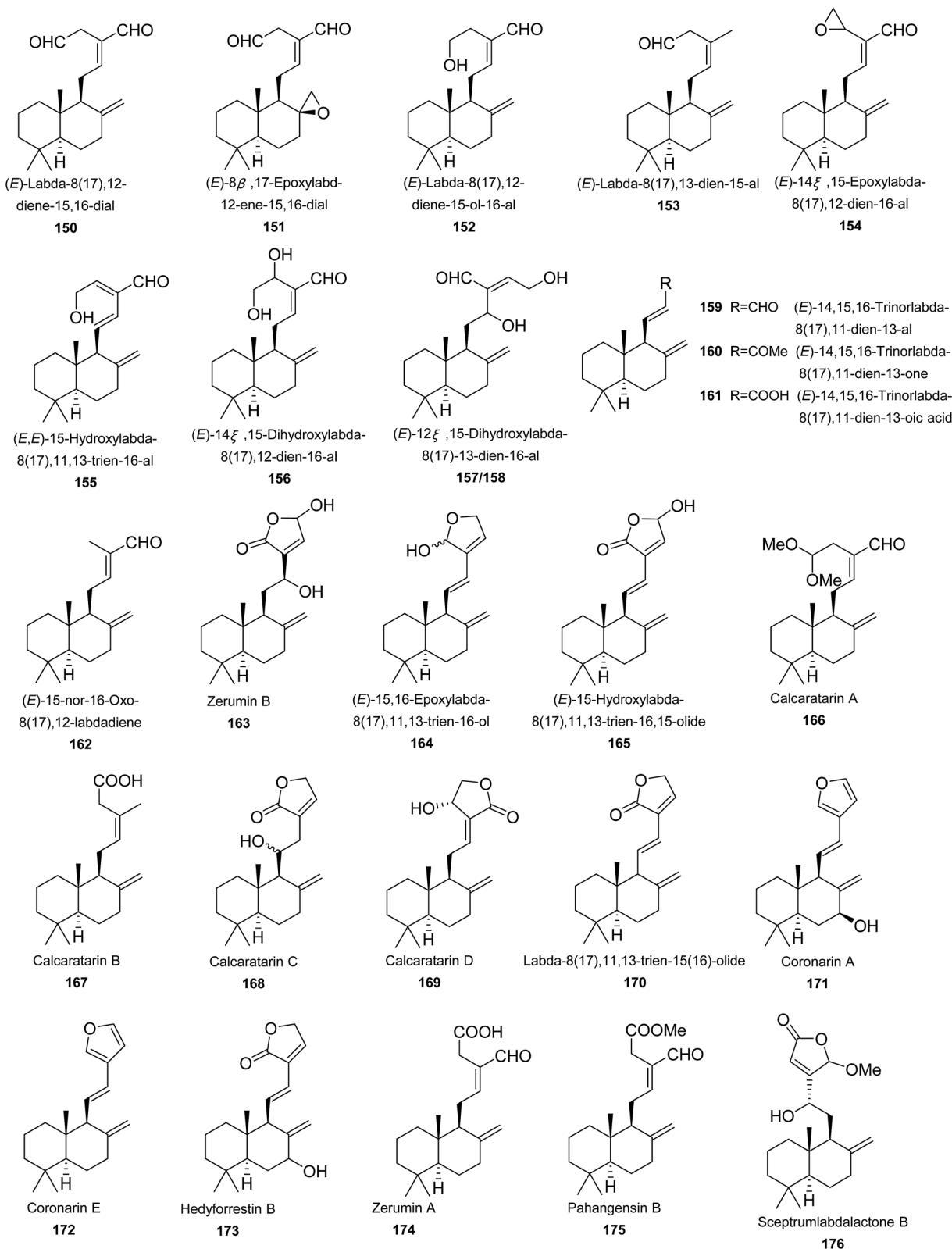
ene-15,16-dial (**151**). It exhibited extensive antibacterial activities, especially against *Candida guilliermondii* and *Candida tropicalis*.<sup>49,91,93,96</sup> Moreover, **151** also showed  $\alpha$ -glucosidase

inhibitory activity with  $IC_{50}$  value between 5  $\mu$ M and 10  $\mu$ M.<sup>70</sup> The  $\alpha$ -glucosidase inhibitory activity of **151** was even much higher than the positive control, acarbose ( $IC_{50} = 400$   $\mu$ M),

Fig. 2 Sesquiterpenoids from *Alpinia* species (continued).

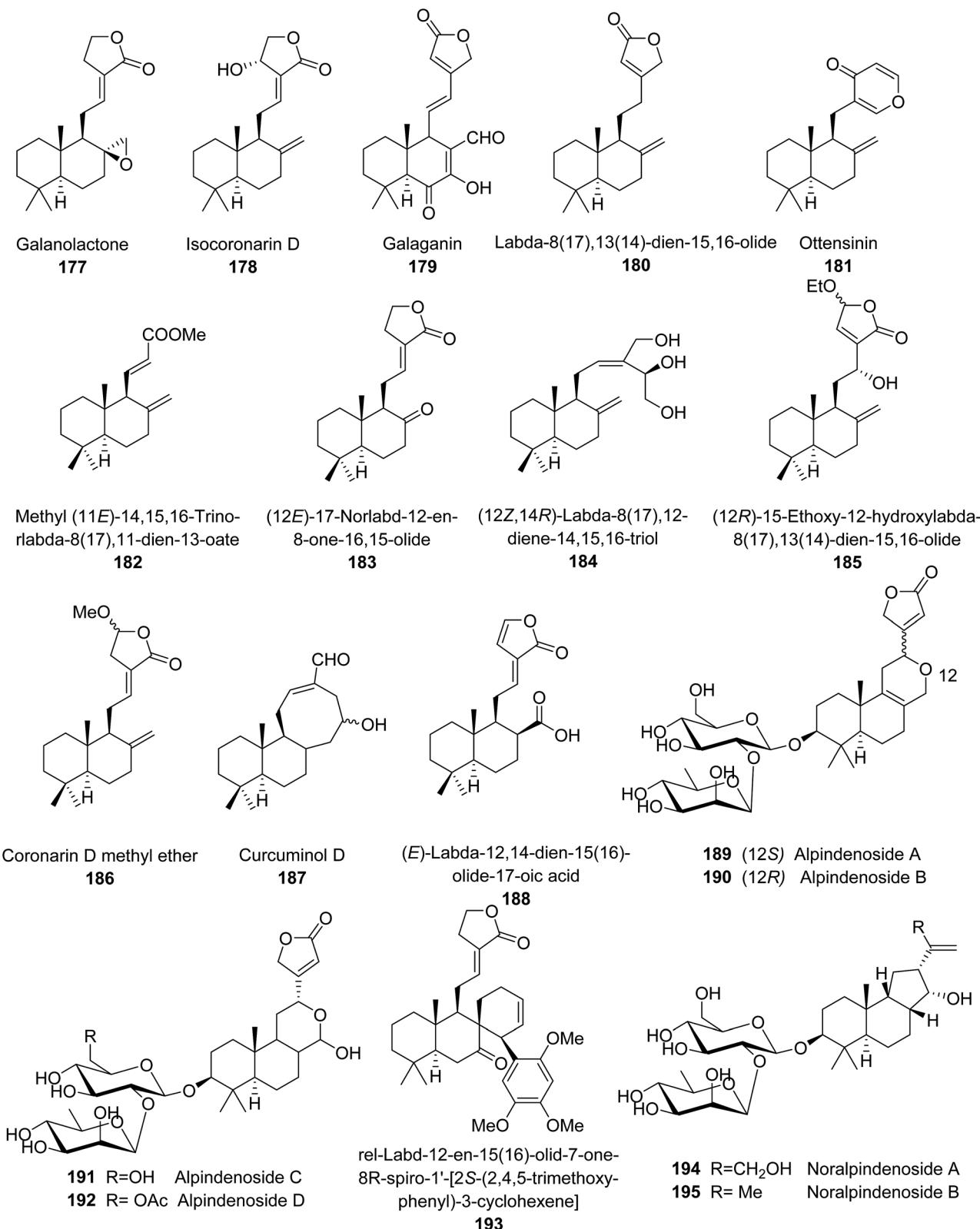
indicating **151** might be a potential candidate as a future anti-diabetic drug.<sup>70</sup> *A. formosana*, *A. calcarata* Rosc., and *A. pahangensis* Ridley provided (*E*)-labda-8(17),12-diene-15-ol-16-al

(**152**),<sup>81,85,96</sup> while (*E*)-labda-8(17),13-dien-15-al (**153**) was only obtained from *A. pahangensis* Ridley.<sup>96</sup> Flowers of *A. chinensis* Rosc. provided compounds **154**–**161**.<sup>81,85,97</sup> *A. tonkinensis*

Fig. 3 Diterpenoids from *Alpinia* species.

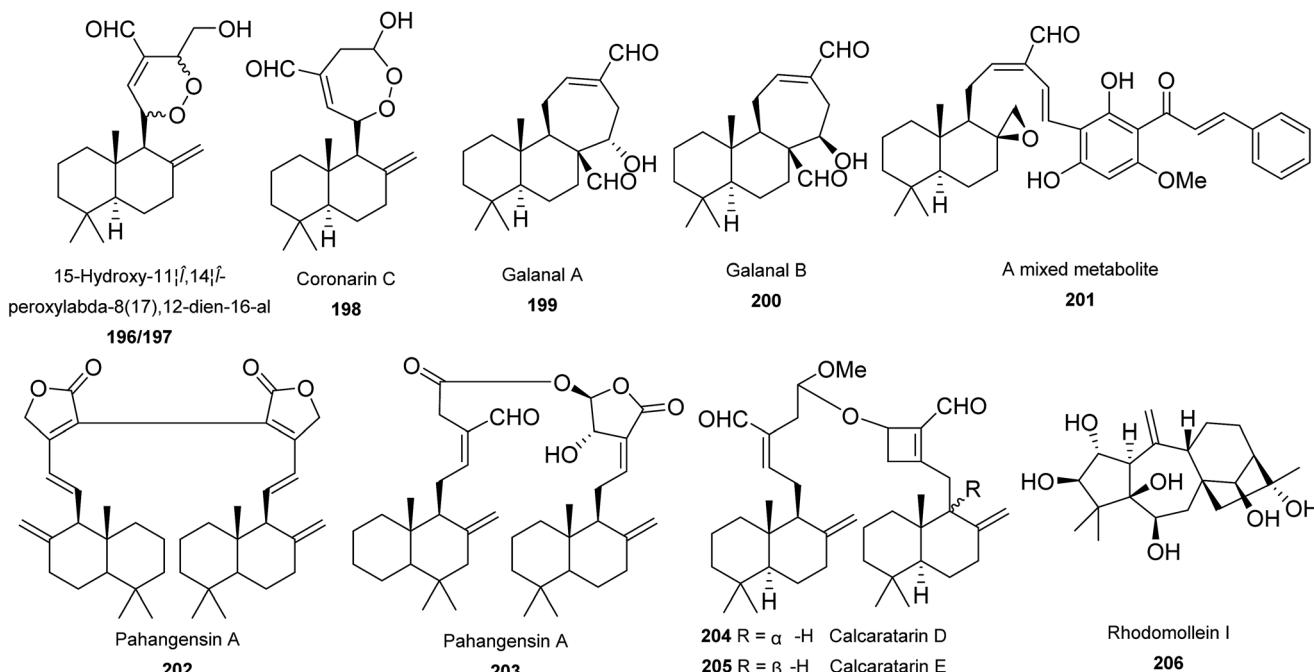
Gagnep. and *A. speciosa* K. Schum. (the accepted name is *A. zerumbet* (Pers.) B. L. Burtt & R. M. Sm.) were the sources of (E)-15-nor-16-oxo-8(17),12-labdadiene (162).<sup>51,98</sup> Both *A. zerumbet*

(Pers.) Burtt and P. M. Smith and *A. pahangensis* Ridley gave birth to zerumin B (163).<sup>96,99</sup> (11E)-15,16-Epoxyabda-8(17),11,13-trien-16-ol (164) and (E)-15-hydroxylabda-8(17),11,13-trien-16,15-olide

Fig. 3 Diterpenoids from *Alpinia* species (continued).

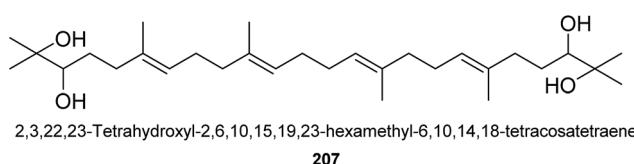
(165) were found in the flowers of *A. chinensis* Rosc.<sup>97</sup> It is noteworthy that 164 was actually a mixture of two epimers. Rhizomes of *A. calcarata* Rosc. produced calcaratarins A–D (166–169) and

labda-8(17),11,13-trien-15(16)-olide (170).<sup>85</sup> Rhizomes of *A. malaccensis* yielded coronarin A (171), coronarin E (172), and hedyforrestin B (173).<sup>100</sup> Coronarin E (172) was also isolated from *A.*

Fig. 3 Diterpenoids from *Alpinia* species (continued).

*zerumbet* (Pers.) Burtt and P. M. Smith, and *A. chinensis* Rosc.<sup>97,99,100</sup> Three antibacterial constituents, zerumin A (174), pahangensin B (175), and sceptrumlabdalactone B (176), were isolated from *A. pahangensis* Ridley.<sup>96</sup> Interestingly, zerumin A (174) was also obtained from *A. calcarata* Rosc. and *A. zerumbet* (Pers.) Burtt and P. M. Smith.<sup>85,99</sup> Compound 175 was also found in *A. japonica* (Thunb.) Miq., with NO production inhibition ( $IC_{50} = 34.3 \mu\text{M}$ ) in LPS-induced RAW264.7 macrophages.<sup>101</sup> Galanolactone (177) was isolated from *A. katsumadai* Hayata and *A. galanga*. It was reported to have moderate antifungal activity to *Candida guilliermondii* PW44 and *Candida tropicalis* PW30 with both MIC values of  $25 \mu\text{g mL}^{-1}$ .<sup>93</sup> Isocoronarin D (178) was found in *A. galanga* (L.) Willd and *A. calcarata* Rosc., which weakly suppressed the proliferation of four cancer cell lines of HeLa, A549, HepG2, and SMMC-7721 in a concentration-dependent way with  $IC_{50}$  values ranging from  $69.1$  to  $87.0 \mu\text{g mL}^{-1}$ .<sup>64,67</sup> Seeds of *A. galanga* yielded galaganin (179), which showed moderate cytotoxicity towards DU145, MCF-7, H522, and k562 cells with  $IC_{50}$  values of  $8.2$ ,  $13.8$ ,  $17.8$ , and  $16.1 \mu\text{M}$ , respectively.<sup>102</sup> Rhizomes of *A. pinnanensis* T. L. Wu et Senjen produced labda-8(17),13(14)-di-en-15,16-olide (180) and ottensinin (181).<sup>96</sup> *A. japonica* provided compounds 182–187, of which 182 and 183 were norlabdanes.<sup>101</sup> Compounds 182, 185, and 186 exhibited

significant NO production inhibitory effects in LPS-induced RAW264.7 macrophages, with respective  $IC_{50}$  values of  $25.9$ ,  $14.6$ , and  $25.6 \mu\text{M}$ , compare to  $39.6 \mu\text{M}$  of the positive control, *N*-monomethyl-L-arginine (L-NMMA).<sup>101</sup> Ethanol extract of *A. oxyphylla* Miq. provided 188, which showed moderate hypoglycemic effect with inhibitory rates of  $10.0\%$  at  $60 \mu\text{M}$ .<sup>77</sup> Ottensinin showed moderate antibacterial activity on the Gram-positive bacteria of *Bacillus cereus* with MIC value of  $0.25 \mu\text{g mL}^{-1}$ .<sup>96</sup> Alpindenosides A–D (189–192) were four labdane glycosides from *A. densespicata* Hayata. They didn't show cytotoxic activities against four human tumor cell lines of HeLa, KB, Doay, and WiDr at  $20 \mu\text{M}$ . Instead, they all exhibited moderate NO inhibitory activities with  $IC_{50}$  ranging from  $30$  to  $49 \mu\text{M}$ .<sup>103</sup> Leaves of *A. flabellata* provided *rel*-labda-12-en-15(16)-olid-7-one-8*R*-spiro-1'-[2S-(2,4,5-trimethoxyphenyl)-3-cyclohexene] (193), a unique labdane diterpene coupled with a phenylbutenoid.<sup>104</sup> Noralpindenosides A (194) and B (195) were two norditerpene glycosides from *A. densespicata* Hayata, both of which showed moderate inhibitory effects on NO production with  $IC_{50}$  values of  $34.2$  and  $49.3 \mu\text{M}$ , respectively.<sup>103</sup> (*E,E*)-15-Hydroxylabda-8(17),11,13-trien-16-al (196) and its diastereoisomer (197) from *A. chinensis* Rosc. may arise by direct oxygenation of (*E,E*)-15-hydroxylabda-8(17),11,13-trien-16-al.<sup>97</sup> From flowers of *A. chinensis* Rosc., coronarin B (198) containing a seven-membered endoperoxide hemiacetal was isolated.<sup>97</sup> It should be noted that although the structure and its NMR and MS spectroscopic data referred to coronarin B (CAS number: 119188-38-4) in the reference, the author gave a wrong name for this compound as coronarin C (CAS number: 119188-35-1) which was previously isolated from *Hedychium coronarium*.<sup>105</sup> Galanals A (199) and B (200) were obtained from *A. galanga* (L.) Willd. Both compounds showed significant antifungal activities against

Fig. 4 Triterpenoids from *Alpinia* species.

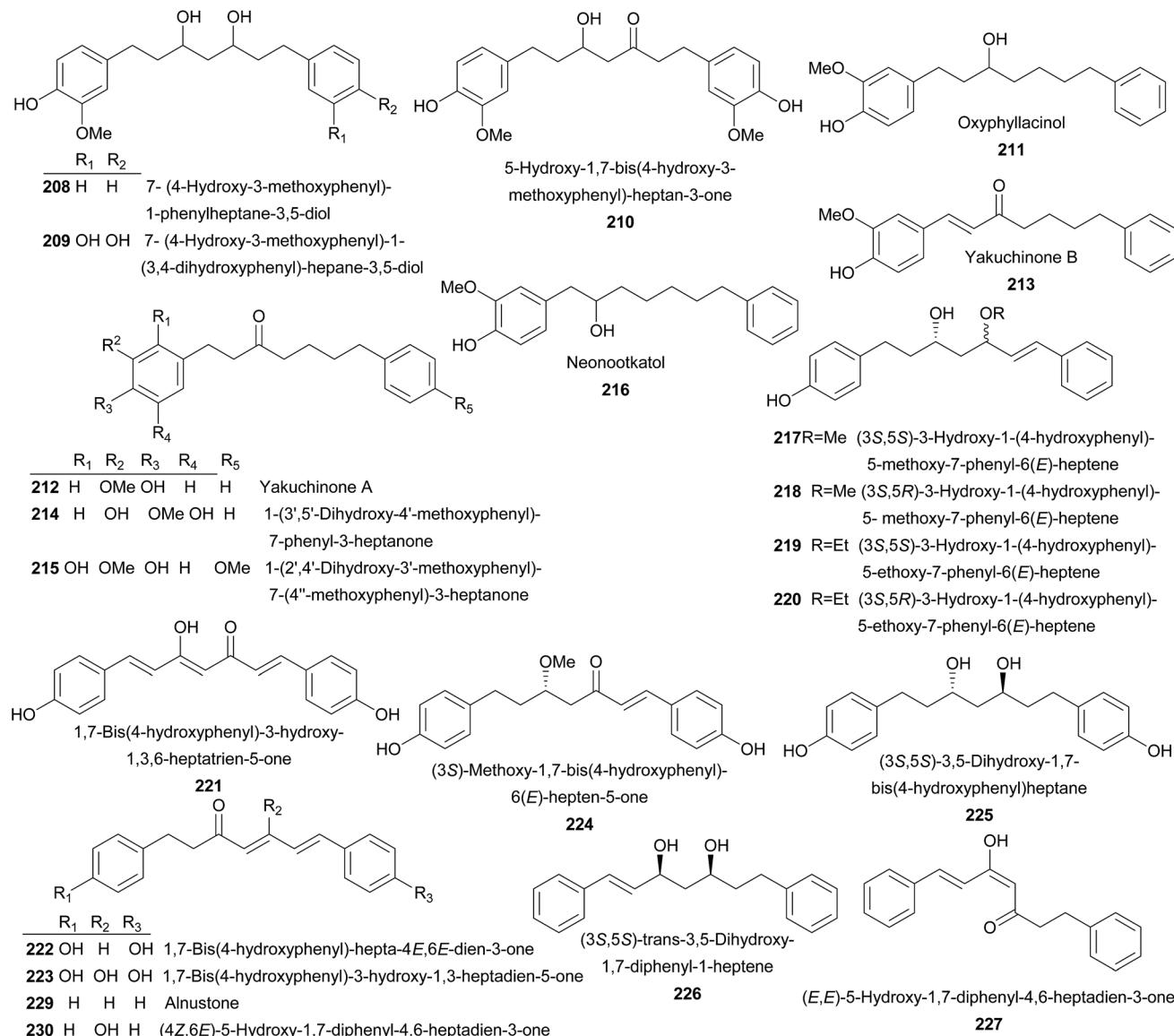


Fig. 5 Diarylheptanoids from *Alpinia* species.

*Candida guilliermondii* PW44 with MIC values of  $12.5 \mu\text{g mL}^{-1}$ . Furthermore, galanal A exhibited potent cytotoxic activity against KB cells ( $\text{IC}_{50} = 3.25 \mu\text{g mL}^{-1}$ ).<sup>7,93</sup> Compound 201 was a novel metabolite conjugated of labdane diterpene with chalcone from aerial parts of *A. katsumadai* Hayata.<sup>49</sup> *A. pahangensis* Ridley provided pahangensins A (202) and C (203) as antibacterial constituents.<sup>96,106</sup> *A. pahangensis* Ridley produced calcaratarins D (204) and E (205), both of which were cytotoxic against human KB cells *in vitro* with  $\text{IC}_{50}$  value of 0.21 and  $0.15 \mu\text{g mL}^{-1}$ , respectively.<sup>107</sup> From seeds of *A. katsumadai* Hayata, a grayanane diterpenoid was isolated and characterized as rhodomollein I (206).<sup>108</sup>

#### 2.4. Triterpenoids

Up to now, only one triterpene was found from this genus (Fig. 4). It was named as 2,3,22,23-tetrahydroxyl-2,6,10,15,19,23-hexamethyl-6,10,14,18-tetracosatetraene (207), an acyclic triterpenoid, isolated

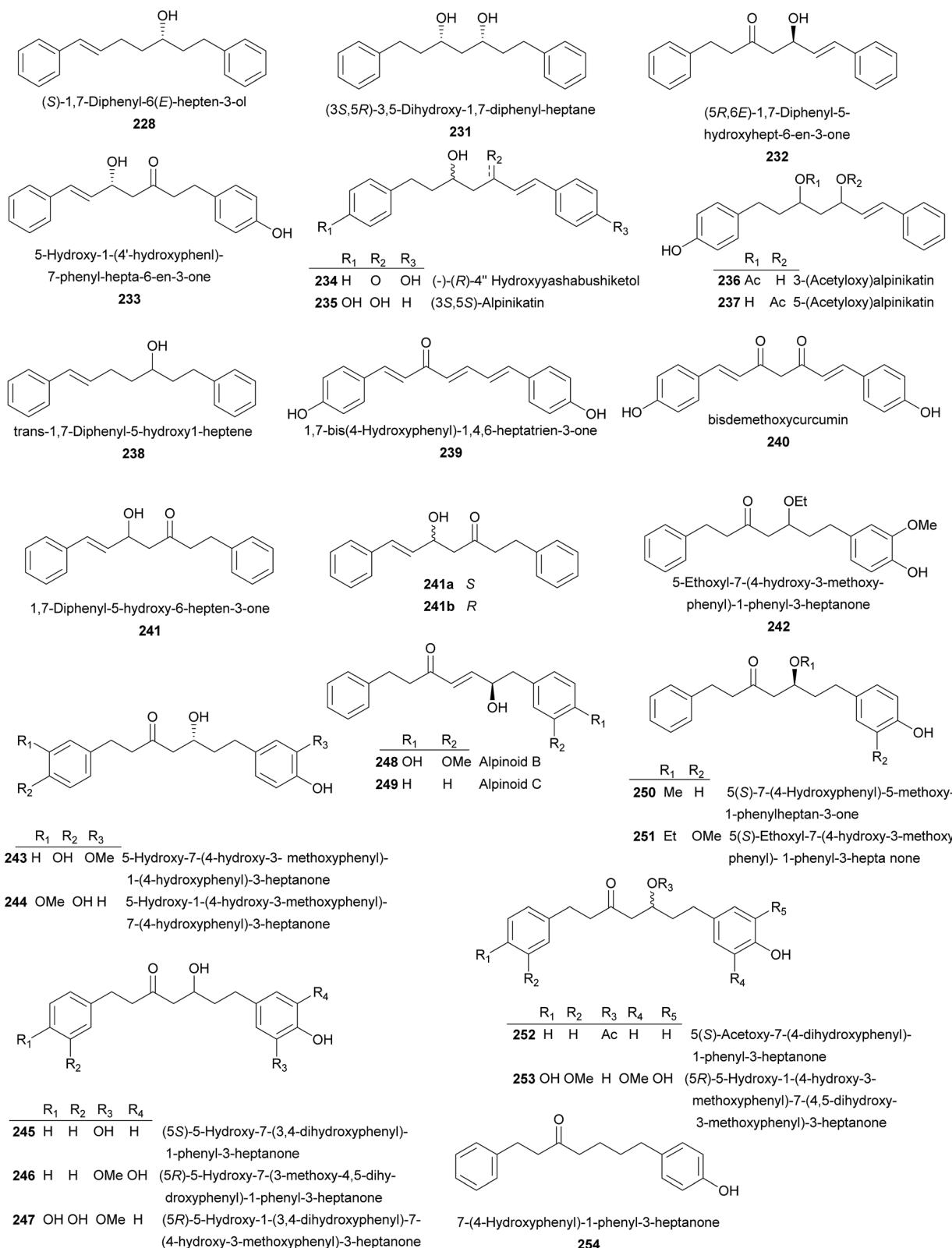
from the seeds of *A. katsumadai* L.<sup>109</sup> It showed weak cholesterol acyltransferase inhibitory activity with  $\text{IC}_{50}$  value of  $47.9 \mu\text{M}$ .<sup>109</sup>

### 3. Diarylheptanoids

A total of 143 diarylheptanoids (208–350, Fig. 5) were isolated from *Alpinia* species, including 66 acyclic diarylheptanoids (208–273), 11 cyclic diarylheptanoids (274–284), 50 diarylheptanoid and flavonoid conjugates (285–334), 10 dimeric diarylheptanoids (335–344), and six others (345–350).

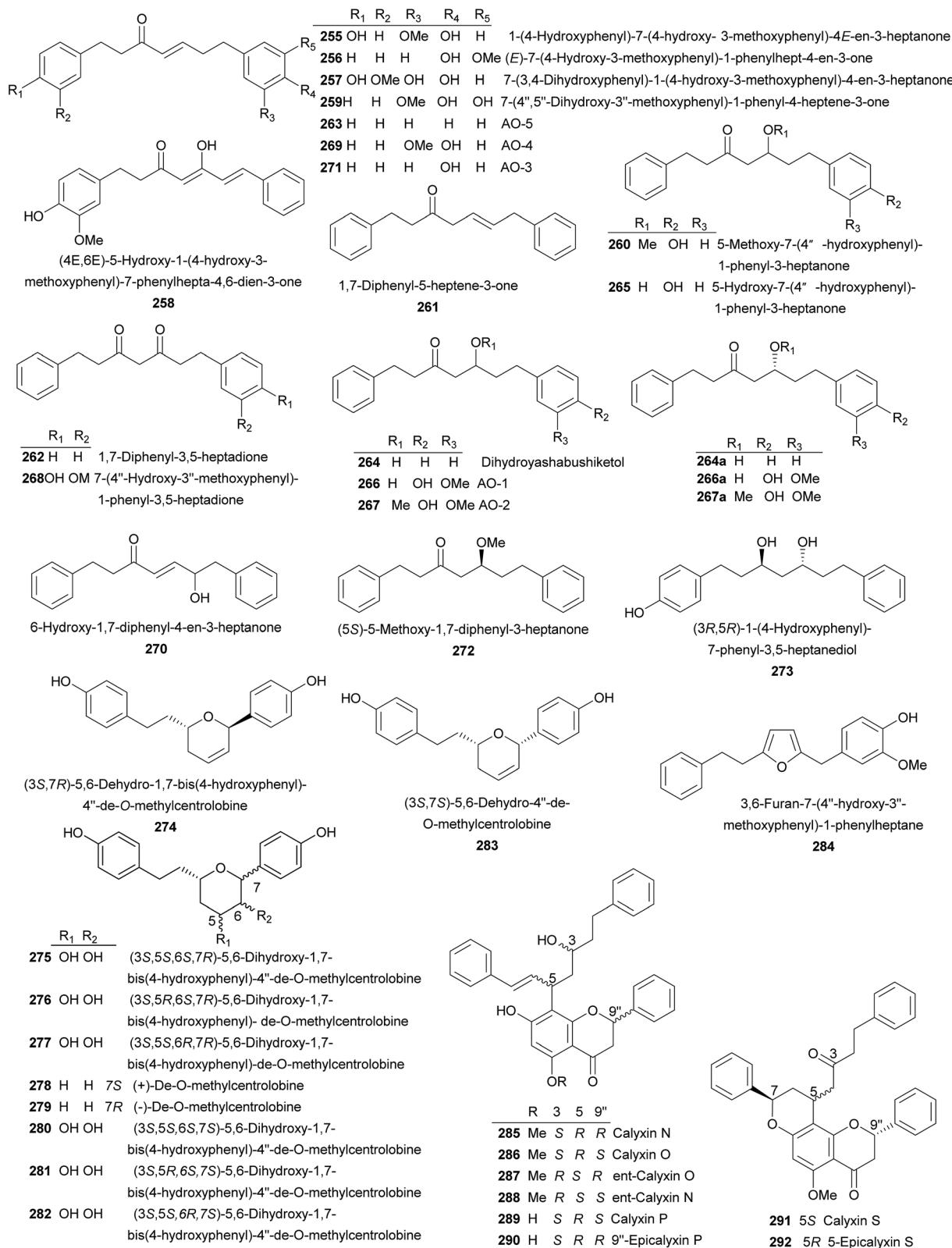
Compounds 208–210 were isolated from rhizomes of *A. officinarum* Hance. They were moderate or weak NO production inhibitors.<sup>110</sup> From fruits of *A. oxyphylla*, oxyphyllacinol (211) and yakuchinones A–B (212–213) were isolated, of which 211 was a NO production inhibitor, while 212 and 213 exhibited anti-tumor activities to human promyelocytic leukemia (HL-60) cells in a concentration-related manner.<sup>32,67</sup> In addition, 212



Fig. 5 Diarylheptanoids from *Alpinia* species (continued).

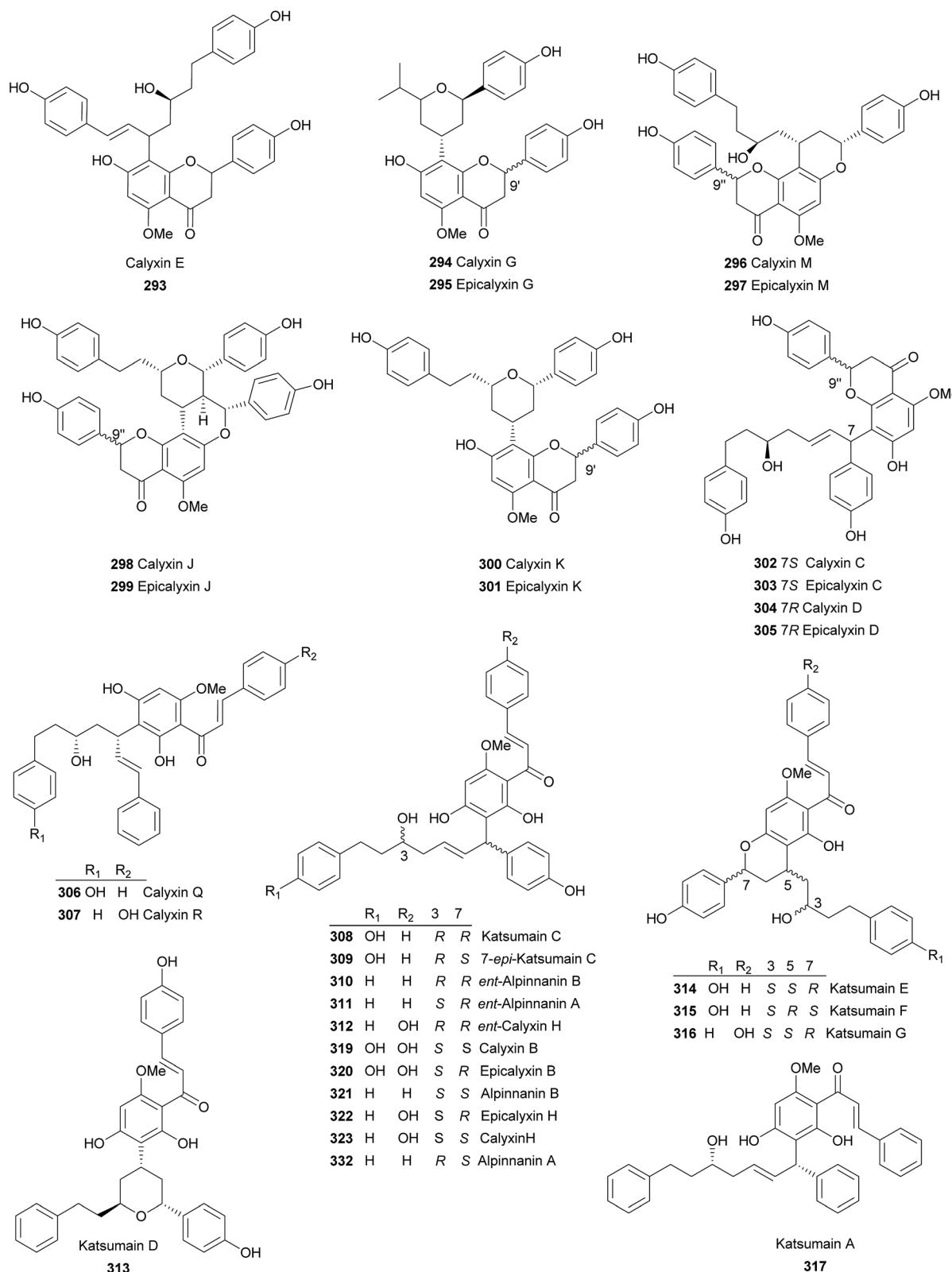
also possessed insecticidal,<sup>36</sup> anti-adipocyte differentiation,<sup>111</sup> NO production inhibitory,<sup>46</sup> and cardiotonic activities.<sup>112</sup> Compounds 213–216 were also yielded by fruits of *A. oxypylalla*.<sup>113,114</sup> Seeds of *A.*

*blepharocalyx* K. Schum. gave birth to 217–225.<sup>115–117</sup> Among these compounds, 1,7-bis(4-hydroxyphenyl)-3-hydroxy-1,3-heptadien-5-one (223) significantly inhibited platelet aggregation induced by

Fig. 5 Diarylheptanoids from *Alpinia* species (continued).

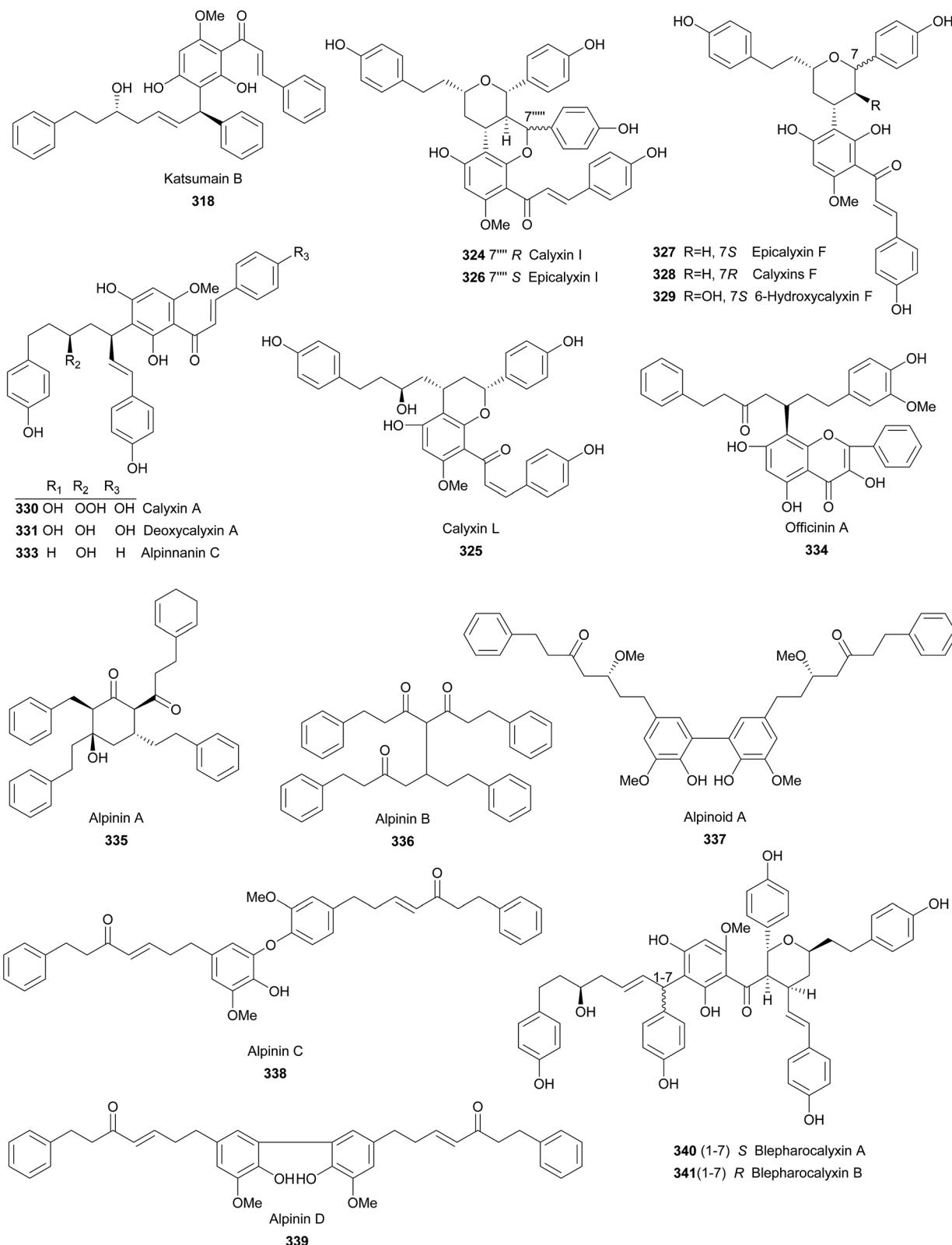
collagen with  $IC_{50}$  value of  $14.7 \mu\text{g mL}^{-1}$ .<sup>117</sup> (*3S,6E*)-Methoxy-1,7-bis(4-hydroxyphenyl)-6-hepten-5-one (224) and (*3S,5S*)-3,5-dihydroxy-1,7-bis(4-hydroxyphenyl)heptane (225) showed

significant antiproliferative activities against murine colon 26-L5 carcinoma and human HT-1080 fibrosarcoma with  $IC_{50}$  values of 5.2 and  $12.8 \mu\text{M}$ , respectively.<sup>115,116</sup> Both *A. pinnanensis*

Fig. 5 Diarylheptanoids from *Alpinia* species (continued).

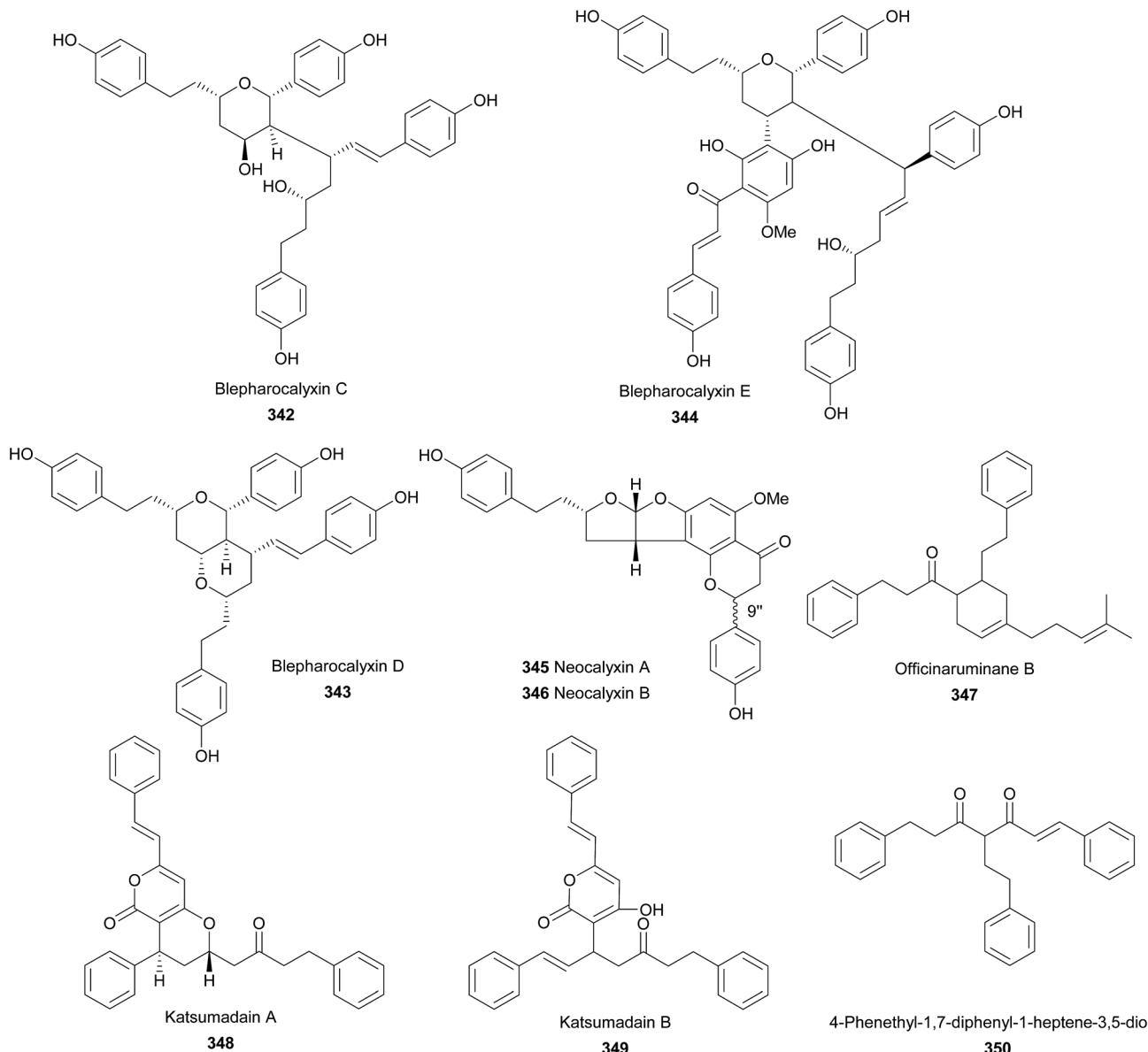
T. L. Wu et Senjen and *A. katsumadai* Hayata provided (*3S,5S*)-*trans*-3,5-dihydroxy-1,7-diphenyl-1-heptene (226).<sup>118,119</sup> It did not showed antimycobacterial activity ( $\text{MIC} \geq 64 \text{ mg L}^{-1}$ ). Instead,

it exhibited weak neuraminidase inhibitory activity ( $\text{IC}_{50} = 29.75 \pm 8.15 \mu\text{M}$ ) *in vitro*.<sup>56,120</sup> (*E,E*)-5-Hydroxy-1,7-diphenyl-4,6-heptadien-3-one (227), (*S*)-1,7-diphenyl-6(*E*)-hepten-3-ol (228),

Fig. 5 Diarylheptanoids from *Alpinia* species (continued).

and alnustone (229) were isolated from *A. katsumadai* Hayata with significantly neuraminidase inhibitory *in vitro* with IC<sub>50</sub> values between 1.0 and 6.1  $\mu$ M.<sup>56</sup> In addition, 229 also possessed

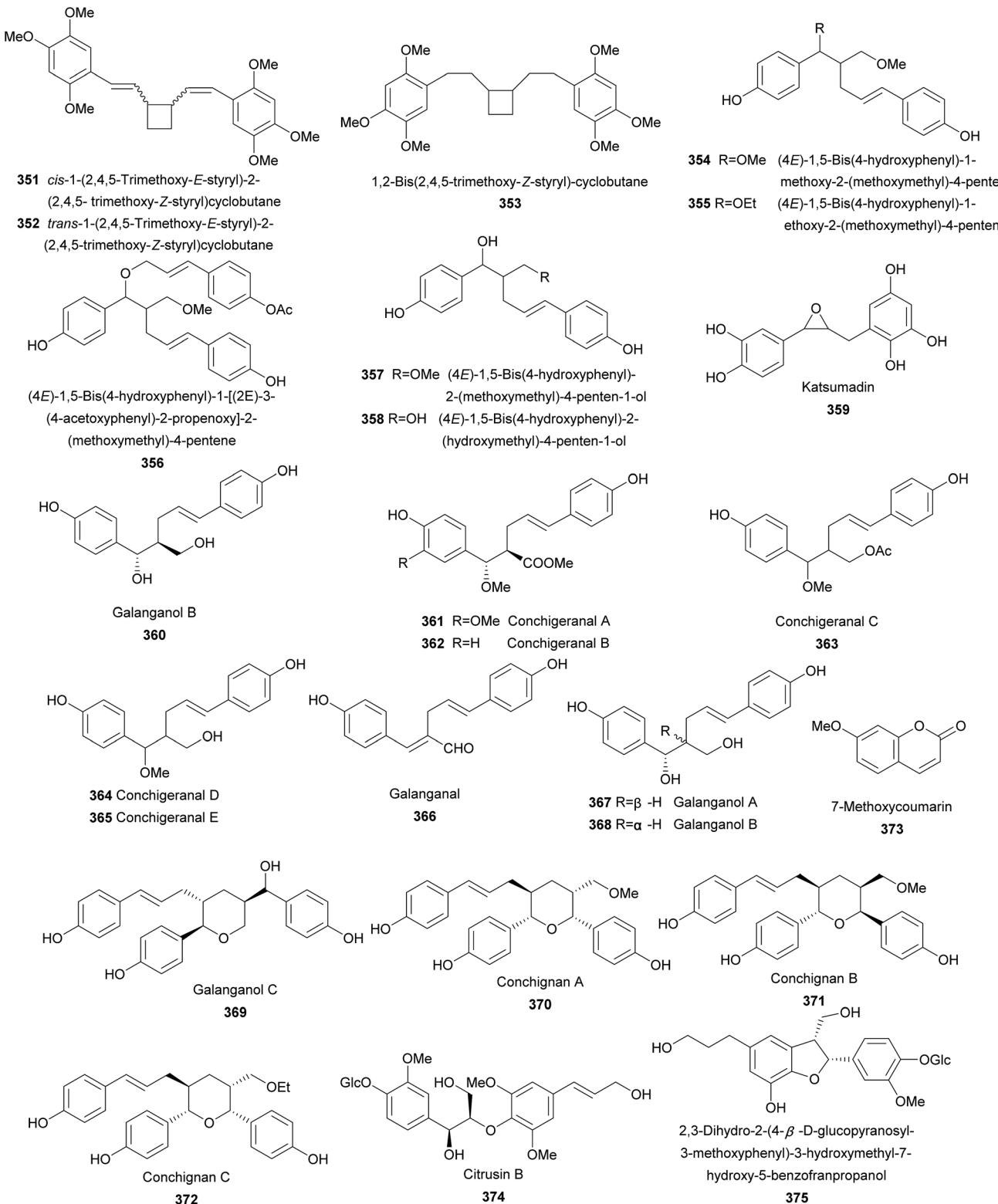
antiemetic,<sup>121</sup> antimycobacterial activities,<sup>120</sup> and significantly inhibited proliferation of Bel 7402 and L0-2 cells.<sup>122</sup> Investigation of *A. katsumadai* Hayata also led the isolation of

Fig. 5 Diarylheptanoids from *Alpinia* species (continued).

compounds 230–238.<sup>49,119,121,123–125</sup> 1,7-Bis(4-hydroxyphenyl)-1,4,6-heptatrien-3-one (239) and bisdemethoxycurcumin (240) were obtained from rhizomes of *A. galanga* (L.) Willd, both of which significantly inhibited the proliferation of melanoma cells and indistinctively inhibited cellular tyrosinase.<sup>126</sup> A planar structure of 1,7-diphenyl-5-hydroxy-6-hepten-3-one (241) was reported from *A. nutans* Rosc.,<sup>127</sup> *A. rafflesiana* Wall.ex.Bak.,<sup>128</sup> and *A. officinarum* Hance.<sup>129</sup> While its enantiomers, 5*S* (241a) and 5*R* (241b) counterparts, were identified from *A. mutica* Roxb.<sup>130</sup> and *A. katsumadai* Hayata,<sup>119</sup> respectively. It was shown that a large amount of diarylheptanoids (242–276) were obtained from the rhizomes of *A. officinarum* Hance.<sup>27,131–136</sup> 7-(3,4-Dihydroxyphenyl)-1-(4-hydroxy-3-methoxyphenyl)-4-en-3-heptanone (257) displayed moderate cytotoxicity against human tumor cell lines of HepG2, MCF-7, and SF-268. While (4*E*,6*E*)-5-hydroxy-1-(4-hydroxy-3-methoxyphenyl)-7-phenylhepta-4,6-dien-3-one (258)

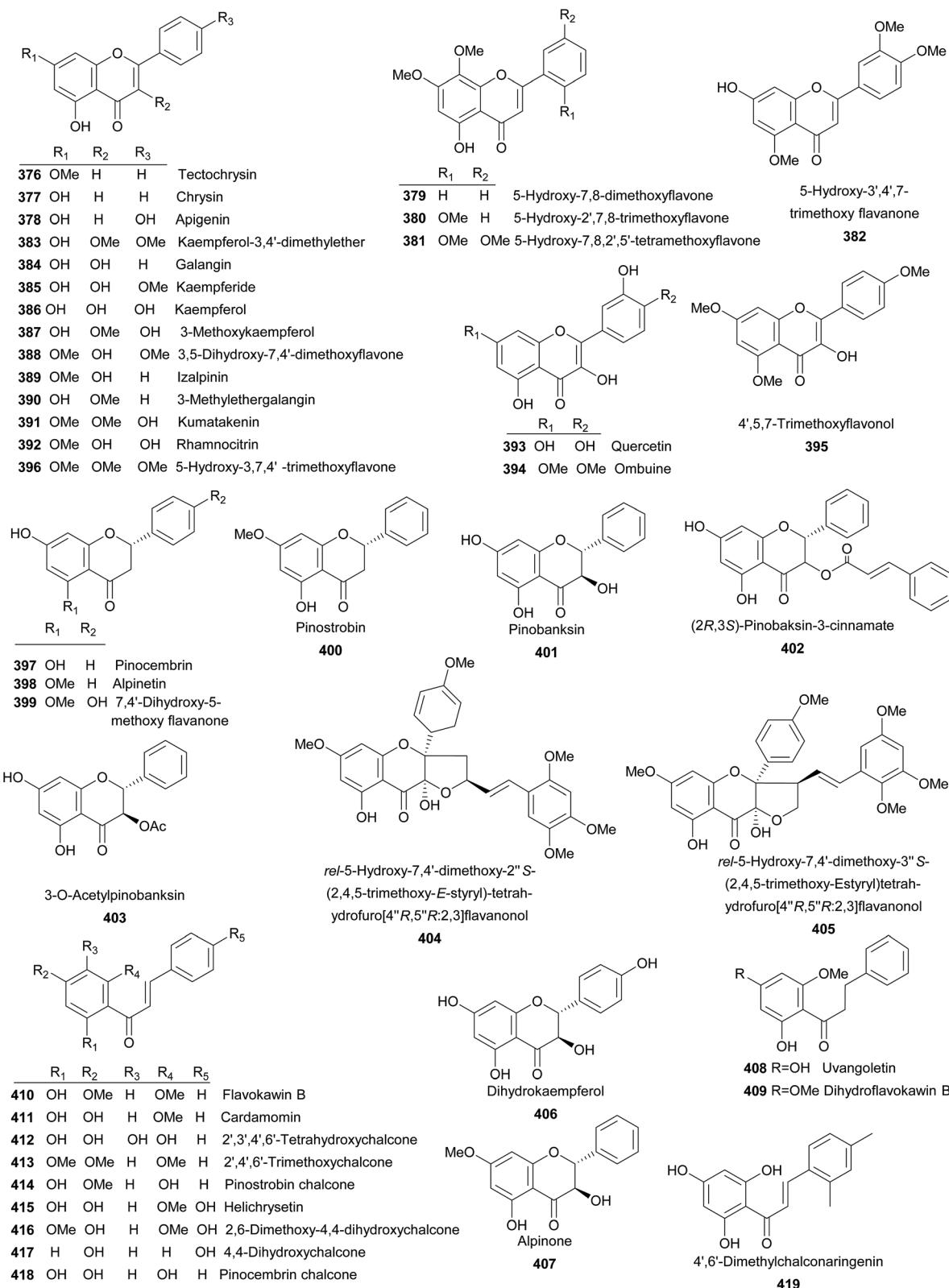
showed weak cytotoxicity against two cancer cell lines of MCF-7 and T98G with  $IC_{50}$  values of 22.68 and 4.44  $\mu$ M, respectively.<sup>135</sup> Meanwhile, 258–267 were proved to be inhibitors of *Helicobacter pylori* (Hp-Sydney and Hp-F44).<sup>129</sup> AO-5 (263) showed anti-inflammatory activity induced by 12-O-tetradecanoylphorbol-13-acetate (TPA), platelet-activating factor (PAF), and NO.<sup>110,136,137</sup> Moreover, it exhibited very weak cytotoxic activity against human glioblastoma T98G cells ( $IC_{50} = 27 \mu$ M).<sup>138</sup> The acetone extract of the rhizomes of *A. officinarum* Hance showed 5*α*-reductase inhibitory effect, which was superior to the drug used in the treatment of androgen-dependent disorders. Therefore, a bioactivity-guided isolation was performed and resulted in the isolation of 263–266 which exerted 5*α*-reductase inhibitory effect with  $IC_{50}$  values ranging from 220 to 390  $\mu$ M, indicating potent usage in treating androgen-dependent diseases.<sup>139</sup> Besides, AO-1 (266) also showed anti-helicobacter pylori, hypolipidemic activities, and NO



Fig. 6 Lignans from *Alpinia* species.

production inhibitory activity.<sup>110,140,141</sup> AO-2 (267) was identified as an inhibitor of prostaglandin (PG) biosynthesis and exerted anti-oxidant activity.<sup>142,143</sup> It is interesting to note that dihydroxyabushiketol (264), AO-1 (266), and AO-2 (267) were firstly reported

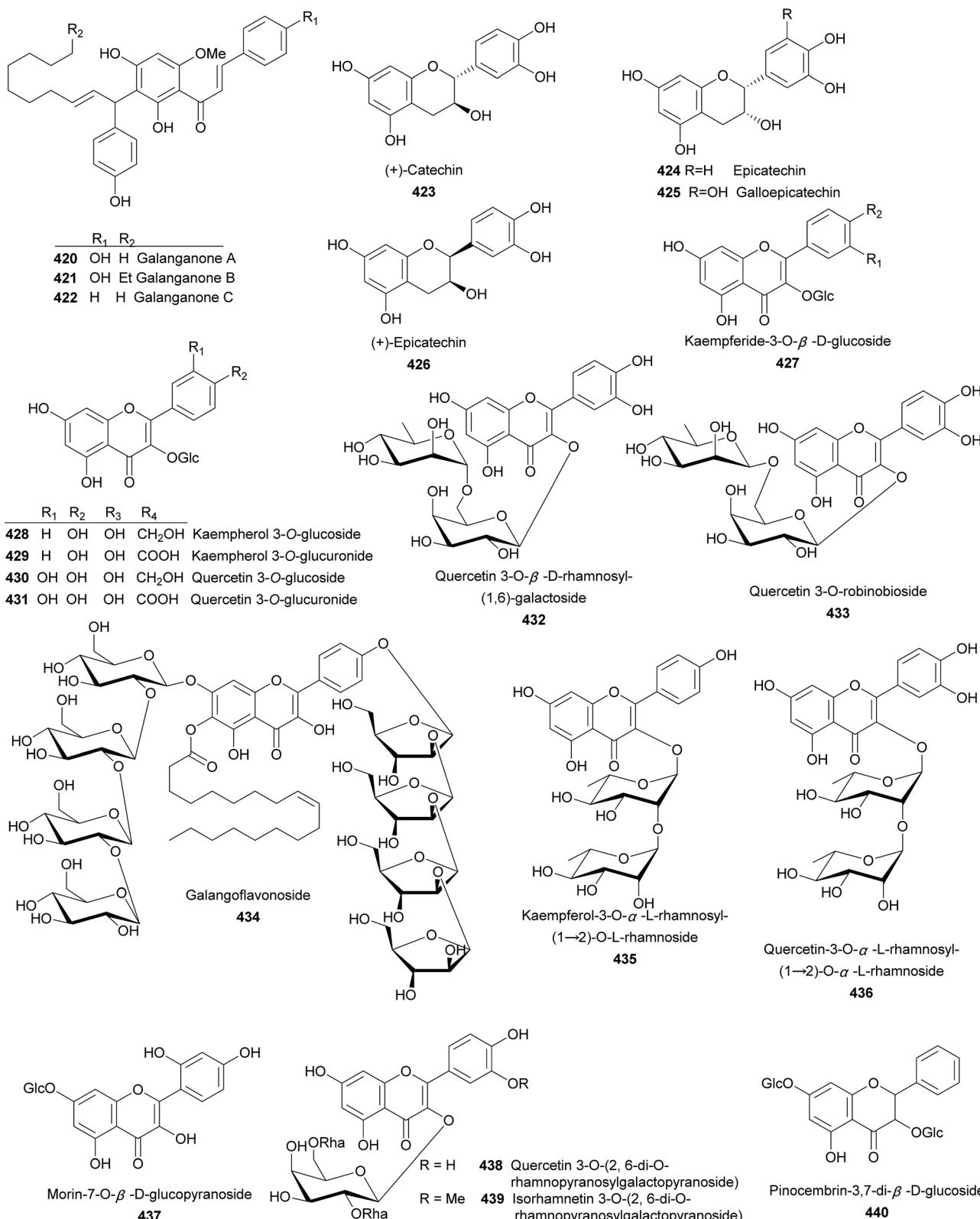
as planar structures, and later, their absolute configurations were established as 264a, 266a, and 267a, respectively.<sup>136,144</sup> 7-(4"-Hydroxy-3"-methoxyphenyl)-1-phenyl-3,5-heptadione (268) also exhibited prostaglandin biosynthesis inhibitory effect

Fig. 7 Flavonoids from *Alpinia* species.

with  $\text{IC}_{50}$  values of 50  $\mu\text{M}$ .<sup>143</sup> AO-4 (269) was found to have marked inhibitory effect on TPA-induced inflammation and antioxidant activity.<sup>142,144</sup> 6-Hydroxy-1,7-diphenyl-4-en-3-heptanone (270) was

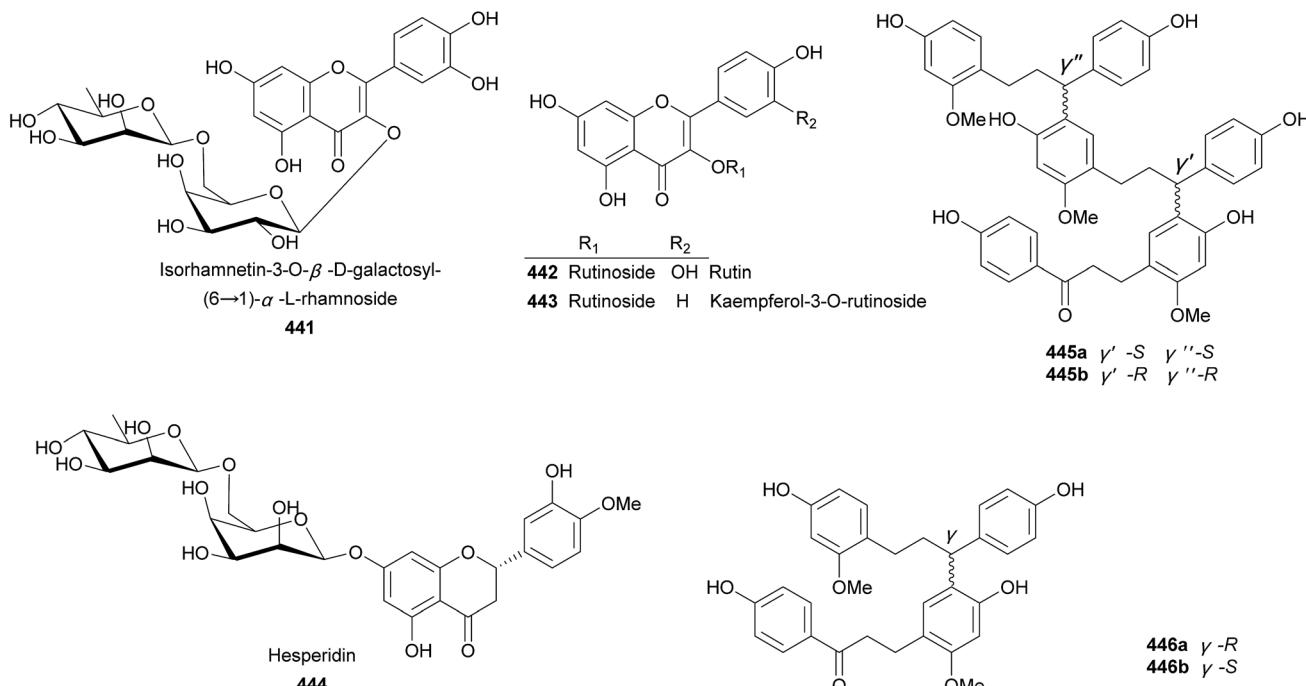
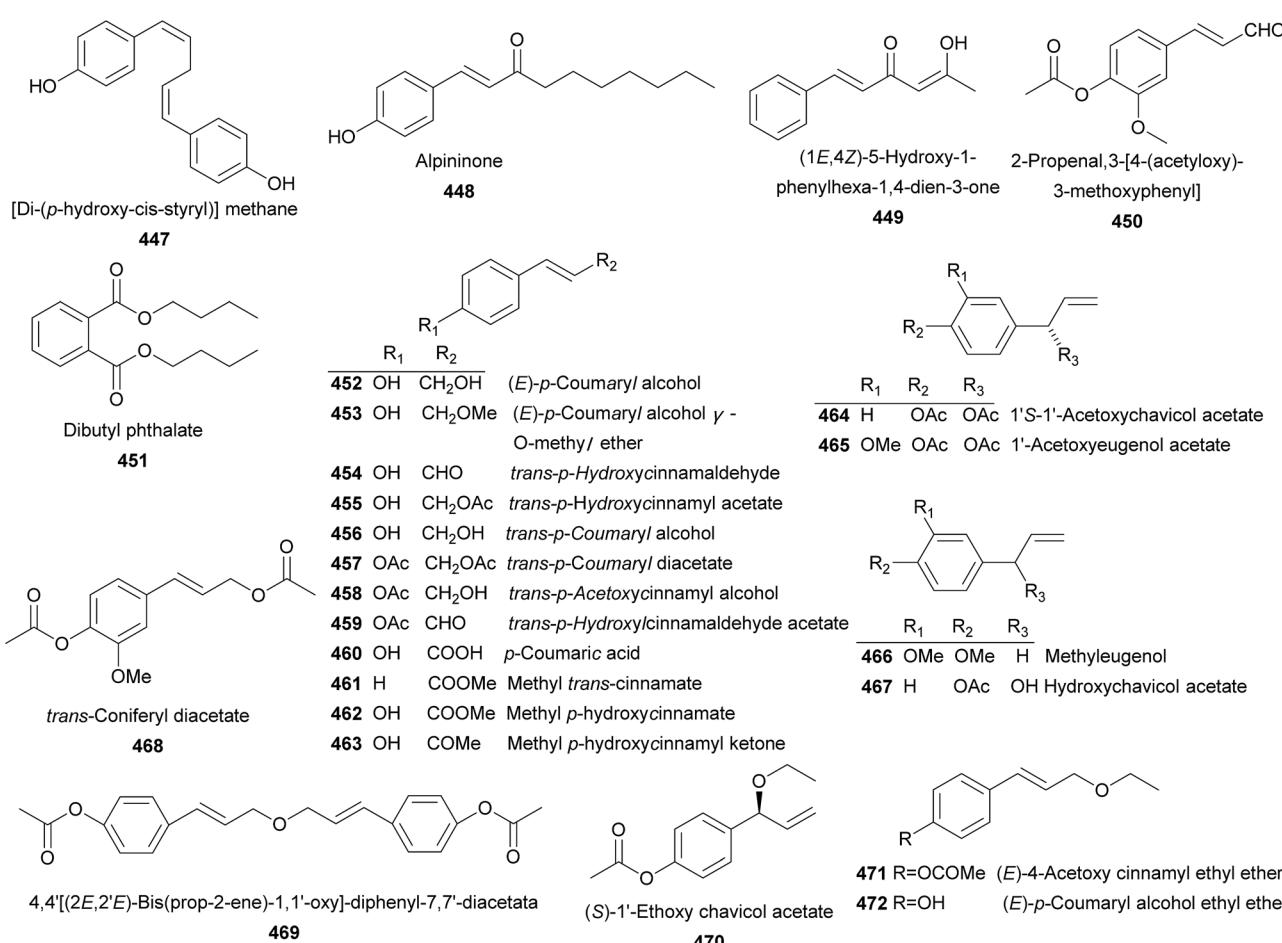
a PAF inhibitor.<sup>137</sup> AO-3 (271) and (5S)-5-methoxy-1,7-diphenyl-3-heptanone (272) displayed potent inhibitory effects on TPA-induced inflammation in mice with 50% of inhibition at a dose

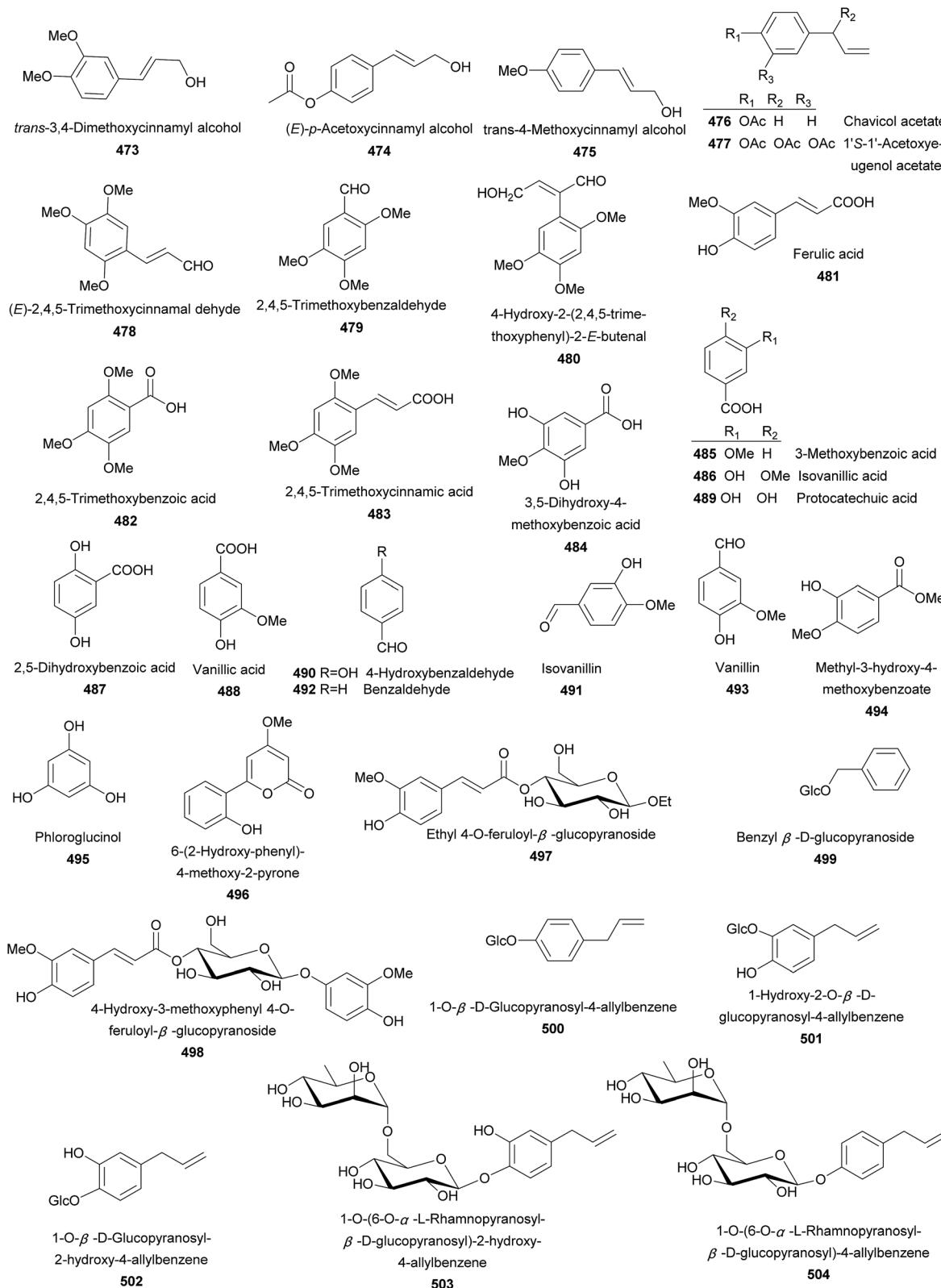


Fig. 7 Flavonoids from *Alpinia* species (continued).

of 0.8–2.7  $\mu$ mol per ear.<sup>144</sup> (3*R*,5*R*)-1-(4-Hydroxyphenyl)-7-phenyl-3,5-heptanediol (273) showed significantly antiemetic effect induced by CuSO<sub>4</sub> with 37.7% inhibition at a dose of 50 mg kg<sup>-1</sup>.<sup>27,145</sup>

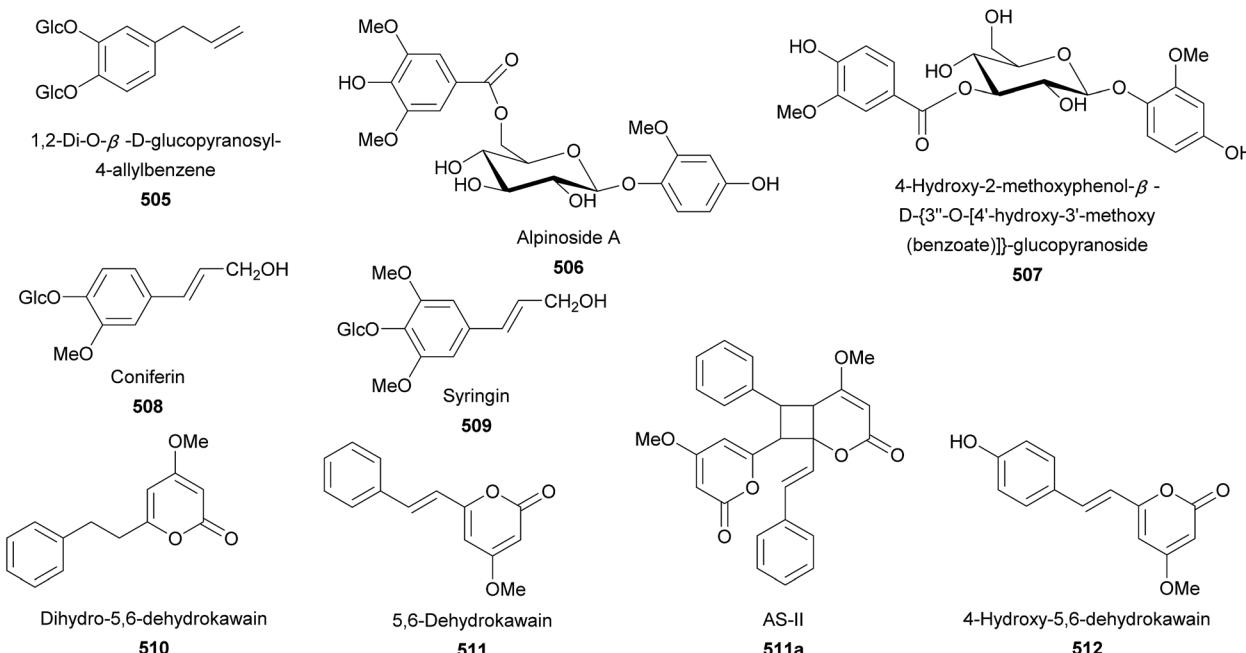
Investigation on seeds of *A. blepharocalyx* K. Schum. led to the isolation of ten cyclic diarylheptanoids (274–283).<sup>115,146–148</sup> Rhizomes of *A. officinarum* Hance provided 3,6-furan-7-(4-

Fig. 7 Flavonoids from *Alpinia* species (continued).Fig. 8 Phenolics from *Alpinia* species.

Fig. 8 Phenolics from *Alpinia* species (continued).

hydroxy-3''-methoxyphenyl)-1-phenylheptane (284).<sup>131</sup> From the seeds of *A. katsumadai*, 285–292 were obtained,<sup>149</sup> three of which (285–287) displayed weak antiproliferative activities against

four cancer cell lines of NCI-H460, HeLa, SMMC-7721, and HCT-116 with IC<sub>50</sub> values of 15.39–42.24 mM.<sup>115,149</sup> *A. blepharocalyx* K. Schum. was the source of 293–305.<sup>115,148,150,151</sup> However,

Fig. 8 Phenolics from *Alpinia* species (continued).

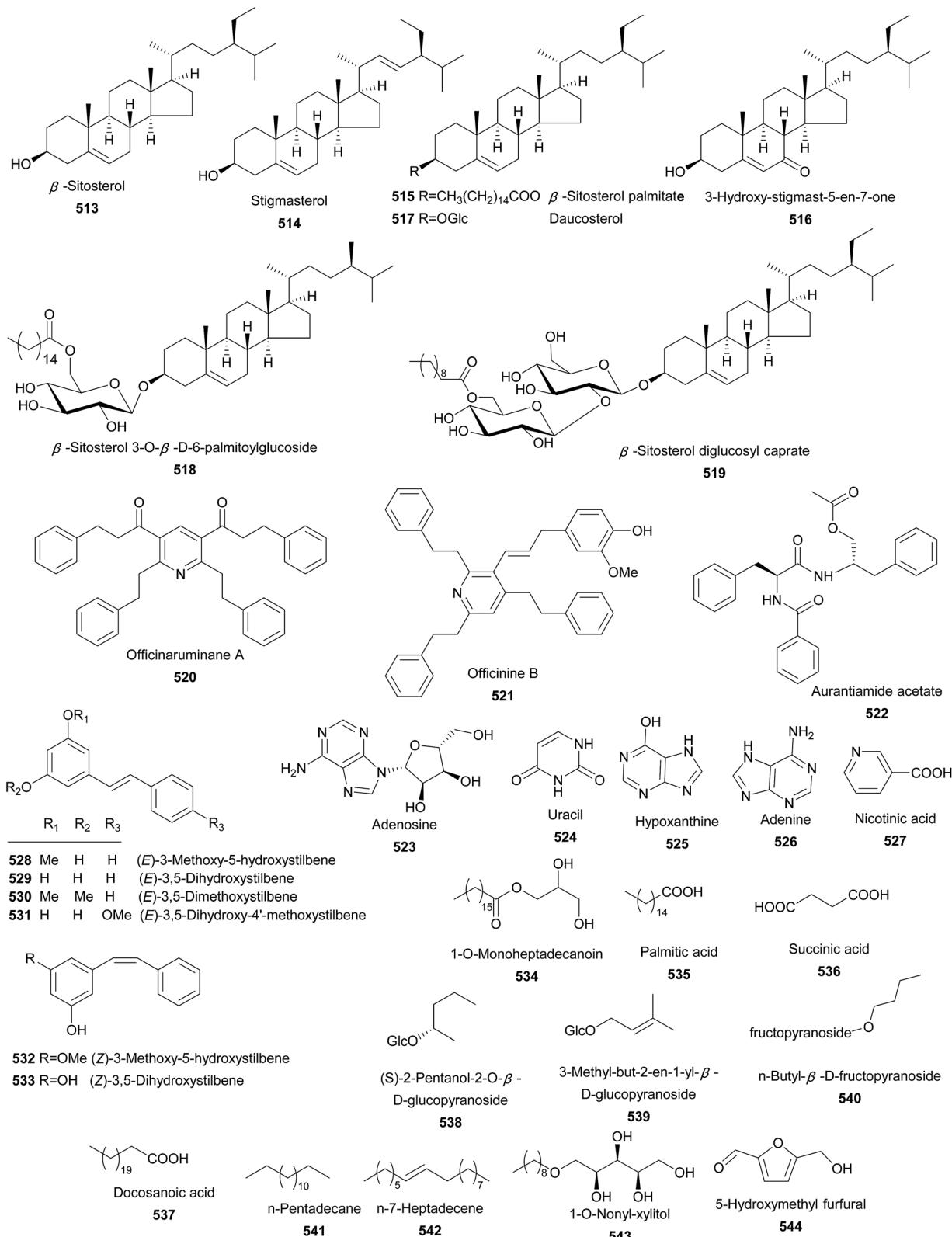
the stereochemistry at C-9" of six stereoisomeric (294/295, 296/297, 298/299, 300/301, 302/303, 304/305) remained unsolved. Calyxin J (298), epicalyxin J (299), calyxin K (300), and epicalyxin K (301) showed marked anti-proliferative activity against human HT-1080 fibrosarcoma cells with  $ED_{50}$  values from 0.3–8.2  $\mu\text{M}$ .<sup>115,152</sup> Compounds 302–305 were proved to inhibit NO production in endotoxin activated murine macrophage J774.1 with 90–94% inhibitory rate at a concentration of 100  $\mu\text{g mL}^{-1}$ .<sup>151</sup> Seeds of *A. katsumadai* Hayata provided 306–318. Calyxins Q (306) and R (307) exerted potent antiproliferative activities against four cancer cell lines of NCI-H460, HeLa, SMMC-7721, and HCT-116 at the level of  $IC_{50}$  values of 15.3–42.2  $\mu\text{M}$ .<sup>149</sup> Calyxin B (319) and epicalyxin B (320) were obtained from *A. blepharocalyx* K. Schum. and *A. pinnanensis* as NO production inhibitors.<sup>115,151</sup> In addition, 319 showed potent anti-proliferative activity against human HT-1080 fibrosarcoma cells with an  $ED_{50}$  value of 0.69  $\mu\text{M}$ .<sup>148</sup> Both *A. pinnanensis* T. L. Wu et Senjen and *A. katsumadai* Hayata yielded alpinnanin B (321).<sup>118,124</sup> From *A. katsumadai* Hayata and *A. blepharocalyx* K. Schum., epicalyxin H (322) and calyxin H (323) were isolated.<sup>118,124,153</sup> Epicalyxin H was identified as NO production inhibitor.<sup>115,153</sup> Seeds of *A. blepharocalyx* yielded 324–330.<sup>115,152,154</sup> It's worth mentioning that all three structures of calyxin L (325), epicalyxin F (327), and calyxins F (328) in the Scifinder were wrong. Out of a series of diarylheptanoids bearing a chalcone or a flavanone moiety, epicalyxins I (326), F (327), and calyxin F (328) were shown to possess strong anti-proliferative activities toward colon 26-L5 carcinoma and HT-1080 fibrosarcoma with  $IC_{50}$  values ranging from 0.5 to 10.1  $\mu\text{M}$ .<sup>115,150</sup> Meanwhile, 326 and 327 were cytotoxic against human fibrosarcoma cells with  $IC_{50}$  values ranging from 0.9 to 12.1  $\mu\text{M}$ .<sup>152</sup> 6-Hydroxycalyxin F (329) and calyxin A (330) demonstrated NO production inhibitory activities with  $IC_{50}$  values of

49 and 62  $\mu\text{M}$ , respectively.<sup>115,150</sup> Rhizomes of *A. pinnanensis* T. L. Wu et Senjen provided deoxycalyxin A (331), alpinnanins A (332), and C (333).<sup>118</sup> In addition, 331 was also found in *A. blepharocalyx* K. Schum.<sup>115</sup> While officinin A (334) was obtained from rhizomes of *A. officinarum* Hance.<sup>155</sup>

Five dimeric diarylheptanoids (335–339) were obtained from rhizomes of *A. officinarum* Hance.<sup>135,136,138,156,157</sup> Only alpinin C (338) displayed selective cytotoxic against MCF-7 ( $IC_{50} = 62.3 \mu\text{M}$ ) and T98G cells ( $IC_{50} = 57.3 \mu\text{M}$ ).<sup>135</sup> Seeds of *A. blepharocalyx* K. Schum. provided 340–344 possessing two diarylheptanoid units and a chalcone moiety.<sup>115,146,153</sup> Both blepharocalyxins A (340) and B (341) showed concentration-dependent inhibition in the range of 1–100  $\mu\text{g mL}^{-1}$  against NO production in endotoxin-activated murine macrophages J774.1.<sup>158</sup> Blepharocalyxins C–E (342–344) were tested for antiproliferative activities against two tested cancer cells, blepharocalyxin D (343) exhibited the strongest effect against highly liver-metastatic murine colon 26-L5 carcinoma cells ( $ED_{50} = 3.6 \mu\text{M}$ ), whereas blepharocalyxin E (344) showed the strongest activity against human HT-1080 fibrosarcoma cells ( $ED_{50} = 9.02 \mu\text{M}$ ).<sup>115,146,159</sup> It is worth mentioning that the stereochemistry at C-I-5 position for 343 in Scifinder was *S*, which was not correct and should be revised as *R*. Moreover, the two diarylheptanoid moieties in 344 were wrongly connected through C-I-6 and C-II-5 by Scifinder. Instead, it should be joined through C-I-6 and C-II-7. Two unusual diarylheptanoid derivatives, neocalyxin A (345) and its epimer neocalyxin B (346), were found from the seeds of *A. blepharocalyx* K. Schum., with the stereochemistry at C-9" undetermined.<sup>115,152</sup>

Rhizomes of *A. officinarum* Hance produced officinaruminane B (347), a diarylheptanoid coupled with a monoterpene unit.<sup>131</sup> Investigation on seeds of *A. katsumadai* Hayata identified two novel anti-emetic diarylheptanoids, katsumadains A (348) and B



Fig. 9 Steroids, alkaloids, stilbenes, and other miscellaneous compounds from *Alpinia* species

(349).<sup>160</sup> Besides, 348 also exerted promising neuraminidase inhibitory effect against human influenza virus A/PR/8/34 (IC<sub>50</sub> = 1.05  $\mu$ M).<sup>56</sup> 4-Phenethyl-1,7-diphenyl-1-heptene-3,5-dione (350)

was isolated from rhizomes of *A. officinarum* Hance. It exhibited weak antibacterial activity against Hp-Sydney and Hp-F44 with the MIC values of 23.6–31.4 and 78.5  $\mu$ M, respectively.<sup>129</sup>

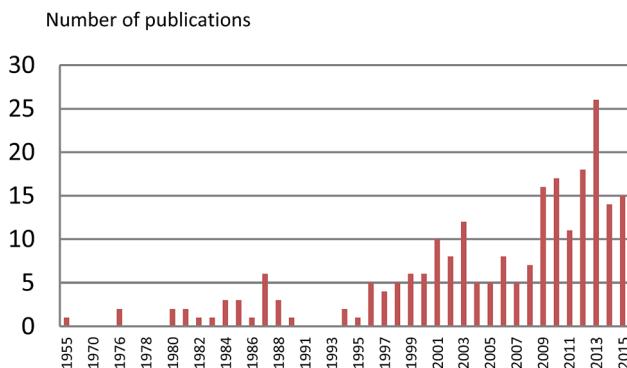


Fig. 10 The number of publications on *Alpinia* since 1955.

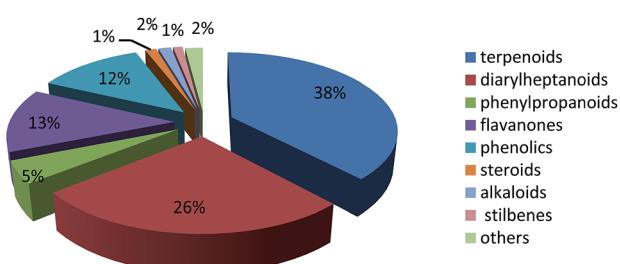
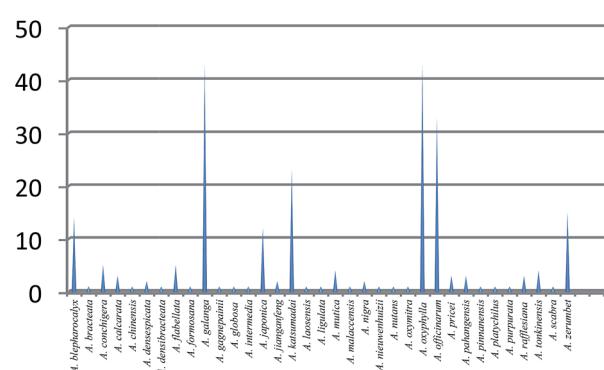


Fig. 11 The percentage of each type of compounds from *Alpinia* species



**Fig. 12** The number of published papers for each investigated *Alpinia* species on chemical constituents and their bioactivities over last six decades since 1955.

#### 4. Lignans

Twenty-four lignans (351–374) were reported from the genus of *Alpinia* (Fig. 6). Separation for leaves of *A. flabellata* Ridley resulted in the isolation of 351–353, three phenylbutanoid dimers bearing a novel tetracyclic moiety.<sup>161,162</sup> *cis*-1-(2,4,5-Tri-methoxy-*E*-styryl)-2-(2,4,5-trimethoxy-*Z*-styryl)cyclobutane (351) and *trans*-1-(2,4,5-trimethoxy-*E*-styryl)-2-(2,4,5-trimethoxy-*Z*-styryl)cyclobutane (352) showed weak antibacterial against *Staphylococcus aureus* with MIC values of 5.0 and 2.5 mM, respectively.<sup>161</sup> Furthermore, 351 significantly decreased the ovalbumin permeability in intestinal cells.<sup>161</sup>

Rhizomes of *A. officinarum* Hance yielded 354–358 containing a rare  $\beta$ - $\gamma$  linkage. All five compounds exhibited weak antioxidant activities against the autoxidation of methyl linoleate in bulk phase.<sup>163</sup> Extracts of seeds of *A. katsumadai* Hayata afforded antiemetic katsumadin (359) with antiemetic activity on CuSO<sub>4</sub>-induced emesis in young quail.<sup>121</sup> Galanganol B (360) was isolated from rhizomes of *A. galanga* (L.) Willd.<sup>164</sup> Investigation on the whole plant of *A. conchigera* afforded eight rare 8-9' linked neolignans 361–368.<sup>165</sup> Although conchigeranals D (364) and E (365) shared the same planar structure, their relative configurations were not be determined. Galanganal (366), galanganols A (367), and B (368) were also found from rhizomes of *A. galanga* (L.) Willd.<sup>166</sup> Compounds 361–367 exhibited significant cytotoxic activity against cancer Hela cells with IC<sub>50</sub> values ranging from 1.5 to 5.29  $\mu\text{g mL}^{-1}$ .<sup>165</sup> Interestingly, 366 and 368 also inhibited NO production in mouse peritoneal macrophages.<sup>166</sup> Galanganol C (369) was obtained from rhizomes of *A. galanga* (L.) Willd as a NO production inhibitor.<sup>166</sup> The whole plant of *A. conchigera* yielded three unusual sesquineolignans, conchignans A–C (370–372) bearing a tetrahydropyran ring.<sup>167</sup> 7-Methoxycoumarin (373) is a coumarin known from *A. calcarata* Rose.<sup>85</sup>

Citrusin B (374) and 2,3-dihydro-2-(4- $\beta$ -D-glucopyranosyl-3-methoxyphenyl)-3-hydroxymethyl-7-hydroxy-5-benzofranpropanol (375) were the only two lignan glycosides isolated from leaves of *A. speciosa*.<sup>168</sup>

## 5. Flavonoids

To date, 71 flavanoides (Fig. 7) were isolated from the *Alpinia* species, including seven flavones (376–382), 14 flavonols (383–396), four flavanones (397–400), seven flavanonols (401–407), two dihydrochalcones (408 and 409), 13 chalcones (410–422), four flavanols (423–426), and 18 flavonoid glycosides (427–444), two flavonoid oligomers (445 and 446).

Tectochrysin (376) and chrysin (377) were isolated from *A. oxyphylla* Miq. and exhibited moderate anti-inflammatory activities against LPS-induced NO production in RAW264.7 macrophage cells.<sup>169</sup> Both *A. bracteata* and *A. officinarum* Hance produced apigenin (378), which displayed moderate activity on scavenging DPPH free radicals ( $EC_{50} = 90 \pm 1.5 \mu\text{M}$ ).<sup>170</sup> *A. galanga* (L.) Willd was the source of 379–381 and *A. tonkinensis* Gagnep. produced 5-hydroxy-3',4',7-trimethoxy flavanone (382).<sup>102,171</sup> Kaempferol-3,4'-dimethylether (383) was afforded by *A. sichuanensis* Z. Y. Zhu.<sup>52</sup> Galangin (384) and kaempferide (385) were the major flavonols distributed in several plants of *Alpinia*, both of which exhibited inhibitory against penicillinase and potent antioxidant activities.<sup>113,172</sup> In addition, galangin effectively inhibited the TPA-induced invasion and migration of HepG2 cells at concentrations of 2.5–5  $\mu\text{M}$ .<sup>173</sup> In 2001, a review summarized anti-genotoxic activity of galangin and demonstrated that galangin was a promising candidate for cancer chemoprevention.<sup>174</sup> Investigation on the whole plant of *A. sichuanensis* Z. Y. Zhu provided kaempferol (386).<sup>52</sup> From *A. speciosa*, *A. galanga* (L.) Willd, *A. katsumadai* Hayata, and *A. tonkinensis* Gagnep., 3-methoxykaempferol (387) was isolated.<sup>175–178</sup> While *A. flabellata* Ridley, *A. oxyphylla*, and *A.*

*tonkinensis* Gagnep. yielded 3,5-dihydroxy-7,4'-dimethoxyflavone (388).<sup>113,161,171</sup> Izalpinin (389) from different parts of *A. oxyphylla* Miq. was a NO production inhibitor and exhibited potent antioxidant activity.<sup>113,176</sup> From rhizomes of *A. officinarum*, 3-methylethergalangin (390) was identified as an inhibitor of pancreatic lipase with an  $IC_{50}$  value of 1.3 mg mL<sup>-1</sup>.<sup>179</sup> Compounds 391–395 were mainly obtained from *A. tonkinensis* Gagnep.<sup>171</sup> 5-Hydroxy-3,7,4'-trimethoxyflavone (396) was yielded by leaves of *A. flabellata* Ridley.<sup>180</sup> Pinocembrin (397) and alpinetin (398) were distributed in several *Alpinia* species and both showed antiemetic activities.<sup>121,181</sup> In addition, 397 also demonstrated several bioactivities, such as cytotoxicity (on human T4 lymphoblastoid cancer cells),<sup>182</sup> anti-inflammation,<sup>169</sup> and antiplatelet aggregation etc.<sup>183</sup> While, 398 was a PAF receptor binding inhibitor.<sup>184</sup> 7,4'-Dihydroxy-5-methoxyflavone (399), pinostrobin (400) were reported from several species.<sup>116,118,128,182</sup> Pinobanksin (401), (2R,3S)-pinobanksin-3-cinnamate (402), and 3-O-acetylpinobanksin (403) were mainly obtained from *A. galanga* (L.) Willd and *A. katsumadai* Hayata.<sup>176,177,185</sup> Compound 402 showed potent neuroprotective effect against PC12 cells.<sup>177,186</sup> Leaves of *A. flabellata* Ridley provided 404 and 405.<sup>180</sup> Dihydrokaempferol (406) were isolated from *A. oxyphylla*.<sup>169</sup> Both *A. japonica* (Thunb.) Miq. and *A. galanga* (L.) Willd were sources for alpinone (407).<sup>176,187</sup> From seeds of *A. katsumadai* Hayata, a dihydrochalcone uvangoletin (408) was isolated.<sup>108</sup> *A. speciosa* K. Schum. and *A. formosana* afforded another dihydrochalcone, dihydroflavokawin B (409).<sup>81,189</sup> Flavokawin B (410) was isolated from several plants and showed strong cytotoxicity against human T4 lymphoblastoid cancer cells ( $IC_{50}$  = 6.5  $\mu$ M) and anti-inflammatory activity.<sup>182,189</sup> Cardamomin (411) distributed in many *Alpinia* species<sup>100,118,123,188,190</sup> and exhibited extensive bioactivities including death receptor 5 (DR5) promotor,<sup>175</sup> antimicrobial,<sup>191</sup> antiemetic,<sup>121</sup> anti-coagulation,<sup>183</sup> and anti-inflammation.<sup>128</sup> Interestingly, it also protected septic mice from acute lung injury by preventing endothelial barrier dysfunction.<sup>192</sup> 2',3',4',6'-Tetrahydroxychalcone (412), which was obtained from *A. rafflesiana* Wall.ex.Bak., was potently active to DPPH free radical scavenging ( $IC_{50}$  = 55  $\mu$ M).<sup>128</sup> Rhizomes of *A. pricei* Hayata yielded 2',4',6'-trimethoxychalcone (413) and pinostrobin chalcone (414).<sup>189</sup> Compounds 415–417 were isolated from the seeds of *A. blepharocalyx* K. Schum.,<sup>116,117</sup> while helichrysetin (415) was also found in *A. katsumadai* Hayata.<sup>108</sup> Pinocembrin chalcone (418) and 4',6'-dimethylchalconaringenin (419) were provided by *A. katsumadai* Hayata and *A. pinnanensis* T. L. Wu et Senjen, respectively.<sup>118,181</sup> Compound 418 was also isolated from *A. platychilus*.<sup>193</sup> Galanganones A–C (420–422) were three novel chalcones bearing a long-chain alkylphenol from *A. galanga*.<sup>194</sup> Whilst *A. katsumadai* Hayata and *A. zerumbet* (Pers.) B. L. Burtt Smith. provided (+)-catechin (423).<sup>195,196</sup> Epicatechin (424) and galloepicatechin (425) were yielded by *A. oxymitra* K. Schum.<sup>75</sup> (+)-Epicatechin (426) was isolated from *A. speciosa* K. Schum. and displayed antioxidant activity.<sup>197</sup> Kaempferide-3-O- $\beta$ -D-glucoside (427) from *A. officinarum* Hance had an weak inhibitory activity against penicillinase.<sup>172</sup> Study on *A. speciosa* K. Schum. lead to the isolation of 428–432.<sup>198</sup> Quercetin 3-O-robinoibioside (433) and galangoflavonoside (434) were obtained

from *A. katsumadai* Hayata and *A. galanga* (L.) Swartz., respectively.<sup>196,199</sup> Compounds 435–437 from *A. densespica* Hayata exhibited moderate NO inhibitory activities.<sup>103</sup> Compounds 438–440 were obtained from the seeds of *A. katsumadai* Hayata and isorhamnetin-3-O- $\beta$ -D-galactosyl-(6 → 1)- $\alpha$ -L-rhamnoside (441) was isolated from rhizomes of *A. tonkinensis* Gagnep.<sup>51,196</sup> Leaves of *A. zerumbet* (Pers.) B. L. Burtt Smith. contained rutin (442) and kaempferol-3-O-rutinoside (443).<sup>195</sup> The whole plant of *A. sichuanensis* Z. Y. Zhu yielded hesperidin (444).<sup>52</sup> Two pairs of enantiomers of flavonoidoligomers (445a and 445b, 446a and 446b) were found from rhizomes of *A. platychilus*. The compounds mixture of 446a and 446b showed anticoagulant activity on the prolongation of both prothrombin times (PT) and the thrombin times (TT) with a dose-effect relationship at 6.25–100 mM.<sup>193</sup>

## 6. Phenolics

A total number of 66 phenolics (447–512) were obtained from *Alpinia* species (Fig. 8). [Di-(*p*-hydroxy-cis-styryl)]methane (447) was obtained from *A. galanga* (L.) Willd.<sup>200</sup> Whist alpininone (448) was isolated from *A. gagnepainii* K. Schum. with antibacterial effect against *E. coli*, *B. subtilis*, and *S. aureus* with the same MIC value of 12.5  $\mu$ g mL<sup>-1</sup>.<sup>191</sup> (1E,4Z)-5-Hydroxy-1-phenylhexa-1,4-dien-3-one (449) and 2-propenal, 3-[4-(acetoxy)-3-methoxyphenyl] (450) were provided by *A. katsumadai* Hayata and *A. galanga* (L.) Willd, respectively.<sup>86,108</sup> From *A. sichuanensi* and *A. oxyphylla*, dibutyl phthalate (451) was isolated.<sup>52,201</sup> Two compounds named as (*E*)-*p*-coumaryl alcohol (452) and (*E*)-*p*-coumaryl alcohol  $\gamma$ -O-methyl ether (453) exhibited potent inhibitory activities against the autoxidation of methyl linoleate in bulk phase.<sup>163</sup> In addition, compound 453 exerted potent cytotoxic activity against the SNU638 cells with  $IC_{50}$  value of 1.62  $\mu$ g mL<sup>-1</sup>.<sup>202</sup>

*A. galanga* (L.) Willd and *A. conchigera* Griff. produced *trans*-*p*-hydroxycinnamaldehyde (454) and *trans*-*p*-hydroxycinnamyl acetate (455).<sup>170,203</sup> Compound 454 displayed weak antiallergic effect,<sup>204</sup> and NO production inhibitory activities ( $IC_{50}$  = 20  $\mu$ M),<sup>166</sup> and 455 exerted no inhibitory activity towards *Staphylococcus aureus* strain VISA (MIC = 203 mM).<sup>82</sup> *trans*-*p*-Coumaryl alcohol (456) was a weak NO production inhibitor from *A. galanga* (L.) Willd ( $IC_{50}$  = 72  $\mu$ M).<sup>166</sup> *trans*-*p*-Coumaryl diacetate (457) from *A. galanga* showed a number of bioactivities, including anti-allergy,<sup>204</sup> efflux pump inhibition,<sup>205</sup> NO production inhibition,<sup>166</sup> xanthine oxidase inhibition,<sup>206</sup> antileishmania,<sup>164</sup> cytotoxicity,<sup>203</sup> and antibacteria.<sup>82</sup> *trans*-*p*-Acetoxy cinnamyl alcohol (458), *trans*-*p*-hydroxycinnamaldehyde acetate (459), and *p*-coumaric acid (460) were obtained from rhizomes of *A. galanga* (L.) Willd.<sup>164,205</sup> In addition, compound 460 was also distributed in *A. galanga* (L.) Willd,<sup>164</sup> *A. sichuanensis* Z. Y. Zhu,<sup>52</sup> *A. speciosa*,<sup>198</sup> *A. blepharocalyx* K. Schum.,<sup>116</sup> and *A. oxyphylla*.<sup>169</sup> Both *A. formosana* and *A. speciosa* K. Schum. were sources of methyl *trans*-cinnamate (461).<sup>81,188</sup> Seeds of *A. blepharocalyx* yielded methyl *p*-hydroxycinnamate (462) and methyl *p*-hydroxycinnamyl ketone (463).<sup>116</sup> From rhizomes of *A. galanga* (L.) Willd, 12 compounds (464–475) were obtained.<sup>33,82,203,207</sup> Among them, 1S-1'-acetoxychavicol acetate (464) and 1-acetoxyeugenol acetate (465) were the most



abundant phenylpropanoids presented in *A. galanga* (L.) Swartz., *A. officinarum* Hance, and *A. conchigera* Griff. They were reported to have anti-ulcer,<sup>24</sup> antileishmanial,<sup>164</sup> and antitumor bioactivities.<sup>33,202,208</sup> Furthermore, **464** also showed antiallergic,<sup>204</sup> efflux pump inhibitory,<sup>205</sup> NO production inhibitory,<sup>166</sup> xanthine oxidase inhibitory,<sup>206</sup> gastroprotective,<sup>209</sup> anti-HIV,<sup>210</sup> anti-cancer,<sup>86</sup> antibacterial,<sup>30,211</sup> plant growth-inhibitory and fungal growth-inhibitory activities.<sup>212</sup> Two compounds, methyleugenol (**466**) and hydroxychavicol acetate (**467**), were isolated from *A. galanga* (L.) Willd.<sup>82,164,166,204,211</sup> It was demonstrated that **467**, a chavicol acetate analogue, suppressed T-bet expression in Th cells.<sup>211</sup> Besides, **467** also showed weak antibacterial activity against *Staphylococcus aureus* strain VISA (MIC = 0.8 mM).<sup>82</sup> *trans*-Confiferyl diacetate (**468**) was proved to be a xanthine oxidase inhibitor.<sup>206</sup> Three new phenolics **469**, **470**, and **471**, along with four known ones **472–475** were also yielded by *A. galang*.<sup>33,203,207</sup> Chavicol acetate (**476**) and 1'S-acetoxeugenol acetate (**477**) were two known phenolics found from *A. conchigera* Griff.<sup>82</sup> Compound **477** possessed antibacterial,<sup>82</sup> xanthine oxidase inhibitory,<sup>206</sup> gastro-protective,<sup>209</sup> and anti-cancer activities.<sup>86</sup> Investigation on leaves of *A. flabellata* Ridley provided **478–480**, with strong antibacterial activities against *Staphylococcus aureus*.<sup>161,162,180</sup> Compounds **481–489** were nine phenolic acids isolated from several *Alpinia* species.<sup>52,89,122,180,198,213,214</sup> Protocatechic acid (**489**) showed potent neuroprotective effect on MPP<sup>+</sup>-induced neurotoxicity and H<sub>2</sub>O<sub>2</sub>-induced oxidative damage in PC12 cells.<sup>215–219</sup> In addition, it also exerted anti-aging effect on spleen and liver antioxidative system of senescent mice.<sup>31</sup> 4-Hydroxybenzaldehyde (**490**), isolated from *A. sichuanensis* Z. Y. Zhu, *A. blepharocalyx* K. Schum., *A. bracteata*, and *A. galanga* (L.) Willd,<sup>52,116,117,166,170</sup> didn't show any DPPH radical-scavenging activity. Instead, it exhibited inhibitory activity on xanthine oxidase (IC<sub>50</sub> = 19.6 μM).<sup>170,206</sup> Compounds **491–496** were provided by several *Alpinia* plants.<sup>52,84,137,167,220</sup>

Ethyl 4-O-feruloyl-β-glucopyranoside (**497**) and 4-hydroxy-3-methoxyphenyl 4-O-feruloyl-β-glucopyranoside (**498**) were two new glucoside esters of ferulic acid from rhizomes of *A. speciosa*, both of which showed antioxidant activities.<sup>197</sup> Investigation on rhizomes of *A. officinarum* Hance yielded **499–506**.<sup>55</sup> While from rhizomes of *A. bracteata*, a new phenolic glycoside (**507**) was isolated and showed moderate antioxidant activity on scavenging DPPH free radicals (EC<sub>50</sub> = 169 ± 4.8 μM).<sup>170</sup> Leaves of *A. speciosa* K. Schum. provided coniferin (**508**) and syringin (**509**).<sup>168</sup>

Dihydro-5,6-dehydrokawain (**510**) and 5,6-dehydrokawain (**511**) were major chemical constituents in several *Alpinia* species.<sup>81,100,128,175,188,221</sup> They showed antiulcerogenic, antithrombotic,<sup>195</sup> antifungal,<sup>191</sup> anti-obesity,<sup>222</sup> and plant growth inhibitory activities.<sup>223</sup> Recently, it was reported that they could strongly inhibit HIV-1 integrase with respective IC<sub>50</sub> values of 4.4 and 3.6 μg mL<sup>-1</sup>. In addition, they exhibited mixed type of inhibition against neuraminidase with both IC<sub>50</sub> values of 25 μM.<sup>95</sup> Furthermore, **511** was also reported as a slow and time-dependent reversible inhibitor of neuraminidase, a moderated antioxidant, a strong inhibitor of skin diseases-related enzymes, and strong antiplatelet inhibitor.<sup>95,127,224</sup> Interestingly, a dimer of 5,6-dehydrokawain, AS-II (**511a**), was an artifact formed by photo-irradiation during the isolation procedure of *A. speciosa*

K. Schum. leaves.<sup>223</sup> 4-Hydroxy-5,6-dehydrokawain (**512**) was an α-pyrone isolated from *A. blepharocalyx* K. Schum. It displayed antiproliferative activity against murine colon 26-L5 carcinoma and human HT-1080 fibrosarcoma with ED<sub>50</sub> 20.7 and 20.1 μM, respectively.<sup>116,117</sup> It also showed inhibitory effect on platelet aggregation induced by collagen, arachidonic acid (AA), adenosine diphosphate, and ristocetin.<sup>96</sup>

## 7. Steroids

Seven steroids (Fig. 9) were isolated from *Alpinia* species including four cholestanes (**513–516**) and three sitosterol glycosides (**517–519**).<sup>27,52,89,116,118,225</sup> As it is the same in plants of the other genera, β-sitosterol (**513**) and stigmasterol (**514**) were also widely distributed in *Alpinia* species.<sup>52,82,89,118,178,191,226–228</sup> β-Sitosterol-3-O-β-D-6-palmitoylglucoside (**518**) showed potent antiemetic activity induced by CuSO<sub>4</sub>.<sup>27</sup>

## 8. Alkaloids

Officinaruminane A (**520**) and officinine B (**521**), two alkaloids of bi-diarylheptanoid connecting by a pyridine ring were contributed by rhizomes of *A. officinarum* Hance.<sup>131,157</sup> A study on seeds of *A. katsumadai* Hayata afforded another six alkaloids (522–527) (Fig. 9).<sup>108,196</sup>

## 9. Stilbenes

Six stilbenes, **528–533** (Fig. 9), were all isolated from aerial parts of *A. katsumadai* Hayata.<sup>29,121</sup>

## 10. Others

One esters (**534**) and three fatty acids, **535–537**, were isolated from several *Alpinia* species.<sup>64,227–229</sup> (S)-2-Pentanol-2-O-β-D-glucopyranoside (**538**), which showed inhibitory effect on NO production from LPS-activated RAW264.7 macrophage cells, was obtained from fruits of *A. oxyphylla*.<sup>89</sup> Two glycosides known as 3-methyl-but-2-en-1-yl-β-D-glucopyranoside (**539**) and *n*-butyl-β-D-fructopyranoside (**540**) were isolated from *A. officinarum* Hance.<sup>55,230</sup> While **541–544** were found in different *Alpinia* species (Fig. 9).<sup>24,51,108,196,201</sup> Interestingly, 5-hydroxymethylfurfural (**544**) exerted memory improvement activity against Alzheimer's disease (AD) by mitigating the degree of neuronal damage.<sup>231</sup>

## 11. Conclusions

The number of publications on the chemical constituents and their bioactivities for *Alpinia* species from 1955 to 2015 are shown in Fig. 10. Before 1999, fewer investigations (less than five per year, except six in 1987) were performed on this genus. However, after 2009, there were more than 10 papers published for each year. In 2013, the number of published articles reached 26, indicating a growing interest in the genus of *Alpinia*.

Till 2015, investigations on chemical constituents of the *Alpinia* species afforded a total of 544 compounds, including 207 terpenoids, 143 diarylheptanoids, 25 phenylpropanoids, 71



Table 1 The number of new compounds isolated from each *Alpinia* species since 1955

Sources of new compounds	Terpenoids	Diarylheptanoids	Lignans	Flavanoids	Phenolics	Sum
<i>A. blepharocalyx</i>	—	45	—	—	—	45
<i>A. bracteata</i>	—	—	—	—	1	1
<i>A. conchigera</i>	—	—	11	—	—	11
<i>A. calcarata</i>	6	—	1	—	—	7
<i>A. chinensis</i>	10	—	—	—	—	10
<i>A. densibracteata</i>	1	—	—	—	—	1
<i>A. densespicata</i>	6	—	—	—	—	6
<i>A. flabellata</i>	1	—	3	2	1	7
<i>A. formosana</i>	1	—	—	—	—	1
<i>A. galanga</i>	4	—	—	5	4	13
<i>A. gagnepainii</i>	—	—	—	—	1	1
<i>A. intermedia</i>	8	—	—	—	—	8
<i>A. japonica</i>	14	—	—	—	—	14
<i>A. katsumadai</i>	2	29	1	—	1	33
<i>A. nigra</i>	1	—	—	—	—	1
<i>A. oxymitra</i>	1	—	—	—	—	1
<i>A. officinarum</i>	1	18	5	—	6	30
<i>A. oxyphylla</i>	41	2	—	—	—	43
<i>A. paengensis</i>	5	—	—	—	—	5
<i>A. pinnanensis</i>	—	2	—	—	—	2
<i>A. rafflesiana</i>	—	—	—	1	—	1
<i>A. tonkinensis</i>	2	—	—	—	—	2
<i>A. zerumbet</i> ( <i>A. speciosa</i> )	2	—	—	—	2	4
Sum	106	96	21	8	16	247

flavanones, 66 phenolics, seven steroids, eight alkaloids, six stilbenes, and 11 others (Fig. 11). Among 207 terpenoids, 17 are monoterpenoids, 132 are sesquiterpenoids, 57 are diterpenoids, and the rest one is a triterpenoid. For sesquiterpenoids, eudesmanes and eremophilanes are undoubtedly predominant with 44 and 21 components, respectively. While for diterpenoids, almost all are labdanes.

Amongst 544 isolated compounds from the genus of *Alpinia*, 247 are new ones (Table 1), including 96 diarylheptanoids and 106 terpenoids. Obviously, diarylheptanoids, especially diarylheptane-flavonoids conjugates, are characteristic components for the genus of *Alpinia*.<sup>149</sup>

The crude extracts of *Alpinia* species and their chemical constituents were found to possess various biological activities. Mainly reported were antiemetic,<sup>26,27</sup> antibacterial,<sup>29–31,37,82,232–236</sup> antioxidant,<sup>127,237–239</sup> anticancer,<sup>32–34,240–245</sup> anti-inflammatory,<sup>189,246,247</sup> insecticidal,<sup>36,164</sup> and neuroprotective bioactivities.<sup>38,39,231,248–250</sup>

In addition, they also showed antiulcer,<sup>25</sup> antiplatelet,<sup>117,183</sup> hepatoprotective,<sup>251</sup> and hypolipidemic effects.<sup>252</sup> Meanwhile, evidences showed that ethanol extract of *A. galanga* can retard lipid oxidation for minced beef, indicating a great potential utility for food storage.<sup>8</sup> What should be aroused considerable interest was the promising anticancer and hepatoprotective properties, which could be a great potential to be developed as herbal medicines.

Although there are about 230 species for the *Alpinia* genus, only 35 were investigated for their chemical constituents and bioactivities (Fig. 12), because *A. jianganfeng* T. L. Wu includes *Alpinia sichuanensis* Z. Y. Zhu, and *A. zerumbet* (Pers.) B. L. Burtt & R. M. Sm. includes *A. speciosa* K. Schum. according to The Plant List. Among these species, *A. galanga*, *A. oxyphylla*, *A.*

*officinarum*, and *A. katsumadai* are four most studied plants with referenced papers of 43, 40, 32, and 23, respectively. While for the rest of 31 species, only very fewer articles were published, most of which were less than five. As a matter of fact, there was even only one paper published for 18 species. Although this genus contributed a diverse array of bioactive compounds, the potential of *Alpinia* species remains virtually untapped. Thus, much attention should be paid to *Alpinia* species on further phytochemical and pharmacological studies, which would produce structurally interesting and biologically active compounds with potential use in agricultural and medicinal applications. In addition, although most of *Alpinia* species were also used as edible plants, the nutritive components and their effects were seldom investigated, which could be a hotspot in the near future.

## Acknowledgements

The project was supported by National Natural Science Foundation of China (41176148, 21372233, 21202080).

## Notes and references

- 1 D. Wu and L. Kai, in *Flora of China*, ed. Z. Y. Wu, Science Press, Beijing, 2000, vol. 24, p. 333.
- 2 T. Wu, *Redai Yaredai Zhiwu Xuebao*, 1994, 2, 1.
- 3 J. Jonczyk, *Acta Pol. Pharm.*, 1970, 27, 155.
- 4 K. D. Kobayashi, J. McEwen and A. J. Kaufman, *Ornamentals and Flowers*, 2007, 37.
- 5 C. Wang, C. Xu, A. Tian, S. Fu and C. Wang, *Color. Technol.*, 2013, 129, 32.



6 X. Yang and R. G. Eilerman, *J. Agric. Food Chem.*, 1999, **47**, 1657.

7 H. Morita and H. Itokawa, *Chem. Lett.*, 1986, **15**, 1205.

8 P. B. Cheah and N. H. Abu Hasim, *J. Sci. Food Agric.*, 2000, **80**, 1565.

9 P. Pripdeevech, N. Nuntawong and S. Wongpornchai, *Chem. Nat. Compd.*, 2009, **45**, 562.

10 J. Qiu, *CN. Pat.*, CN101711590A, 2010.

11 Z. Wei, *CN. Pat.*, CN1385106A, 2002.

12 L. Xu and X. Gao, *CN. Pat.*, CN1481713A, 2004.

13 H. Yan, *CN. Pat.*, CN102084999A, 2011.

14 H. Yang, *CN. Pat.*, CN103156116A, 2013.

15 B. Zhu, *CN. Pat.*, CN101380121A, 2009.

16 J. Wang, W. Hou and Z. Zhang, *CN. Pat.*, CN104893910A, 2015.

17 Z. Cheng, *CN. Pat.*, CN102429267A, 2012.

18 Y. Dou, *CN. Pat.*, CN101088380A, 2007.

19 H. Zhao, *CN. Pat.*, CN1138960A, 1997.

20 L. X. Du, Z. T. Jiang and R. Li, *Zhongguo Tiaoweipin*, 2012, **37**, 22.

21 H. Itokawa, H. Morita, K. Watanabe, S. Mihashi and Y. Iitaka, *Chem. Pharm. Bull.*, 1985, **33**, 1148.

22 H. Itokawa, H. Morita, I. Midorikawa, R. Aiyama and M. Morita, *Chem. Pharm. Bull.*, 1985, **33**, 4889.

23 B. Roy and A. Swargiary, *J. Parasit. Dis.*, 2009, **33**, 48.

24 S. Mitsui, S. Kobayashi, H. Nagahori and A. Ogiso, *Chem. Pharm. Bull.*, 1976, **24**, 2377.

25 M. A. Al-Yahya, S. Rafatullah, J. S. Mossa, A. M. Ageel, M. S. Al-Said and M. Tariq, *Phytother. Res.*, 1990, **4**, 112.

26 Y. Yang, K. Kinoshita, K. Koyama, K. Takahashi, T. Tai, Y. Nunoura and K. Watanabe, *Nat. Prod. Sci.*, 1999, **5**, 20.

27 D. Shin, K. Kinoshita, K. Koyama and K. Takahashi, *J. Nat. Prod.*, 2002, **65**, 1315.

28 Y. Yang, K. Kinoshita, K. Koyama, K. Takahashi, S. Kondo and K. Watanabe, *Phytomedicine*, 2002, **9**, 146.

29 K. Rao, B. Ch, L. Narasu and A. Giri, *Appl. Biochem. Biotechnol.*, 2010, **162**, 871.

30 P. Niyomkam, S. Kaewbumrung, S. Kaewnpparat and P. Panichayupakaranant, *Pharm. Biol.*, 2010, **48**, 375.

31 X. Zhang, G. F. Shi, X. Z. Liu, L. J. An and S. Guan, *Cell Biochem. Funct.*, 2011, **29**, 342.

32 K. S. Chun, Y. Sohn, H. S. Kim, O. H. Kim, K. K. Park, J. M. Lee, J. Lee, J. Y. Lee, A. Moon, S. S. Lee and Y. J. Surh, *Mutat. Res.*, 1999, **428**, 49.

33 H. Itokawa, H. Morita, T. Sumitomo, N. Totsuka and K. Takeya, *Planta Med.*, 1987, **53**, 32.

34 S. Samarghandian, M. A. R. Hadjzadeh, J. Afshari and M. Hosseini, *BMC Complementary Altern. Med.*, 2014, **14**, 1.

35 R. Rajasekar, K. Manokaran, N. Rajasekaran, G. Duraisamy and D. Kanakasabapathi, *J. Diabetes Metab. Disord.*, 2014, **13**, 1.

36 Y. M. Chang, C. T. Tsai, C. C. R. Wang, Y. S. Chen, Y. M. Lin, C. H. Kuo, B. S. Tzang, R. J. Chen, F. J. Tsai and C. Y. Huang, *Biosci. Biotechnol., Biochem.*, 2013, **77**, 229.

37 K. Klahan, N. Nantapong and N. Chudapongse, *Planta Med.*, 2011, **77**, PM27.

38 S. H. Shi, X. Zhao, A. J. Liu, B. Liu, H. Li, B. Wu, K. S. Bi and Y. Jia, *Physiol. Behav.*, 2015, **139**, 13.

39 X. Z. Li, S. N. Zhang, S. M. Liu and F. Lu, *Fitoterapia*, 2013, **84**, 273.

40 D. P. de Sousa, P. de Almeida Soares Hocayen, L. N. Andrade and R. Andreatini, *Molecules*, 2015, **20**, 18620.

41 H. Lv and G. She, *Nat. Prod. Commun.*, 2010, **5**, 1687.

42 D. Kaushik, J. Yadav, P. Kaushik, D. Sacher and R. Rani, *Zhongxiyi Jiehe Xuebao*, 2011, **9**, 1061.

43 J. W. Nam and E. K. Seo, *Nat. Prod. Commun.*, 2012, **7**, 795.

44 P. Chen, P. P. Wang, Z. Z. Jiao and L. Xiang, *Xiandai Yaowu Yu Linchuang*, 2013, **28**, 617.

45 M. A. Rahman and M. S. Islam, *Pharmacogn. Rev.*, 2015, **9**, 55.

46 E. W. Chan and S. K. Wong, *J. Integr. Med.*, 2015, **13**, 368.

47 T. D. Xuan and R. Teschke, *Molecules*, 2015, **20**, 16306.

48 S. Z. Hua, J. G. Luo, X. B. Wang, J. S. Wang and L. Y. Kong, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 2728.

49 K. S. Ngo and G. D. Brown, *Phytochemistry*, 1998, **47**, 1117.

50 L. K. Sy and G. D. Brown, *Phytochemistry*, 1997, **45**, 537.

51 J. Zhang and L. Y. Kong, *J. Asian Nat. Prod. Res.*, 2004, **6**, 199.

52 D. Liu, W. Qu and J. Y. Liang, *Biochem. Syst. Ecol.*, 2013, **46**, 127.

53 J. J. Xu, N. H. Tan, Y. S. Chen, X. L. Pan, G. Z. Zeng, H. J. Han, C. J. Ji and M. J. Zhu, *Helv. Chim. Acta*, 2009, **92**, 1621.

54 Y. Someya, A. Kobayashi and K. Kubota, *Biosci. Biotechnol., Biochem.*, 2001, **65**, 950.

55 T. N. Ly, R. Yamauchi, M. Shimoyamada and K. Kato, *J. Agric. Food Chem.*, 2002, **50**, 4919.

56 U. Grienke, M. Schmidtke, J. Kirchmair, K. Pfarr, P. Wutzler, R. Dürrwald, G. Wolber, K. R. Liedl, H. Stuppner and J. M. Rollinger, *J. Nat. Med.*, 2010, **53**, 778.

57 M. Morita, H. Nakanishi, H. Morita, S. Mihashi and H. Itokawa, *Chem. Pharm. Bull.*, 1996, **44**, 1603.

58 M. Miyazawa, Y. Nakamura and Y. Ishikawa, *J. Agric. Food Chem.*, 2000, **48**, 3639.

59 P. Chen, P. P. Wang, Z. Z. Jiao and L. Xiang, *Helv. Chim. Acta*, 2014, **97**, 388.

60 P. Chen, L. Qu, L. Tian, P. P. Wang and L. Xiang, *Helv. Chim. Acta*, 2013, **96**, 1163.

61 H. Itokawa, H. Morita, T. Kobayashi, K. Watanabe and Y. Iitaka, *Chem. Pharm. Bull.*, 1987, **35**, 2860.

62 S. Ando, H. Matsuda, T. Morikawa and M. Yoshikawa, *Bioorg. Med. Chem.*, 2005, **13**, 3289.

63 X. Luo, J. Yu, L. Xu, K. Li, P. Tan and J. Feng, *Yaoxue Xuebao*, 2000, **35**, 204.

64 L. Hou, X. X. Lu, B. B. Xie, W. H. Huang, J. G. Yu and B. L. Guo, *Tianran Chanwu Yanjiu Yu Kaifa*, 2013, **25**, 878.

65 J. Luo, X. Lv, X. Wang and L. Kong, *Phytochem. Lett.*, 2012, **5**, 134.

66 J. J. Xu, N. H. Tan, J. Xiong, A. H. Adebayo, H. J. Han, G. Z. Zeng, C. J. Ji, Y. M. Zhang and M. J. Zhu, *Chin. Chem. Lett.*, 2009, **20**, 945.

67 D. H. Park, J. W. Lee, Q. Jin, W. K. Jeon, M. K. Lee and B. Y. Hwang, *Bull. Korean Chem. Soc.*, 2014, **35**, 1565.

68 X. Q. Lv, J. G. Luo, X. B. Wang, J. S. Wang, J. Luo and L. Y. Kong, *Chem. Pharm. Bull.*, 2011, **59**, 402.

69 H. Itokawa, H. Morita and K. Watanabe, *Chem. Pharm. Bull.*, 1987, **35**, 1460.



70 H. Itokawa, K. Watanabe, S. Mihashi and Y. Iitaka, *Chem. Pharm. Bull.*, 1980, **28**, 681.

71 B. Jiang, W. J. Wang, M. P. Li, X. J. Huang, F. Huang, H. Gao, P. H. Sun, M. F. He, Z. J. Jiang, X. Q. Zhang and W. C. Ye, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 3879.

72 O. Muraoka, M. Fujimoto, G. Tanabe, M. Kubo, T. Minematsu, H. Matsuda, T. Morikawa, I. Toguchida and M. Yoshikawa, *Bioorg. Med. Chem. Lett.*, 2001, **11**, 2217.

73 J. Xu, C. Ji, Y. Zhang, J. Su, Y. Li and N. Tan, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 1660.

74 J. J. Xu, N. H. Tan, G. Z. Zeng, H. J. Han and Y. F. Peng, *Chin. J. Nat. Med.*, 2010, **8**, 6.

75 K. Jitsaeng, W. De-Eknamkul and B. Schneider, *Rec. Nat. Prod.*, 2009, **3**, 110.

76 S. M. Xu, X. J. Huang, Y. Wang and W. C. Ye, *Zhongguo Tianran Yaowu*, 2012, **10**, 374.

77 L. Hou, G. Ding, B. L. Guo, W. H. Huang, X. J. Zhang, Z. Y. Sun and X. F. Shi, *Molecules*, 2015, **20**, 1551.

78 H. Itokawa, H. Morita, K. Watanabe, A. Takase and Y. Iitaka, *Chem. Lett.*, 1984, **13**, 1687.

79 H. Itokawa, H. Morita, K. Osawa, K. Watanabe and Y. Iitaka, *Chem. Pharm. Bull.*, 1987, **35**, 2849.

80 H. Itokawa, K. Watanabe, H. Morita, S. Mihashi and Y. Iitaka, *Chem. Pharm. Bull.*, 1985, **33**, 2023.

81 H. Itokawa, S. Yoshimoto and H. Morita, *Phytochemistry*, 1988, **27**, 435.

82 A. N. Aziz, H. Ibrahim, D. Rosmy Syamsir, M. Mohtar, J. Vejayan and K. Awang, *J. Ethnopharmacol.*, 2013, **145**, 798.

83 Q. M. Li, J. G. Luo, X. B. Wang, M. H. Yang and L. Y. Kong, *Fitoterapia*, 2013, **86**, 29.

84 J. Xu, N. Tan, G. Zeng, H. Han, H. Huang, C. Ji, M. Zhu and Y. Zhang, *Zhongguo Zhongyao Zazhi*, 2009, **34**, 990.

85 L. Y. Kong, M. J. Qin and M. Niwa, *J. Nat. Prod.*, 2000, **63**, 939.

86 Q. H. Zeng, C. L. Lu, X. W. Zhang and J. G. Jiang, *Food Funct.*, 2015, **6**, 431.

87 H. Itokawa, H. Morita, K. Watanabe and Y. Iitaka, *Chem. Lett.*, 1984, **13**, 451.

88 T. Morikawa, H. Matsuda, I. Toguchida, K. Ueda and M. Yoshikawa, *J. Nat. Prod.*, 2002, **65**, 1468.

89 Z. J. Qing, Y. Wang, L. Y. Hui, L. W. Yong, L. H. Long, D. J. Ao and P. L. Xia, *Arch. Pharmacal Res.*, 2012, **35**, 2143.

90 J. Xu, J. Su, Y. Li and N. Tan, *Chem. Nat. Compd.*, 2013, **49**, 457.

91 S. Ghosh, K. Indukuri, S. Bondalapati, A. K. Saikia and L. Rangan, *Eur. J. Med. Chem.*, 2013, **66**, 101.

92 S. Ghosh and L. Rangan, *Appl. Biochem. Biotechnol.*, 2015, **175**, 1477.

93 H. Morita and H. Itokawa, *Planta Med.*, 1988, **54**, 117.

94 J. Chompoo, A. Upadhyay, W. Kishimoto, T. Makise and S. Tawata, *Food Chem.*, 2011, **129**, 709.

95 A. Upadhyay, J. Chompoo, W. Kishimoto, T. Makise and S. Tawata, *J. Agric. Food Chem.*, 2011, **59**, 2857.

96 Y. Sivasothy, H. Ibrahim, A. S. Paliyan, S. A. Alias, N. R. Md Nor and K. Awang, *Planta Med.*, 2013, **79**, 1775.

97 L. K. Sy and G. D. Brown, *J. Nat. Prod.*, 1997, **60**, 904.

98 H. Itokawa, M. Morita and S. Mihashi, *Chem. Pharm. Bull.*, 1980, **28**, 3452.

99 H. X. Xu, H. Dong and K. Y. Sim, *Phytochemistry*, 1996, **42**, 149.

100 N. Nuntawong and A. Suksamrarn, *Biochem. Syst. Ecol.*, 2008, **36**, 661.

101 Q. M. Li, J. G. Luo, M. H. Yang and L. Y. Kong, *Chem. Biodiversity*, 2015, **12**, 388.

102 V. S. Chauhan, M. Swapna and A. Singh, *Int. J. Appl. Biol. Pharm. Technol.*, 2014, **5**, 186.

103 Y. J. Kuo, P. C. Hsiao, L. J. Zhang, M. D. Wu, Y. H. Liang, H. O. Ho and Y. H. Kuo, *J. Nat. Prod.*, 2009, **72**, 1097.

104 S. Tesaki, H. Kikuzaki, S. Yonemori and N. Nakatani, *J. Nat. Prod.*, 2001, **64**, 515.

105 H. Itokawa, H. Morita, I. Katou, K. Takeya, A. J. Cavalheiro, R. C. B. de Oliveira, M. Ishige and M. Motidome, *Planta Med.*, 1988, **54**, 311.

106 Y. Sivasothy, H. Ibrahim, A. S. Paliyan, S. A. Alias and K. Awang, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 6280.

107 L. Y. Kong, M. J. Qin and M. Niwa, *Planta Med.*, 2002, **68**, 813.

108 X. B. Wang, C. S. Yang, S. Z. Hua and L. Y. Kong, *Zhongguo Tianran Yaowu*, 2010, **8**, 419.

109 S. Y. Choi, M. H. Lee, J. H. Choi and Y. K. Kim, *Biol. Pharm. Bull.*, 2012, **35**, 2092.

110 H. J. Lee, J. S. Kim and J. H. Ryu, *Planta Med.*, 2006, **72**, 68.

111 R. J. Lin, C. M. Yen, T. H. Chou, F. Y. Chiang, G. H. Wang, Y. P. Tseng, L. Wang, T. W. Huang, H. C. Wang, L. P. Chan, H. Y. Ding and C. H. Liang, *BMC Complementary Altern. Med.*, 2013, **13**, 1.

112 N. Shoji, A. Umeyama, T. Takemoto and Y. Ohizumi, *Planta Med.*, 1984, **50**, 186.

113 Q. Y. Bian, S. Y. Wang, L. J. Xu, C. O. Chan, D. K. W. Mok and S. B. Chen, *J. Asian Nat. Prod. Res.*, 2013, **15**, 1094.

114 Q. Zhang, S. Luo, H. Wang and D. Fan, *Zhongcaoyao*, 1997, **28**, 131.

115 S. Kadota, Y. Tezuka, J. K. Prasain, M. S. Ali and A. H. Banskota, *Curr. Top. Med. Chem.*, 2003, **3**, 203.

116 M. S. Ali, Y. Tezuka, S. Awale, A. H. Banskota and S. Kadota, *J. Nat. Prod.*, 2001, **64**, 289.

117 H. Dong, S. X. Chen, H. X. Xu, S. Kadota and T. Namba, *J. Nat. Prod.*, 1998, **61**, 142.

118 P. M. Giang, P. T. Son, K. Matsunami and H. Otsuka, *Chem. Pharm. Bull.*, 2005, **53**, 1335.

119 M. Kuroyanagi, T. Noro, S. Fukushima, R. Aiyama, A. Ikuta, H. Itokawa and M. Morita, *Chem. Pharm. Bull.*, 1983, **31**, 1544.

120 B. Groebelacher, O. Kunert and F. Bucar, *Bioorg. Med. Chem.*, 2012, **20**, 2701.

121 W. Z. Huang, C. F. Zhang, M. Zhang and Z. T. Wang, *J. Chin. Chem. Soc.*, 2007, **54**, 1553.

122 Y. Li, L. Yang, C. Wang, G. Chou and Z. Wang, *Shanghai Zhongyiyao Daxue Xuebao*, 2010, **24**, 72.

123 Y. Y. Li, G. X. Chou and Z. T. Wang, *Helv. Chim. Acta*, 2010, **93**, 382.

124 J. W. Nam, G. Y. Kang, A. R. Han, D. Lee, Y. S. Lee and E. K. Seo, *J. Nat. Prod.*, 2011, **74**, 2109.

125 J. W. Nam and E. K. Seo, *Helv. Chim. Acta*, 2013, **96**, 1670.

126 C. Y. Lo, P. L. Liu, L. C. Lin, Y. T. Chen, Y. C. Hsue, Z. H. Wen and H. M. Wang, *Sci. World J.*, 2013, 186505, DOI: 10.1155/2013/186505.



127 M. Habsah, N. H. Lajis, A. M. Ali, M. A. Sukari, Y. Y. Hin, H. Kikuzaki and N. Nakatani, *Pharm. Biol.*, 2003, **41**, 7.

128 H. Mohamad, F. Abas, D. Permana, N. H. Lajis, A. M. Ali, M. A. Sukari, T. Y. Y. Hin, H. Kikuzaki and N. Nakatani, *Z. Naturforsch. C: J. Biosci.*, 2004, **59**, 811.

129 B. B. Zhang, Y. Dai, Z. X. Liao and L. S. Ding, *Fitoterapia*, 2010, **81**, 948.

130 H. M. Sirat, A. A. Rahman, H. Itokawa and H. Morita, *Planta Med.*, 1996, **62**, 188.

131 N. An, H. W. Zhang, L. Z. Xu, S. L. Yang and Z. M. Zou, *Food Chem.*, 2010, **119**, 513.

132 L. Zhao, W. Qu, J. Q. Fu and J. Y. Liang, *Chin. J. Nat. Med.*, 2010, **8**, 241.

133 N. An, Z. M. Zou, Z. Tian, X. Z. Luo, S. L. Yang and L. Z. Xu, *Fitoterapia*, 2007, **79**, 27.

134 Y. Sun, K. Tabata, H. Matsubara, S. Kitanaka, T. Suzuki and K. Yasukawa, *Planta Med.*, 2008, **74**, 427.

135 D. Liu, Y. W. Liu, F. Q. Guan and J. Y. Liang, *Fitoterapia*, 2014, **96**, 76.

136 Y. Sun, H. Matsubara, S. Kitanaka and K. Yasukawa, *Helv. Chim. Acta*, 2008, **91**, 118.

137 G. J. Fan, Y. H. Kang, Y. N. Han and B. H. Han, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 6720.

138 D. Liu, W. Qu, L. Zhao, F. Q. Guan and J. Y. Liang, *Chin. J. Nat. Med.*, 2014, **12**, 139.

139 Y. U. Kim, H. K. Son, H. K. Song, M. J. Ahn, S. S. Lee and S. K. Lee, *Planta Med.*, 2003, **69**, 72.

140 H. B. Lee, H. K. Lee, J. R. Kim and Y. J. Ahn, *J. Korean Soc. Appl. Biol. Chem.*, 2009, **52**, 367.

141 J. E. Shin, M. J. Han, M. C. Song, N. I. Baek and D. H. Kim, *Biol. Pharm. Bull.*, 2004, **27**, 138.

142 Z. Liu, M. M. Rafi, N. Zhu, K. Ryu, S. Sang, C. T. Ho and R. T. Rosen, *ACS Symp. Ser.*, 2003, **851**, 369.

143 F. Kiuchi, M. Shibuya and U. Sankawa, *Chem. Pharm. Bull.*, 1982, **30**, 2279.

144 K. Yasukawa, Y. Sun, S. Kitanaka, N. Tomizawa, M. Miura and S. Motohashi, *J. Nat. Med.*, 2008, **62**, 374.

145 S. I. Uehara, I. Yasuda, K. Akiyama, H. Morita, K. Takeya and H. Itokawa, *Chem. Pharm. Bull.*, 1987, **35**, 3298.

146 M. S. Ali, Y. Tezuka, A. H. Banskota and S. Kadota, *J. Nat. Prod.*, 2001, **64**, 491.

147 J. K. Prasain, Y. Tezuka, J. X. Li, K. Tanaka, P. Basnet, H. Dong, T. Namba and S. Kadota, *Planta Med.*, 1999, **65**, 196.

148 M. S. Ali, A. H. Banskota, Y. Tezuka, I. Saiki and S. Kadota, *Biol. Pharm. Bull.*, 2001, **24**, 525.

149 X. B. Wang, C. S. Yang, C. Zhang, J. Luo, M. H. Yang, J. G. Luo, W. Y. Yu and L. Y. Kong, *Tetrahedron*, 2014, **70**, 8714.

150 J. Kumar Prasain, Y. Tezuka, J.-X. Li, K. Tanaka, P. Basnet, H. Dong, T. Namba and S. Kadota, *J. Chem. Res., Synop.*, 1998, **22**, DOI: 10.1039/A706250H.

151 J. Kumar Prasain, Y. Tezuka, J. Xin Li, K. Tanaka, P. Basnet, H. Dong, T. Namba and S. Kadota, *Tetrahedron*, 1997, **53**, 7833.

152 Y. Tezuka, M. B. Gewali, M. S. Ali, A. H. Banskota and S. Kadota, *J. Nat. Prod.*, 2001, **64**, 208.

153 J. K. Prasain, J. X. Li, Y. Tezuka, K. Tanaka, P. Basnet, H. Dong, T. Namba and S. Kadota, *J. Nat. Prod.*, 1998, **61**, 212.

154 M. B. Gewali, Y. Tezuka, A. H. Banskota, M. S. Ali, I. Saiki, H. Dong and S. Kadota, *Org. Lett.*, 1999, **1**, 1733.

155 L. Zhao, J. Y. Liang, J. Y. Zhang and Y. Chen, *Chin. Chem. Lett.*, 2010, **21**, 194.

156 D. Liu, W. Qu, L. Zhao and J. Y. Liang, *Chin. Chem. Lett.*, 2012, **23**, 189.

157 L. Zhao, J. Y. Liang and W. Qu, *Chem. Nat. Compd.*, 2012, **48**, 836.

158 S. Kadota, J. K. Prasain, J. X. Li, P. Basnet, H. Dong, T. Tani and T. Namba, *Tetrahedron Lett.*, 1996, **37**, 7283.

159 Y. Tezuka, M. S. Ali, A. H. Banskota and S. Kadota, *Tetrahedron Lett.*, 2000, **41**, 5903.

160 Y. Yang, K. Kinoshita, K. Koyama, K. Takahashi, T. Tai, Y. Nunoura and K. Watanabe, *J. Nat. Prod.*, 1999, **62**, 1672.

161 S. Tesaki, H. Kikuzaki, S. Tanabe, M. Watanabe and N. Nakatani, *ITE Lett. Batteries, New Technol. Med.*, 2001, **2**, 106.

162 H. Kikuzaki, S. Tesaki, S. Yonemori and N. Nakatani, *Phytochemistry*, 2001, **56**, 109.

163 T. N. Ly, M. Shimoyamada, K. Kato and R. Yamauchi, *J. Agric. Food Chem.*, 2003, **51**, 4924.

164 A. Kaur, R. Singh, C. S. Dey, S. S. Sharma, K. K. Bhutani and I. P. Singh, *Indian J. Exp. Biol.*, 2010, **48**, 314.

165 J. J. Xu, G. Z. Zeng, S. C. Yang, Y. Shen and N. H. Tan, *Fitoterapia*, 2013, **91**, 82.

166 T. Morikawa, S. Ando, H. Matsuda, S. Kataoka, O. Muraoka and M. Yoshikawa, *Chem. Pharm. Bull.*, 2005, **53**, 625.

167 J. J. Xu, H. M. Zhao, Y. Shen, J. H. Chen, Y. Li, N. H. Tan and S. C. Yang, *J. Asian Nat. Prod. Res.*, 2013, **15**, 833.

168 T. Obata, A. Sawabe, M. Morita, N. Yamashita and Y. Matsubara, *J. Jpn. Oil Chem. Soc.*, 1995, **44**, 1012.

169 N. Wei, Y. Wang, H. F. Li, J. Q. Zhang and Y. H. Li, *Chem. Nat. Compd.*, 2013, **49**, 934.

170 L. Liu, J. G. Luo and L. Y. Kong, *Chem. Nat. Compd.*, 2012, **48**, 785.

171 J. Zhang, Q. H. Guo and L. Y. Kong, *Zhongguo Zhongyao Zazhi*, 2003, **28**, 41.

172 G. Eumkeb, S. Siriwong, S. Phitaktim, N. Rojtinakorn and S. Sakdarat, *J. Appl. Microbiol.*, 2012, **112**, 55.

173 S. T. Chien, M. D. Shi, Y. C. Lee, C. C. Te and Y. W. Shih, *Cancer Cell Int.*, 2015, **15**, 1.

174 M. Y. Heo, S. J. Sohn and W. W. Au, *Mutat. Res.*, 2001, **488**, 135.

175 T. Ohtsuki, H. Kikuchi, T. Koyano, T. Kowithayakorn, T. Sakai and M. Ishibashi, *Bioorg. Med. Chem.*, 2009, **17**, 6748.

176 M. Q. Bian, H. Q. Wang, J. Kang, R. Y. Chen, Y. F. Yang and H. Z. Wu, *Yaoxue Xuebao*, 2014, **49**, 359.

177 B. R. Xin, S. J. Ren and J. Li, *Zhongguo Zhongyao Zazhi*, 2014, **39**, 2674.

178 J. Zhang, X. He, J. Gao and L. Kong, *Zhongguo Yaoxue Zazhi*, 2003, **38**, 502.

179 J. E. Shin, M. J. Han and D. H. Kim, *Biol. Pharm. Bull.*, 2003, **26**, 854.



180 H. Kikuzaki and S. Tesaki, *J. Nat. Prod.*, 2002, **65**, 389.

181 X. Xiao, X. Si, X. Tong and G. Li, *Sep. Purif. Technol.*, 2011, **81**, 265.

182 N. A. Mustahil, M. A. Sukari, A. B. Abdul, N. A. Ali and G. E. C. Lian, *Pak. J. Pharm. Sci.*, 2013, **26**, 391.

183 I. Jantan, S. M. Raweh, H. M. Sirat, S. Jamil, Y. H. Mohd Yasin, J. Jalil and J. A. Jamal, *Phytomedicine*, 2008, **15**, 306.

184 I. Jantan, M. Pisar, H. M. Sirat, N. Basar, S. Jamil, R. M. Ali and J. Jalil, *Phytother. Res.*, 2004, **18**, 1005.

185 H. Zhang, L. X. Xu, P. Wu and X. Y. Wei, *Redai Yaredai Zhiwu Xuebao*, 2014, **22**, 89.

186 B. R. Xin, J. F. Liu, J. Kang and W. P. Chan, *Mol. Cell. Toxicol.*, 2014, **10**, 165.

187 J. Gripenberg, E. Honkanen and K. Silander, *Acta Chem. Scand.*, 1956, **10**, 393.

188 H. Itokawa, M. Morita and S. Mihashi, *Phytochemistry*, 1981, **20**, 2503.

189 C. T. Lin, K. J. Senthil Kumar, Y. H. Tseng, Z. J. Wang, M. Y. Pan, J. H. Xiao, S. C. Chien and S. Y. Wang, *J. Agric. Food Chem.*, 2009, **57**, 6060.

190 S. Ahmad, D. A. Israf, N. H. Lajis, K. Shaari, H. Mohamed, A. A. Wahab, K. T. Ariffin, W. Y. Hoo, N. A. Aziz, A. A. Kadir, M. R. Sulaiman and M. N. Somchit, *Eur. J. Pharmacol.*, 2006, **538**, 188.

191 H. T. Le, M. G. Phan and T. S. Phan, *Tap Chi Hoa Hoc*, 2007, **45**, 126.

192 Z. Wei, J. Yang, Y. F. Xia, W. Z. Huang, Z. T. Wang and Y. Dai, *J. Biochem. Mol. Toxicol.*, 2012, **26**, 282.

193 C. P. Shen, J. G. Luo, M. H. Yang and L. Y. Kong, *Fitoterapia*, 2015, **106**, 153.

194 W. Q. Yang, Y. Gao, M. Li, D. R. Miao and F. Wang, *J. Asian Nat. Prod. Res.*, 2015, **17**, 783.

195 M. A. Mpalantinos, R. S. De Moura, J. P. Parente and R. M. Kuster, *Phytother. Res.*, 1998, **12**, 442.

196 Y. Y. Li, G. X. Chou and Z. T. Wang, *Zhongguo Tianran Yaowu*, 2009, **7**, 417.

197 T. Masuda, S. Mizuguchi, T. Tanaka, K. Iritani, Y. Takeda and S. Yonemori, *J. Agric. Food Chem.*, 2000, **48**, 1479.

198 N. Nakatani, *Abstracts of Papers, 223rd ACS National Meeting*, AGFD, Orlando, FL, United States, April 7-11, 2002, p. 2002.

199 S. B. Jaju, N. H. Indurwade, D. M. Sakarkar, N. K. Fuloria, M. D. Ali, S. Das and S. P. Basu, *Trop. J. Pharm. Res.*, 2009, **8**, 545.

200 B. R. Barik, A. B. Kundu and A. K. Dey, *Phytochemistry*, 1987, **26**, 2126.

201 S. H. Shi, C. N. Zhang, A. J. Liu, H. Li, K. S. Bi and Y. Jia, *Zhongguo Shiyuan Fangjixue Zazhi*, 2013, **19**, 97.

202 J. W. Nam, S. J. Kim, A. R. Han, S. K. Lee and E. K. Seo, *J. Appl. Pharmacol.*, 2005, **13**, 263.

203 L. Zhao, L. Y. Chen and J. Y. Liang, *Chin. J. Nat. Med.*, 2012, **10**, 370.

204 H. Matsuda, T. Morikawa, H. Managi and M. Yoshikawa, *Bioorg. Med. Chem. Lett.*, 2003, **13**, 3197.

205 S. K. Roy, S. Pahwa, H. Nandanwar and S. M. Jachak, *Fitoterapia*, 2012, **83**, 1248.

206 T. Noro, T. Sekiya, M. Katoh, Y. Oda, T. Miyase, M. Kuroyanagi, A. Ueno and S. Fukushima, *Chem. Pharm. Bull.*, 1988, **36**, 244.

207 N. Sukhirun, W. Pluempanupat, V. Bullangpoti and O. Koul, *J. Econ. Entomol.*, 2011, **104**, 1534.

208 N. Hasima, L. I. L. Aun, M. N. Azmi, A. N. Aziz, E. Thirthagiri, H. Ibrahim and K. Awang, *Phytomedicine*, 2010, **17**, 935.

209 H. Matsuda, Y. Pongpiriyadacha, T. Morikawa, M. Ochi and M. Yoshikawa, *Eur. J. Pharmacol.*, 2003, **471**, 59.

210 Y. Ye and B. Li, *J. Gen. Virol.*, 2006, **87**, 2047.

211 H. J. Min, J. W. Nam, E. S. Yu, J. H. Hong, E. K. Seo and E. S. Hwang, *Int. Immunopharmacol.*, 2009, **9**, 448.

212 R. Mongkol, W. Chavasiri, M. Ishida, K. Matsuda and M. Morimoto, *Weed Biol. Manage.*, 2015, **15**, 87.

213 H. C. Wang, J. X. Li, H. Li, L. W. Yang, M. H. Gao and H. A. Chen, *Yaoxue Yanjiu*, 2013, **32**, 559.

214 W. Wang, S. Qi, H. Zhong and Q. Yao, *Shipin Yu Yaopin*, 2012, **14**, 88.

215 S. Guan, Y.-M. Bao, B. Jiang and L.-J. An, *Eur. J. Pharmacol.*, 2006, **538**, 73.

216 S. Guan, B. Jiang, Y. M. Bao and L. J. An, *Food Chem. Toxicol.*, 2006, **44**, 1659.

217 L. J. An, S. Guan, G. F. Shi, Y. M. Bao, Y. L. Duan and B. Jiang, *Food Chem. Toxicol.*, 2006, **44**, 436.

218 G. F. Shi, L. J. An, B. Jiang, S. Guan and Y. M. Bao, *Neurosci. Lett.*, 2006, **403**, 206.

219 Y. M. Liu, B. Jiang, Y. M. Bao and L. J. An, *Toxicol. in Vitro*, 2008, **22**, 430.

220 H. Zhao, J. Xu and S. Yang, *Yunnan Nongye Daxue Xuebao*, 2014, **29**, 468.

221 M. G. Phan and T. S. Phan, *Tap Chi Hoa Hoc*, 2004, **42**, 376.

222 P. T. B. Tu and S. Tawata, *Molecules*, 2014, **19**, 16656.

223 T. Fujita, H. Nishimura, K. Kaburagi and J. Mizutani, *Phytochemistry*, 1994, **36**, 23.

224 J. Chompoo, A. Upadhyay, M. Fukuta and S. Tawata, *BMC Complementary Altern. Med.*, 2012, **12**, 106.

225 S. B. Jaju, N. H. Indurwade, D. M. Sakarkar, N. K. Fuloria, M. D. Ali and S. P. Basu, *Pharmacogn. Res.*, 2010, **2**, 264.

226 X. Wang, X. Yang and J. Li, *Zhongyaocai*, 2008, **31**, 853.

227 C. Qiao, X. Hao, Z. Wang and L. Xu, *Zhongguo Zhongyao Zazhi*, 2002, **27**, 130.

228 L. Di, Z. Wang, Z. Wang, N. Li and K. Wang, *Zhiwu Ziyuan Yu Huanjing Xuebao*, 2011, **20**, 94.

229 S. Qi, F. Ji and Q. Yao, *Shipin Yu Yaopin*, 2010, **12**, 39.

230 N. An, J. Lin, S. Yang, Z. Zou and L. Xu, *Yaoxue Xuebao*, 2006, **41**, 233.

231 A. Liu, X. Zhao, H. Li, Z. Liu, B. Liu, X. Mao, L. Guo, K. Bi and Y. Jia, *Int. Immunopharmacol.*, 2014, **23**, 719.

232 P. G. Ray and S. K. Majumdar, *Indian J. Exp. Biol.*, 1976, **14**, 712.

233 J. J. C. Scheffer, A. Gani and A. Baerheim Svendsen, *Planta Med.*, 1981, **42**, 140.

234 H. Haraguchi, Y. Kuwata, K. Inada, K. Shingu, K. Miyahara, M. Nagao and A. Yagi, *Planta Med.*, 1996, **62**, 308.

235 K. Chukanhom, P. Borisuthpeth and K. Hatai, *Biocontrol Sci.*, 2005, **10**, 105.



236 M. M. Yusoff, H. Ibrahim and N. A. Hamid, *Chem. Biodiversity*, 2011, **8**, 916.

237 S. E. Lee, H. T. Shin, H. J. Hwang and J. H. Kim, *Phytother. Res.*, 2003, **17**, 1041.

238 W. Y. Hsu, A. Simonne, A. Weissman and J. M. Kim, *Food Sci. Biotechnol.*, 2010, **19**, 873.

239 C. A. Raj, P. Ragavendran, D. Sophia, T. Starlin, M. A. Rathi and V. K. Gopalakrishnan, *Chin. J. Integr. Med.*, 2014, **1**, DOI: 10.1007/s11655-014-1762-1.

240 P. Muangnoi, M. Lu, J. Lee, A. Thepouyporn, R. Mirzayans, X. C. Le, M. Weinfield and S. Changbumrung, *Planta Med.*, 2007, **73**, 748.

241 C. W. Phang, S. Malek and H. Ibrahim, *BMC Complementary Altern. Med.*, 2013, **13**, 1.

242 C. L. Hsu, Y. S. Yu and G. C. Yen, *J. Agric. Food Chem.*, 2009, **58**, 2201.

243 A. Reddy, S. Malek, H. Ibrahim and K. Sim, *BMC Complementary Altern. Med.*, 2013, **13**, 1.

244 T. P. T. Be, J. Chompoon and S. Tawata, *Drug Discoveries Ther.*, 2015, **9**, 197.

245 W. Fan, C. Z. Wang, X. L. Bao, H. H. Yuan and M. B. Lan, *Asian J. Chem.*, 2015, **27**, 532.

246 M. Y. Lee, C. S. Seo, J. A. Lee, I. S. Shin, S. J. Kim, H. K. Ha and H. K. Shin, *Inflammation*, 2012, **35**, 746.

247 Y. S. Yu, C. L. Hsu and G. C. Yen, *J. Agric. Food Chem.*, 2009, **57**, 7673.

248 X. Yu, Y. W. Ceballos, H. Zhao and Z. Xu, *Neurosci. Res. Commun.*, 2003, **33**, 105.

249 J. C. Hanish Singh, V. Alagarsamy, P. V. Diwan, S. Sathesh Kumar, J. C. Nisha and Y. Narsimha Reddy, *J. Ethnopharmacol.*, 2011, **138**, 85.

250 Z. J. Zhang, L. C. V. Cheang, M. W. Wang, G. H. Li, I. Chu, Z. X. Lin and S. M. Y. Lee, *Cell. Mol. Neurobiol.*, 2012, **32**, 27.

251 F. M. Hammouda, S. S. El-Hawary, H. A. Kassem, W. A. Tawfik, A. A. Abdel Motaal, N. M. Nazif and S. S. El-Shamy, *Res. J. Pharm., Biol. Chem. Sci.*, 2015, **6**, 448.

252 C. R. Achuthan and J. Padikkala, *Indian J. Clin. Biochem.*, 1997, **12**, 55.

