**Stereoselective synthesis of 2′-modified nucleosides by using ortho-alkynyl benzoate as a gold(i)-catalyzed removable neighboring participation group†**

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In the present paper, we report a novel strategy for highly efficient stereoselective synthesis of 2′-modified nucleosides by using ortho-alkynyl benzoate as neighboring participation group. Subsequently, ortho-alkynyl benzoate can be removed smoothly in the presence of 5 mol% Ph3PAuCl as neighboring participation group. Subsequently, ortho-alkynyl benzoate can be removed smoothly in the presence of 5 mol% Ph3PAuCl–AgOTf in dichloromethane with H2O (1 eq.) and ethanol (6 eq.) to afford 2′-OH nucleosides in high yields and selectivity.

**Introduction**

In the past decades, tremendous efforts have been devoted to the synthesis of novel nucleosides and the evaluation of their biological activities. Accordingly, a large number of nucleosides have been successfully developed as antiviral and antitumor drugs.1 Among these chemical entities, C-2′ substituted nucleosides have showed special importance.2–5 For instance, clofarabine, gemcitabine, nelarabine, clofarabine and most recently FDA approved sofosbuvir for chronic HCV treatment all contain the C-2′ substituted nucleoside core structure (Fig. 1).5-* On the other hand, 2′-modified nucleosides have also been used as biochemical probes to investigate the structure and function of nucleic acid.5–11

Currently, there are generally two synthetic methodologies to access 2′-modified ribonucleosides, namely the convergent approach and the linear approach (Fig. 2). In the convergent approach, the nucleoside was produced by glycosylation of nucleobase with the corresponding sugar moiety. In the linear approach, the nucleoside was prepared by chemical modification of commercially available natural nucleosides or related compounds. From the synthetic point of view, the linear approach offers a relatively convenient way because of it could avoid the glycosylation step, which is often cumbersome. However, compared with the linear approach, the convergent approach is potentially more flexible. It could provide abundant structural diversity by using Vorbrüggen glycosylation of a variety of nucleobase with modified carbohydrate. But in absence of neighboring group participation (NGP), Vorbrüggen glycosylation always generated a mixture of α and β isomer in low selectivity and yield. Moreover, the separation is often time-consuming and labor intensive. Therefore, a general and efficient synthetic approach for C-2′ substituted nucleosides is highly desired.

In the literature, some examples were reported using acyl group as neighboring participation group, which could be removed selectively by using NH2NH2, CH3NH2 or NH2CH2-CH2NH2 etc.12 Nevertheless, these approaches' reproducibilities were far from satisfied by our evaluation. At the meantime, acetyl transfer from 3′-OH to 2′-OH was inevitable, which will make the purification troublesome and sometimes impossible. So a general strategy is highly desired to solve this potential problem.

Gold(i) complexes are mild electrophiles and have wide usage in homogeneous catalysis. In recent years, the application of gold(i)-catalyzed electrophilic activation of alkynes has been
nucleophilic attack from group to form dioxolanylium with anomeric cation, which led to the desired 2

Under the similar condition, a series of nucleosides have participated the reaction. In our experiment, we reasoned that residue H2O in EtOH could improve the solubility of H2O in DCM.

Table 1: Optimization of gold(i)-catalysed deprotection of ortho-alkynyl benzoate

<table>
<thead>
<tr>
<th>Entry</th>
<th>Catalyst (mol%)</th>
<th>Solvent</th>
<th>Time (h)</th>
<th>Yield(^d) (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>AuCl (5%)</td>
<td>DCM</td>
<td>5</td>
<td>70%</td>
</tr>
<tr>
<td>2</td>
<td>Ph3PAuCl (5%)</td>
<td>DCM</td>
<td>5</td>
<td>Trace</td>
</tr>
<tr>
<td>3</td>
<td>AgOTf (10%)</td>
<td>DCM</td>
<td>5</td>
<td>NR</td>
</tr>
<tr>
<td>4</td>
<td>Ph3PAuCl-AgOTf (1.0%)</td>
<td>DCM</td>
<td>5</td>
<td>49%</td>
</tr>
<tr>
<td>5</td>
<td>Ph3PAuCl-AgOTf (2.5%)</td>
<td>DCM</td>
<td>5</td>
<td>81%</td>
</tr>
<tr>
<td>6</td>
<td>Ph3PAuCl-AgOTf (5.0%)</td>
<td>DCM</td>
<td>5</td>
<td>96%</td>
</tr>
<tr>
<td>7</td>
<td>Ph3PAuCl-AgNTf (5.0%)</td>
<td>DCM</td>
<td>5</td>
<td>31%</td>
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<tr>
<td>8</td>
<td>Ph3PAuCl-AgOTf (5.0%)</td>
<td>Toluene</td>
<td>5</td>
<td>Trace</td>
</tr>
<tr>
<td>9</td>
<td>Ph3PAuCl-AgOTf (5.0%)</td>
<td>DMF</td>
<td>5</td>
<td>Trace</td>
</tr>
<tr>
<td>10</td>
<td>Ph3PAuCl-AgNTf (5.0%)</td>
<td>CH3CN</td>
<td>5</td>
<td>32</td>
</tr>
<tr>
<td>11</td>
<td>Ph3PAuCl-AgOTf (5.0%)</td>
<td>THF</td>
<td>5</td>
<td>26</td>
</tr>
<tr>
<td>12</td>
<td>Ph3PAuCl-AgOTf (5.0%)</td>
<td>EtOH</td>
<td>5</td>
<td>49</td>
</tr>
</tbody>
</table>

\(^a\) Each reaction was carried out with \(\beta\)-nucleoside 3a (0.20 mmol) with \(\text{H}_2\text{O} (1 \text{ eq.})\) and ethanol (6 eq.) in the presence of catalyst at room temperature under \(\text{N}_2\) atmosphere for 5 hours. \(^b\) Isolated yield.
Ph3PAuCl–AgOTf (entry 6). But Ph3PAuCl–AgNTf afforded 4a in low yield (31%) (entry 7). When the catalysis loading of Ph3-PAuCl–AgOTf was decreased to 2.5 mol% and 1 mol%, the corresponding yields were reduced to 81% and 49% respectively (entries 4–5).

After 5 mol% Ph3PAuCl–AgOTf was indentified as the best catalyst loading, the solvents effect was also investigated by performing the reaction in a series of organic solvents under room temperature conditions (entries 8–12). It was observed that toluene and dimethylformamide (DMF) only gave a trace amount of product 4a (~5%). The reaction also proceeded less efficiently in THF, acetonitrile or ethanol. DCM was found to be the optimum solvent.

With the optimized condition in hand, the synthesized β-nucleoside substrates 3a–i were subjected to 5 mol% Ph3PAuCl–AgOTf, ethanol (6 eq.) and H2O (1 eq.) in DCM. The results are summarized in Scheme 2. Except for cytidine derivate 4c, the reaction of pyrimidine nucleosides (3b, 3d–e), purine nucleosides (3a, 3f–h), and 7-deazaguanine nucleoside (3i) all proceeded smoothly and the desired products (4a–b, 4d–i) were obtained in nearly quantitative yield. For cytidine derivate 4c, it was speculated that the unprotected N-4 primary amine group may chelate with gold(i) ion to deactivate its catalytic activity. The speculation was further proved that the reaction can afford corresponding nucleoside 4d in high yield after its N-4 amine was protected by benzoate. For nucleosides 3g and 3h, the electron-withdrawing substituents (F and Cl) in purine base might reduce the nucleophilicity of N-6 amine, but they did not interfere the gold(i) catalyst’s activity. Therefore, nucleosides 4g and 4h could be obtained successfully. HPLC analysis of 4a–b and 4d–i showed that the no transesterification of 3′-benzoate to 2′-OH was noticed, which was crucial for following synthesis of 2′-modified ribonucleosides.

In order to further investigate the reaction mechanism, H2O18 was used in the control reaction. After reaction accomplished, O18-labeled isocoumarin G was obtained, which was confirmed by HRMS. According to the above evidences, a plausible catalytic cycle is proposed in Scheme 3. The coordination of the carbon–carbon triple bond of nucleoside A to the gold(i) catalyst improved the electrophilicity of alkyne (B). Subsequently, the carbonyl oxygen could attack the electron-deficient alkyne to form the intermediate C. While H2O addition to the formed onium ion would generate the intermediate D. After hydrogen atom transferred, nucleoside E would be released along with the generation of the intermediate F. Finally, isocoumarin G was yielded and the corresponding active gold(i) species were liberated to participate in the next catalytic cycle.

In order to testify this strategy to prepare 2′-modified nucleosides, disaccharide nucleoside 6 (9-(2′-O-β-D-ribofuranosyl-β-D-ribofuranosyl)-adenine) was synthesized. Disaccharide nucleoside is an important family of natural compounds, which is widely found in t-RNA, antibiotics, and other physiologically

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**Scheme 2** Gold(i)-catalyzed selective deprotection of β-nucleosides (4a–i).

**Scheme 3** Proposed catalytic cycle for the gold(i)-catalysed removing ortho-alkynyl benzoate.

**Scheme 4** Synthesis of 2′-O-β-D-ribofuranosyladenosine: (i) 1-O-acetyl-2,3,5-tri-O-benzoyl-β-D-ribofuranose, SnCl4, 1.2-dichloroethane, −10 °C, 8 hours, 42%; (ii) NH3, MeOH, 60 °C, 5 hours, 92%.
active compounds.\textsuperscript{16,17} Several synthetic approaches were reported. As shown in Scheme 4, glycosylation of nucleoside 4a with a little excess of 1-O-acetyl-2,3,5-tri-O-benzoyl-\beta-D-ribofuranose gave the corresponding disaccharide nucleoside in 42\% yield. After deprotection of all ester groups, nucleoside 6 (10 grams) was obtained in high purity. All the characterization spectra were identified with the reported data.

To further extend the application of this methodology, we also attempted to develop a new approach for synthesis of antitumor drug clofarabine 9.\textsuperscript{18,19} As presented in Scheme 5, the reaction of nucleoside 4h with trifluoromethanesulfonic anhydride (Tf$_2$O) in pyridine gave corresponding trifluoromethanesulfonic ester of 2'-OH in almost quantitative yield. After fluorination with Et$_3$N-HF, deacetylation with ammonia in methanol gave clofarabine (16 grams) in 56\% overall yield. HPLC analysis showed the purity is above 99\% and coincidence with reference standard from Sigma Aldrich.

In summary, ortho-alkynyl benzoate was proved to be an efficient neighboring participation group in stereoselective synthesis of nucleosides by using Vorbrüggen glycosylation. It could be removed as isocoumarin 5 using gold(1)-catalysis to afford 2'-OH nucleosides in high yield and selectivity. The powerfulness of present strategy was further demonstrated by the synthesis of 9-(2-O-benzoyl-b-ribofuranosyl-b-ribofuranosyl)-adenine 6 and antitumor drug clofarabine 9 in high overall yields. This novel protocol could be used as a general alternative approach for the synthesis of 2'-modified nucleosides. Its application in carbohydrate synthesis is under way.

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Notes and references


