

Cite this: *RSC Adv.*, 2017, 7, 12144

Antibacterial and catalytic activities of biosynthesized silver nanoparticles prepared by using an aqueous extract of green coffee bean as a reducing agent

Mengmeng Wang,^a Wenjing Zhang,^a Xuesong Zheng^{*b} and Peizhi Zhu^{*a}

Spherical biogenic silver nanoparticles (AgNPs) were synthesized using aqueous green coffee bean extract as a reducing agent. The green coffee bean extract was used for the time to reduce silver ions *via* a one-step, green, and rapid approach. The prepared AgNPs were characterized *via* transmission electron microscopy (TEM), atomic force microscopy (AFM), and UV-visible spectroscopy. Dynamic light scattering (DLS) was performed to measure the particle size. These AgNPs exhibit strong antibacterial activity against both Gram-negative and Gram-positive bacteria and the minimum inhibitory concentration (MIC) of AgNPs for *Escherichia coli* and *Staphylococcus aureus* are 0.05 $\mu\text{mol L}^{-1}$ and 0.1 $\mu\text{mol L}^{-1}$, respectively. Furthermore, AgNPs could serve as an effective catalyst for the reduction of 4-nitrophenol to 4-aminophenol in the presence of NaBH_4 .

Received 3rd December 2016

Accepted 4th February 2017

DOI: 10.1039/c6ra27706c

rsc.li/rsc-advances

Introduction

In recent years, nanoparticles (NPs) have attracted extensive attention due to their size effect, surface effect, quantum size effect, macroscopic quantum tunnelling effect, and dielectric confinement effect. NPs have been widely used as magnetic materials,¹ ceramic materials,² sensing elements,³ semiconductors,⁴ catalysts,⁵ and even in biomedicine.⁶ As a noble metal with excellent performance, silver nanoparticles (AgNPs) exhibit broad applications in catalysis,⁷ biomedicine,⁸ as an antimicrobial agent,⁹ in chemical and biological sensings,¹⁰ and cancer therapy.¹¹ The conventional method for preparing silver nanoparticles, which is based on the chemical reduction of a silver salt, is simple and effective, but the removal of the agents used is both a costly and time-consuming process. Furthermore, the residual agents show biological toxicity.^{12,13}

Thus, there is an increased interest in using biodegradable and biocompatible reducing agents to prepare AgNPs. Over the last decade, polysaccharide-based biopolymers such as alginate, starch, and chitosan^{14–16} have been used as green alternatives^{17–20} to conventional agents due to their low-cost, non-toxicity, and environment-friendly properties. In most cases, the chemical dispersants used for controlling the growth of metallic nanoparticles were replaced by these biopolymers due to their low-toxicity to animal cells.²¹ Nano-sized metal catalysts

play a key role in the chemical industries; among these, silver nanoparticles have gained significant attention owing to their unique electronic properties and high surface to volume ratio.²² In addition, silver nanocatalysts are cost-effective, efficient, and environmentally friendly. In particular, silver nanoparticles present highly efficient catalytic activity in the oxidation of methanol and ethylene,^{23,24} as well as in the reduction of nitric oxides (NO_x).²⁵

For the synthesis of silver nanoparticles, a variety of methods have been employed such as chemical reduction,²⁶ electrochemical process,²⁷ microemulsion method,²⁸ laser ablation,²⁹ and hydrothermal method.³⁰ Chemical reduction is still the most common method. Recently, inspired by the concept of green chemistry, several studies have been focussed on the development of novel eco-friendly reducing agents for synthesizing AgNPs. Many interesting biomolecules including those obtained from plant extract,^{31–33} bacteria,³⁴ viruses,³⁵ DNA,³⁶ and peptides^{37–39} have been used to successfully synthesize AgNPs.⁴⁰

In this study, we report the synthesis of silver nanoparticles using green coffee bean extract as a reducing agent for the first time. Green coffee bean extracts were obtained from the seeds of *Coffea arabica*, *Coffea canephora*, and *Coffea liberica*. Green coffee bean extracts have been used as a healthy food supplement for long time because the main ingredient chlorogenic acid is a native antioxidant and can reduce blood pressure. We used TEM, AFM, and DLS to characterize the morphology and particle size of the silver nanoparticles synthesized using green coffee bean extracts. The formation of silver nanoparticles was studied by UV-visible spectroscopy. In antibacterial experiments, *Escherichia coli* and *Staphylococcus aureus* were used to

^aSchool of Chemistry and Chemical Engineering, Yangzhou University, Jiangsu 225002, PR China

^bSchool of Perfume and Aroma Technology, Shanghai Institute of Technology, China. E-mail: xuesongzheng10@gmail.com



evaluate the antimicrobial activity of AgNPs against Gram-negative and Gram-positive bacteria, respectively. Moreover, the catalytic activity of the as-prepared silver nanoparticles for the reduction of 4-nitrophenol in the presence of NaBH_4 was also studied.

Materials and methods

Chemicals and materials

Silver nitrate (AgNO_3 , 99.8%), 4-nitrophenol (4-NP), sodium borohydride (NaBH_4 , 98%) were purchased from Sinopharm Chemical Reagent Co. Ltd (Shanghai, China). Green coffee bean extracts were purchased from Xi'an Zebang Biological Technology Co. Ltd. Chlorogenic acid (CGA, 98%) was purchased from Bioduly Biochemical Reagent (Nanjing, China). All the chemicals were of analytical reagent grade and were used without further purification. All the solutions were freshly prepared using deionized double-distilled water.

Preparation of silver nanoparticles

For the biosynthesized silver nanoparticles, 0.2175 g silver nitrate was dissolved in 15 mL deionized double-distilled water. Then, 25 mL of a 1 mM aqueous green coffee bean extract solution was added dropwise to a silver nitrate solution under stirring at 30 °C. To ensure a complete mixing, stirring was maintained for 5 minutes. The color of the as-prepared silver nanoparticles appeared similar to that of the green coffee bean extracts solution. The synthesis of CGA-AgNPs was performed in a water-based system; 2.8 mL of 0.5 mM AgNO_3 was placed in a glass vial with a cap and heated to 80 °C on a hot plate under stirring for 3 min. A 1.2 mL of 0.05 mM CGA was added dropwise under stirring on a hot plate within 2 min. Then, the reaction mixture was further incubated in an oven for 10 h.

TEM analysis of the silver nanoparticles

Transmission electron microscopy (TEM, Tecnai C2 F30 S-Twin, FEI, USA) was used to observe the morphologies and particle sizes of AgNPs, and selected-area electron diffraction (SAED) patterns were obtained *via* high-resolution transmission electron microscopy (HRTEM). Transmission electron microscopy was performed by fixation on a 200-mesh carbon-coated copper grid.

UV-visible spectra analysis

The absorbance spectra of AgNPs and CaP-coated AgNPs were obtained by UV-Vis spectroscopy (Varian Cary 500, USA).

Dynamic light scattering measurement

The particle size distribution of the biosynthesized AgNPs was analyzed using a Zetasizer Nanoseries apparatus (Malvern) with laser He-Ne (4 mW) at 632.8 nm.

Atomic force microscope analysis

AFM (Bruker, Santa Barbara, CA) images were obtained using Shimadzu nano search microscope SFT-3500 in QNM

(quantitative nanomechanical mapping) mode. The typical resonance frequency of the cantilever with an integrated silicon tip (spring constant 40 N m⁻¹) was between 283.8 and 360 kHz.

Antibacterial test

The antibacterial activity of the synthesized AgNPs was tested by growth inhibition curve method, and the minimum inhibitory concentration (MIC) was also determined. The assay was performed on *Escherichia coli* (ATCC 25922) and *Staphylococcus aureus* (ATCC 25923), which acted as the representatives for Gram-negative and Gram-positive bacteria, respectively.

First, the suspension solutions of *E. coli* and *S. aureus* were prepared by shaking in an incubator at 37 °C for 8 h. The AgNPs solutions of various concentrations were prepared by diluting the nanosilver solution of 50 $\mu\text{mol L}^{-1}$ with double-distilled H_2O . Then, 100 μL of AgNPs solutions and 900 μL of LB were completely mixed in 1.5 mL microcentrifuge tubes. The tube of dd H_2O in place of the AgNPs solution in the mixture was used as the control. Therefore, the final concentration of AgNPs in the media were 0.5 $\mu\text{mol L}^{-1}$, 0.2 $\mu\text{mol L}^{-1}$, 0.1 $\mu\text{mol L}^{-1}$, 0.05 $\mu\text{mol L}^{-1}$, 0.04 $\mu\text{mol L}^{-1}$, 0.03 $\mu\text{mol L}^{-1}$, 0.02 $\mu\text{mol L}^{-1}$, 0.01 $\mu\text{mol L}^{-1}$, and 0.005 $\mu\text{mol L}^{-1}$. Two hundred microliters of the mixture was transferred with pipette into each well of a Corning 96-well microtiter plate and 2 μL of a bacteria suspension solution was inoculated in each well. Immediately, this plate was incubated at 37 °C for 180 min in Multimode Plate-Reader (Tecan infinite 200PRO). The OD₆₀₀ values were measured at the intervals of 5 min, following 20 s plate-shaking every time.

Catalytic reduction of 4-nitrophenol

In a typical run for the reduction of 4-NP by NaBH_4 , 0.2 mL of biosynthesized silver nanoparticles (1 mM) was added to 5 mL reaction solution containing 0.05 mmol fresh solution of 4-NP and 0.2 mmol NaBH_4 solution. To set a blank group, the reaction system was kept same as the abovementioned system except that the prepared silver nanoparticles were replaced by ultra-pure water. The reaction in the presence of NaBH_4 was immediately monitored by UV-visible spectrophotometric monitoring instrument over a 210–600 nm wavelength.

Results and discussion

Characterization of the biosynthesized silver nanoparticles

TEM was carried out to investigate the morphology and size of the obtained AgNPs. Fig. 1a, d and e shows typical TEM images of the AgNPs and most AgNPs nanoparticles were well-dispersed in the solution of green coffee bean extract. Fig. 1a and b shows that the nanoparticles were spherical in shape. The high-resolution TEM image, as shown in Fig. 1c, shows polycrystalline structure of the particles. From Fig. 1d, clear lattice fringes can be easily observed, and the interplanar spacing of these was measured to be 0.209 nm that corresponds to the Ag (200) plane.³¹ The selected-area electron diffraction (SAED) pattern, as shown in Fig. 1e, proves the polycrystalline structure of the synthesized AgNPs, which is consistent with the TEM result. The AFM topography image (Fig. 1f and g) shows that the



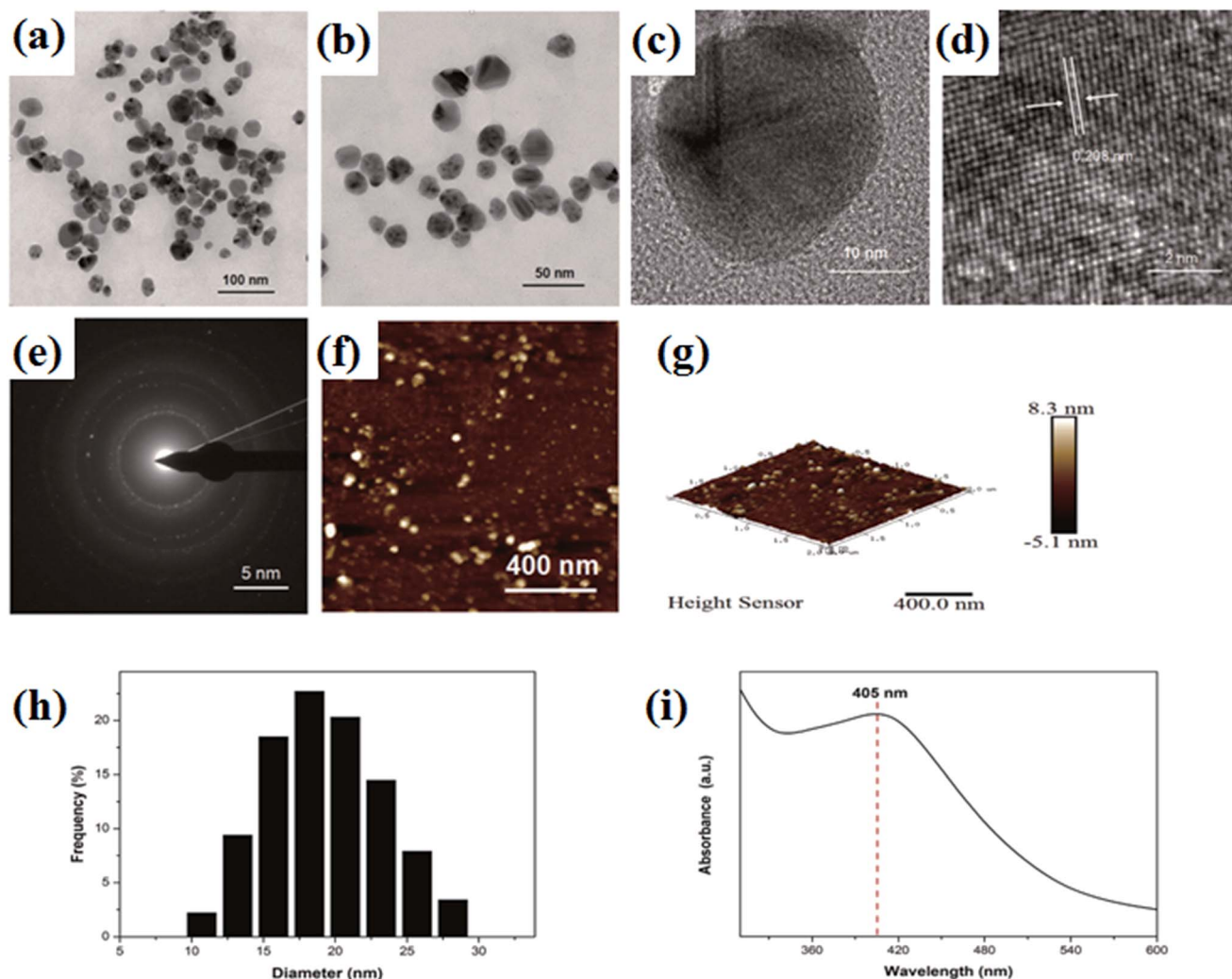


Fig. 1 HRTEM (a–d) images and SAED pattern (e) of the AgNPs. AFM (f and g) images of the biosynthesized silver nanoparticles. (h) Size-distribution histogram of the biosynthesized AgNPs. (i) UV-vis spectrum of the biosynthesized AgNPs.

size of AgNPs ranges from 10 to 30 nm in diameter, which is consistent with the DLS histogram (Fig. 1h). Fig. 1i displays the UV-vis absorption spectra of the as-prepared AgNPs. An obvious absorption peak at 405 nm, as shown in Fig. 1i, confirmed the formation of AgNPs since the characteristic silver SPR (surface plasmon resonance) bands are around 400–450 nm.⁴¹

Antibacterial properties of the biosynthesized AgNPs

The 180 min growth curve for *E. coli* and *S. aureus* in LB media containing various concentrations of AgNPs are displayed in Fig. 2. Based on these curves, the excellent antibacterial activity of the AgNPs against both Gram-negative and Gram-positive bacteria can be concluded, and the minimum inhibitory concentration (MIC) (MIC = 0.04 $\mu\text{mol L}^{-1}$ for *E. coli* and 0.2 $\mu\text{mol L}^{-1}$ for *S. aureus*) can be determined. For *E. coli* (Fig. 2a), the concentration of 0.02 $\mu\text{mol L}^{-1}$ AgNPs exhibited a weak growth inhibition. During 180 min of the assay, 0.05 $\mu\text{mol L}^{-1}$ AgNPs absolutely repressed the bacterial growth. However, for *S. aureus* (Fig. 2b), 0.05 $\mu\text{mol L}^{-1}$ AgNPs just slightly repressed

the growth and the absolute inhibition concentration was approximately 0.1 $\mu\text{mol L}^{-1}$.

This difference may be due to the difference in the bacterial cell wall structure. Shrivastava *et al.* elucidated in their study that the different cell wall structure leads to variety in the penetration of AgNPs.⁴² The cell wall of the Gram-positive bacteria is composed of a thick layer of peptidoglycan that constitutes linear polysaccharides chains cross-linked by short peptides, thus forming a rigid structure. This leads to difficulty in the penetration of AgNPs compared to that for Gram-negative bacteria, where the cell wall possesses a much thinner layer of peptidoglycan.

Catalytic reduction of 4-nitrophenol

The reduction of 4-nitrophenol by NaBH_4 was taken as a model reaction to examine the catalytic activity of the AgNPs. The absorption peak of 4-NP with a light yellow color shifted from 317 nm to 400 nm after the addition of freshly prepared NaBH_4 solution. This is due to the presence of NaBH_4 that changed the



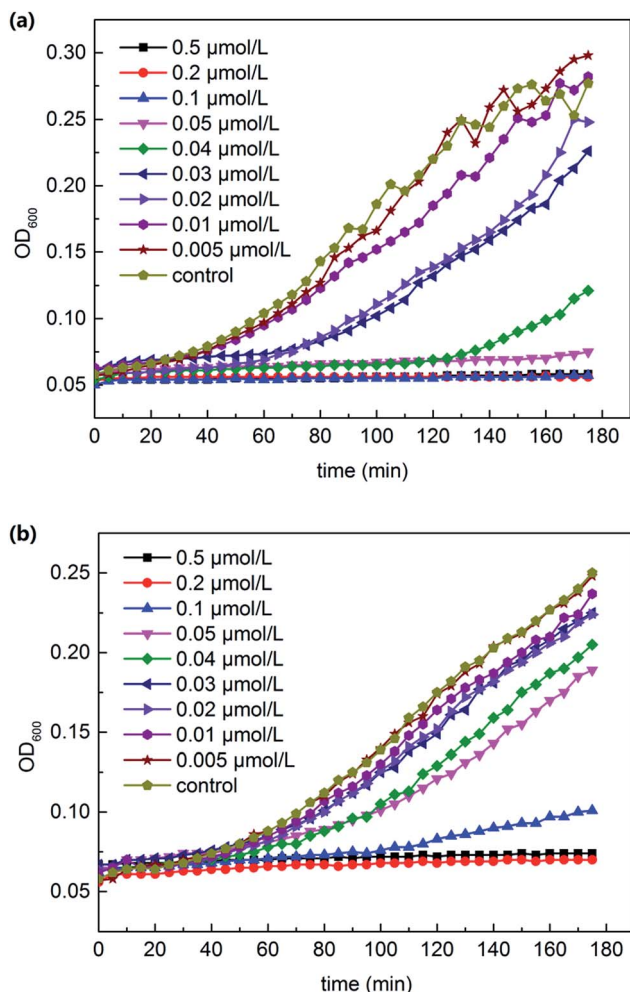


Fig. 2 180 min growth curve for *E. coli* (a) and *S. aureus* (b) in LB media containing various concentrations of AgNPs.

pH of the solution.⁴³ Moreover, the light yellow color turned to intense yellow, indicating the formation of 4-nitrophenolate ion.

The catalytic process of this reaction was monitored by UV-vis spectroscopy; the absorption intensity at 400 nm for 4-nitrophenolate ion remained unvaried without the silver nanocatalyst. As shown in Fig. 3a, a new band at around 300 nm emerged after adding 2 mL AgNPs as a catalyst, indicating the reduction of 4-NP to 4-AP by NaBH₄. Until the end of the reaction, the intensity of the absorption peak at 400 nm gradually decreased with time, whereas the absorption peak at 300 nm simultaneously increased. The kinetic process of the reduction was monitored by measuring the extinction of the solution at 400 nm as a function of time. A good linear correlation is shown in Fig. 3b and the apparent rate constant (K) of this catalytic reaction was 0.0456 s^{-1} , suggesting that the biosynthesized AgNPs possessing a larger surface area exhibit high-effective catalytic activity for the reduction of 4-NP.

The main content of the green coffee bean is chlorogenic acid (CGA). However, we could not synthesize silver nanoparticles using only chlorogenic acid as a reducing agent at

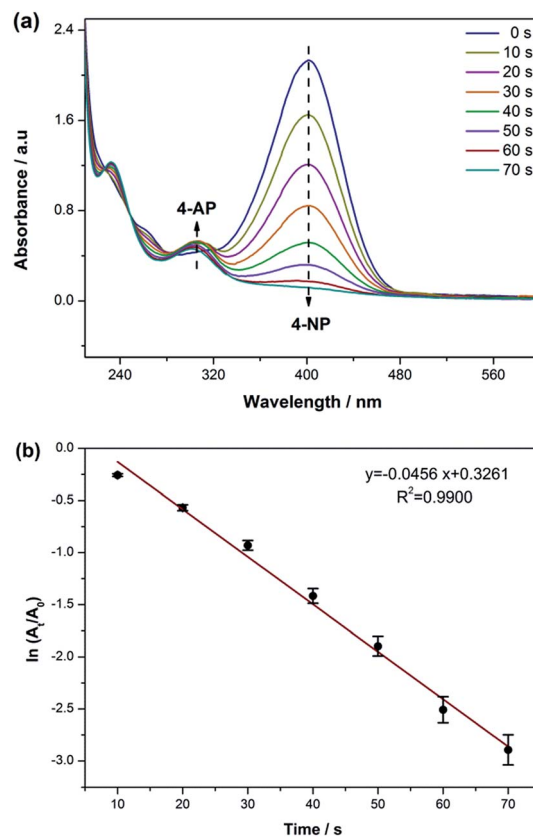


Fig. 3 (a) UV-vis spectra of the 4-NP reduction in the presence of NaBH₄ and 0.2 mL biosynthesized AgNPs and (b) catalytic degradation curves of 4-NP over the biosynthesized AgNPs.

30 °C. Then, reaction temperature was increased by a step of every 10 °C. The CGA-AgNPs were synthesized at 80 °C, which is consistent with the results of previous report.⁴⁴ TEM images showed that the CGA-AgNPs were well-dispersed nanoparticles. The average particle sizes of the CGA-AgNPs were smaller than those of the AgNPs synthesized using aqueous green coffee bean extracts (Fig. 4). The synthesis of CGA-AgNPs showed that the reducing agent in the aqueous green coffee bean extracts was mainly chlorogenic acid. Green coffee bean extracts contain a variety of components such as caffeine, sucrose, citric acid, and trigonelline, many isomers of chlorogenic acid, amino

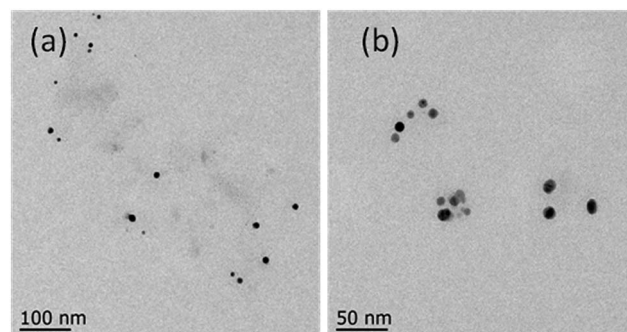


Fig. 4 TEM images of CGA-AgNPs.



acids, *etc.*⁴⁵ Chlorogenic acid is the main component in the green coffee bean extracts, which is accountable for the reduction of Ag^+ to Ag^0 for the green synthesis of AgNPs. We hypothesized that other components, such as citric acid, can also act as reducing agents and the synergistic effect of several reducing agents lowered the energy barrier for the reduction of silver ions. These chemicals also act as capping agents to stabilize the synthesized silver nanoparticles. Since green coffee bean extract is much cheaper than chlorogenic acid, silver nanoparticles can be synthesized using a green coffee bean extract as a reducing agent at 30 °C, which is a much lower temperature than that required by chlorogenic acid only. This environmentally friendly approach is suitable for the large-scale biosynthesis of silver nanoparticles due to its operational simplicity, low-cost, and green conditions. These well-capped silver nanoparticles produced by the green coffee bean extract are more stable and can strongly inhibit the growth of both Gram-negative and Gram-positive bacteria. In addition, these silver nanoparticles have potential in the simple and safe reduction of phenol compounds in water.

Conclusions

In this study, biogenic spherical AgNPs were successfully synthesized using aqueous green coffee bean extracts for the first time. The approach was reproducible, cost-effective, and eco-friendly. TEM and DLS analysis showed that the particle size of the as-prepared silver nanoparticles ranged from 12 nm to 31 nm. These small AgNPs exhibit strong antibacterial activity against *E. coli* and *S. aureus* and its MICs were evaluated to be 0.05 $\mu\text{mol L}^{-1}$ and 0.1 $\mu\text{mol L}^{-1}$, respectively. Moreover, biosynthesized silver nanoparticles could be an effective catalyst for the reduction of 4-nitrophenol to 4-aminophenol in the presence of NaBH_4 .

Acknowledgements

We would like to thank Jiangsu Province for specially appointed professorship to Professor Peizhi Zhu and Testing Center of Yangzhou University for the support. This work was also supported by the Priority Academic Program Development of Jiangsu Higher Education Institutions.

Notes and references

- H. Lin, R. R. Zhang and Q. W. Chen, *Nanoscale*, 2014, **6**, 14064–14105.
- F. Hua, T. H. Cui and Y. M. Lvov, *Nano Lett.*, 2004, **4**, 823–825.
- P. Ginzburg, N. Berkovitch, A. Nevet, I. Shor and M. Orenstein, *Nano Lett.*, 2011, **11**, 2329–2333.
- X. L. Hu, G. S. Li and J. C. Yu, *Langmuir*, 2010, **26**, 3031–3039.
- W. Wang, T. Y. Chiang, D. Velego and T. E. Mallouk, *J. Am. Chem. Soc.*, 2013, **135**, 10557–10565.
- B. H. Dong, C. Y. Li, G. C. Chen, Y. J. Zhang, Y. Zhang, M. J. Deng and Q. B. Wang, *Chem. Mater.*, 2013, **25**, 2503–2509.
- M. X. Zhang, Y. H. Cao, Y. Chong, Y. F. Ma, H. L. Zhang, Z. W. Deng, C. H. Hu and Z. J. Zhang, *ACS Appl. Mater. Interfaces*, 2013, **5**, 13325–13332.
- S. Dasa and B. B. Dhar, *RSC Adv.*, 2014, **4**, 46285–46292.
- M. S. Noh, B. H. Jun, S. Kim, H. Kang, M. A. Woo, A. Minai-Tehrani, J. E. Kima, J. Kim, J. Y. Park, H. T. Lim, S. C. Park, T. Hyeon, Y. K. Kim, D. H. Jeong, Y. S. Lee and M. H. Cho, *Biomaterials*, 2009, **30**, 3915–3925.
- M. R. Reithofer, A. Lakshmanan, A. T. K. Ping and J. M. Chin, *Biomaterials*, 2014, **35**, 7535–7542.
- D. Kagan, P. Calvo-Marzal, S. Balasubramanian, S. Sattayasamitsathit, K. M. Manesh, G. U. Flechsig and J. Wang, *J. Am. Chem. Soc.*, 2009, **131**, 12082–12083.
- Y. Yang, T. Liu, L. Cheng, G. S. Song, Z. Liu and M. W. Chen, *ACS Appl. Mater. Interfaces*, 2015, **7**, 7526–7533.
- P. Sanpui, A. Chattopadhyay and S. S. Ghosh, *ACS Appl. Mater. Interfaces*, 2011, **3**, 218–228.
- E. Y. Furuya and F. D. Lowy, *Nat. Rev. Microbiol.*, 2006, **4**, 36–45.
- S. A. Jones, P. G. Bowler, M. Walker and D. Parsons, *Wound Repair Regen.*, 2004, **12**, 288–294.
- V. Prabhu, P. Karthick, A. Rajendran, D. Natarajan, M. S. Kiran and D. K. Pattanayak, *RSC Adv.*, 2015, **5**, 50767–50777.
- Z. Abdali, H. Yeganeh, A. Solouk, R. Gharibi and M. Sorayya, *RSC Adv.*, 2015, **5**, 66024–66036.
- R. S. Chen, H. W. Ni, H. S. Zhang, G. Yue, W. T. Zhan and P. Y. Xiong, *Vacuum*, 2013, **89**, 249–253.
- A. M. Mittelman, D. S. Lantagne, J. Rayner and K. D. Pennell, *Environ. Sci. Technol.*, 2015, **49**, 8515–8522.
- H. J. Klasen, *Burns*, 2000, **33**, 117–138.
- W. Lee, K. J. Kim and D. G. Lee, *BioMetals*, 2014, **27**, 1191–1201.
- L. Sintubin, W. D. Windt, J. Dick, J. Mast, D. vanderHa, W. Verstraete and N. Boon, *Appl. Microbiol. Biotechnol.*, 2009, **84**, 741–749.
- E. Qayyum, V. A. Castillo, K. Warrington, M. A. Barakat and J. N. Kuhn, *Catal. Commun.*, 2012, **28**, 128–133.
- M. J. Lippits and B. E. Nieuwenhuys, *Catal. Today*, 2010, **154**, 127–132.
- S. Wunder, F. Polzer, Y. Lu, Y. Mei and M. Ballauff, *J. Phys. Chem. C*, 2010, **114**, 8814–8820.
- Y. Sun and Y. Xia, *Science*, 2002, **298**, 2176–2179.
- C. D. Gu and T. Y. Zhang, *Langmuir*, 2008, **24**, 12010–12016.
- S. Pal, E. J. Yoon, Y. K. Tak, E. C. Choi and J. M. Song, *J. Am. Chem. Soc.*, 2009, **131**, 16147–16155.
- H. S. Shin, H. J. Yang, S. B. Kim and M. S. Lee, *J. Colloid Interface Sci.*, 2004, **274**, 89–94.
- M. Feng, M. Zhang, J. M. Song, X. G. Li and S. H. Yu, *ACS Nano*, 2011, **5**, 6726–6735.
- A. A. Kajani, A. K. Bordbar, S. H. Z. Esfahani, A. R. Khosropour and A. Razmjou, *RSC Adv.*, 2014, **4**, 61394.
- S. Irvani, *Green Chem.*, 2011, **13**, 2638–2650.
- S. K. Li, Y. H. Shen, A. J. Xie, X. R. Yu, L. G. Qiu, L. Zhang and Q. F. Zhang, *Green Chem.*, 2007, **9**, 852–858.
- H. Zhang, Q. Li, Y. Lu, D. Sun, X. Lin, X. Deng, N. He and S. Zheng, *J. Chem. Technol. Biotechnol.*, 2005, **80**, 285–290.



- 35 C. X. Yang, S. Jung and H. Yi, *Biochem. Eng. J.*, 2014, **89**, 10–20.
- 36 S. Kundu, *Phys. Chem. Chem. Phys.*, 2013, **15**, 1107–14119.
- 37 K. T. Nam, Y. J. Lee, E. M. Krauland, S. T. Kottmann and A. M. Belcher, *ACS Nano*, 2008, **2**, 1480–1486.
- 38 G. Upert, F. Bouillere and H. Wennemers, *Angew. Chem., Int. Ed.*, 2012, **51**, 4231–4234.
- 39 H. Wennemers, *J. Pept. Sci.*, 2012, **18**, 437–441.
- 40 P. R. Rathi Sre, M. Reka, R. Poovazhagi, M. Arul Kumar and K. Murugesan, *Spectrochim. Acta, Part A*, 2015, **135**, 1137–1144.
- 41 E. C. Njagi, H. Huang, L. Stafford, H. Genuino, H. M. Galindo, J. B. Collins, G. E. Hoag and S. L. Suib, *Langmuir*, 2011, **27**, 264–271.
- 42 S. Shrivastava, T. Bera, A. Roy, G. Singh, P. Ramachandrarao and D. Dash, *Nanotechnol.*, 2007, **18**, 225103–225122.
- 43 B. Baruah, G. J. Gabriel, M. J. Akbashev and M. E. Booher, *Langmuir*, 2013, **29**, 4225–4234.
- 44 H. J. Noh, H. S. Kim, S. H. Jun, Y. H. Kang, S. Cho and Y. Park, *J. Nanosci. Nanotechnol.*, 2013, **13**, 5787–5793.
- 45 F. Wei, K. Furihata, F. Hu, T. Miyakawa and M. Tanokura, *Magn. Reson. Chem.*, 2010, **48**, 857–865.

