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A smart NIR fluorescent probe for the highly selective detection of palladium†

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A tailor-made colorimetric and NIR fluorescent probe for Pd⁰ was developed by introducing allyl chloroformate into the methylene blue (MB) fluorophore. The probe exhibited vivid color change and significant fluorescence enhancement towards Pd⁰, accompanied with excellent selectivity and sensitivity. No absorption and fluorescence response could be observed towards other metal ions. The fluorescence intensity of this probe showed a linear response to Pd⁰ in the concentration range of 0–2 μM with a detection limit of 5.7 nM. Moreover, living cell imaging results indicated that this probe holds promising application prospects for detecting intracellular Pd⁰ species based on its low cytotoxicity and specific turn-on NIR emission.

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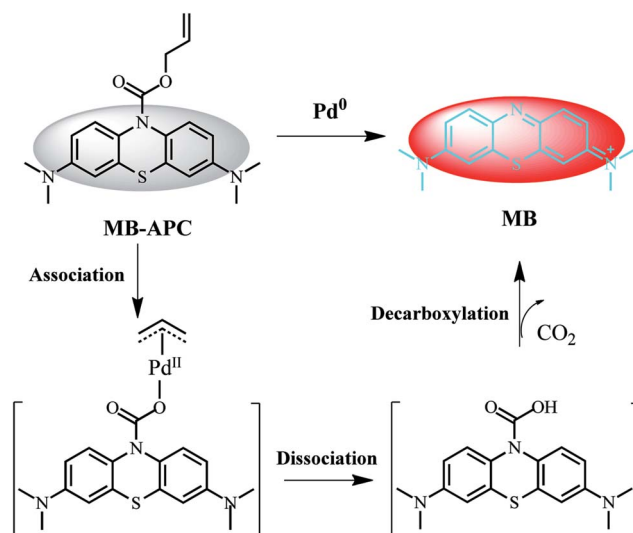
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Introduction

Along with the rapid development of modern industry, heavy and transition metals have been widely applied in many fields such as chemistry, biology and environmental science. Palladium, as an important transition metal, plays an important role in various fields such as glass, fine chemical, electronics, and petroleum industries, as well as in automobiles.^{1–3} However, the high level of residual palladium due to the wide exploitation of palladium species has raised palladium contamination, which could cause severe adverse health effects.^{4,5} Therefore, it is urgent to develop highly selective and sensitive methods for palladium detection in environmental and biological samples. Atomic absorption spectroscopy (AAS), inductively coupled plasma mass spectroscopy (ICP-MS), and plasma emission spectroscopy are the conventional approaches for palladium detection, which can achieve accurate and sensitive detection, but expensive facilities, well-controlled experimental conditions and complicated sample-pretreatment procedures are required.^{6–8} In recent years, optical probes, including colorimetric or fluorescent probes, emerged as powerful alternatives to monitor and sense palladium species because of their convenience, on-site detection, ease of manipulation and

biological applications.^{9–13} In particular, fluorescent probes with near-infrared (NIR) emission are extremely favourable for bioimaging because of deep penetration and minimum autofluorescence.^{14,15} However, most of the reported probes for palladium display absorbance and emissions in the visible region, which greatly restricts their applications in biological imaging. To this end, it is highly desired and urgent to develop NIR fluorescent probes for selective and sensitive detection of palladium.

Methylene blue (MB), an oxidised phenothiazin compound already FDA-approved for several indications, was widely used as a medication and stain in both clinical and basic research field.^{16–18} Recently, MB was also applied as NIR imaging agent in



Scheme 1 Chemical structure and proposed sensing process of MB-APC for Pd⁰ based on Tsuji–Trost reaction.

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the image-guided surgery.^{19–22} However, the development of new fluorescent probes based on this NIR fluorophore has rarely been reported.²³ It is well-known that MB could be converted to a colorless and non-emissive form called leucomethylene blue under reducing or acidic condition. To obtain the NIR fluorescent probe for Pd⁰, the allyl chloroformate moiety was introduced into the leucomethylene blue through a carbamate bond, leading to a stable colorless and non-emissive probe **MB-APC**. We speculated that **MB-APC** would be uncaged through the Tsuji–Trost reaction in the presence of Pd⁰,²⁴ leading to the releasing of free MB, accompanied with color change and turn-on NIR emission (Scheme 1).

Results and discussion

The time-dependent absorption and fluorescence intensity changes of probe **MB-APC** after adding 3 equivalent of Pd(PPh₃)₄ were evaluated to investigate its spectral properties and response to Pd⁰. As shown in Fig. 1A, **MB-APC** exhibited almost no absorbance in the visible region in the EtOH-PBS (9 : 1, v/v) solution. An absorption peak appeared at around 657 nm after adding 3 equivalent of Pd(PPh₃)₄ solution, which increased gradually with incubation time and reached the maximum values after incubation for 30 min at room temperature. In the fluorescence spectra, no emission could be observed for **MB-APC** solution (Fig. 1B), which was consistent

with leucomethylene blue. After adding 3 equivalent of Pd(PPh₃)₄ solution, an emission peak at approximately 676 nm appeared and was significantly enhanced with incubation time, accompanied with slight red-shift. The emission peak was shifted to 681 nm and the fluorescence intensity reached a plateau in 30 min at room temperature (Fig. 1B). The turn-on fluorescence may be attributed to the releasing of free MB moiety through the broken of the carbamate bond, which was confirmed by almost the same absorbance and emissive peak with MB from commercial source (Fig. S4†), and the HRMS result (Fig. S5†). It is also noteworthy that both the excitation and emission of **MB-APC** reach the NIR range, which is very attractive for intracellular sensing and imaging.

To further explore the sensing ability of **MB-APC** to Pd⁰, UV-vis and fluorescence titration experiments were carried out. As shown in Fig. 2A, the absorption band at 657 nm enhanced with the increasing Pd⁰ concentrations concomitantly. The significant absorbance enhancement (about 393 fold increase) was accompanied by a marked and vivid color change from achromatic to blue in ambient light (inset of Fig. 2A), demonstrating that **MB-APC** could be utilized as an on-site and naked-eye indicator for Pd⁰. Meanwhile, the linear equation was obtained. As shown in Fig. S6A,† the plots of absorption fitted

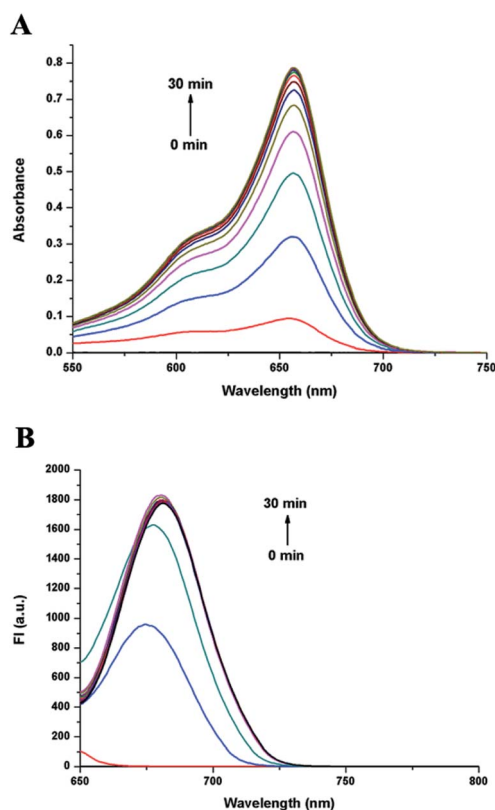


Fig. 1 UV-vis (A) and fluorescence spectral changes (B) of MB-APC (10 μM) against time in the presence of Pd(PPh₃)₄ (3 equiv.) in EtOH-PBS (9 : 1, v/v) at room temperature.

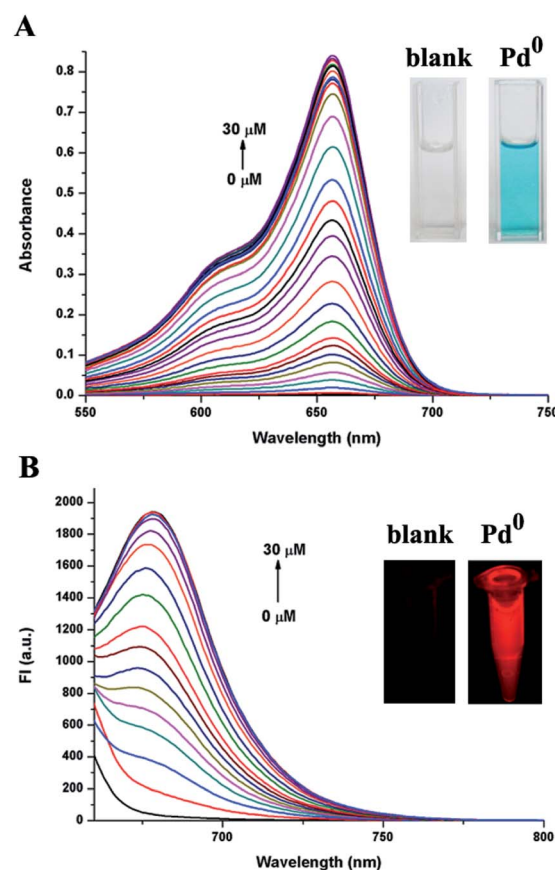


Fig. 2 UV-vis (A) and fluorescent (B) spectroscopic titration of MB-APC by stepwise addition of Pd(PPh₃)₄. Conditions: [MB-APC] = 10 μM; V_{EtOH} : V_{PBS} = 9 : 1, 10 mM PBS buffer, PH 7.4, λ_{ex} = 650 nm. The spectra were recorded at 30 min intervals. Inset: color and emission changes of MB-APC upon addition of Pd(PPh₃)₄.



linearly with Pd^0 concentration range of 0–6 μM with a correlation coefficient of 0.994. The detection limit of absorption for Pd^0 was as low as 12.48 nM. Moreover, the enhancement (about 55 fold increase) of fluorescence intensity can be easily found upon addition of increasing amounts of Pd^0 , which could also be observed by the naked eye under the UV irradiation (Fig. 2B). As shown in Fig. S6B,[†] the plots of fluorescence fit linearly with Pd^0 concentration range of 0–2 μM with a correlation coefficient of 0.994. The detection limit of fluorescence for Pd^0 was deduced to be as low as 5.7 nM. The result indicated that **MB-APC** had low detection limit in both absorption and fluorescence, which illustrated that the probe can be utilized as an efficient tool for sensing traces of palladium in both colorimetry and fluorescence signal with high sensitivity. Compared with other reported fluorescent probes for Pd^0 (Table S1[†]), NIR excitation and emission, and high sensitivity of **MB-APC** makes it a robust tool for sensing in both environment and biology samples.

To clarify the specificity of **MB-APC** towards Pd^0 over other metal ions, the absorption and the fluorescent spectra response were further investigated upon addition of various metal ions to EtOH-PBS solution of **MB-APC**, respectively. 3 equivalent of Pd^0 and 10 equivalent of other metal ions were added to the detecting system. As expected, only Pd^0 caused such remarkable signal changes in the both absorption and fluorescence spectra of **MB-APC** (Fig. 3). In contrast, other noble metal with catalysis such as Ru^0 , and other metal ions such as Ag^+ , Fe^{3+} , Hg^{2+} , K^+ , Na^+ , Pb^{2+} , Ca^{2+} , Mg^{2+} , Cu^{2+} , Ni^{2+} , Co^{2+} , Ba^{2+} , Zn^{2+} , Mn^{2+} , Pt^{2+} , Cd^{2+} , Al^{3+} , Cr^{3+} , showed almost negligible disturbance to the spectra and color (Fig. 3), even up to 10 equivalent concentrations, indicating that **MB-APC** showed a notable selectivity to Pd^0 . **MB-APC** could efficiently discriminate Pd^0 from other metal ions by the visual readout (Fig. S7[†]). Moreover, metal chelators such as EDTA have little influence on this detecting system (Fig. S8[†]). All these experimental results suggested that

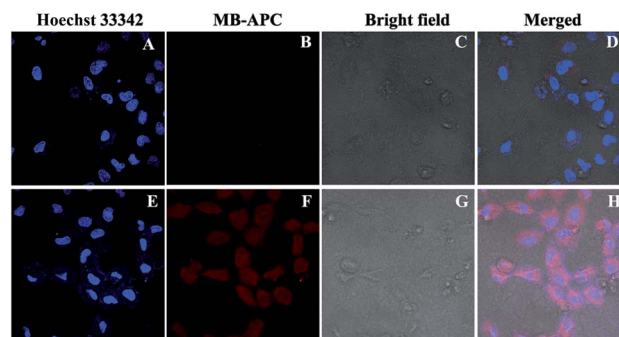


Fig. 4 Confocal fluorescence images of living HeLa cells using a 635 nm laser. HeLa cells were co-stained with Hoechst 33342 (10 μM) and MB-APC (10 μM), without (A–D) and with (E–H) incubation of palladium (30 μM) for 30 min.

this probe **MB-APC** had an excellent selectivity for Pd^0 over other metal ions.

Encouraged by the above findings, the potential intracellular applications of **MB-APC** were investigated using confocal laser scanning microscopy. We first studied the cytotoxicity of this probe toward using a standard MTT assay. The results revealed that exhibited very low cytotoxicity to living cells for 48 h even up to 100 μM (Fig. S9[†]). Living cell imaging assay was conducted using HeLa cells. As shown in Fig. 4A–D, co-staining living cells using Hoechst 33342 revealed that free **MB-APC** exhibited almost no fluorescence signal, which was consistent with fluorescent spectra studies. By contrast, after incubation with Pd^0 for 30 min, a distinct strong emission response could be observed (Fig. 4E–H). The results demonstrated that **MB-APC** is both cell-permeable and capable of sensing palladium in living cells, which makes it a versatile tool for the detection of palladium species in environmental samples and in living cells.

Conclusions

In conclusion, a colorimetric and NIR fluorescent probe **MB-APC** for Pd^0 was rationally developed. The distinct color change and turn-on NIR emission make it an on-site and visual indicator for Pd^0 . **MB-APC** also exhibited high specificity and sensitivity, with the detection limit of 5.7 nM. In addition, **MB-APC** has been successfully applied in detecting and imaging of Pd^0 in living cells, which revealed that **MB-APC** has the potential to track intracellular palladium species taking advantage of its specific turn-on NIR emission. All these results featured its promising application prospects for palladium sensing in both environment and biology field.

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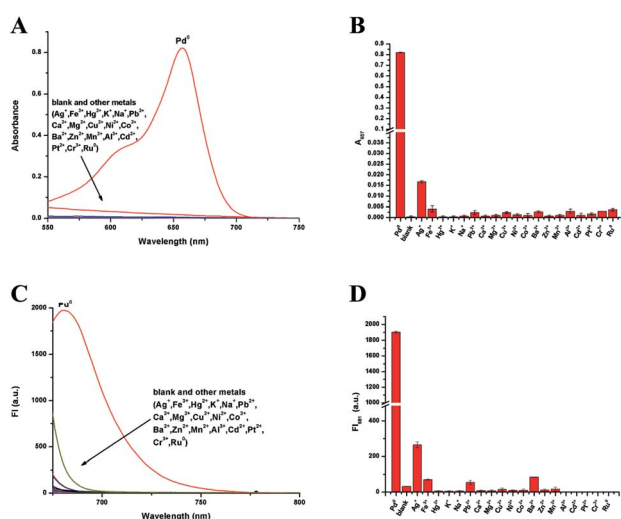


Fig. 3 UV-vis (A) and fluorescent spectra (C), absorbance at 657 nm (B) and fluorescence intensity at 681 nm (D) of MB-APC (10 μM) in the presence of $\text{Pd}(\text{PPh}_3)_4$ (3 equiv.), $\text{Ru}_3(\text{CO})_{12}$ (3 equiv.) and the excess of representative metal ions (10 equiv.) in EtOH-PBS (9 : 1, v/v) at room temperature. Each spectrum was acquired after mixing for 30 min.



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Notes and references

- 1 R. R. Barefoot, *Trends Anal. Chem.*, 1999, **18**, 702–707.
- 2 K. Pyrznska, *J. Environ. Monit.*, 2000, **2**, 99N–103N.
- 3 K. Ravindra, L. Bencs and R. Van Grieken, *Sci. Total Environ.*, 2004, **318**, 1–43.
- 4 R. Merget and G. Rosner, *Sci. Total Environ.*, 2001, **270**, 165–173.
- 5 J. Kielhorn, C. Melber, D. Keller and I. Mangelsdorf, *Int. J. Hyg. Environ. Health*, 2002, **205**, 417–432.
- 6 B. Dimitrova, K. Benkhedda, E. Ivanova and F. Adams, *J. Anal. At. Spectrom.*, 2004, **19**, 1394–1396.
- 7 C. Locatelli, D. Melucci and G. Torsi, *Anal. Bioanal. Chem.*, 2005, **382**, 1567–1573.
- 8 K. Van Meel, A. Smekens, M. Behets, P. Kazandjian and R. Van Grieken, *Anal. Chem.*, 2007, **79**, 6383–6389.
- 9 H. Li, J. Fan and X. Peng, *Chem. Soc. Rev.*, 2013, **42**, 7943–7962.
- 10 M. P. Tracey, D. Pham and K. Koide, *Chem. Soc. Rev.*, 2015, **44**, 4769–4791.
- 11 J.-w. Yan, X.-l. Wang, Q.-f. Tan, P.-f. Yao, J.-h. Tan and L. Zhang, *Analyst*, 2016, **141**, 2376–2379.
- 12 J. Zhang, L. Zhang, Y. Zhou, T. Ma and J. Niu, *Microchim. Acta*, 2013, **180**, 211–217.
- 13 Q. Huang, Y. Zhou, Q. Zhang, E. Wang, Y. Min, H. Qiao, J. Zhang and T. Ma, *Sens. Actuators, B*, 2015, **208**, 22–29.
- 14 L. Yuan, W. Lin, K. Zheng, L. He and W. Huang, *Chem. Soc. Rev.*, 2013, **42**, 622–661.
- 15 Z. Guo, S. Park, J. Yoon and I. Shin, *Chem. Soc. Rev.*, 2014, **43**, 16–29.
- 16 N. Ormeci, B. Savas, S. Coban, M. Palabiyıkoğlu, A. Ensari, I. Kuzu and N. Kursun, *Surg. Endosc.*, 2008, **22**, 693–700.
- 17 J. P. Tardivo, A. Del Giglio, C. S. de Oliveira, D. S. Gabrielli, H. C. Junqueira, D. B. Tada, D. Severino, R. de Fátima Turchiello and M. S. Baptista, *Photodiagn. Photodyn. Ther.*, 2005, **2**, 175–191.
- 18 M. Wagner, E. R. Suarez, T. R. Theodoro, C. D. A. S. Machado Filho, M. F. M. Gama, J. P. Tardivo, F. M. Paschoal and M. A. S. Pinhal, *Clin. Exp. Dermatol.*, 2012, **37**, 527–533.
- 19 Y. Ashitate, B. T. Lee, R. G. Laurence, E. Lunsford, M. Hutteman, R. Oketokoun, H. S. Choi and J. V. Frangioni, *Ann. Plast. Surg.*, 2013, **70**, 360–365.
- 20 A. Matsui, E. Tanaka, H. S. Choi, V. Kianzad, S. Gioux, S. J. Lomnes and J. V. Frangioni, *Surgery*, 2010, **148**, 78–86.
- 21 F. P. R. Verbeek, J. R. van der Vorst, B. E. Schaafsma, R.-J. Swijnenburg, K. N. Gaarenstroom, H. W. Elzevier, C. J. H. van de Velde, J. V. Frangioni and A. L. Vahrmeijer, *J. Urology*, 2013, **190**, 574–579.
- 22 E. Tanaka, F. Y. Chen, R. Flaumenhaft, G. J. Graham, R. G. Laurence and J. V. Frangioni, *J. Thorac. Cardiovasc. Surg.*, 2009, **138**, 133–140.
- 23 J. Bae, L. E. McNamara, M. A. Nael, F. Mahdi, R. J. Doerksen, G. L. Bidwell, N. I. Hammer and S. Jo, *Chem. Commun.*, 2015, **51**, 12787–12790.
- 24 F. Song, A. L. Garner and K. Koide, *J. Am. Chem. Soc.*, 2007, **129**, 12354–12355.

