



CrossMark
 click for updates

Cite this: *RSC Adv.*, 2017, 7, 2573

A novel ratiometric fluorescent probe for selective detection of bisulfite in living cells†

Zhen Chen, Fengzao Chen, Yuanchao Sun, Heng Liu,* Hanping He, Xiuhua Zhang and Shengfu Wang

A new type of ratiometric fluorescent probe, 8-(diethylamino)-2-methyl-4-oxo-4*H*-pyrano[2,3-*b*]chromen-10-ylum tetrafluoroborate (APCT), is reported. A new type of ratiometric fluorescent probe, 8-(diethylamino)-2-methyl-4-oxo-4*H*-pyrano[2,3-*b*]chromen-10-ylum tetrafluoroborate (APCT), is reported. This probe can successfully detect bisulfite anions with a detection limit of 6.1×10^{-7} M based on Michael-type addition reaction in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v). The response time of the probe is approximately 5 min. Upon treating with bisulfite anions, the probe exhibits an obvious blue shift from 560 nm to 510 nm and the fluorescence intensity ratio at 510 nm and 560 nm (I_{510}/I_{560}) displays a good linear relationship with the concentration of HSO_3^- in the range of 20–120 μM . Additionally, the preliminary cell imaging experiments using HeLa cells demonstrate that the probe can be used to detect intracellular bisulfite anions.

Received 25th November 2016
 Accepted 21st December 2016

DOI: 10.1039/c6ra27347e

www.rsc.org/advances

Introduction

As one kind of main environmental air pollutant, sulfur dioxide (SO_2) is mainly derived from human activities such as burning of fossil fuel, paper pulp manufacturing and metal processing, causing mortal threats to human health.¹ SO_2 inhaled by human beings will be hydrated to the corresponding bisulfite (HSO_3^-) and sulfite (SO_3^{2-}) derivatives in the neutral physiological environment.² In daily life, bisulfite and sulfite ($\text{HSO}_3^-/\text{SO}_3^{2-}$) play an important role in food, beverages, and pharmaceutical products, because they can prevent oxidation, and microbial reactions during production and storage.³ Epidemiological studies have indicated that frequent exposure to SO_2 , not only gives rise to a variety of respiratory tract reaction,⁴ but also is related to cardiovascular diseases,⁵ lung cancer and many kinds of neurological disorders.⁶ However, as an important gaseous signal molecule, endogenous SO_2 , which can be produced by sulfur-containing amino acid, can together with nitric oxide (NO) as vasodilator substance to regulate the vascular smooth muscle.⁷ Taking into account the close relationship between SO_2 derivatives and human health, there is urgently needed to develop simple, high selectivity and selectivity method for detecting of $\text{HSO}_3^-/\text{SO}_3^{2-}$ ion and make further efforts to elucidate their detailed biological and pharmacological functions.

Compared with traditional detection method, fluorescent probes and imaging techniques have a number of significant advantages including simple sample pretreatment, rapid response time, high sensitivity and suitable for high-throughput screening application.⁸ Therefore, in recent years, fluorescent probes are increasingly considered as one of efficient tools for monitoring cations, anions and biomolecules.⁹ To date, a large quantity of fluorescent probes capable of selective response to SO_2 and its derivatives have been reported based on three disparate approaches, including aldehydes¹⁰ or Michael-type addition reaction,¹¹ levulinate esters cleavage reaction¹² and coordinative interactions.¹³ However, some of the reported SO_2 -specific fluorescent probes suffer from low selectivity over reactive sulfur especially for biothiols and slow response time, which greatly limits their biological applications. Additionally, some SO_2 -specific fluorescent probes are developed on the basis of fluorescence enhancement or quenching of a single fluorophore and have many limitations such as probe concentration, environmental effects, excitation power and detector sensitivity.¹⁴ In order to get over the limitations, ratiometric fluorescent probes are desirable because they can effectively eliminate external interferences by self-calibration two emission bands.¹⁵ As a consequence, developing novel ratiometric fluorescent probes for detection of $\text{HSO}_3^-/\text{SO}_3^{2-}$ is of great importance.

In this paper, we described the synthesis of a novel ratiometric fluorescent probe (APCT) for $\text{HSO}_3^-/\text{SO}_3^{2-}$ based on Michael-type addition reaction. The probe displayed high selectivity for detection of $\text{HSO}_3^-/\text{SO}_3^{2-}$ over other interfering species with detection limit down to 6.1×10^{-7} M. Meanwhile, upon treatment with HSO_3^- , the fluorescence emission peak of

Hubei Collaborative Innovation Center for Advanced Organic Chemical Materials, Ministry of Education Key Laboratory for the Synthesis and Application of Organic Functional Molecules & College of Chemistry and Chemical Engineering, Hubei University, Wuhan 430062, PR China. E-mail: liuheng11b@hubei.edu.cn

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c6ra27347e



the probe exhibited an obvious blue shift from 560 nm to 510 nm. The sensing mechanism research showed that the conjugate structure of probe-APCT was destroyed. Preliminary cell experiments also confirmed that probe-APCT could be used for intracellular HSO_3^- detection.

Experimental

Chemicals and apparatus

All reagents and solvents were purchased from commercial suppliers and used without further purification. Distilled water was used in the experiment after passing through a water ultra-purification system. Reactions were monitored by TLC. Column chromatographic were carried out using 200–300 mesh silica with the proper solvent according to TLC using UV light to visualize the reaction components. The fluorescence spectra and relative fluorescence intensity were measured by a Shimadzu RF-5301 spectrofluorimeter, the excitation wavelength was 465 nm for all fluorescence measurements, the excitation slit width and the emission slit width set at 3 nm. UV-Vis absorption spectra were measured using a Shimadzu UV-2700 spectrophotometer. ^1H NMR and ^{13}C NMR spectra were recorded on a BRUKER 400 spectrometer. The pH was measured with a PHS-3C acidometer. Electrospray mass spectra (ESI-MS) were recorded on Agilent 1100 Series. Cells images were obtained using CLSM TCS SP5 (Leica, Germany).

Synthesis of APCT

In a 50 mL round-bottom flask, 4-hydroxy-6-methyl-2H-pyran-2-one (1.26 g, 10 mmol) and 4-(diethylamino)-2-hydroxybenzaldehyde (2.32 g, 12 mmol) were dissolved in ethanol (20 mL) and the resulting mixture was stirred at 80 °C for 4 h. After cooling to room temperature, the red solid was collected by suction filtration and then recrystallized from ethanol to afford the desired product C1 as red powder (2.45 g, yield 70%).

Compound C1 (602 mg, 2 mmol) and boron trifluoride ethyl etherate (283.5 mg, 2 mmol) were mixed in component solvent (acetic acid/acetic anhydride = 3 : 1, 9 mL/3 mL, v/v), and the resulting solution was refluxed until completion of the reaction (monitored by TLC). The red solid precipitated upon cooling was collected by suction filtration and then washed with methyl *tert*-butyl ether to afford the target fluorescent probe-APCT (515 mg, yield 60%). ^1H NMR (400 MHz, d_6 -DMSO): δ 8.87 (s, 1H), 7.83 (d, J = 9.3 Hz, 1H), 7.34 (s, 1H), 6.92 (dd, J = 9.2, 2.4 Hz, 1H), 6.67 (d, J = 2.2 Hz, 1H), 3.57 (q, J = 7.1 Hz, 4H), 2.37 (s, 3H), 1.17 (t, J = 7.1 Hz, 6H); ^{13}C NMR (100 MHz, d_6 -DMSO): δ 190.3, 176.3, 158.8, 158.5, 155.1, 149.2, 133.9, 111.5, 109.2, 105.8, 98.9, 96.1, 44.9, 24.3, 12.4; LC-MS TOF: calcd for $\text{C}_{17}\text{H}_{18}\text{BF}_4\text{NO}_3$ [$\text{M} + \text{H}$] $^+$: 371.1; found: 372.1.

General procedure for the spectra measurement

The stock solution of probe (5.0 mM) was prepared in DMSO. Solutions of various testing species (ClO^- , HCO_3^- , CH_3COO^- , PO_4^{3-} , CO_3^{2-} , SO_4^{2-} , HSO_4^- , H_2PO_4^- , NO_2^- , F^- , Cl^- , Br^- , I^- , HPO_4^{2-} , SCN^- , S^{2-} , H_2O_2 , GSH, Cys, Hcy, SO_3^{2-} , HSO_3^-) were

prepared in twice distilled water. All of anions were prepared from their sodium salts. The resulting solution was kept at room temperature (25 °C) and then the absorption or fluorescence spectra were recorded.

Cell culture and imaging

HeLa cells were cultured in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% (v/v) fetal bovine serum (FBS), 100 units per mL of penicillin and 100 units per mL of streptomycin at 37 °C in a CO_2 incubator, and culture media were replaced with fresh media every day. The HeLa cells were pre-treated with 100 μM HSO_3^- , and then further incubated with 10 μM of probe-APCT for 30 min at 37 °C. In the control experiment, HeLa cells were incubated with 10 μM of probe-APCT for 30 min at 37 °C. The cells were washed three times with PBS buffer before cell fluorescence imaging experiments with confocal laser scanning microscopy.

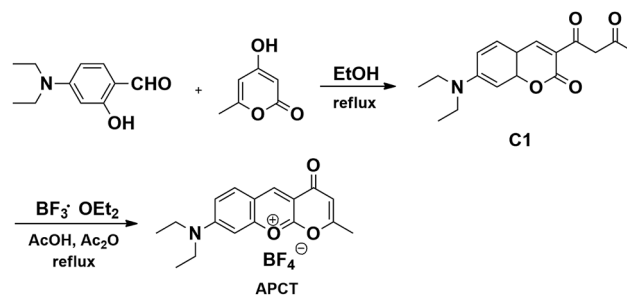
Results and discussion

The synthesis of the target fluorescent probe (APCT) was shown in Scheme 1. According to the designed route, compound C1 was synthesized *via* the condensation of 4-(diethylamino)-2-hydroxybenzaldehyde and 4-hydroxy-6-methyl-2H-pyran-2-one. Followed by the hemiketal, dehydration, aromatization with boron trifluoride diethyl etherate, a new heterocyclic molecule APCT was prepared.

Spectroscopic properties

For practical applications, a short response time was an important parameter for the target probe. Fig. 1 displayed time-dependent fluorescence emission spectra of probe-APCT in the presence of HSO_3^- . It can be clearly seen that stable fluorescence signal of the probe at 510 nm and 560 nm was obtained in approximately 5 min. In view of time-dependent fluorescence emission characteristics of the probe, the following fluorescence measurements were performed about 5 min after addition of HSO_3^- .

With the probe in hand, we first carried out the UV-Vis and fluorescence titration experiment in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v). As demonstrated in Fig. 2A, upon increasing of HSO_3^- amount (6 equiv.), the peak at 510 nm gradually decreased and the low-energy region of 300–



Scheme 1 Synthesis of the target fluorescent probe-APCT.



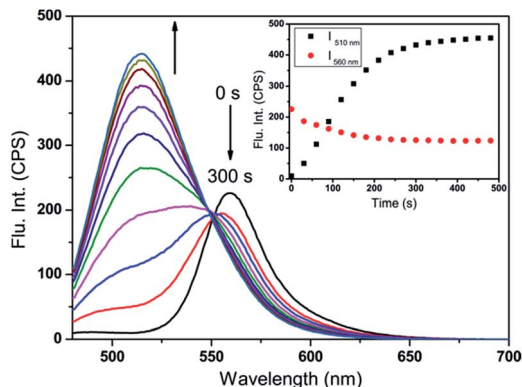


Fig. 1 The time-dependent fluorescence emission spectra ($\lambda_{\text{ex}} = 465$ nm) of probe-APCT upon addition of $120 \mu\text{M}$ HSO_3^- in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v) at 25°C . Slit width: $d_{\text{ex}} = d_{\text{em}} = 3$ nm.

375 nm increased accompanied by the emergence of a well-defined isosbestic point at 445 nm, as well as a noticeable color change from orange-red to yellow. The appearance of new isosbestic point suggested that the interaction of HSO_3^- and the probe was one-to-one conversion to form a new compound. Subsequently, upon the addition of HSO_3^- , the marked fluorescence enhancement of the peak at 510 nm at the expense of the peak at 560 nm was observed in Fig. 2B. The color of the solution gradually turned from yellow to green under a 365 nm UV lamp. Fluorescence analysis revealed that the fluorescence quantum yield changed from 0.14 to 0.06 (using FITC as a reference, $\Phi_{\text{FL}} = 0.92$ in 0.1 M NaOH) before and after the addition of 6 equiv. HSO_3^- . All the results showed that the as-prepared probe could be suitable for colorimetric detection of HSO_3^- by naked-eyes.

Moreover, we could see from the fluorescence titration experiment that the ratiometric responses (I_{510}/I_{560}) of probe-APCT kept a balance and unchanged after addition of 6 equiv. HSO_3^- (Fig. S1†). As shown in Fig. 3, a good linear relationship between I_{510}/I_{560} and the concentration of HSO_3^- in the range of 20–120 μM ($R^2 = 0.983$) was displayed. The detection limit ($S/N = 3$) of 6.1×10^{-7} M of the probe was determined according to the fluorescence titration results, which was comparable to most of the reported SO_2 -specific fluorescent probes (Table S1†).

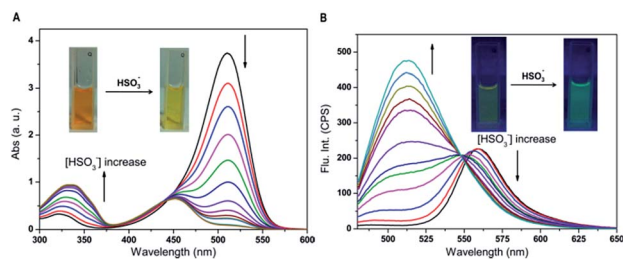


Fig. 2 UV-Vis absorption (A) and fluorescence emission spectra ($\lambda_{\text{ex}} = 465$ nm) (B) of probe-APCT ($20 \mu\text{M}$) upon gradual addition of HSO_3^- ($120 \mu\text{M}$) in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v) at 25°C . Slit width: $d_{\text{ex}} = d_{\text{em}} = 3$ nm.

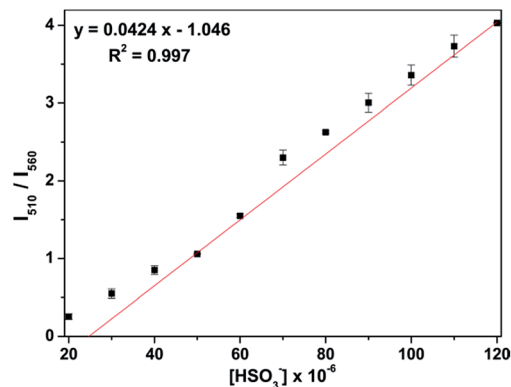


Fig. 3 The ratios of fluorescence intensities (I_{510}/I_{560}) of probe-APCT ($20 \mu\text{M}$) as a function of the concentration of HSO_3^- from $20 \mu\text{M}$ to $120 \mu\text{M}$ in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v) at 25°C . $\lambda_{\text{ex}} = 465$ nm, slit width: $d_{\text{ex}} = d_{\text{em}} = 3$ nm.

The selectivity

In order to evaluate the selectivity of probe-APCT toward HSO_3^- , a series of common physiologically important species including ClO^- , HCO_3^- , CH_3COO^- , PO_4^{3-} , CO_3^{2-} , SO_4^{2-} , HSO_4^- , H_2PO_4^- , NO_2^- , F^- , Cl^- , Br^- , I^- , HPO_4^{2-} , SCN^- , S^{2-} , H_2O_2 , GSH, Cys, Hcy, were added to the solution of the probe in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v) (Fig. S2†). As illustrated in Fig. S3,† upon addition of HSO_3^- , changes of fluorescence intensity ratio (I_{510}/I_{560}) reached up to 35-fold compared to the free probe. In the same test condition, the addition of interfering species only caused a negligible increase in fluorescence intensity ratio (I_{510}/I_{560}) except a moderate value was detected for SH^- . Furthermore, the competitive experiment also carried out in the presence of 6 equiv. HSO_3^- and 6 equiv. interfering species. It can be clearly seen from Fig. 4 that the ratiometric fluorescence signal response of the probe almost unaffected by interfering species. The above results suggested that the as-prepared probe could highly selective recognize $\text{HSO}_3^-/\text{SO}_3^{2-}$ over other interfering species.

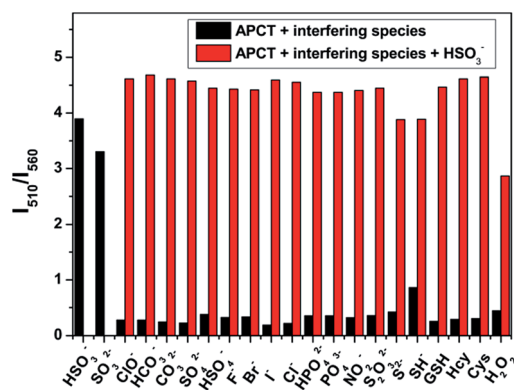


Fig. 4 The selectivity of probe-APCT for HSO_3^- . Fluorescence intensity ratio (I_{510}/I_{560}) of probe-APCT ($20 \mu\text{M}$) toward 6 equiv. HSO_3^- and 6 equiv. interfering species after incubation in pH = 7.4 Tris buffer solution containing 70% DMSO (v/v) at 25°C . $\lambda_{\text{ex}} = 465$ nm, slit width: $d_{\text{ex}} = d_{\text{em}} = 3$ nm.



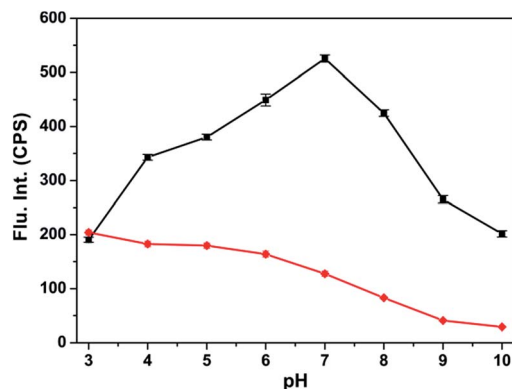


Fig. 5 The fluorescence intensities at 560 nm and 510 nm of probe-APCT (20 μM) in the presence of 6 equiv. HSO_3^- at different pH values.

pH effect

The effects of pH on the fluorescence intensities of probe-APCT were investigated at different pH values. We first evaluated the stability of the probe at pH from 3.0 to 10.0. As can be seen from Fig. S4,[†] there was virtually no change in the fluorescence intensities at 560 nm and 510 nm in the range of pH 3.0–7.0. The fluorescence intensity at 560 nm reduced dramatically under alkaline conditions, probably due to the nucleophilic addition of OH^- . In the presence of 6 equiv. HSO_3^- , the fluorescence intensity at 510 nm appeared remarkable enhancement and achieved maximum at pH 7.4 (Fig. 5). Overall, probe-APCT had the capability to be used for detection of HSO_3^- in physiological environment.

Sensing mechanism

To gain further insight into the reaction mechanism of probe-APCT and HSO_3^- , ^1H NMR titration experiment was carried out. As shown in Fig. 6B, upon addition of excessive HSO_3^- , the NMR signal of H_a at 8.87 ppm up-field shifted to 4.98 ppm (H'_a) and other signals of hydrogen were also shifted to high-field with small shift. Such obvious up-field shift of H_a suggested that a typical nucleophilic addition have been taken place. Another strong evidence of reaction mechanism was from the

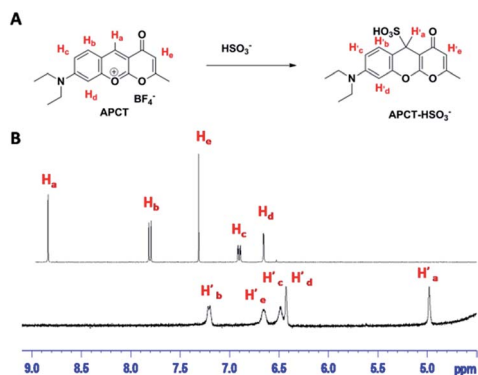


Fig. 6 The mechanism (A) and ^1H NMR titration (B) of the reaction between APCT and HSO_3^- in $\text{d}_6\text{-DMSO}/\text{D}_2\text{O} = 9/1$ (v/v).

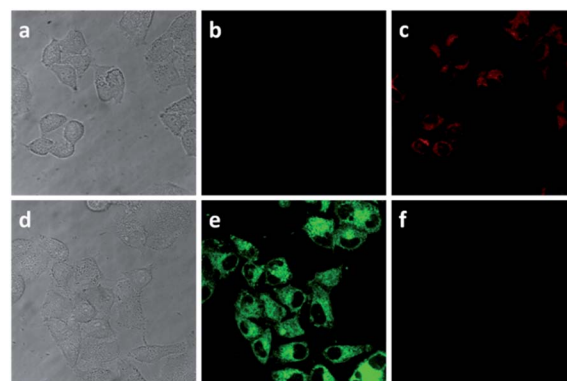


Fig. 7 The top: CLSM fluorescence images ($\lambda_{\text{ex}} = 405$ nm) of HeLa cells incubated with probe-APCT for 30 min, (a) bright field; (b) green channel (475–510 nm); (c) red channel (570–625 nm). The bottom: CLSM fluorescence images of HeLa cells pre-treated with HSO_3^- , and then incubated with probe-APCT for 30 min, (d) bright field; (e) green channel; (f) red channel.

data of LC-MS in Fig. S5.[†] Specifically, probe-APCT displayed a main peak at 372.0 before addition of sodium bisulfate, originating from the species $[\text{APCT} + \text{H}]^+$. Upon addition of excessive sodium bisulfate, a new peak at 365.1 matched well with the species $[\text{APCT} + \text{SO}_3^{2-} + \text{H}]^+$. Therefore, all these experiment results of ^1H NMR and LC-MS indicated that the sensing mechanism of the as-prepared probe-APCT and sodium bisulfate most likely followed the Michael-type addition reaction as shown in Fig. 6A.

Cellular imaging

Encouraged by the above experiment results, we further investigated the practicability of the probe for intracellular HSO_3^- . Firstly, the cytotoxicity of the probe was investigated by a standard CCK-8 assay.¹⁶ The experiment results suggested that the probe exhibited low toxicity on HeLa cells (Fig. S6[†]). Finally, the applicability of probe-APCT for detection of HSO_3^- in living cells *via* confocal laser scanning microscopy (CLSM) was confirmed. The fluorescence images data of HeLa cells incubated with probe-APCT was obtained in the absence and presence of HSO_3^- (Fig. 7). In a typical imaging experiment, HeLa cells incubated with 10 μM free probe emitted moderate red fluorescence and faint green fluorescence with exciting of 405 nm laser, suggesting that the as-prepared probe have a good cell permeability and suitable for intracellular HSO_3^- sensing. When HeLa cells was pre-treated with 100 μM HSO_3^- , and then further incubated with 10 μM probe-APCT for 30 min, green fluorescence emission markedly enhanced and red fluorescence visibly diminished at the excitation wavelength of 405 nm. The preliminary cell experiment results demonstrated the usefulness of probe-APCT in detecting intracellular HSO_3^- .

Conclusions

In conclusion, we have developed a new type of ratiometric fluorescent probe, namely APCT, for HSO_3^- *via* the Michael-type



addition reaction. The probe can discriminate HSO_3^- from other interfering species with high selectivity and sensitivity. The detection limit of the probe for HSO_3^- is 6.1×10^{-7} M. Additionally, in live cell imaging experiment, the probe give rapid fluorescence changes in red and green channels, revealing the capability of the probe for intracellular HSO_3^- sensing.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (NSFC.21602051), the Natural Science Foundation of Hubei Province (2016CFB200) and the Key Project of the Education Department of Hubei Province (No. Q2015010).

References

- 1 X. Shi, *J. Inorg. Biochem.*, 1994, **56**, 155.
- 2 Z. Meng, G. Qin, B. Zhang and J. Bai, *Mutagenesis*, 2004, **19**, 465.
- 3 (a) R. F. McFeeters, *J. Food Prot.*, 1998, **61**, 885; (b) T. Fazio and C. R. Warner, *Food Addit. Contam.*, 1990, **7**, 433; (c) S. L. Taylor, N. A. Higley and R. K. Bush, in *Advances in Food Research*, ed. E. M. M. C. O. Chichester and B. S. Schweigert, Academic Press, 1986, vol. 30, pp. 1–76.
- 4 S. Iwasawa, Y. Kikuchi, Y. Nishiwaki, M. Nakano, T. Michikawa, T. Tsuboi, S. Tanaka, T. Uemura, A. Ishigami, H. Nakashima, T. Takebayashi, M. Adachi, A. Morikawa, K. Maruyama, S. Kudo, I. Uchiyama and K. Omae, *J. Occup. Health*, 2009, **51**, 38.
- 5 G. Li and N. Sang, *Ecotoxicol. Environ. Saf.*, 2009, **72**, 236.
- 6 N. Sang, Y. Yun, H. Li, L. Hou, M. Han and G. Li, *Toxicol. Sci.*, 2010, **114**, 226–236.
- 7 (a) M. H. Stipanuk and I. Ueki, *J. Inherited Metab. Dis.*, 2011, **34**, 17; (b) N. D. Mathew, D. I. Schlipalius and P. R. Ebert, *J. Toxicol.*, 2011, **2011**, 14; (c) J. Li, R. Li and Z. Meng, *Eur. J. Pharmacol.*, 2010, **645**, 143.
- 8 (a) Z. Xu and L. Xu, *Chem. Commun.*, 2016, **52**, 1094; (b) S. Christoph and M. B. Sergey, *Methods Appl. Fluoresc.*, 2015, **3**, 042005.
- 9 (a) M. H. Lee, J. S. Kim and J. L. Sessler, *Chem. Soc. Rev.*, 2015, **44**, 4185; (b) G. R. Hamilton, S. K. Sahoo, S. Kamila, N. Singh, N. Kaur, B. W. Hyland and J. F. Callan, *Chem. Soc. Rev.*, 2015, **44**, 4415.
- 10 (a) X. Liu, Q. Yang, W. Chen, L. Mo, S. Chen, J. Kang and X. Song, *Org. Biomol. Chem.*, 2015, **13**, 8663; (b) C. Yu, M. Luo, F. Zeng and S. Wu, *Anal. Methods*, 2012, **4**, 2638.
- 11 (a) Y. Zhang, L. Guan, H. Yu, Y. Yan, L. Du, Y. Liu, M. Sun, D. Huang and S. Wang, *Anal. Chem.*, 2016, **88**, 4426; (b) J. Yang, K. Li, J.-T. Hou, L. L. Li, C. Y. Lu, Y. M. Xie, X. Wang and X. Q. Yu, *ACS Sens.*, 2016, **1**, 166; (c) J. Xu, J. Pan, X. Jiang, C. Qin, L. Zeng, H. Zhang and J. F. Zhang, *Biosens. Bioelectron.*, 2016, **77**, 725; (d) W. L. Wu, Z. Y. Wang, X. Dai, J. Y. Miao and B. X. Zhao, *Sci. Rep.*, 2016, **6**, 25315; (e) G. Wang, H. Chen, X. Chen and Y. Xie, *RSC Adv.*, 2016, **6**, 18662; (f) H. Li, Q. Yao, J. Fan, C. Hu, F. Xu, J. Du, J. Wang and X. Peng, *Ind. Eng. Chem. Res.*, 2016, **55**, 1477; (g) Q. Zhang, Y. Zhang, S. Ding, H. Zhang and G. Feng, *Sens. Actuators, B*, 2015, **211**, 377; (h) H. Zhang, Z. Huang and G. Feng, *Anal. Chim. Acta*, 2016, **920**, 72.
- 12 (a) H. Zhang, S. Xue and G. Feng, *Sens. Actuators, B*, 2016, **231**, 752; (b) C. Liu, H. Wu, W. Yang and X. Zhang, *Anal. Sci.*, 2014, **30**, 589; (c) P. Hou, S. Chen, K. Voitchovsky and X. Song, *Luminescence*, 2014, **29**, 749; (d) X. Ma, C. Liu, Q. Shan, G. Wei, D. Wei and Y. Du, *Sens. Actuators, B*, 2013, **188**, 1196; (e) S. Chen, P. Hou, J. Wang and X. Song, *RSC Adv.*, 2012, **2**, 10869; (f) X. Gu, C. Liu, Y.-C. Zhu and Y. Z. Zhu, *J. Agric. Food Chem.*, 2011, **59**, 11935; (g) M. G. Choi, J. Hwang, S. Eor and S. K. Chang, *Org. Lett.*, 2010, **12**, 5624.
- 13 (a) C. Wang, S. Feng, L. Wu, S. Yan, C. Zhong, P. Guo, R. Huang, X. Weng and X. Zhou, *Sens. Actuators, B*, 2014, **190**, 792; (b) Y. Sun, C. Zhong, R. Gong, H. Mu and E. Fu, *J. Org. Chem.*, 2009, **74**, 7943.
- 14 R. Y. Tsien and M. Poenie, *Trends Biochem. Sci.*, 1986, **11**, 450.
- 15 (a) L. Yuan, W. Lin, K. Zheng, L. He and W. Huang, *Chem. Soc. Rev.*, 2013, **42**, 622; (b) J. Fan, M. Hu, P. Zhan and X. Peng, *Chem. Soc. Rev.*, 2013, **42**, 29; (c) G. Chen, F. Song, X. Xiong and X. Peng, *Ind. Eng. Chem. Res.*, 2013, **52**, 1122.
- 16 Y. Lv, P. Liu, H. Ding, Y. Wu, Y. Yan, H. Liu, X. Wang, F. Huang, Y. Zhao and Z. Tian, *ACS Appl. Mater. Interfaces*, 2015, **7**, 20640–20648.

