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Hyperbranched polyglycerol conjugated fluorescent carbon dots with improved *in vitro* toxicity and red blood cell compatibility for bioimaging

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Fluorescent carbon dots (CDs) have a variety of biomedical applications such as bio-imaging. Successful application of CDs in clinic greatly depends on their biocompatibility and favorable bio-functions. In the present study, we report a facile and efficient method to synthesize a CDs-based nanohybrid bearing a multi-hydroxy hyperbranched polyglycerol (HPG) shell by combing a "grafting from" method with an anionic ring-opening polymerization technique. The resultant CDs-*g*-HPG had high water-dispersibility, low cytotoxicity, improved hemocompatibility and strong green fluorescence. The results demonstrate that the CDs-*g*-HPG possesses promising potential for bio-imaging.

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1 Introduction

Fluorescent carbon dots (CDs) as a new type of fluorescence nanomaterial, have shown promising potential in biomedical applications such as bio-imaging, disease diagnosis, and nanoprobe.^{1,2} However, bio-safety concerns of CDs for biomedical applications were pointed out in previous literature. It is necessary to modify CDs to improve the biocompatibility and preferably increase more surface functional groups on CDs surface for further functionalization. Recently, a number of materials have been used to functionalize CDs by chemical covalent bonds or physical interactions, such as Fe₃O₄@SiO₂,³ ethylenediamine,⁴ L-tyrosine methyl ester,⁵ hyaluronic acid,⁶ PEG200N,⁷ polyethylenimine,^{8,9} cellulose nanofibrils,¹⁰ and glucose.¹¹ The fluorescence strength, quantum yield, biocompatibility and/or application range of the functionalized CDs were improved to different extents. However, most of these modification materials have no abundant functional groups or only possess charged functional groups that may cause complicated interactions with biomacromolecules and cells in biomedical applications *in vivo*. Therefore, it is necessary to functionalize the CDs with neutral functional groups like hydroxyl groups.

In this work, we modified CDs with hyperbranched polyglycerol (HPG). HPG is a highly branched hydrophilic polymer with excellent biocompatibility, multifunctionality, good solubility, controllable synthesis and long circulation half-time *in vivo*.^{12–14} Hence, HPG has attracted increasing attention as an

ideal material for functionalization or modification of nanomaterials. A large number of studies reported HPG-grafted nanomaterials such as Fe₃O₄,¹⁵ multiwalled carbon nanotubes,¹⁶ gold,¹⁷ Fe₃O₄@SiO₂ (ref. 18) and gadolinium.¹⁹ Nevertheless, there is no reports on surface modification and functionalization of CDs with HPG. Here, we presented a simple and effective method to prepare HPG-conjugated CDs (CDs-*g*-HPG) with CDs as an inner core and HPG as an outer shell. The advantages of the CDs-*g*-HPG include: (1) HPG with numerous functional groups and three-dimensional structure could further increase functionalization potential of the CDs; (2) the thickness of the HPG shell is controllable; (3) HPG with low toxicity could improve the biocompatibility of CDs. These expected properties of the CDs-*g*-HPG were further examined in this work.

2 Materials and methods

2.1 Materials

α -Cyclodextrin ($\geq 98\%$) was purchased from Sigma-Aldrich (USA). KH₂PO₄, methanol and alcohol was purchased from Guangzhou Chemical Reagent Factory (Guangzhou, China). Quinine sulfate dehydrate, Amberlite 732 cation exchange resin, potassium methoxide and glycidol were purchased from Aladdin (Shanghai, China).

2.2 Synthesis of fluorescent CDs

Fluorescent CDs were synthesized according to our previously reported method,²⁰ and were dried in vacuum oven at 40 °C overnight.

2.3 Synthesis of CDs-*g*-HPG

Polymerization of glycidol was carried out in a dry round bottom flask under an atmosphere of nitrogen and in dark

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unless otherwise stated. Briefly, 340 mg of as-prepared CDs were added to the flask followed by 1 mL of potassium methylate solution. The mixture was stirred and heated to 75 °C until the CDs dissolved. The resulting methanol was removed. The round bottom flask was kept in an oil bath at 95 °C and 17.4 mL glycidol was dropwise added using a syringe pump (1 mL h⁻¹). After completion of the monomer addition, the reaction continued for an additional 2 h with constant stirring, and then the mixture was cooled to room temperature. Methanol (35 mL) was added and the product was passed through an Amberlite 732 cation exchange resin to remove the potassium ions. The resin was removed by vacuum filtration and the methanol was removed by rotary evaporation. Then, the product was dialysed for three days against water by using cellulose acetate dialysis tubing (MWCO 500 g mol⁻¹). Finally, CDs-g-HPG was obtained by freeze drying. The overall synthesis scheme is illustrated in Fig. 1.

2.4 Characterization of the CDs and CDs-g-HPG

The CDs or CDs-g-HPG solution in water was dropped on copper grids. After dried, the CDs or CDs-g-HPG were observed with a JEOL-2100F high-resolution transmission electron microscope (HRTEM). The zeta potentials of the CDs and CDs-g-HPG in aqueous solution were measured using a Malvern Zetasizer Nano ZS apparatus. Fourier transform infrared (FTIR) spectra of the CDs and CDs-g-HPG were obtained by using a Bruke Vertex 70 FTIR spectrometer (KBr disk). ¹H nuclear magnetic resonance (¹H NMR) spectra of the CDs and CDs-g-HPG were conducted on a Bruker Avance 300 MHz spectrometer with D₂O as the solvent. The molecular weights of the CDs and CDs-g-HPG were measured

by gel permeation chromatography (GPC, Viscotek 270 max) with polyethylene oxide as the standard and the mixed solution of 0.2 M sodium nitrate and 0.003 M sodium azide as the eluent at a flow rate of 1 mL min⁻¹. The UV-visible absorption spectra of the CDs and CDs-g-HPG were recorded using a UV-2550 spectrophotometer (Shimadzu Corporation, Japan). The fluorescence emission spectra of the CDs and CDs-g-HPG were recorded on a Hitachi F-7000 fluorescence spectrophotometer (Hitachi High-Technologies Corp., Tokyo, Japan).

2.5 Cell imaging

2.5.1 Cell imaging by confocal laser scanning microscopy (CLSM). A549 cell line was obtained from American Type Culture Collection (ATCC, USA). The cells were seeded into 24-well (1 × 10⁵ cells per well) chamber slides and cultured in DMEM supplemented with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin at 37 °C and 5% CO₂. After overnight growth, 500 μL of the CDs (0.1 mg mL⁻¹) or CDs-g-HPG (10 mg mL⁻¹) solutions in DMEM were added to each wells and incubated for 5 h. Next, the cells were washed with fresh phosphate buffered saline (PBS, pH 7.4) for three times, fixed with 4% formaldehyde for 15 min at room temperature, mounted with glycerol and then observed with a CLSM (LSM 700, Zeiss, Germany).

2.5.2 Cell quantification by flow cytometry. For quantitative analysis, the cellular uptake of the CDs and CDs-g-HPG was measured using a flow cytometry (BD Accuri™ C6). A549 cells were seeded in a 24-well plate (1 × 10⁵ cells per well), and cultured with complete medium overnight. The medium was then replaced

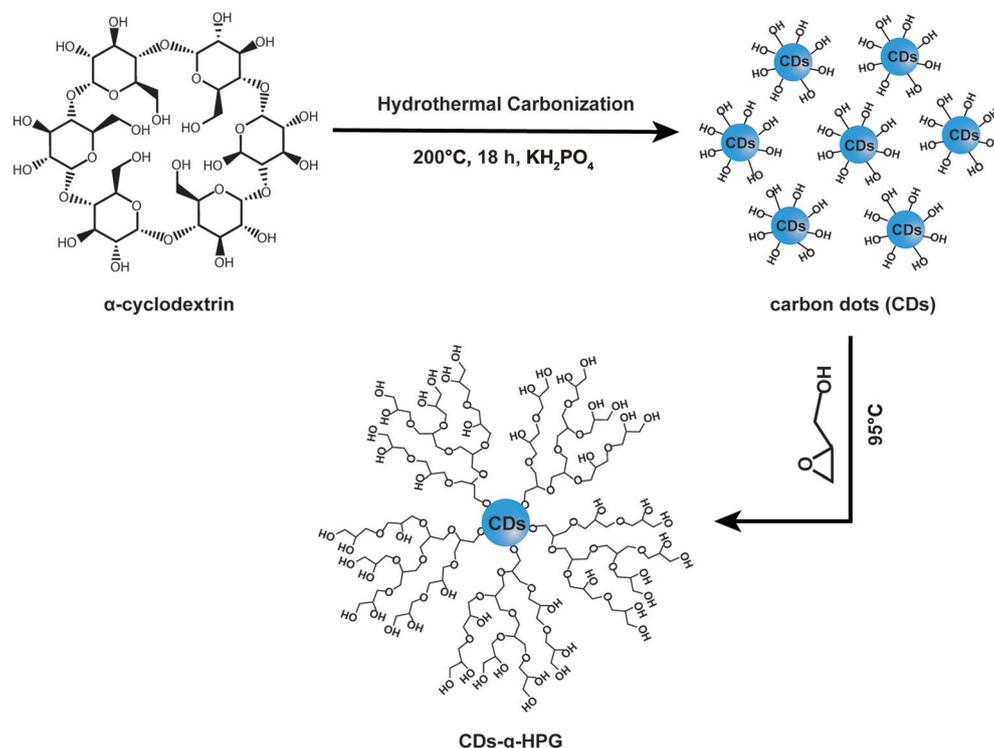


Fig. 1 Synthesis route of CDs-g-HPG.



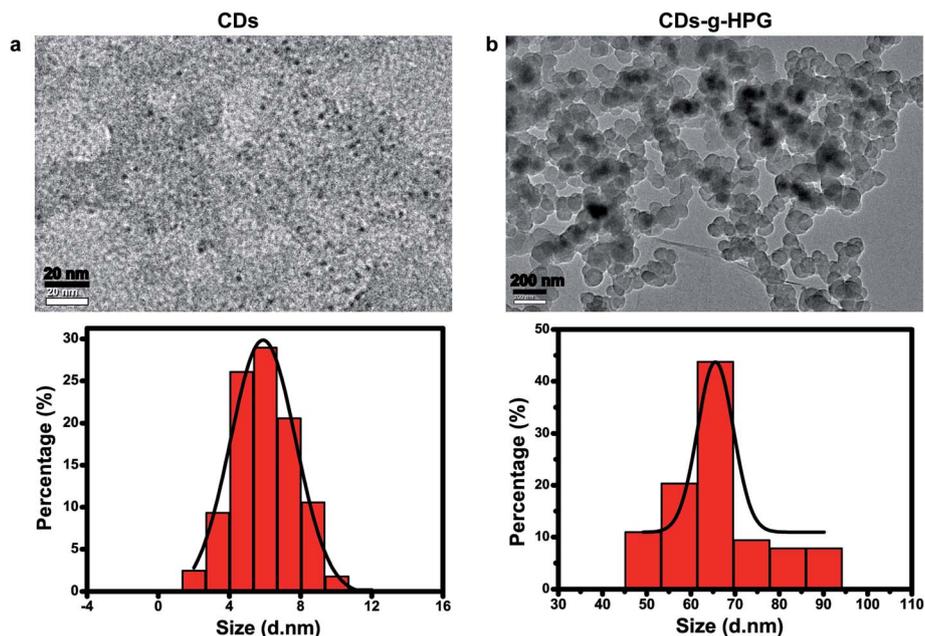


Fig. 2 HRTEM images and size distribution histograms of (a) CDs and (b) CDs-g-HPG.

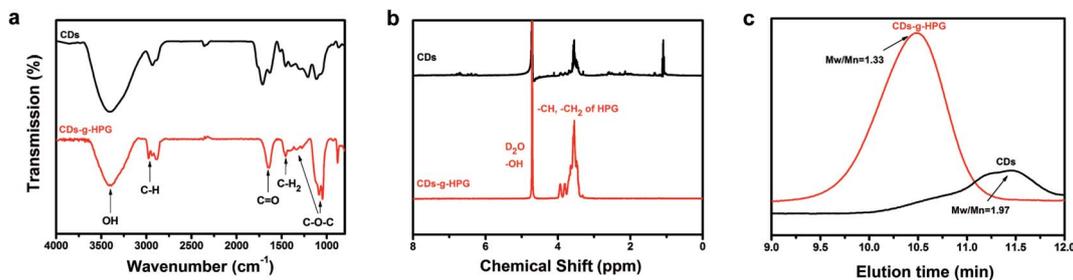


Fig. 3 Characterization of CDs and CDs-g-HPG by (a) FTIR, (b) ^1H NMR, and (c) GPC.

with different concentrations of the CDs and CDs-g-HPG dissolved in DMEM. The media containing the materials were removed after 5 h and the cells were washed with PBS three times, trypsinized, centrifuged (1000 rpm, 5 min), and resuspended with PBS for flow cytometry analysis. A total of 10 000 viable cells were counted and the data were analyzed using FlowJo 7.6.

2.6 Cytotoxicity

A549 cells were seeded into 96-well plates (1×10^4 cells per well) at 37°C for 24 h. Then, the cells were treated with different concentrations of the materials dissolved in DMEM for 24 h. Subsequently, the cells were washed with PBS three times followed by addition of 100 μL of the DMEM plus 10% (v/v) Cell

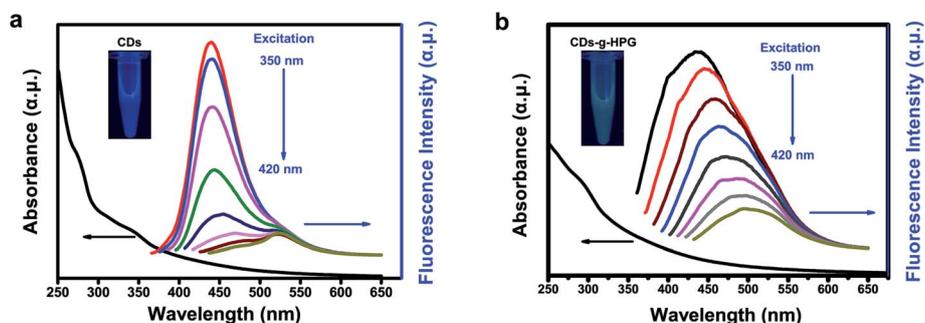


Fig. 4 Optical properties of (a) CDs and (b) CDs-g-HPG. UV-visible absorption spectra (black line) and fluorescence emission spectra (color lines) excited from 350 to 420 nm. Inset: photograph of CDs or CDs-g-HPG aqueous solution under UV light at 365 nm, respectively.



Table 1 Quantum yield of the CDs and CDs-g-HPG

Samples	Integrated PL intensity (<i>I</i>)	Absorbance intensity at 350 nm (<i>A</i>)	Refractive index (ϵ)	Quantum yield (ϕ)
Quinine sulfate	9965	0.065	1.33	0.54 ^a
CDs	384.2	0.089	1.33	0.015
CDs-g-HPG	259.1	0.077	1.33	0.012

^a A known parameter from reference.

Counting Kit8 reagent (CCK8, Dojindo Laboratories) to each well and incubated for 4 h. The absorbance was measured at 450 nm by a microplate reader (Multiskan MK3; Thermo Lab-systems, Finland). The relative cell viability was calculated with the negative control cells as 100% cell viability.

2.7 RBC morphology

Freshly collected citrate-anticoagulated whole blood donated by healthy consenting volunteers was centrifuged at $1000 \times g$ for 10 min to obtain RBC pellets followed by washing with PBS. The RBC pellet was incubated with different concentrations of the CDs or CDs-g-HPG solutions for 10 min at room temperature. The RBCs incubated with PBS were used as a negative control. Subsequently, the RBCs were fixed with 4% formaldehyde for

2 h and then dehydrated with 75, 85, 95 and 100% (v/v) ethanol for 10 min respectively. After air drying, the RBCs were coated with gold and observed with a scanning electron microscope (SEM, Philips XL-30, Holland).

2.8 RBC lysis

Fresh RBC suspension (16% v/v in PBS, 50 μ L) was incubated with 1 mL of different concentrations of the CDs and CDs-g-HPG solutions dissolved in PBS at 37 °C. Fifty microliters of the RBC suspension was added to 1 mL of water to yield complete hemolysis used as a positive control (100% hemolysis). After incubation for a certain time, the RBC suspensions were centrifuged at $1000 \times g$ for 5 min, and the supernatants were collected. The absorbance values of the supernatants (200 μ L) at 540 nm were measured with the microplate reader. The hemolysis of the test samples were calculated by comparing the absorbance values to that of the positive control (100% hemolysis).

2.9 Statistical analysis

Data were expressed as mean values \pm standard deviation (SD). Statistical analysis was performed by using one-way ANOVA with SPSS 16.0. Difference is considered significant when $p < 0.05$ (*) and $P < 0.01$ (**).

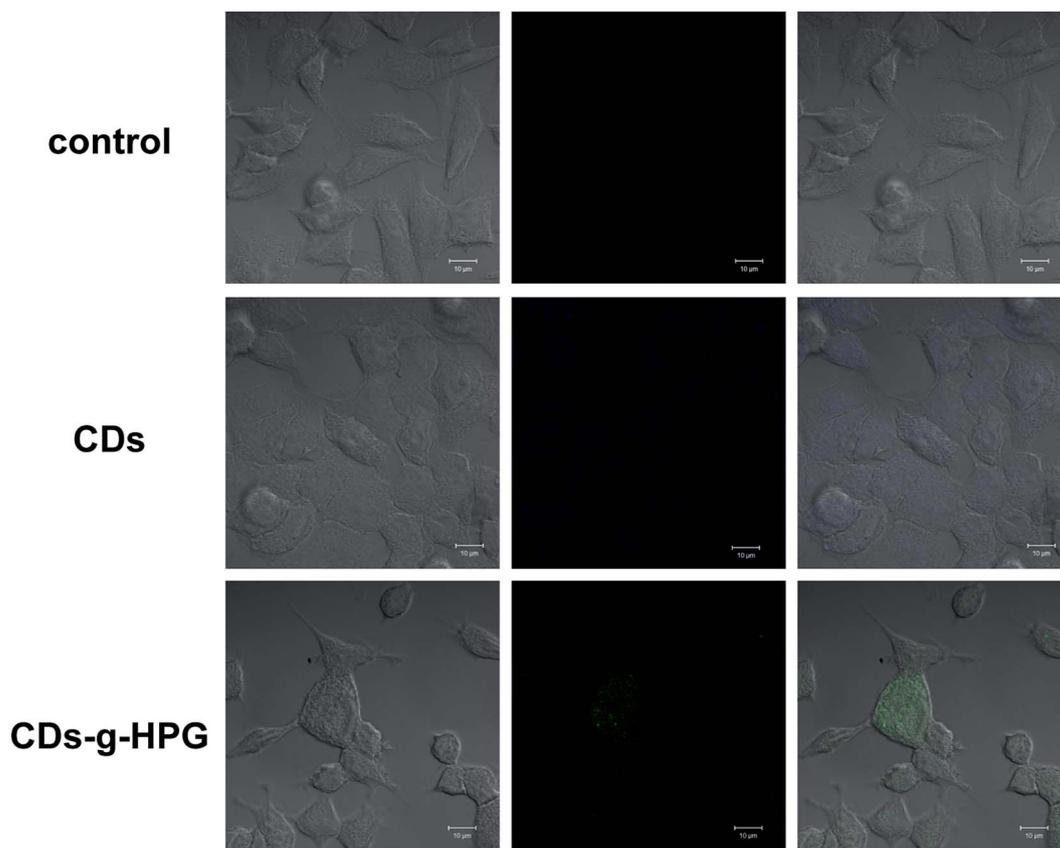


Fig. 5 CLSM images of A549 cells cultured in the presence of CDs (excited at 405 nm) or CDs-g-HPG (excited at 488 nm). Left are bright field images, middle are fluorescence images and right are overlap images. All of the images have a scale ball of 10 μ m.



3 Results and discussion

3.1 Synthesis and characterization of CDs and CDs-g-HPG

Fluorescent CDs were reported to have abundant carboxyl and hydroxyl groups on the surface.²¹ In this work, we prepared the CDs with abundant hydroxyl groups by hydrothermal treatment of α -cyclodextrin. Subsequently, the obtained CDs with hydroxyl groups were used as an initiator to initiate the anionic ring-opening polymerization of glycidol on the surface of the CDs to obtain CDs-g-HPG. The synthesis procedure of the CDs-g-HPG is shown in Fig. 1. The as-prepared CDs and CDs-g-HPG were near spherical in morphology and well-dispersed, as shown in their HRTEM images (Fig. 2a and b). The sizes of the CDs and CDs-g-HPG were 6.8 ± 1.7 nm and 70.8 ± 11.3 nm respectively according to their size distribution histograms (statistical analysis by ImageJ). The zeta potential of the CDs and CDs-g-HPG was -14.6 ± 1.0 mV and -15.0 ± 1.5 mV, respectively. The negative charges of the CDs and CDs-g-HPG nanoparticles were due to the existence of abundant -COOH groups on the surfaces, leading to the good dispersity and water-solubility of the two materials.

The chemical structures and molecular weights of the CDs and CDs-g-HPG were respectively examined by FTIR, ¹H NMR and GPC (Fig. 3). The FTIR spectrum of CDs shows C-H stretching at $2970\text{--}2880$ cm^{-1} , C-H₂ bending at 1460 cm^{-1} , and a strong band at 3411 cm^{-1} referring to surface hydroxyl groups of the CDs. The FTIR spectrum of CDs-g-HPG shows the typical signatures of HPG:^{22,23} stretching vibration of O-H, C-H, and C-O-C at 3405 , 2982 , and $1042\text{--}1089$ cm^{-1} , respectively. The presence of HPG on the surface of the CDs and its chemical structure were further confirmed by ¹H NMR (Fig. 3b). The characteristic proton resonances of -CH₂O- and -CHO- units of HPG in the range of $3.5\text{--}4.0$ ppm were observed, and the solvent peak of the D₂O was appeared at $4.4\text{--}4.7$ ppm.²³ The FTIR and ¹H NMR results indicate that HPG was successfully grafted onto the surface of the CDs. From the GPC result (Fig. 3c), the eluting time was used to compare the difference of the molecular weights of the CDs and CDs-g-HPG. From the eluting times, the molecular weight of CDs-g-HPG was higher than that of the CDs, implying that the HPG was conjugated to the CDs.

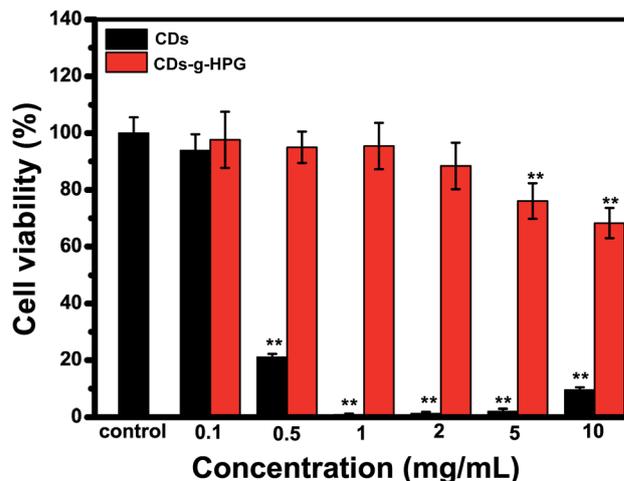


Fig. 7 Cell viability of A549 cells after incubation with CDs or CDs-g-HPG at different concentrations for 24 h.

Optical properties of the CDs and CDs-g-HPG were detected by UV-visible and fluorescence emission spectra excited by changing excitation wavelengths (Fig. 4a and b). The UV-visible absorption spectrum exhibits a peak around 289 nm, which is attributable to the $\pi\text{--}\pi^*$ transition of aromatic sp^2 domains.^{21,24} At an excitation wavelength of 350 nm, a strong emission peak at 435 nm is observed in the fluorescence emission spectra of the CDs or CDs-g-HPG. Along with the excitation wavelength increase from 350 to 420 nm, the fluorescence emission intensity decreased remarkably, and the emission peaks of the CDs shifted from 430 to 520 nm while that of CDs-g-HPG did not shift much with a maxima red shift of ~ 60 nm from 435 to 500 nm. The result indicates that the obtained CDs-g-HPG displayed a strong excitation-dependence of fluorescence emission like CDs. The inset images in Fig. 4 show that the prepared CDs and CDs-g-HPG exhibited excellent water-dispersibility and emitted bright blue (CDs) and green (CDs-g-HPG) fluorescence under UV lamp (365 nm), respectively. The CDs and CDs-g-HPG emitted different fluorescence colors, which is attributed to the poly-aromatic sp^2 carbon nanostructures in the materials as well as the diverse size distribution and surface functional

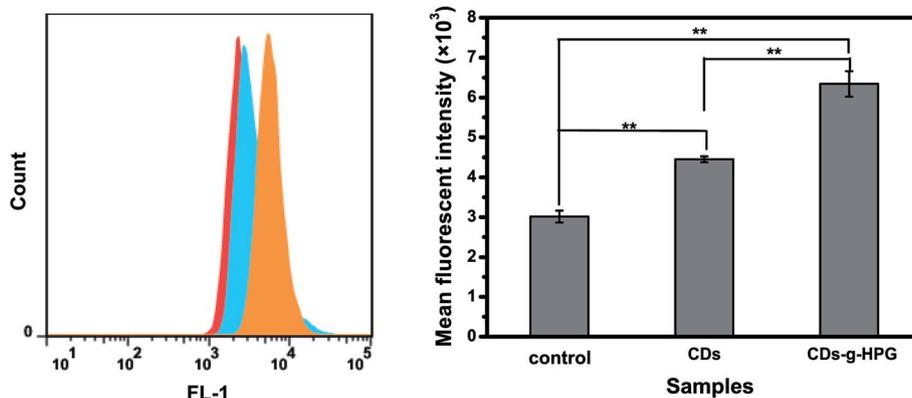


Fig. 6 Flow cytometry analysis of the cellular uptake of the CDs or CDs-g-HPG by A549 cells after cultured for 5 h.



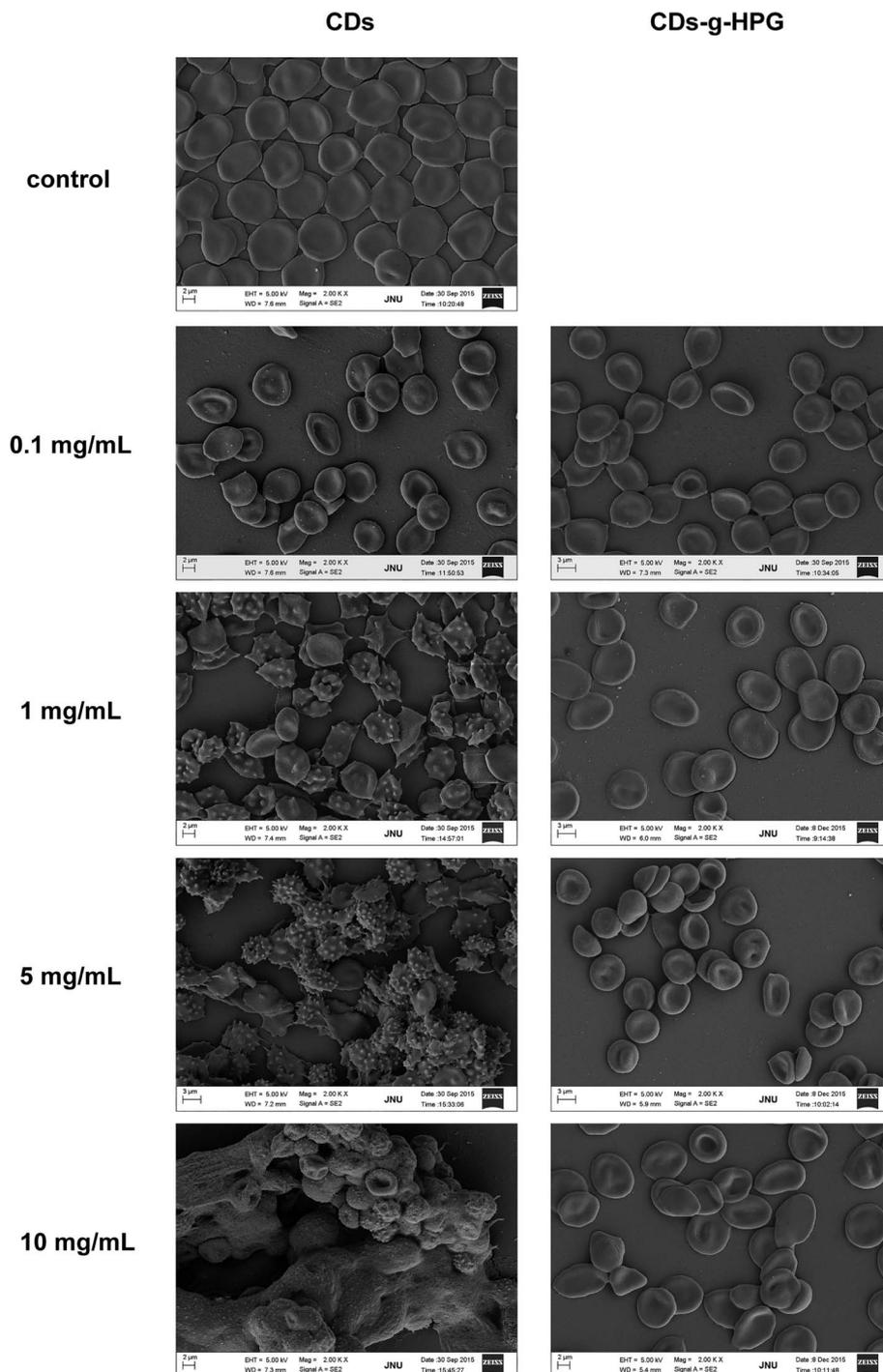


Fig. 8 Morphology and aggregation of the RBCs in the presence of different concentrations of CDs or CDs-g-HPG.

groups.^{1,7} Quinine sulfate (quantum yield 54%)²⁵ was used as a standard sample to calculate the quantum yield of the CDs and CDs-g-HPG (Table 1). The quantum yields were 1.5% for CDs and 1.2% for CDs-g-HPG, respectively. Although the fluorescence emission mechanism of CDs has not been well elucidated, there is increasing evidence that the emission arises from the radioactive recombination of excitations located at the surface energy trap.²⁵ The quantum yield of the CDs-g-HPG was

less than that of the CDs, probably because the HPG modification layer reduced the surface energy trap of the CDs.

3.2 Cell imaging

3.2.1 Cell imaging by CLSM. To ensure the feasibility of the CDs-g-HPG for cell imaging, we firstly examined the ability of the CDs-g-HPG for A549 cells imaging *in vitro*, as presented in Fig. 5. In detail, 10 mg mL⁻¹ of the CDs-g-HPG solution (in



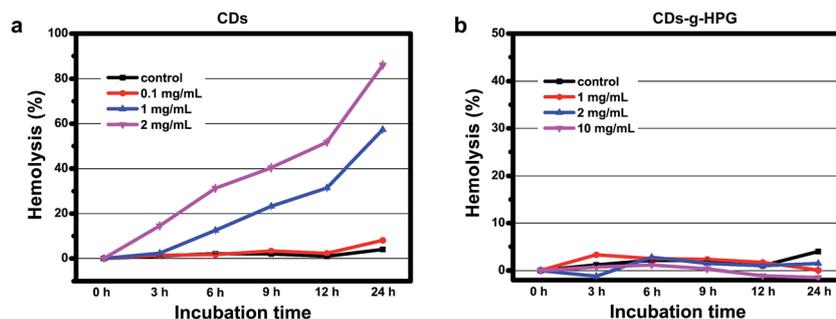


Fig. 9 Lysis of the RBCs after incubated with different concentrations of (a) CDs or (b) CDs-g-HPG at room temperature.

DMEM) was incubated with A549 cells for 5 h, and strong green fluorescence was observed when excited at 488 nm. This indicates that the CDs-g-HPG can be quickly internalized by the cells, confirming the potential application of the CDs-g-HPG as an imaging agent. In addition, 0.1 mg mL⁻¹ of the CDs solution (in DMEM) was incubated with A549 for 5 h, and then blue emission fluorescence was observed when excited at 405 nm. The results suggest that the fluorescence emission of the CDs was adjusted from blue to green after HPG grafting on the surface of the CDs. This phenomenon may be contributed to the change of the size and surface groups of the CDs before and after grafting HPG.

3.2.2 Cell imaging by flow cytometry. The cell imaging ability of the CDs and CDs-g-HPG was further quantitatively analyzed by using flow cytometry. Briefly, A549 cells were incubated with 10 mg mL⁻¹ of CDs-g-HPG or 0.1 mg mL⁻¹ of CDs for 5 h. Then, the fluorescence intensity of the cells was examined by using flow cytometry, as shown in Fig. 6. The result further suggests that CDs or CDs-g-HPG was quickly taken up into A549 cells, which could label the cells.

3.3 Cytotoxicity

The cytotoxicity of the CDs and CDs-g-HPG from 0.1 to 10 mg mL⁻¹ on A549 cells was detected by CCK-8 assay. Cell viability was normalized to control cells (no materials). As shown in Fig. 7, the cell viability was significantly lower than the control when treated with the CDs as low as 0.5 mg mL⁻¹. By contrast, the cell viability had no significant difference from the control when treated with the CDs-g-HPG as high as 2 mg mL⁻¹. This suggests that the CDs-g-HPG had much lower cytotoxicity than the CDs did, indicating that the conjugation with HPG layer improved the bio-safety of the CDs.

3.4 RBC morphology

RBCs are the most abundant blood cells in the blood tissue and have important transport function for oxygen and carbon dioxide. Moreover, RBCs, due to their typical morphology and easy resource, are generally used as a membrane model to study the complicated interactions between foreign materials with mammalian cell membranes.²⁶ RBC morphology is considered as a main criterion to evaluate the hemocompatibility of foreign materials.²⁷ It is unavoidable for the CDs and CDs-g-HPG to

contact RBCs in their blood-contacting biomedical applications. Here, we explored the effect of the CDs and CDs-g-HPG on RBCs morphology and aggregation.

Fig. 8 shows the RBC morphology and aggregation in the presence of the CDs and CDs-g-HPG at different concentrations. It clearly displays that the CDs from 1 mg mL⁻¹ altered RBC morphology from normally biconcave to crenated, and that the CDs at 5 and 10 mg mL⁻¹ even caused RBC aggregation like big lumps. This suggests that the CDs had strong interaction with RBC membrane, which was presumed to be mediated by their hydrophobic interaction.²⁰ By comparison, the CDs-g-HPG as high as 10 mg mL⁻¹ did not cause the morphological change and aggregation of RBCs. It indicates that the hydrophobic interaction between the CDs and RBC membrane was prevented by the HPG layer, and that the hemocompatibility of the CDs was improved after grafted with HPG on their surface.

3.5 RBC lysis

We further evaluated the hemolytic properties of the CDs and CDs-g-HPG, in order to have a better understanding of the extent of the membrane damage after contact with the two materials. Hemolysis is a common index to indicate the degree of the RBC membrane damage by foreign materials. Significant hemolysis is caused when the percentage of hemolysis is above 5%.²⁷ The hemolysis percentages of the RBCs incubated with the CDs and CDs-g-HPG are shown in Fig. 9. The hemolysis by the CDs clearly depended on their concentration and incubation time. Obviously, the CDs at 1 and 2 mg mL⁻¹ gradually caused serious hemolysis along with incubation time. By contrast, even 10 mg mL⁻¹ of CDs-g-HPG did not cause significant hemolysis after 24 h of incubation. This further demonstrates that the HPG layer blocked the hydrophobic interaction between the CDs and RBC membrane.

4 Conclusion

In this work, a fluorescent CDs-g-HPG nanohybrid composed of a CDs core and a HPG shell was synthesized by a surface-initiated anionic ROP technique. The HPG shell not only enhanced the biocompatibility of the CDs but also added numerous favorable surface functional groups (-OH) for further functionalization. Combining their convenient synthesis, unique chemical structure, favorable optical properties, good water-dispersibility and



much improved bio-safety, the CDs-g-HPG has promising applications for bio-imaging, biosensors, etc.

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