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nitrogen heterocycle moiety†

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Design, synthesis and antitubercular evaluation of

benzothiazinones containing an oximido or amino

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A series of 8-nitro-6-(trifluoromethyl)-1,3-benzothiazin-4-ones (BTZs) bearing an oximido or amino nitrogen heterocycle moiety through modifications at the C-2 position of BTZ043 and BPTZ169 were designed and synthesized as new antitubercular agents. Many of the target compounds demonstrate excellent *in vitro* activity (MIC: <0.016–0.088  $\mu$ g mL<sup>-1</sup>) against the drug susceptive H37Rv strain and two clinically isolated multidrug-resistant *Mycobacterium tuberculosis* (MTB) strains. Compound 10a displays acceptable safety, aqueous solubility and pharmacokinetic properties, opening up a new possibility for further development.

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Tuberculosis (TB), one of the world's major causes of illness and death, is caused mainly by *Mycobacterium tuberculosis* (MTB).¹ The World Health Organization (WHO) estimated that approximately one third of the world population is infected with MTB, and 9.6 million people were infected and 1.5 million died from TB worldwide in 2014.² In particular, the spread of multidrugresistant (MDR) TB and the emergence of extensively drugresistant (XDR) TB have revitalized drug discovery efforts in search of novel drugs recently.³-5 It is encouraging that bedaquiline and delamanid have been approved for the treatment of MDR-TB, after a huge gap of over 40 years.⁵,7 However, there are only two new chemical entities, Q203 and TBA-354, in phase 1 clinical trials at present. Therefore, it is urgent to develop new anti-TB drugs.⁵,9

8-Nitro-6-(trifluoromethyl)-1,3-benzothiazin-4-ones (BTZs), a novel class of TB agents targeting DprE1, $^{10-12}$  were reported to have potent activity in multiple models. $^{13}$  BTZ043 (Fig. 1) and PBTZ169 (Fig. 1) are being studied preclinically. Compared with BTZ043, PBTZ169 is slightly more potent and not being stereoselective which makes it easier and cheaper to synthesize. $^{14}$  The structure–activity relationship (SAR) studies of BTZs show that  $-\text{CF}_3$  and  $-\text{NO}_2$  are the optimal groups at the C-6 and C-8

positions, respectively, so there is only one possible structural modifications at the C-2 position.

On the other hand, some of fluoroquinolones (FQs), a class of important second-line anti-TB drugs, such as ciprofloxacin, ofloxacin and levofloxacin, are frequently used for management of TB including MDR-TB.<sup>15</sup> Pyrrolidinyl, piperazinyl and piperidinyl are the most common groups at the C-7 position of FQs. It is interesting that recently the discovery of gemifloxacin and IMB-070593 highlights the importance of oxime-functionalized nitrogen heterocycles with respect to antibacterial activity.<sup>16-18</sup>

Inspired by the above research results, we intended to replace the spiroketal moiety of BTZ043 with nitrogen cycloketone oximes and shift the terminal nitrogen on the piperazine ring of BPTZ169 outside the ring (Fig. 1), which were expected to explore SAR of BTZs though variations on the sizes of the heterocycle and the alkyl group of the oxime. Thus, a series of novel BTZ derivatives containing an oximido or amino nitrogen heterocycle moiety were designed and synthesized in this study. Our primary objective was to identify alternative groups at the

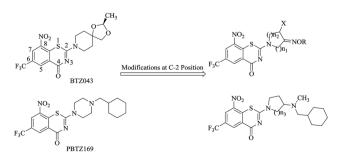


Fig. 1 Design of the new molecules.

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Scheme 1 Synthesis of target compounds 10a-i and 11a, 11b. Reagents and conditions: (i) RONH₂·HCl, K₂CO₃, C₂H₅OH, rt, 40-65%; (ii) TFA, CH<sub>2</sub>Cl<sub>2</sub>, rt, 50–52%; (iii) c-C<sub>6</sub>H<sub>11</sub>CH<sub>2</sub>Br, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C, 50–52%; (iv) SOCl<sub>2</sub>, NH<sub>3</sub>·H<sub>2</sub>O, reflux, 76%; (v) CS<sub>2</sub>, CH<sub>3</sub>I, DMSO, rt, 41%; (vi) (C<sub>2</sub>H<sub>5</sub>)<sub>3</sub>N, C<sub>2</sub>H<sub>5</sub>OH, 60 °C, 45-59%.

C-2 position of BTZs with potent antimycobacterial activity and facilitate the further development of BTZs.

Detailed synthetic pathways to novel BTZ derivatives 10a-i and 11a, 11b are outlined in Scheme 1. 4-7-Membered nitrogen cycloketones 1a-i were smoothly converted to the oximes 2a by reaction with O-alkylhydroxyamines in ethanol at room temperature. The N-Boc protecting group on 2a-i was removed with trifluoroacetic acid (TFA) in dichloromethane to afford the nitrogen cycloketone oximes 3a-i in 40-65% yield.

Compounds 6a, 6b were easily prepared from the corresponding tert-butyl 3-(methylamino) pyrrolidine/piperidine-1carboxylate 4a, 4b via nucleophilic substitution reaction with (bromomethyl)cyclohexane in the presence of K2CO3 in dimethyl formamide (DMF) at 80 °C, and then the resulting cyclohexylmethylates 2a, 2b were treated with TFA.

Amidation of 2-chloro-3-nitro-5-(trifluoromethyl) benzoic acid 7 gave amide 8, which was condensated with carbon disulfide and methyl iodide yielded BTZ core compound 9.10 Derivatives 10a-i and 11a, 11b were conveniently obtained from 9 by nucleophilic substitution with side chain compounds 3a-i and 6a, 6b, respectively.

The synthesized compounds 10a-i and 11a, 11b were preliminarily screened for in vitro activity against MTB H37Rv ATCC 27294 strain, using the Microplate Alamar Blue Assay (MABA).<sup>19</sup> The minimum inhibitory concentration (MIC) is defined as the lowest concentration effecting a reduction in fluorescence of >90% relative to the mean of replicate bacterium-only controls. The minimum inhibitory concentration (MIC) values of the compounds along with PBTZ169, isoniazid (INH) and rifampicin (RFP) for comparison are presented in Table 1.

The data reveal that with a few exceptions (10d, 10f, 10i), all of the BTZ derivatives have potent in vitro activity against this strain (MIC:  $<0.1 \,\mu\mathrm{g \, mL^{-1}}$ ), which is better than **BPTZ169** (MIC: 0.116  $\mu g \text{ mL}^{-1}$ ). In particular, the most active compounds 10b, **10c** and **10g** (MIC:  $<0.016 \mu g \text{ mL}^{-1}$ ) were found to be >3->7 fold

more potent than BPTZ169, INH and RFP (MIC: 0.049-0.116 μg mL<sup>-1</sup>). The potency of the oxime BTZ derivatives (10a-h) in this study is related to both of the sizes of the heterocycle and alkyl group (R). Generally, 4- and 6-membered heterocycles show the best activity, followed by 7- and 5-membered ones with the same R group ( $CH_3$ ) successively (10a vs. 10d vs. 10e vs. 10h). Moreover, the contribution of the alkyl groups of the oxime moiety to the activity is relevant to the heterocycles. The activity of the R groups is as follows: benzyl = ethyl > methyl for azetidyl-based BTZs (10a vs. 10b vs. 10c), and but benzyl > methyl \( \rightarrow \) ethyl for piperidinyl-based ones (10e vs. 10f vs. 10g). Further investigation also suggests that introduction of an amino group on the heterocycles is detrimental. For instance, 3amino-4-(methoxyimino)piperidin-1-yl derivative (10i) displays MIC value of 0.436 μg mL<sup>-1</sup> which is 15-fold less potent than **10e** (MIC:  $0.029 \mu g \text{ mL}^{-1}$ ).

On the other hand, when the nitrogen atom of the piperazine ring of PBTZ169 was converted to an exocyclic tertiary amino group, the resulting compound 11a demonstrates increased activity (MIC: 0.051 µg mL<sup>-1</sup>). It is clear that the piperidin-1-yl group (11a) could be displaced by a pyrrolidin-1-yl moiety (11b) without obviously affecting the potency. These results indicate that 4-aminopiperidine and 3-aminopyrrolidine rings are preferred over piperazine.

The compounds 10a-i and 11a, 11b were examined for toxicity (CC<sub>50</sub>) in human lung adenoma A549 cell lines by MTT assay and the results are reported in Table 1. All of them (CC<sub>50</sub>: 57.66–707.33 μg mL<sup>-1</sup>) are more cytotoxic than **PBTZ169** (CC<sub>50</sub>: 784.20  $\mu$ g mL<sup>-1</sup>), but the selectivity index (SI: 7667–>44 208) of compounds 10b, 10c, 10g, 11a and 11b are bigger than PBTZ169 (SI: 6760) for MTB H37Rv ATCC 27294.

Encouraged by their strong potency against the drugsensitive MTB H37Rv strain, these BTZ derivatives except compounds 10d, 10f and 10i were further evaluated against two clinical isolated MDR-MTB strains 16892 (resistant to both of INH and RFP) and 16802 (resistant to INH, RFP,

Table 1 Structures and *in vitro* activity of **10a–i** and **11a**, **11b** against MTB H37Rv ATCC 2729<sup>a</sup>

$NO_2$	
s <sub>~</sub>	J
F <sub>3</sub> C N	
, II	

	0				
Compd	W	MIC (μg mL <sup>-1</sup> )	CC <sub>50</sub> <sup>b</sup> (μg mL <sup>-1</sup> )	SI <sup>c</sup>	
10a	-{-NOCH₃	0.030	$91.37 \pm 35.18$	3046	
10b	$-\xi$ -NOC <sub>2</sub> H <sub>5</sub>	<0.016	707.33 $\pm$ 133.77	>44 208	
10c	-ξ-N NOBn	<0.016	$157.74 \pm 66.57$	>9858	
10d	NOCH <sub>3</sub>	0.116	$57.66 \pm 11.34$	497	
10e	-{-NOCH₃	0.029	$102.07 \pm 7.13$	3520	
10f	-{-N-NOC <sub>2</sub> H <sub>5</sub>	0.232	$191.29 \pm 33.94$	825	
10g	-ξ-N NOBn	<0.016	$440.69 \pm 102.73$	>27 543	
10h	NOCH <sub>3</sub>	0.060	$58.30\pm3.78$	972	
10i	$-\xi-N \longrightarrow NH_2$ $-NOCH_3$	0.436	$241.49 \pm 47.5$	554	
11a	-{-N_CH <sub>3</sub>	0.051	$530.97 \pm 180.70$	10 411	
11b	$\sum_{\mathbf{Z}_{2}^{N}} - \sum_{\mathbf{CH}_{3}}$	0.088	$67.46 \pm 19.96$	7667	
PBTZ169	-{-N_N-	0.116	$784.20 \pm 185.8$	6760	
INH RFP		0.050 0.049			

 $<sup>^</sup>a$  MTB H37Rv ATCC 2729 was acquired from ATCC.  $^b$  CC<sub>50</sub>: 50% cytotoxic concentration.  $^c$  SI: selectivity index for MTB H37Rv ATCC 27294, CC<sub>50</sub>/MIC; **INH**: isoniazid: **RFP**: rifampicin.

streptomycin, ethambutol and levofloxacin) (Table 2). The results indicate that all of **10a-c**, **10e**, **10g**, **10h** and **11a**, **11b** have excellent activity against both of the two strains with similar MIC values (0.016–0.031  $\mu g$  mL<sup>-1</sup>) to **PBTZ169** (MIC: 0.016  $\mu g$  mL<sup>-1</sup>), suggesting their promising potential for both drug-sensitive (MIC: <0.1  $\mu g$  mL<sup>-1</sup>) and resistant MTB strains (Tables 1 and 2).

The BTZ derivatives were initially evaluated for their water solubility which was determined by HPLC measurement of the concentration of a micromembrane filtered saturated solution. Compared with other oxime derivatives (0.02–0.05 mg mL $^{-1}$ , clog P: 3.28–4.98), compound **10a** (clog P: 2.83) has much

**Table 2** *In vitro* activity against MDR-MTB strains, solubility and metabolic stability of selected compounds $^a$ 

	MIC (μg mL <sup>-1</sup> )			
Compd	MDR-MTB 16892 <sup>b</sup>	$\begin{array}{c} \text{MDR-MTB} \\ 16802^b \end{array}$	Water Solubility (mg m $L^{-1}$ )	Metabolic stability $t_{1/2}$ (min)
10a	0.016	0.031	0.24	>120
10b	0.016	0.016	0.03	NT
10c	0.016	0.016	0.05	NT
10e	0.016	0.016	0.02	NT
10g	0.016	0.016	0.05	NT
10h	0.031	0.016	0.03	NT
11a	0.016	0.016	4.05	49.1
11b	0.016	0.016	3.11	24.3
PBTZ169	0.016	0.016	0.30	19.5
INH	>40	2.5	NT	NT
RFP	>40	20	NT	NT
	10a 10b 10c 10e 10g 10h 11a 11b PBTZ169	MDR-MTB 16892 <sup>b</sup> 10a 0.016 10b 0.016 10c 0.016 10e 0.016 10h 0.031 11a 0.016 11b 0.016 PBTZ169 0.016 INH >40	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$

 $<sup>^</sup>a$  NT: not tested.  $^b$  MDR-MTB 16892 and MDR-MTB 16802 were isolated from patients in Beijing Chest Hospital.

**Table 3** PK Profiles of **10a**, **11a** and **11b** dosed orally in mice<sup>a</sup> at 50 mg  $kq^{-1}$  (n = 3)

Compd	10a	11a	11b	PBTZ169
$C_{ m max}$	$5767\pm3190$	$173\pm76.8$	$327\pm137$	$1512\pm696$
$(ng mL^{-1})$				
$T_{\max}$ (h)	$\textbf{0.333} \pm \textbf{0.144}$	$\textbf{1.000} \pm \textbf{0.866}$	$0.250\pm0$	$\textbf{0.583} \pm \textbf{0.382}$
$AUC_{0-\infty}$	$7678\pm4395$	$627 \pm ND$	$646 \pm 460$	$5681 \pm 1756$
$(h ng mL^{-1})$				
$t_{1/2}$ (h)	$\textbf{1.15} \pm \textbf{0.540}$	$1.76 \pm ND$	$\textbf{1.69} \pm \textbf{0.483}$	$\textbf{3.31} \pm \textbf{0.187}$
MRT (h)	$\textbf{1.85} \pm \textbf{0.634}$	$2.92 \pm ND$	$\textbf{2.42} \pm \textbf{0.171}$	$\textbf{4.48} \pm \textbf{0.484}$

<sup>&</sup>lt;sup>a</sup> Male CD-1 mice was acquired from WuXi AppTec (Shanghai) CO., Ltd; ND: not determined (parameters not determined due to inadequately defined terminal elimination phase).

greater solubility (0.24 mg mL<sup>-1</sup>) which is only slightly smaller than **PBTZ169** (0.30 mg mL<sup>-1</sup>). As expected, exocyclic tertiary amino derivatives **11a**, **11b** possess excellent aqueous solubility (3.11–4.05 mg mL<sup>-1</sup>) which is more than ten times that of **BPTZ169** (Table 2). Moreover, all of **10a**, **11a** and **11b** show better metabolic stability ( $t_{1/2}$ : 24.3–>120 min) in human liver microsomes compared to **BPTZ169** ( $t_{1/2}$ : 19.5 min).

Based on the measured activity levels against the tested strains, solubility and metabolic stability, **10a**, **11a** and **11b** were further tested for *in vivo* pharmacokinetic (PK) profiles in mice after a single oral administration of 50 mg kg<sup>-1</sup>. As shown in Table 3, compound **10a**, with a 3-(methoxyimino) azetidine moiety, displays good PK properties, with  $C_{\rm max}$  of 5767 ng mL<sup>-1</sup>, AUC<sub>0-\infty</sub> of 7678 h ng mL<sup>-1</sup> and  $t_{1/2}$  of 1.15 h. Compounds **11a** and **11b** have a relatively longer  $t_{1/2}$  of 1.69–1.76 h and MRT of 2.42–2.92 h, but a poor  $C_{\rm max}$  of 173–327 ng mL<sup>-1</sup> and AUC<sub>0-\infty</sub> of 627–646 h ng mL<sup>-1</sup>. Unexpectedly, all of the three compounds have a shorter  $t_{1/2}$  of 1.15–1.76 h compared to **BPTZ169** ( $t_{1/2}$ : 3.31 h). The *in vitro* and *in vivo* half-times of inconsistency may be due to differences in species between human liver microsomes and mice.

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## Conclusions

A series of novel BTZ derivatives bearing an oximido or amino nitrogen heterocycle moiety were designed as new antitubercular agents through modifications at the C-2 position of BTZ043 and BPTZ169. Many of them exhibit excellent *in vitro* inhibitory activity against both drug-sensitive MTB strain H37Rv and drug-resistant clinical isolates. Compound 10a displays acceptable safety, aqueous solubility and PK properties, and it may serve as a new and promising lead compound for further antitubercular drug discovery. Studies to determine the *in vivo* efficacy of 10a are currently underway.

#### Ethical statement

Animals were maintained in accordance with the guidelines of the Chinese Association for Laboratory Animal Sciences, Beijing, China, and approved by the Institutional Ethical Committee (IEC) of Peking Union Medical College.

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