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Fabrication and characterization of electrospun feather keratin/poly(vinyl alcohol) composite nanofibers

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Feathers, which contain more than 90% of keratin, are valuable natural protein resources. Moreover, recycling waste feathers to develop biomaterials contributes to environmental protection. The aim of this study was to fabricate and characterize both random and aligned feather keratin (FK)/PVA composite nanofibers through an electrospinning process. The morphology, molecular interaction, crystallization behavior, and tensile properties of the nanofibers were investigated. The electrospinning process appeared stable and successful for solutions containing no more than 40 wt% of FK. Smooth and bead-free FK/PVA random nanofibers were obtained for all the experimental samples. The fiber average diameter decreased for the random nanofibers and the degree of fiber orientation increased for the aligned nanofibers as the FK content increased. The crystallinity of the nanofibers reduced with the incorporation of FK but the orientation of nanofibers was favorable to crystallization. Random nanofibers exhibited an increased tensile strength when the FK to PVA ratio increased. Tensile strength and elongation at break in the direction of the aligned fibers were improved compared to the random nanofibers at the same FK content. This work demonstrates the potential of expanding the application of nanofiber-based products in the biomaterial field.

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1 Introduction

Electrospinning is a material-processing method widely used in the fabrication of polymeric nanofibers (NFs) by depositing a charged polymer jet onto a grounded collector. Due to their specific structural characteristics and functional properties, the nanofibers have shown great potential in various applications, including wound dressing, drug delivery, vascular tissue engineering, heavy metal ion adsorption, and antimicrobial active packaging.^{1–5} The natural polymers have cell-binding sites and biomolecular signatures that can mimic natural tissue. While the synthetic materials usually have good mechanical properties and limited cell affinity compared to the natural polymers. Therefore, in recent years the combination of natural and synthetic polymers for the production of nanofibers by electrospinning has gained increased attention. Shalumon *et al.*⁶ successfully electrospun chitosan (CS) together with poly(*ε*-caprolactone) (PCL) into nanofibers from a novel solvent mixture composed of formic acid and acetone. The authors

demonstrated that fibers obtained from a solution containing 1% CS and 8% PCL at a 1 : 1 ratio resulted in slightly beaded fibers presenting a diameter of 116 ± 27 nm. In contrast, they obtained fine nanofibers exhibiting a diameter of 102 ± 24 nm when they used a 1 : 3 ratio composition. Besides, Vega-Lugo *et al.*⁷ studied the positive effects of poly(ethylene oxide) (PEO) on the electrospinning of whey protein isolate (WPI) solutions under different pH conditions, and revealed that the electrospinning-enabling properties of PEO on aqueous WPI solutions were attributed to physical chain entanglement between the two polymers, rather than specific polymer–polymer interactions. Moreover, Cho *et al.*⁸ found that the diameter and mechanical strength of the electrospun soy protein isolate (SPI)/poly(vinyl alcohol) (PVA) hybrid nanofibers decreased gradually as the SPI content increased, but the biodegradation rate of the hybrid nanofibers increased with the incorporation of SPI.

Among the numerous natural polymers available, keratin is considered as one of the most abundant proteins in nature and can be used in various medical applications due to its biocompatibility and biodegradability.^{9–11} In addition, the poultry industry generates important annual volumes of by-produced feathers,^{12,13} representing a significant resource as they contain more than 90% in weight of keratin.¹⁴ Unfortunately, most of the feathers are discarded as solid waste in landfill sites without any pretreatment, causing severe environmental pollution. Therefore, recycling and utilizing feather

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keratin (FK) for the production of biomaterials could also contribute to the protection of the environment.

To date, many studies reported on electrospun nanofibers containing keratin from hair and wool, and the keratin used in these researches was generally extracted by a reduction method employing sodium metabisulfite or 2-mercaptoethanol.^{3,15-19} In the present study, feather keratin (FK) was extracted by partially oxidizing feathers with peracetic acid. The resulting FK was easily dispersed in water but could not be directly used for electrospinning due to the lack of sufficient entanglement or interactions. To overcome this issue, considering the hydrophilicity of partially oxidized keratin, we suggested blending FK with a water-soluble electrospinnable polymer, namely polyvinyl alcohol (PVA). It was expected that the affinity between PVA and FK would facilitate the fiber formation during the spinning process. The main objective of this work was to evaluate the feasibility of using FK and PVA from blended aqueous solutions to produce both random and aligned electrospun composite nanofibers by altering the rotating speed of the collector. Morphologies of the composite nanofibers and the interaction between FK and PVA were investigated by scanning electron microscopy (SEM) and Fourier transform infrared (FT-IR) spectroscopy. Thermal behaviors and tensile properties of the fibers with different ratios of FK and PVA were evaluated.

2 Experimental

2.1 Materials

The chicken feathers were collected from markets in Guangzhou city. Poly(vinyl alcohol) (PVA, degree of alcoholysis: 87–89%) was purchased from Aladdin industrial corporation (China). Peracetic acid was purchased from Tianjin Damao chemical reagents factory (China). All the chemicals were used without further purification. Double-distilled water was used as solvent.

2.2 Extraction of feather keratin

The chicken feathers collected from markets were firstly immersed in ethanol for 24 h, and then thoroughly washed with detergents and water respectively. Then the wet feathers were dried in an oven at 60 °C for 48 h. Next, FK was extracted according to the literature reported by Timmons *et al.* with slight modifications.^{20,21} The cleaned chicken feathers were added to a 16 wt% solution of freshly prepared peracetic acid (1 : 12, w/w), and the extraction was performed at 60 °C for

100 min under constant stirring. The resultant solution were passed through a nylon mesh (150 and 75 µm, respectively). The filtrates were subsequently dialyzed in dialysis tubes (Oso-T8280, 8000-14000, Union Carbide, USA) against distilled water at room temperature for a week, and the water was changed every 12 hours. The desired FK precipitated through dialysis. After filtration, washing and drying, FK was ground and stored in a sealed bag until further usage.

2.3 Preparation of polymer solutions for electrospinning

Polymer solutions for electrospinning were prepared by dissolving FK and PVA at different weight ratios in distilled water and maintaining the total weight of polymers in solution constant at 14 wt%. FK powders were dispersed in water and completely dissolved by slowly dropping a 1 mol L⁻¹ NaOH solution under continuous stirring at 50 °C. Different weight ratios of PVA was added to the resulting FK solution and the pH value of the mixture was adjusted to 9.5 using a 0.1 mol L⁻¹ NaOH solution. A series of FK/PVA solutions labeled FK₀PVA₁₀, FK₁PVA₉, FK₂PVA₈, FK₃PVA₇, and FK₄PVA₆ were prepared with FK : PVA weight ratios of 0 : 10, 1 : 9, 2 : 8, 3 : 7, and 4 : 6, respectively.

2.4 Characterization of the polymer solutions

The viscosity of the polymer solutions was determined by a digital viscometer (SNB-2, Shanghai Precision & Scientific Instrument Co. Ltd., China) at 25 °C. In addition, electrical conductivity of the solutions was measured by a digital conductivity meter (DDS-11A, Shanghai Leici-Chuangyi Instrument and Meter Co. Ltd., China).

2.5 Electrospinning of FK/PVA composite nanofibers

FK/PVA nanofibers were prepared using a electrospinning machine (ET-2535DC, Beijing Yongkang Leye Technology Development Co. Ltd., China) and the schematic representation is showed in Fig. 1. 10 mL of each polymer solution was loaded into a syringe equipped with a 22-gauge blunt-tipped needle. A rotating cylindrical mandrel wrapped with an aluminum foil was used as the fiber collector. The electrospinning process was performed at 9 kV applied voltage, 10 cm needle tip-to-collector distance, and 0.5 mL h⁻¹ flow rate. The rotating speed of the collector was 80 rpm for the random nanofibers (RNFs) and 2800 rpm for aligned the nanofibers (ANFs).

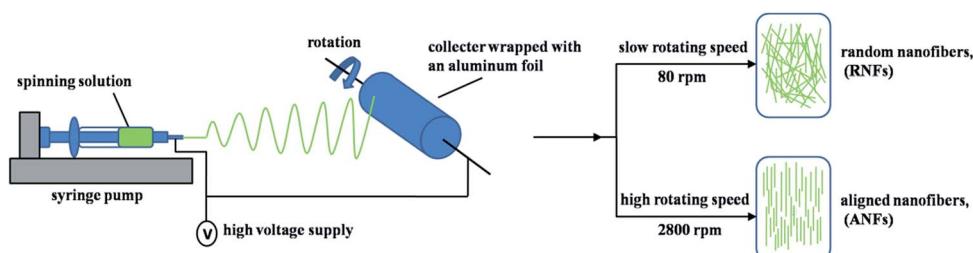


Fig. 1 Schematic representation of the applied electrospinning process.



2.6 Scanning electron microscopy (SEM)

The morphologies of the electrospun nanofibers were observed by scanning electron microscopy (EVO 18, Carl Zeiss, Germany) at 10 kV accelerating voltage. The average fiber diameter and diameter distribution were determined by measuring 100 fibers selected randomly from each sample.

2.7 Attenuated total reflectance infrared spectroscopy (ATR-FTIR)

The FTIR spectra of the nanofiber samples were recorded with an infrared spectrometer (spectrum 100, Perkin-Elmer, USA), using attenuated total reflectance mode (ATR). A range of wavenumber from 4000–650 cm⁻¹ and accumulation of 8 scans were used for each analysis.

2.8 Differential scanning calorimetry (DSC)

The thermal properties of the electrospun nanofibers were performed using a differential scanning calorimeter (DSC1, Mettler Toledo, Switzerland). One sample of each formulation was analyzed by being heated from 25 to 250 °C at a heating rate of 10 °C min⁻¹.

2.9 Tensile properties

The tensile properties of the nanofibers including tensile strength (TS) and elongation at break (EAB) were determined using a universal testing machine (CMT6503, Shenzhen MTS Test Machine Company Ltd., China). A strain rate of 10 mm min⁻¹ was used throughout the experiment, according to the ASTM standard D638. The nanofibers were cut into a size of 75 mm × 10 mm and the thicknesses of the samples were measured by a micrometer. Three replications of each formulation were performed and average values of the measurements were used. The schematic representation of the tensile performance test for aligned fibers is displayed in Fig. 2.

3 Results and discussion

3.1 Physicochemical properties of the polymer solutions

It has been widely approved that the physicochemical properties of an electrospinning solution such as viscosity and conductivity play a decisive role in the formation of continuous fibers.^{22–24} These parameters determine the morphology and size of the fibers. The solution properties of the various FK/PVA compositions are represented in Fig. 3. During electrospinning,

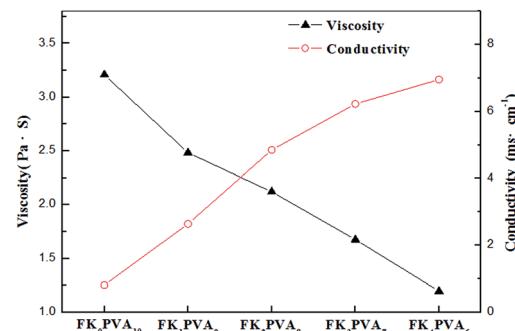


Fig. 3 Viscosity, and conductivity of the spinning solution as function of FK/PVA ratio.

an appropriate viscosity of the solution is essential for the formation of fibers. A viscosity below a minimum threshold would form beads, while one above a maximum limit would not permit to achieve electro ejection. It could be observed that at a fixed total polymer concentration of 14 wt%, the pure PVA solution (FK₀PVA₁₀) presented the highest viscosity (3.210 Pa s). The viscosity of the electrospinning solution decreased when the FK proportion increased. The lowest viscosity of 1.195 Pa s was achieved with sample FK₄PVA₆.

Electrical conductivity determines the ability to transfer charge to the surface of the pendant droplet, and it directly affects the buildup of an electrostatic repulsion force that is critical to initiate jetting. As shown in Fig. 3, conductivity increased as the amount of FK increased inside the blend solution. This observation could be attributed to the presence of many polar groups such as amide and carboxylic groups existing in keratin, as reported for the case of keratin/poly(lactic acid) blends in the literature.²⁵ The results indicated that the variation of conductivity with the increasing FK content was not linear, which could be due to the existence of contact resistance between both polymers.²³

3.2 Morphologies

The electrospinning process appeared stable and successful for all the experimental groups at a total polymer concentration of 14 wt%. The SEM images of electrospun random nanofibers (RNFs) prepared at different ratio of FK and PVA are displayed in Fig. 4. The RNFs exhibited common features consisting of a porous and fibrous structure. The fibers were smooth, bead-free, and randomly oriented for all the PVA fibers and FK/PVA fibers. These observations implied that no obvious phase separation between FK and PVA occurred during the electrospinning process, supporting the favorable compatibility between the two phases. However, the addition of FK led to modifications in the surface morphologies of the fibers. Indeed, pure PVA nanofibers (RNFs–FK₀PVA₁₀) were straight with circular cross sections, while FK-incorporated RNFs exhibited more curved and belt-like morphologies and reduced inter-fiber space. These belt-like morphologies, which were also observed in the electrospinning of wool keratin in formic acid at 20 wt% and fibroin-based nanofibers from solutions presenting

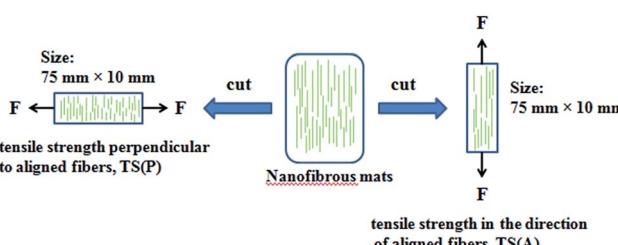


Fig. 2 Schematic representation of the tensile performance test applied on the aligned fibers.



relatively high viscosity (about 0.5 Pa s),^{26,27} could be the consequence of an uneven evaporation of the solvent from the skin and the core of the jet during drawing, causing the collapse of the filament.²⁸ Moreover, branching of the fibers appeared in the FK/PVA RNFs, which could be attributed to the uneven distribution of the charge carried by the jets, caused by jet elongation/bending and solvent evaporation. Indeed, jets could reduce their local surface charge density by ejecting smaller jets from the surface of the primary jets or by splitting into multiple smaller jets.²² The conductivity of the electrospinning solutions increased as the content of FK increased (Fig. 3), potentially leading to more uneven jet surface charge distribution, which consequently resulted in higher probability of jet branching.

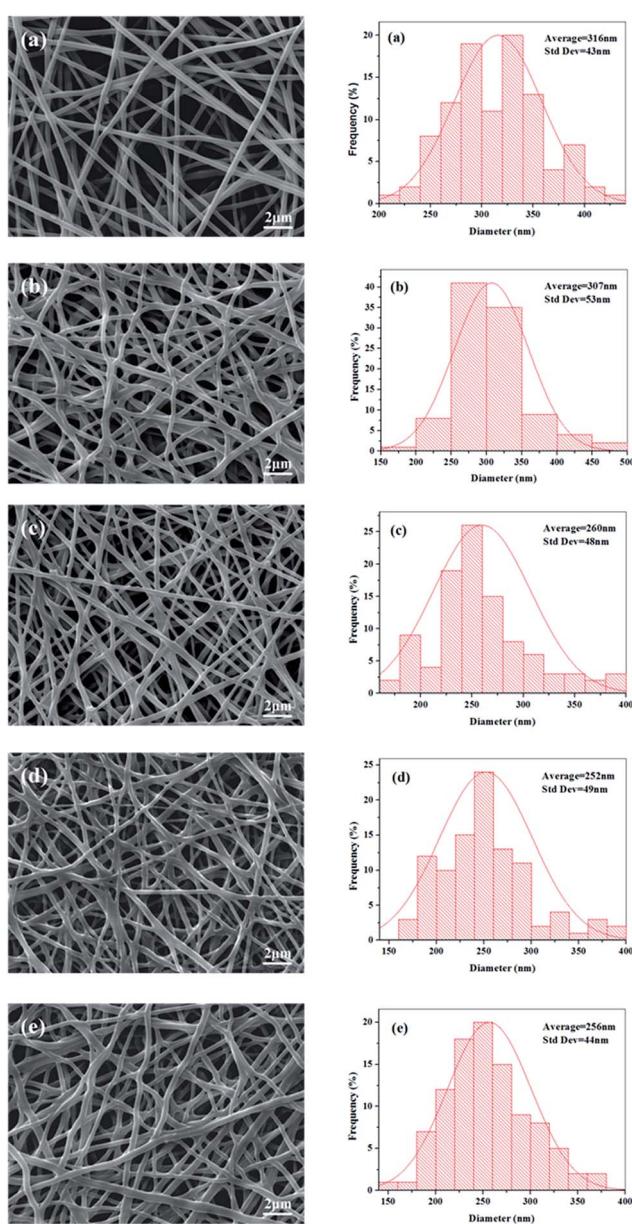


Fig. 4 SEM images and diameter distributions of electrospun random nanofibers (RNFs) with various blending ratios: (a) FK₀PVA₁₀, (b) FK₁PVA₉, (c) FK₂PVA₈, (d) FK₃PVA₇, (e) FK₄PVA₆.

We recorded the micrographs of the RNFs prepared from various blending FK/PVA ratios by SEM and analyzed their diameter distributions as illustrated in Fig. 4. The results showed that pure PVA nanofibers (NFS-FK₀PVA₁₀) presented the highest fiber average diameter and the lowest standard deviation (316 ± 43 nm) among all the tested fibers. Upon the addition of FK into the spinning solutions, we could observe a general trend of decreased average fiber diameter and wide distribution. The viscosity of the solutions decreased with the incorporation of FK, thus the interactions and entanglements between molecular chains were weakened. Furthermore, the conductivity increased as the ratio of FK to PVA increased. As a consequence, the jets generated by the blend solutions became less resistant to the stretching repulsive forces from the charge during the electrospinning process, resulting in the decrease in diameter of the composite electrospun fibers. Similar results were also obtained in the study of human hair keratin/polyurethane nanofibrous mats and soy protein isolate/PVA hybrid nanofibers.^{8,15}

It should be pointed out that to obtain satisfactory fibers, the ratio of FK in the blend solution had to be less than or equal to 40 wt% (FK₄PVA₆). Electrospinning was not successful when the FK content increased further. Poor morphologies of fibers exhibiting many large beads were obtained and the fibers were difficult to handle (data not presented). Similarly, Yen *et al.*²⁹ also found that the fibrous structure was not constructed by electrospinning keratin/fibroin blend solutions when the keratin content exceeded 35%. Therefore, only electrospun fibers from solutions containing no more than 40 wt% of FK were further considered in the present study.

The aligned nanofibers (ANFs) were reported to be able to instruct the cultured cells directionally growth, resulting in the elongation and alignment of cells along the major axis of the nanofibers.³⁰⁻³² Thus the orientation of nanofibers provide nanofibers a broad range of applications in the biomaterial field. Several researchers have found out that the orientation of fibers could be modulated by the rotational speed of the collector and that by increasing the rotating rate, more uniform fibers were produced.^{33,34} Fig. 5 displays the SEM images of electrospun ANFs which were collected by rotating cylindrical mandrel at the speed of 2800 rpm. It can be seen that ANFs were

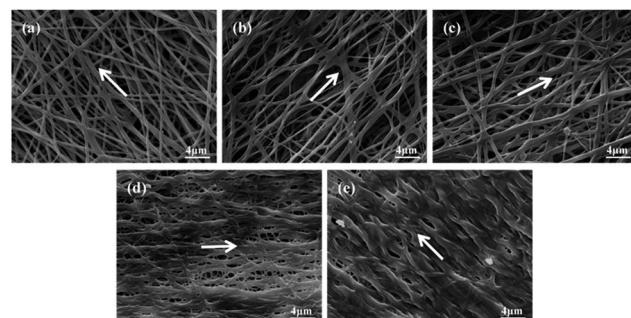


Fig. 5 SEM images of electrospun aligned nanofibers (ANFs) with various blending ratios: (a) FK₀PVA₁₀, (b) FK₁PVA₉, (c) FK₂PVA₈, (d) FK₃PVA₇, (e) FK₄PVA₆. Notes: the white arrow indicates the orientation of the ANFs.





fabricated by the higher rotating speed of collector, and the degree of the ANFs orientation increased as the FK content increased in the blends. In addition, it was interesting that when the weight fraction of FK in the solution was higher than 20 wt% (FK_3PVA_7 and FK_4PVA_6), the ANFs were found to be densely packed as the interstice between the fibers reduced. This result was probably due to the plasticizing effect of FK during the electrospinning process, facilitating orientation and flow of PVA by uncoiling and wrapping around PVA chains. A similar contribution was reported for pullulan in the pullulan/API electrospun fibers and PEO in the PEO/chitosan blends.^{24,35}

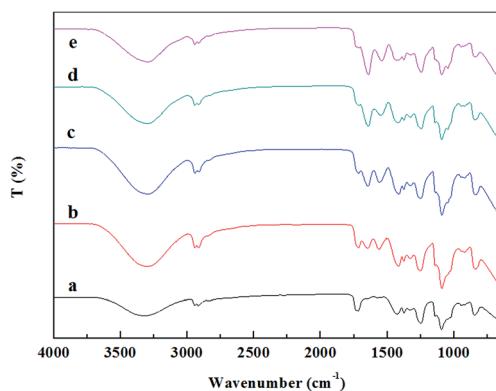


Fig. 6 FTIR spectra of the RNFs with various blending ratios: (a) RNFs– $\text{FK}_0\text{PVA}_{10}$, (b) RNFs– FK_1PVA_9 , (c) RNFs– FK_2PVA_8 , (d) RNFs– FK_3PVA_7 , (e) RNFs– FK_4PVA_6 .

3.3 FT-IR spectra

The molecular interactions between FK and PVA, together with the secondary structure of the keratin in the composite nanofibers were investigated by FTIR spectroscopy. As shown in Fig. 6, the characteristic absorption band of PVA, attributed to the O–H stretching vibration, was observed at 3319 cm^{-1} in the spectra of $\text{FK}_0\text{PVA}_{10}$.³⁶ When FK was added into the spinning system, spectra of the resultant composite nanofibers exhibited new absorption peaks of amide I (C=O stretching vibration) and amide II (N–H bending and C–H stretching vibration), which were located at $1700\text{--}1600$ and $1580\text{--}1480\text{ cm}^{-1}$, respectively.³⁷ These bands confirmed the introduction of keratin inside the composite nanofibers. It was found that the wavenumbers of amide I and amide II shifted to a lower wavenumber and the intensities gradually increased as the proportion of FK increased. In addition, the characteristic absorption band of PVA at 3319 cm^{-1} also shifted to a lower wavenumber in the composite nanofibers. These results strongly indicated that the main interaction between FK and PVA in the fibers would be through the formation of hydrogen bonds, which improved the electrospinnability of FK/PVA composite nanofibers.

The amide I ($1700\text{--}1600\text{ cm}^{-1}$) band is known to be especially sensitive to the secondary structure of the proteins.³⁸ On the basis of the above spectral analysis, the position of the amide I transmission band was not disturbed by the absorption peaks generated by PVA. Therefore, the amide I band could characterize the secondary structure of the keratin inside the composite nanofibers. A closer look into the region of $1700\text{--}1600\text{ cm}^{-1}$

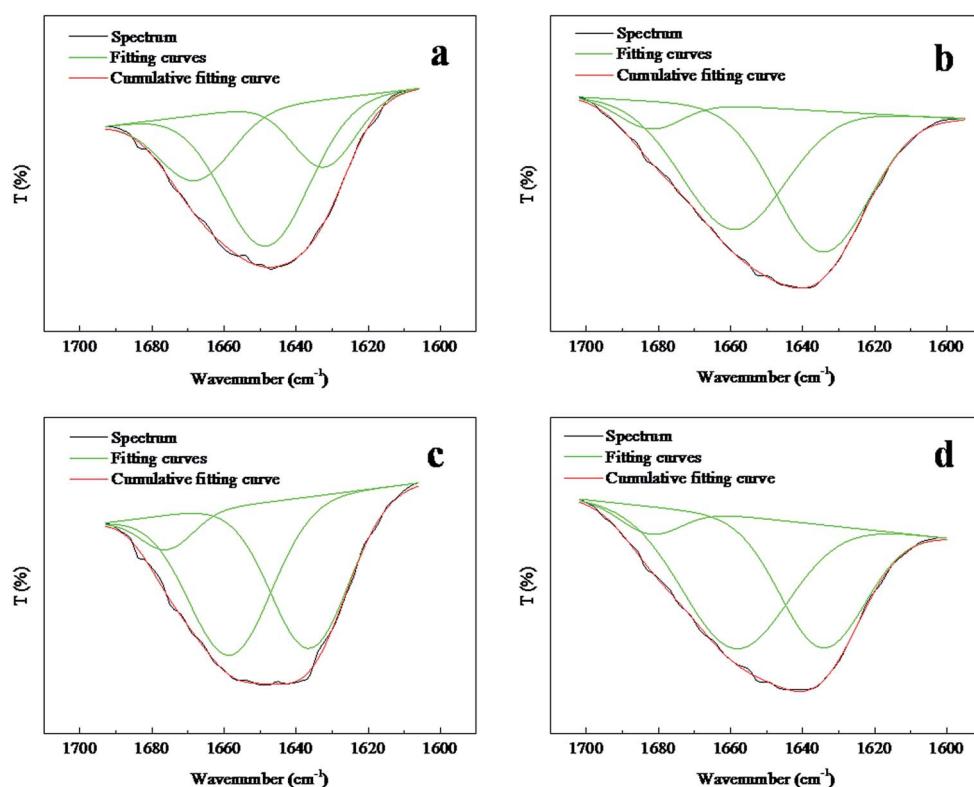


Fig. 7 Amide I region spectra of random and aligned nanofibers: (a) RNFs– FK_1PVA_9 , (b) RNFs– FK_4PVA_6 , (c) ANFs– FK_1PVA_9 , (d) ANFs– FK_4PVA_6 .

1600 cm^{-1} for both selected random and aligned FK/PVA nanofibers are presented in Fig. 7. The absorption band at $\sim 1650 \text{ cm}^{-1}$ is characteristic of α -helix structures in keratin chains, whereas the peaks between 1610–1633 cm^{-1} are attributed to β -sheet structures.^{39–42} The areas of the peak's corresponding structure were calculated and summarized in Table 1. The results indicated that the ANFs–FK₄PVA₆ showed a lower band area of the β -sheet structure and a higher area of the α -helix structure, compared to the ANFs–FK₁PVA₉. This observation suggested that the molecular chains of keratin could be hindered by the PVA chains during the process of fiber alignment. A similar effect was reported for the keratin/poly(lactic acid) nanofiber mats.²⁵ However, the contrary behavior was observed for the α -helix and β -sheet structure variation of the random nanofibers.

3.4 DSC analysis

The DSC curves together with the related peak temperatures and enthalpy values (ΔH_m) of the random and aligned nanofibers are presented in Fig. 8 and Table 2. The endothermic peak observed at about 200 °C for all the samples was attributed to the crystalline melting (T_m) of the nanofibers, and the peak area represented the crystallinity of the nanofibers.⁴³ The RNFs exhibited a melting peak (T_m) at approximately 198 °C, and the ΔH_m value decreased from 35.29 (RNFs–FK₀PVA₁₀) to 18.81 J g^{−1} (RNFs–FK₄PVA₆) after adding 40 wt% of keratin. This result suggested

that the crystallinity of the nanofibers reduced upon the incorporation of FK. Similar observation was reported for the case of keratin/PVA films by the casting process.⁴⁴ The ΔH_m values of the ANFs were markedly higher than that of the RNFs when compared at the same amount of keratin, indicating that the orientation of the nanofibers was favorable to crystallization.

3.5 Tensile properties

The results of the tensile tests in terms of tensile strength (TS) and elongation at break (EAB) of the nanofibers were summarized in Table 3 and 4. In the literature, different results are reported on the effect of protein content on the tensile properties of composite nanofibers. Li *et al.*⁴⁵ reported that PCL/keratin composite nanofibrous mats showed an enhanced tensile strength when keratin was added. This observation was explained by the compatibility of keratin and PCL in a limited range or by the rigid-filler role of hard keratin.⁴⁶ While Cho *et al.*⁸ found that the increase in the soy protein isolate (SPI) content induced the sharp reduction of both the breaking strength and elongation of the SPI/PVA hybrid nanofibers. This result corresponded with the fact that SPI protein could not produce strong fibers due to its globular structure. In the present study, the TS values of the FK/PVA composite RNFs were higher than that of the pure PVA fibers (RNFs–FK₀PVA₁₀). The increased TS was naturally coupled with a decrease of extensibility (EAB value). Reinforced strength was possibly related to the favorable compatibility and strong interactions between FK and PVA (Fig. 4 and 6), as well as the increased content of β -sheet structure in the keratin at higher FK content (Table 1). It is known that β -sheet structure could provide a more rigid structure than α -helix, and lead to high strength and small displacement characteristic of the materials.²⁵

With respect to the aligned nanofibers, the TS in the direction of the aligned fibers was higher and the TS perpendicular to the aligned fibers was lower, compared to the RNFs prepared at the same FK ratio. Moreover, extensibility (EAB value) in the

Table 1 Characteristics of the amide I bands of FK/PVA nanofibers

Samples	Secondary structure	Area (%)
RNFs–FK ₁ PVA ₉	α -Helix	57
	β -Sheet	20
RNFs–FK ₄ PVA ₆	α -Helix	46
	β -Sheet	48
ANFs–FK ₁ PVA ₉	α -Helix	47
	β -Sheet	46
ANFs–FK ₄ PVA ₆	α -Helix	54
	β -Sheet	40

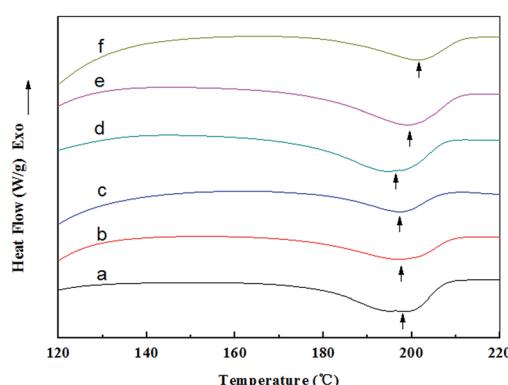


Fig. 8 DSC curves of random and aligned nanofibers: (a) RNFs–FK₀PVA₁₀, (b) RNFs–FK₁PVA₉, (c) RNFs–FK₄PVA₆, (d) ANFs–FK₀PVA₁₀, (e) ANFs–FK₁PVA₉, (f) ANFs–FK₄PVA₆.

Table 2 Peak temperatures and enthalpy values of the DSC analysis

Samples	T_m (°C)	ΔH_m (J g ^{−1})
RNFs–FK ₀ PVA ₁₀	198.55	35.29
RNFs–FK ₁ PVA ₉	197.20	28.45
RNFs–FK ₄ PVA ₆	197.18	18.81
ANFs–FK ₀ PVA ₁₀	194.59	41.79
ANFs–FK ₁ PVA ₉	198.89	39.47
ANFs–FK ₄ PVA ₆	201.23	24.61

Table 3 Tensile properties of the RNFs

Samples	TS (MPa)	EAB (%)
RNFs–FK ₀ PVA ₁₀	5.59 ± 1.18	50.17 ± 6.05
RNFs–FK ₁ PVA ₉	6.15 ± 2.12	34.33 ± 7.09
RNFs–FK ₂ PVA ₈	6.87 ± 0.38	23.17 ± 5.97
RNFs–FK ₃ PVA ₇	6.82 ± 0.92	17.83 ± 5.35
RNFs–FK ₄ PVA ₆	6.71 ± 0.56	14.33 ± 3.51



Table 4 Tensile properties of the ANFs

Samples	TS (MPa)		EAB (%)	
	In the direction of aligned fibers	Perpendicular to aligned fibers	In the direction of aligned fibers	Perpendicular to aligned fibers
ANFs-FK ₀ PVA ₁₀	8.54 ± 0.95	1.52 ± 0.28	154.17 ± 12.57	42.33 ± 4.51
ANFs-FK ₁ PVA ₉	8.93 ± 1.58	1.18 ± 0.33	77.00 ± 11.63	33.83 ± 7.82
ANFs-FK ₂ PVA ₈	9.44 ± 1.10	0.95 ± 0.18	87.83 ± 16.00	31.00 ± 4.36
ANFs-FK ₃ PVA ₇	10.53 ± 1.35	1.28 ± 0.16	110.67 ± 14.57	21.67 ± 6.51
ANFs-FK ₄ PVA ₆	10.71 ± 1.36	1.31 ± 0.31	111.10 ± 15.70	17.50 ± 3.77

direction of the aligned fibers was also significantly improved. These results could be explained by the orientation of the fibers, which is represented by a white arrow in Fig. 5.

4 Conclusion

Feather keratin (FK) extracted by partial oxidization using peracetic acid was blended with PVA at different weight fraction, while maintaining the total weight of polymer in the solution constant (14 wt%). The viscosity decreased and the electrical conductivity increased when the weight fraction of keratin in the blend solutions increased. The electrospinning process appeared stable and successful for FK/PVA blend solutions containing no more than 40 wt% of FK. Both random and aligned composite nanofibers (RNFs and ANFs) were fabricated by changing the rotating speed of the collector. All the nanofibers were found to be smooth and bead-free with favorable compatibility between FK and PVA. The FK/PVA RNFs exhibited a lower fiber average diameter and higher standard deviation compared to the pure PVA fibers. The degree of fiber orientation increased with the increase of the FK content in the blends for the ANFs. Formation of hydrogen bonds between the two components was confirmed by FTIR spectroscopy. The crystallinity of the nanofibers reduced with the incorporation of FK but the orientation of nanofibers was favorable to crystallization. The FK/PVA RNFs exhibited an increased tensile strength coupled with a decrease of extensibility when increasing the FK content. Tensile strength and elongation at break in the direction of the ANFs were higher compared to the RNFs at the same FK ratio. Overall, the present work demonstrated the possibility to incorporate FK into nanofibers that have the potential to be applied in the biomaterial field.

Acknowledgements

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Notes and references

1 C. S. Ki, E. H. Gang, I. C. Um and Y. H. Park, *J. Membr. Sci.*, 2007, **302**, 20–26.

- 2 J. Li, Y. Li, L. Li, A. F. T. Mak, F. Ko and L. Qin, *Polym. Degrad. Stab.*, 2009, **94**, 1800–1807.
- 3 H.-L. Zhang, J. Wang, N. Yu and J.-S. Liu, *J. Polym. Res.*, 2014, **21**, 329–336.
- 4 M. Boakye, N. Rijal, U. Adhikari and N. Bhattacharai, *Materials*, 2015, **8**, 4080.
- 5 D. G. Yu, G. R. Williams, X. Wang, X. K. Liu, H. L. Li and S. W. A. Bligh, *RSC Adv.*, 2013, **3**, 4652–4658.
- 6 K. T. Shalumon, K. H. Anulekha, C. M. Girish, R. Prasanth, S. V. Nair and R. Jayakumar, *Carbohydr. Polym.*, 2010, **80**, 413–419.
- 7 A.-C. Vega-Lugo and L.-T. Lim, *J. Polym. Sci., Part B: Polym. Phys.*, 2012, **50**, 1188–1197.
- 8 D. Cho, A. N. Netravali and Y. L. Joo, *Polym. Degrad. Stab.*, 2012, **97**, 747–754.
- 9 R. Li and D. Wang, *J. Appl. Polym. Sci.*, 2013, **127**, 2648–2653.
- 10 Y. W. Wu, C. Y. Han, J. H. Yang, S. X. Jia and S. G. Wang, *Surf. Coat. Technol.*, 2011, **206**, 506–510.
- 11 S. Reichl, *Biomaterials*, 2009, **30**, 6854–6866.
- 12 E. Jin, N. Reddy, Z. Zhu and Y. Yang, *J. Agric. Food Chem.*, 2011, **59**, 1729–1738.
- 13 L. Zhao, H. Zhou and J. Hua, *China Leather*, 2011, **40**, 36–40.
- 14 N.-B. Song, J.-H. Lee, M. Al Mijan and K. B. Song, *LWT-Food Sci. Technol.*, 2014, **57**, 453–460.
- 15 Y. Wang, P. Li, P. Xiang, J. Lu, J. Yuan and J. Shen, *J. Mater. Chem. B*, 2016, **4**, 635–648.
- 16 J. Fan, T.-D. Lei, J. Li, P.-Y. Zhai, Y.-H. Wang, F.-Y. Cao and Y. Liu, *Mater. Des.*, 2016, **104**, 60–67.
- 17 C. Jawun, G. Panthi, L. Yanan, K. Jongwan, C. Su-Hyeong, L. Chohye, P. Mira and K. Hak-Yong, *Polymer*, 2015, **58**, 146–152.
- 18 S. Li and X.-H. Yang, *Adv. Mater. Sci. Eng.*, 2014, **2014**, 1–7.
- 19 A. Aluigi, C. Tonetti, C. Vineis, C. Tonin and G. Mazzuchetti, *Eur. Polym. J.*, 2011, **47**, 1756–1764.
- 20 S. F. Timmons, C. R. Blanchard and R. A. Smith, *US pat. 006124265A*, 2000.
- 21 S. F. Timmons, C. R. Blanchard and R. A. Smith, *EP pat. 1033953B1*, 2008.
- 22 X. Xu, L. Jiang, Z. Zhou, X. Wu and Y. Wang, *ACS Appl. Mater. Interfaces*, 2012, **4**, 4331–4337.
- 23 J. Colin-Orozco, M. Zapata-Torres, G. Rodriguez-Gattorno and R. Pedroza-Islands, *Food Biophys.*, 2015, **10**, 134–144.
- 24 M. Aceituno-Medina, S. Mendoza, J. M. Lagaron and A. López-Rubio, *Food Res. Int.*, 2013, **54**, 667–674.



25 S. Isarankura Na Ayutthaya, S. Tanpichai and J. Wootthikanokkhan, *J. Polym. Environ.*, 2015, **23**, 506–516.

26 A. Aluigi, A. Corbellini, F. Rombaldoni, M. Zoccola and M. Canetti, *Int. J. Biol. Macromol.*, 2013, **57**, 30–37.

27 D. H. Baek, C. S. Ki, I. C. Um and Y. H. Park, *Fibers Polym.*, 2007, **8**, 271–277.

28 S. Koombhongse, W. X. Liu and D. H. Reneker, *J. Polym. Sci., Part B: Polym. Phys.*, 2001, **39**, 2598–2606.

29 K.-C. Yen, C.-Y. Chen, J.-Y. Huang, W.-T. Kuo and F.-H. Lin, *J. Mater. Chem. B*, 2016, **4**, 237–244.

30 J. Hu, L. L. Tian, M. P. Prabhakaran, X. Ding and S. Ramakrishna, *Polymers*, 2016, **8**, 54–70.

31 W. Zhu, F. Masood, J. O'Brien and L. G. Zhang, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2015, **11**, 693–704.

32 K. H. Zhang, C. Y. Wang, C. Y. Fan and X. M. Mo, *J. Biomed. Mater. Res., Part A*, 2014, **102**, 2680–2691.

33 Z. Wei, F. Masood, J. O'Brien and L. G. Zhang, *Nanomedicine Nanotechnology Biology & Medicine*, 2015, **11**, 693–704.

34 N. Zander, J. A. Orlicki, A. M. Rawlett and T. Beebe, *Biointerphases*, 2010, **5**, 149–158.

35 C. Kriegel, K. Kit, D. J. McClements and J. Weiss, *Polymer*, 2009, **50**, 189–200.

36 M. Park, H. K. Shin, G. Panthi, M. M. Rabbani, A.-M. Alam, J. Choi, H.-J. Chung, S.-T. Hong and H.-Y. Kim, *Int. J. Biol. Macromol.*, 2015, **76**, 45–48.

37 F. Selmin, F. Cilurzo, A. Aluigi, S. Franzè and P. Minghetti, *Results Pharma Sci.*, 2012, **2**, 72–78.

38 W. K. Surewicz, H. H. Mantsch and D. Chapman, *Biochemistry*, 1993, **32**, 389–394.

39 S. I. N. Ayutthaya, S. Tanpichai, W. Sangkhun and J. Wootthikanokkhan, *Int. J. Biol. Macromol.*, 2016, **85**, 585–595.

40 Y. Liu, X. Yu, J. Li, J. Fan, M. Wang, T.-D. Lei, J. Liu and D. Huang, *J. Nanomater.*, 2015, **2015**, 1–7.

41 M. Zoccola, A. Aluigi, C. Vineis, C. Tonin, F. Ferrero and M. G. Piacentino, *Biomacromolecules*, 2008, **9**, 2819–2825.

42 A. Aluigi, C. Vineis, A. Varesano, G. Mazzuchetti, F. Ferrero and C. Tonin, *Eur. Polym. J.*, 2008, **44**, 2465–2475.

43 Y. Nakano, Y. Bin, M. Bando, T. Nakashima, T. Okuno, H. Kurosu and M. Matsuo, *Macromol. Symp.*, 2007, **258**, 63–81.

44 Y. Dou, B. Zhang, M. He, G. Yin, Y. Cui and I. N. Savina, *Polymers*, 2015, **7**, 580–591.

45 Y. Li, Y. Wang, J. Ye, J. Yuan and Y. Xiao, *Mater. Sci. Eng., C*, 2016, **68**, 177–183.

46 J. Lee, G. Tae, Y. H. Kim, I. S. Park, S. H. Kim and S. H. Kim, *Biomaterials*, 2008, **29**, 1872–1879.

