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Sesquiterpenoids and monoterpenoid coumarins from *Parasenecio rubescens*[†]

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Ten new sesquiterpenoids (1–2, 5–9, 12–14) and four new monoterpenoid coumarins (15–18), along with four known sesquiterpenoids (3, 4, 10, 11) and one known monoterpenoid coumarin (19), were isolated from the whole plant of *Parasenecio rubescens*. The structures of the isolates were determined through comprehensive spectroscopic analyses and HRESIMS data, with the isolated sesquiterpenoids being classified into oplopane-type (1–4) and bisabolane-type (5–14). The absolute configuration of **1** was established by single-crystal X-ray diffraction analysis with Cu K α radiation. The cytotoxic effects against three cancer cell lines (B16 mouse melanoma cells, HepG2 human hepatocellular carcinoma cells, and MCF7 human breast adenocarcinoma cells) of the isolated compounds, as well as their antimicrobial effects against *Staphylococcus aureus*, *Escherichia coli*, and *Monilia albicans*, were evaluated. This study demonstrates that the unusual monoterpenoid coumarins, highly oxygenated oplopane-type and bisabolane-type sesquiterpenoids could have an important chemotaxonomic significance for the genus *Parasenecio*.

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Introduction

Consisting of more than 60 species (43 endemic in China), the genus *Parasenecio* of the family Compositae finds its main distribution in East Asia, the Sino-Himalayan region, and Russia.¹ More than 20 species of this genus have been used as Chinese folk medicines for the treatment of swelling, inflammation, dysentery, and so on. According to the International Code of Botanical Nomenclature (ICBN), the genus *Parasenecio* had been misnamed as European herb *Cacalia* L., the type species of which was first classified as *Cacalia alpina* L., and then transferred to *Adenostyles alpina* (L.) Bluff & Fingerh. The former species of *Cacalia* are now divided into a few different genera, such as *Parasenecio*, *Arnoglossum*, and *Adenostyles*.² Although the first investigation on *Parasenecio* was undertaken over decades ago,³ plants of the genus haven't received much interest in recent years due to their limited new bioactive secondary metabolites reported. So far, phytochemical studies have been carried out on more than a dozen species of this genus with the described chemical constituents mainly being eremophilanes.^{4–9} *Parasenecio rubescens*, a perennial herb, is mainly distributed in forest understories and margins, and thickets in valleys of the Anhui, Fujian, Hunan, and Jiangxi

Provinces of China. This herb has long been employed as a folk medicinal herb for the treatment of tussis, tonsillitis, and tuberculosis.¹⁰

In our previous research paper on *P. rubescens*,¹¹ we described the isolation, structural elucidation and bioactivities of 22 new highly oxygenated bisabolane-type sesquiterpenoids, which was the first isolation of bisabolanes from the genus *Parasenecio*. Inspired by those works, and also to continue the search for diverse bioactive compounds from Chinese herbs, we have kept studying on the whole plant of *P. rubescens*. As a result, ten new sesquiterpenoids (1–2, 5–9, 12–14) and four new monoterpenoid coumarins (15–18), along with four known sesquiterpenoids (3, 4, 10, 11) and one known monoterpenoid coumarin (19), were isolated. The isolated sesquiterpenoids were classified into oplopane-type (1–4) and bisabolane-type (5–14). This paper describes the continued work of the identification and characterization of secondary metabolites from *P. rubescens* and the evaluation of their cytotoxic and antimicrobial effects.

Results and discussion

The air-dried whole plant of *P. rubescens* (18.4 kg) was grounded into powder, and then extracted with 90% acetone at room temperature. The whole extract was sequentially partitioned with petroleum ether and EtOAc. The EtOAc-soluble fraction was subjected to column chromatography involving MCI gel, silica gel, reversed-phase C18 silica gel, Sephadex LH-20, and then purified by preparative HPLC to yield compounds 1–19 (Fig. 1).

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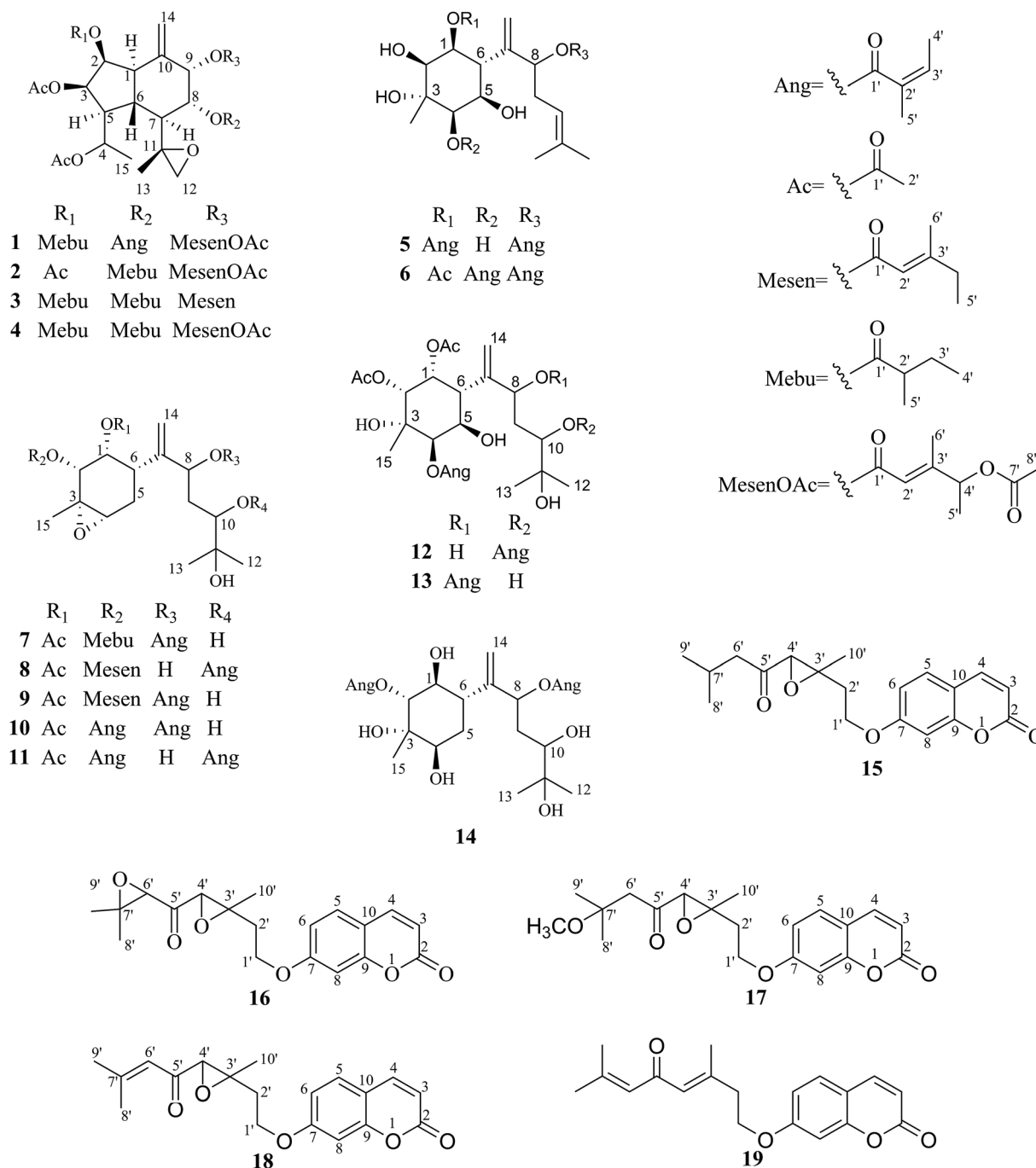


Fig. 1 Structures of compounds 1–19.

Oplopane-type sesquiterpenoids

Pararunine A (**1**) was isolated as colorless crystals. Its molecular formula was established as $C_{37}H_{52}O_{13}$, with 12 indices of hydrogen deficiency by the HRESIMS and ^{13}C NMR data. The 1H and ^{13}C NMR data (Tables 1 and 2) displayed characteristic signals for one (2-methylbutanoyl)oxy group [δ_H 2.43 (1H, m, H-2'), 1.49 (1H, m, H-3'), 1.78 (1H, m, H-3'), 0.98 (3H, t, $J = 7.2$, H-4'), 1.21 (3H, d, $J = 7.2$, H-5'); δ_C 176.2 (C-1'), 41.9 (C-2'), 26.7 (C-3'), 12.1 (C-4'), 17.7 (C-5')], two acetoxy groups [δ_H 2.04 (3H, s, H-2''), δ_C 170.0 (C-1''), 21.2 (C-2'')], [δ_H 2.09 (3H, s, H-2'''), δ_C 170.0

(C-1'''), 21.3 (C-2''')], one angeloyloxy group [δ_H 6.09 (1H, qq, $J = 7.0$, 1.4 Hz, H-3'''), 1.98 (3H, dq, $J = 7.0$, 1.4 Hz, H-4'''), 1.88 (3H, q, $J = 1.4$ Hz, H-5'''); δ_C 167.2 (C-1'''), 127.6 (C-2'''), 139.4 (C-3'''), 15.9 (C-4'''), 20.4 (C-5''')], one (4-acetoxy-4-methylseneciyl)oxy group [δ_H 5.81 (1H, brs, H-2'''), 5.23 (1H, q, $J = 6.7$ Hz, H-4'''), 1.33 (3H, d, $J = 6.7$ Hz, H-5'''), 2.08 (3H, brs, H-6'''), 1.97 (3H, s, H-8'''); δ_C 165.0 (C-1'''), 114.9 (C-2'''), 158.3 (C-3'''), 73.7 (C-4'''), 19.3 (C-5'''), 15.4 (C-6'''), 170.4 (C-7'''), 21.2 (C-8''')], one epoxy group [δ_H 2.74 (1H, s, H-12a), 2.83 (1H, s, H-12b); δ_C 55.3 (C-11), 53.2 (C-12)], and a terminal double bond



Table 1 ^{13}C NMR data for compounds **1**, **2**, **5–9**, **12–18** (100 MHz)

No.	1 ^a	2 ^a	No.	5 ^a	6 ^a	7 ^b	8 ^b	9 ^b	12 ^b	13 ^b	14 ^b	No.	15 ^a	16 ^a	17 ^a	18 ^a
1	44.9	44.5	1	71.9	72.5	70.3	71.5	70.5	72.9	71.5	75.2	2	161.2	161.2	161.3	161.1
2	71.2	71.7	2	75.2	75.0	73.5	72.8	72.8	73.8	73.7	78.6	3	113.6	113.6	113.5	113.2
3	72.3	72.6	3	71.9	72.5	58.2	58.3	58.3	74.5	74.3	75.7	4	143.4	143.4	143.5	143.4
4	69.9	69.9	4	77.4	75.7	61.4	61.6	61.3	78.6	78.3	75.2	5	129.0	129.0	129.0	128.9
5	45.0	44.9	5	68.5	72.6	27.0	27.2	27.0	66.2	66.4	35.8	6	112.8	112.9	112.8	112.7
6	43.8	43.8	6	44.5	41.9	38.7	36.7	38.8	41.0	42.0	43.5	7	161.8	161.8	161.8	161.8
7	49.4	49.2	7	144.5	146.9	149.0	150.3	149.3	146.4	144.1	151.3	8	101.8	101.7	101.8	101.5
8	72.2	72.5	8	74.3	78.8	75.9	74.7	75.7	75.8	77.8	74.3	9	156.0	156.1	156.1	155.8
9	73.6	73.4	9	34.0	31.6	37.3	36.5	37.3	35.2	37.0	37.8	10	113.0	113.1	112.9	112.7
10	137.9	137.9	10	119.5	119.8	75.5	77.8	75.5	77.8	75.7	75.7	1'	65.3	65.1	65.4	65.0
11	55.3	55.3	11	134.6	134.4	73.5	72.6	73.5	72.8	73.5	73.5	2'	31.5	31.4	31.3	31.8
12	53.2	53.2	12	25.9	25.9	25.0	26.8	25.0	25.4	25.5	25.2	3'	62.2	63.1	62.5	61.9
13	16.7	16.5	13	25.2	18.1	25.7	26.8	25.7	26.8	25.0	25.2	4'	65.5	63.8	66.5	66.1
14	115.5	115.3	14	118.1	110.3	114.8	114.9	114.6	116.7	117.8	109.8	5'	205.7	201.8	204.7	194.7
15	16.4	16.3	15	18.1	24.4	19.8	19.8	19.8	23.8	23.7	23.7	6'	50.4	65.1	52.5	121.2
1'	176.2	169.9	1'	167.2	170.5	172.2	172.6	172.3	168.2	168.0	168.7	7'	24.6	62.1	75.1	159.3
2'	41.9	21.4	2'	127.8	21.1	20.9	20.9	20.8	128.9	128.9	128.9	8'	22.8	19.0	24.8	21.4
3'	26.7		3'	139.0					140.2	140.2	137.4	9'	22.8	25.1	25.2	28.1
4'	12.1		4'	16.0					16.3	16.3	16.0	10'	22.7	22.5	22.8	22.7
5'	17.7		5'	20.6					20.7	21.1	20.8	OCH ₃			49.6	
1''	170.0	169.8	1''	168.6	166.4	177.0	167.2	167.1	172.0	171.8	168.1					
2''	21.2	21.2	2''	127.8	127.1	42.5	114.6	114.6	21.1	20.6	128.3					
1'''	170.0	170.2	3''	139.7	140.0	27.5	164.6	164.6			138.7					
2'''	21.3	21.1	4''	16.0	16.1	12.0	34.7	34.7			16.0					
1''''	167.2	176.3	5''	20.6	20.9	17.2	12.4	12.3			20.8					
2''''	127.6	41.3	6''				19.1	19.1								
3''''	139.4	26.4	1'''		168.5	168.5	169.0	168.6	172.9	172.2						
4''''	15.9	11.7	2'''		127.5	129.1	129.4	129.1	21.1	20.9						
5''''	20.4	16.4	3'''		140.3	139.5	138.9	139.5								
1'''''	165.0	165.0	4'''		16.0	16.2	16.0	16.2								
2'''''	114.9	115.0	5'''		20.6	20.9	20.8	20.9								
3'''''	158.3	158.3	1''''						169.3	168.5						
4'''''	73.7	73.7	2''''						129.4	128.9						
5'''''	19.3	19.3	3''''						138.8	140.1						
6'''''	15.4	15.4	4''''						16.0	16.3						
7'''''	170.4	170.4	5''''						20.9	20.9						
8'''''	21.2	21.1														

^a Data were measured in CDCl₃. ^b Data were measured in CD₃OD.

[δ_{H} 4.87 (1H, s, H-14a), 5.27 (1H, s, H-14b); δ_{C} 137.9 (C-10), 115.5 (C-14)], which accounted for 10 of 12 indices of hydrogen deficiency, as a result, compound **1** was proposed as having a dual-ring structure.

The ^1H - ^1H COSY experiment of **1** showed clear signals for two main structural sequences, $-\text{CH}(\text{H}-1)-\text{CH}(\text{H}-2)-\text{CH}(\text{H}-3)-\text{CH}(\text{H}-4)-\text{CH}(\text{H}-5)-\text{CH}(\text{H}-6)-\text{CH}_3(\text{H}-15)-$ and $-\text{CH}(\text{H}-7)-\text{CH}(\text{H}-8)-\text{CH}(\text{H}-9)-$ (Fig. 2), which were further extended by the following HMBC correlations (Fig. 2): H-1, H-9/C-14 and H-13/C-11. The above evidences suggested that **1** was an oplopane-type sesquiterpenoid.¹² The attachment of one (2-methylbutanoyl) oxy, two acetoxy, one angeloyloxy, and one (4-acetoxy-4-methylseneciyl)oxy groups could also be determined by the HMBC correlations of H-2/C-1', H-3/C-1'', H-4/C-1''', H-8/C-1''', and H-9/C-1''', respectively.

The relative configuration of the dual-ring moiety of **1** was established by the NOESY correlations (Fig. 2) and ^1H NMR coupling constants. When H-1 is assumed to be α -oriented,

obvious NOESY correlations of H-3/H-1, H-1/H-7, and H-7/H-5 indicated that H-1, H-3, H-5, and H-7 were all oriented on the same side of the molecule and were assigned to be α -oriented. H-2 should be α -equatorial, based on the small coupling constant of H-1/H-2 ($J_{1,2} = 3.2$ Hz), while H-6 should be β -axial, since the large coupling constant of H-1/H-6 ($J_{1,6} = 11.5$ Hz) was observed. For another, NOESY correlations of H-6/H-8 revealed that H-8 was β -oriented, and H-9 must be β -equatorial due to the small coupling constant ($J_{8,9} = 3.2$ Hz) observed. The absolute configuration of **1** was established as 1*R*,2*S*,3*R*,4*R*,5*S*,6*R*,7*S*,8*R*,9*S*,11*S*,2'*S*,2'''*Z*,2''''*E*,4''''*S* by single-crystal X-ray diffraction using Cu K α radiation (Fig. 3). Thus, the chemical structure of **1** was established as shown and given the name pararunine A.

Pararunine B (**2**), a yellow oil, showed a molecular formula of C₃₄H₄₈O₁₃ based on its HRESIMS and ^{13}C NMR data. Its ^1H and ^{13}C NMR data (Tables 1 and 2) resembled those of compound **1**, which implied that **2** was also an oplopane-type



Table 2 ^1H NMR data for compounds 1, 2 and 5–9 (400 MHz, J in Hz)

No.	1 ^a	2 ^a	No.	5 ^a	6 ^a	7 ^b	8 ^b	9 ^b
1	2.52 dd (11.5, 3.2)	2.50 dd (11.9, 2.9)	1	5.25 dd (11.7, 2.7)	5.48 dd (11.3, 3.2)	5.25 dd (4.5, 1.5)	5.35 dd (4.4, 1.2)	5.26 dd (4.5, 1.4)
2	5.69 dd (4.4, 3.2)	5.62 dd (4.2, 2.9)	2	3.86 d (2.7)	3.79 d (3.2)	5.35 d (4.5)	5.36 d (4.4)	5.35 d (4.5)
3	5.56 dd (10.5, 4.4)	5.54 dd (10.3, 4.2)	4	3.88 d (2.8)	5.38 d (3.2)	3.26 dd (3.9, 1.8)	3.25 d (5.1)	3.26 dd (4.9, 1.9)
4	5.23 m	5.18 m	5	4.40 dd (11.7, 2.8)	4.17 dd (11.3, 3.2)	2.12 m	2.09, overlap	2.12 overlap
5	2.76 overlap	2.74 overlap	6	3.06 t (11.7)	2.71 t (11.3)	2.65 dd (10.9, 7.1)	2.61 dd (11.5, 6.5)	2.65 dd (11.3, 7.0)
6	2.01 overlap	2.00 overlap	8	5.14 overlap	4.83 m	5.48 dd (11.0, 2.3)	4.07 dd (8.5, 5.0)	5.47 dd (10.7, 2.0)
7	1.80 m	1.78 m	9	2.29 m	2.23 m, 2.60 m	1.54 m, 2.07 m	1.83 m, 2.06 m	1.55 m, 2.05 m
8	5.18 overlap	5.08 m	10	5.13 overlap	5.16 dd overlap	3.36 dd (10.5, 1.3)	4.84 overlap	3.36 dd (10.7, 1.1)
9	5.71 brd (3.2)	5.68 brd (3.1)	12	1.66 s	1.69 s	1.15 s	1.17 s	1.15 s
12	2.74 d (4.0), 2.83 d (4.0)	2.75 d (3.7), 2.83 d (3.7)	13	1.59 s	1.64 s	1.17 s	1.19 s	1.17 s
13	1.25 s	1.21 s	14	5.26 s, 5.33 s	5.15 s, 5.19 s	5.21 s, 5.01 s	4.99 s, 5.06 s	5.02 s, 5.21 s
14	4.87 s, 5.27 s	4.87 s, 5.26 s	15	1.60 s	1.43 s	1.28 s	1.28 s	1.28 s
15	1.46 d (6.7)	1.44 d (6.8)	2'		2.03 s	2.01 s	2.02 s	2.00 s
2'	2.43 m	2.13 s	3'	6.07 qq (7.3, 1.2)				
3'	1.49 m, 1.78 m		4'	1.98 dq (7.3, 1.2)				
4'	0.98 t (7.2)		5'	1.87 q (1.2)				
5'	1.21 d (7.2)		2''			2.36 m	5.65 m	5.65 m
2''	2.04 s	2.08 s	3''	6.13 qq (7.3, 1.2)	6.14 qq (7.2, 1.4)	1.46 m, 1.69 m		
2'''	2.09 s	2.16 s	4''	1.98 dq (7.3, 1.2)	2.05 dq (7.2, 1.4)	0.91 t (7.5)	2.20 q (7.4)	2.21 q (7.4)
2''''		2.39 m	5''	1.89 q (1.2)	1.95 q (1.4)	1.14 d (7.0)	1.09 t (7.4)	1.08 t (7.4)
3''''	6.09 qq (7.0, 1.4)	1.46 m, 1.76 m	6''				2.16 s	2.16 s
4''''	1.98 dq (7.0, 1.4)	0.89 t (7.4)	3'''		6.14 qq (7.2, 1.4)	6.13 qq (7.3, 1.5)	6.12 qq (7.3, 1.4)	6.13 qq (7.3, 1.5)
5''''	1.88 q (1.4)	1.15 d (7.0)	4'''		1.97 dq (7.2, 1.4)	1.98 dq (7.3, 1.4)	1.98 dq (7.3, 1.4)	1.98 dq (7.3, 1.5)
2''''	5.81 brs	5.82 brs	5'''		1.88 q (1.4)	1.91 q (1.5)	1.91 q (1.4)	1.91 q (1.5)
4''''	5.23 q (6.7)	5.24 overlap						
5''''	1.33 d (6.7)	1.34 d (6.8)						
6''''	2.08 brs	2.09 brs						
8''''	1.97 s	1.98 s						

^a Data were measured in CDCl_3 . ^b Data were measured in CD_3OD .

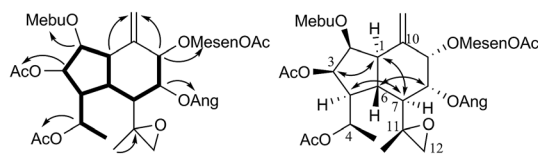


Fig. 2 Key HMBC (\rightarrow), ^1H - ^1H COSY (thick lines) and NOESY (\leftrightarrow) correlations of compound 1.

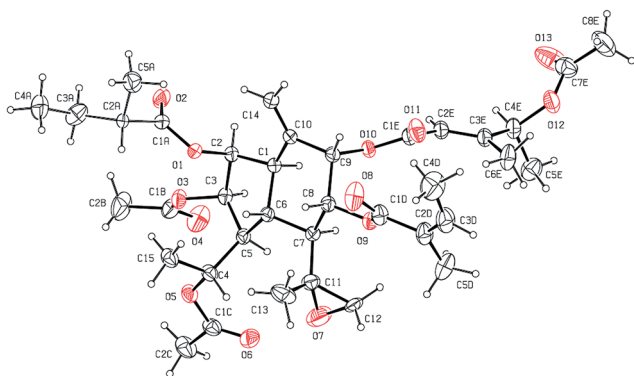


Fig. 3 ORTEP drawing of compound 1.

sesquiterpenoid, with the differences being the types and positions of some substituent groups. The three acetoxy, one (2-methylbutanoyl)oxy, and one (4-acetoxy-4-methylseneciyl)oxy groups of 2 were located at C-2, C-3, C-4, C-8, and C-9 *via* the HMBC correlations of H-2/C-1', H-3/C-1'', H-4/C-1''', H-8/C-1''', and H-9/C-1''', respectively.

The relative configuration of the dual-ring moiety of 2 was deduced using the same methods as those applied for 1. Thus, the structure of 2 was defined as shown, named pararunine B.

Compounds 3 and 4 were respectively confirmed as 2 β ,8 α -dimethylbutyryloxy-3 β ,4-diacetoxy-9 α -[(4''''-methylseneciyl)oxy]-11,12-epoxyoplop-10(14)-en,¹³ and 2 β ,8 α -dimethylbutyryloxy-3 β ,4-diacetoxy-9 α -[(4''''-acetoxy-4''''-methylseneciyl)oxy]-11,12-epoxyoplop-10(14)-en,¹⁴ by comparing their spectroscopic data with published literature data.

Bisabolane-type sesquiterpenoids

Pararunine C (5) was obtained as a yellow oil. The molecular formula $\text{C}_{25}\text{H}_{38}\text{O}_8$ (7 degrees of unsaturation) was established by HRESIMS analysis and NMR spectroscopic data (Tables 1 and 2). Two angeloyloxy groups were present in 5, as supported by the observed ^1H and ^{13}C NMR data: [δ_{H} 6.07 (1H, qq, $J = 7.3$, 1.2 Hz, H-3'), 1.98 (3H, dq, $J = 7.3$, 1.2 Hz, H-4'), 1.87 (3H, q, $J = 1.2$ Hz, H-5'); δ_{C} 167.2 (C-1'), 127.8 (C-2'), 139.0 (C-3'), 16.0 (C-4'),



20.6 (C-5''), and [δ_{H} 6.13 (1H, qq, $J = 7.3, 1.2$ Hz, H-3''), 1.98 (3H, dq, $J = 7.3, 1.2$ Hz, H-4''), 1.89 (3H, q, $J = 1.2$ Hz, H-5''); δ_{C} 168.6 (C-1''), 127.8 (C-2''), 139.7 (C-3''), 16.0 (C-4''), 20.6 (C-5'')]. Apart from the two angeloyloxy groups, the ^{13}C NMR and DEPT spectra of **5** showed fifteen more carbon signals, consisting of two double bonds, one quaternary carbon, three methyl groups, one methylene group, and six methine groups including five oxygenated ones. The aforementioned data were similar to those acquired for the known compound pararubin A, which was previously isolated from the same plant. The ^1H - ^1H COSY correlations (Fig. 4) of **5** exhibited two main structural sequences, $-\text{CH}(\text{H}-2)-\text{CH}(\text{H}-1)-\text{CH}(\text{H}-6)-\text{CH}(\text{H}-5)-\text{CH}(\text{H}-4)-$ and $-\text{CH}(\text{H}-8)-\text{CH}_2(\text{H}-9)-\text{CH}(\text{H}-10)-$, which were connected to disclose the presence of a bisabolane sesquiterpenoid skeleton by the following HMBC correlations (Fig. 4): H-1, H-5/C-7; H-2, H-4/C-15; H-6, H-8/C-14; H-10/C-12; and H-12/C-13. Furthermore, according to the HMBC correlations of H-1/C-1' and H-8/C-1'', the positions of the two angeloyloxy groups were attached to C-1 and C-8, respectively.

5 shared the same relative configuration of the cyclohexane ring with pararubin A,¹¹ which was established by the ^1H NMR coupling constants: when H-6 is assumed to be β -oriented, H-1 and H-5 should be α -axial, based on the large coupling constants of H-1/H-6 ($J_{1,6} = 11.7$ Hz) and H-5/H-6 ($J_{5,6} = 11.7$ Hz). H-2 and H-4 must be α -equatorial due to the small coupling constants ($J_{1,2}, J_{4,5} = 2.7$ Hz). Thus, the structure of **5** was established as shown and given the name pararubine C.

Pararubine D (**6**), a yellow oil, possessed a molecular formula of $\text{C}_{27}\text{H}_{40}\text{O}_9$ according to the ^{13}C NMR and HRESIMS data. The ^1H and ^{13}C NMR data (Tables 1 and 2) were similar to those of **5**. However, one more acetoxy group was observed, indicating that **6** was a bisabolane-type sesquiterpenoid with an acetoxy and two angeloyloxy groups, the positions of which were determined by the HMBC correlations of H-1/C-1', H-4/C-1'', and H-8/C-1''', respectively. Using the same method applied in compound **5** permitted the assignment of the relative configuration of **6**. The structure of **6** was therefore elucidated, and named pararubine D.

Pararubine E (**7**), a yellow oil, possessed a molecular formula of $\text{C}_{27}\text{H}_{42}\text{O}_9$ based on its ^{13}C NMR and HRESIMS data. The ^1H and ^{13}C NMR data (Tables 1 and 2) of **7** showed the presence of one acetoxy group [δ_{H} 2.01 (3H, s, H-2'); δ_{C} 172.2 (C-1'), 20.9 (C-2')], one (2-methylbutanoyl)oxy group [δ_{H} 2.36 (1H, m, H-2''), 1.46 (1H, m, H-3''), 1.69 (1H, m, H-3''), 0.91 (3H, t, $J = 7.5$ Hz, H-4''), 1.14 (3H, d, $J = 7.0$ Hz, H-5''); δ_{C} 177.0 (C-1''), 42.5 (C-2''), 27.5 (C-3''), 12.0 (C-4''), 17.2 (C-5'')], one angeloyloxy group [δ_{H} 6.13 (1H, qq, $J = 7.3, 1.5$ Hz, H-3'''), 1.98 (3H, dq, $J = 7.3, 1.5$ Hz, H-4'''), 1.91 (3H, m, H-5'''); δ_{C} 168.5 (C-1'''), 129.1 (C-2'''), 139.5

(C-3'''), 16.2 (C-4'''), 20.9 (C-5''')], and one epoxy group [δ_{H} 3.26 (1H, m, H-4); δ_{C} 58.2 (C-3), 61.4 (C-4)]. The 1D and 2D NMR data of **7** showed similarity to those of 2 β ,8-diangeloyloxy-3 β ,4 β -epoxy-1 β ,10,11-trihydroxybisabol-7(14)-ene,¹⁵ which was a bisabolane-type sesquiterpene derived from the roots of *Ligularia cymbulifera*. The difference between them lies primarily in the positions of an acetoxy group, a (2-methylbutanoyl)oxy group, and an angeloyloxy group, which were deduced from the HMBC correlations of H-1/C-1', H-2/C-1'', and H-8/C-1''', respectively.

The relative configuration of the six-membered ring in **7** was established from the ^1H NMR coupling constants and the NOESY correlations (Fig. 5). When H-6 is assumed to be β -oriented, H-1 should be β -oriented due to the small coupling constant of H-1/H-6 ($J_{1,6} = 1.5$ Hz). H-2 was also determined to be β -oriented by the NOESY correlation between H-2 and H-6. Considering the fact that the six-membered ring should have a twist-boat conformation because of the epoxy group, H-4 was supposed to have a β -equatorial position due to the NOESY correlation between H-4 and H-6. H-15 was deduced in β -oriented position based on the NOESY correlations of H-2, H-4/Me-15. Thus, the relative configuration within the cyclohexane ring was established and the structure of **7** was then defined.

Pararubine F (**8**), a yellow oil, exhibited a molecular formula of $\text{C}_{28}\text{H}_{42}\text{O}_9$ as determined by the HRESIMS and ^{13}C NMR data, requiring 8 indices of hydrogen deficiency. The NMR spectral data (Tables 1 and 2) of **8** were similar to those of **7**, except that a (2-methylbutanoyl)oxy group in **7** was replaced by a (4-methylseneciyl)oxy group in **8**, and the angeloyloxy group in **8** was attached at C-10 by the HMBC correlations of H-10/C-1'''. The relative configuration of the six-membered ring in **8** was determined using the same methods as that applied for **7**. Thus, the structure of **8** was elucidated as shown.

Pararubine G (**9**) appeared as a yellow oil. Its molecular formula was established as $\text{C}_{28}\text{H}_{42}\text{O}_9$, which was the same as **8**, by the HRESIMS and ^{13}C NMR data, requiring 8 indices of hydrogen deficiency. The NMR spectral data (Tables 1 and 2) of **9** were generally similar to those of **8**, with the only difference being that the angeloyloxy group in **9** was attached at C-8 by the HMBC correlations of H-8/C-1'''. The relative configuration of the six-membered ring in **9** was established by methods similar to those used for **7**. Therefore, the structure of **9** was deduced as shown.

Pararubine H (**12**), a yellow oil, was assigned the molecular formula of $\text{C}_{29}\text{H}_{44}\text{O}_{12}$ with 8 indices of hydrogen deficiency according to the HRESIMS and ^{13}C NMR data. The 1D NMR data (Tables 1 and 3) of **12** were very similar to those of 4 α -acetyl-2 β ,5 α ,8-triisobutyryl-1 β ,3 α ,10,11-tetrahydroxybisabolene,¹⁶

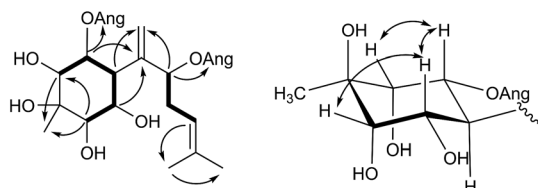


Fig. 4 Key HMBC (\rightarrow), ^1H - ^1H COSY (thick lines) and NOESY (\leftrightarrow) correlations of compound **5**.

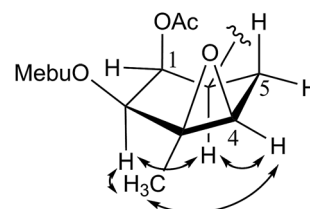


Fig. 5 Key NOESY (\leftrightarrow) correlations of compound **7**.



Table 3 ^1H NMR data for compounds **12**–**18** (400 MHz, J in Hz)

No.	12^b	13^b	14^b	No.	15^a	16^a	17^a	18^a
1	5.42 dd (3.9, 2.7)	5.24 overlap	4.07 d (10.9, 10.2)	3	6.26 d (9.5)	6.26 d (9.5)	6.26 d (9.6)	6.23 d (9.5)
2	4.95 d (3.9)	4.90 d (3.7)	5.18 d (10.2)	4	7.63 d (9.5)	7.63 d (9.5)	7.63 d (9.6)	7.62 d (9.5)
4	5.35 d (3.0)	5.36 d (2.8)	3.61 dd (3.5, 1.7)	5	7.37 d (8.5)	7.37 d (8.6)	7.37 d (8.5)	7.35 d (8.6)
5	4.60 dd (11.5, 3.0)	4.59 dd (11.3, 2.8)	1.92 overlap	6	6.81 dd (8.5, 2.3)	6.86 dd (8.6, 2.3)	6.82 dd (8.5, 2.4)	6.79 dd (8.6, 2.4)
6	2.99 dd (11.5, 2.7)	3.02 dd (11.3, 2.6)	2.58 ddd (11.0, 10.9, 5.2)	8	6.79 d (2.3)	6.82 d (2.3)	6.80 d (2.4)	6.74 d (2.4)
8	4.08 dd (8.9, 5.0)	5.59 dd (9.5, 3.8)	5.34 m	1'	4.13 m	4.15 m	4.15 m	4.11 m
9	1.78 m, 2.11 m	1.57 m, 2.07 overlap	1.87 m	2'	2.02 m	2.11 m	2.02 m	2.06 m
10	4.96 overlap	3.34 overlap	3.42 m	4'	3.45 s	3.73 s	3.67 s	3.41 s
12	1.11 s	1.13 s	1.16 s	6'	2.47 d (6.9)	3.52 s	2.74 d (4.4)	6.31 s
13	1.11 s	1.11 s	1.17 s	7'	2.20 m			
14	5.16 s, 5.19 s	5.24 s, 5.43 s	5.07 s, 5.17 s	8'	0.95 d (4.5)	1.40 s	1.30 s	2.19 s
15	1.11 s	1.11 s	1.23 s	9'	0.97 d (4.5)	1.49 s	1.28 s	1.95 s
3'	6.19 qq (7.3, 1.4)	6.21 qq (7.5, 1.4)	6.08 qq (7.2, 1.3)	10'	1.52 s	1.54 s	1.53 s	1.50 s
4'	1.96 dq (7.3, 1.4)	2.04 dq (7.5, 1.4)	1.98 dq (7.2, 1.3)	OCH ₃			3.24 s	
5'	1.87 q (1.4)	1.97 q (1.4)	1.97 q (1.3)					
2''	2.02 s	2.00 s						
3''			6.14 qq (7.2, 1.3)					
4''			1.98 dq (7.2, 1.3)					
5''			1.93 q (1.3)					
2'''	2.06 s	2.03 s						
3'''	6.11 qq (7.3, 1.4)	6.16 qq (7.5, 1.4)						
4'''	1.98 dq (7.3, 1.4)	2.00 dq (7.5, 1.4)						
5'''	1.91 q (1.4)	1.89 q (1.4)						

^a Data were measured in CDCl₃. ^b Data were measured in CD₃OD.

except for the different substituent groups. Accordingly, **12** was presented to be a bisabolane-type sesquiterpenoid with two acetoxy and two angeloyloxy groups, the positions of which were determined by the HMBC correlations of H-1/C-1', H-2/C-1'', H-4/C-1''', and H-10/C-1''', respectively.

The relative configuration of the six-membered ring in **12** was identical to that of 4 α -acetyl-2 β ,5 α ,8-triisobutyl-1 β ,3 α ,10,11-tetrahydroxybisabolene on the basis of its ^1H NMR coupling constants and NOESY correlations. When H-6 is assumed to be in a β -orientation, H-1 should be β -equatorial due to the small coupling constant of H-1/H-6 ($J_{1,6} = 2.7$ Hz), H-5 must be α -axial due to the large coupling constant of H-5/H-6 ($J_{5,6} = 11.5$ Hz), and H-4 should be α -equatorial due to the small coupling constant of H-4/H-5 ($J_{4,5} = 3.0$ Hz). The NOESY correlation of H-2/H-6 indicated a β -orientation for H-2. **12** was therefore elucidated as shown.

Pararunine I (**13**), obtained as a yellow oil, was designated with a molecular formula of C₂₉H₄₄O₁₂ with 8 indices of hydrogen deficiency on the basis of its HRESIMS and ^{13}C NMR data. The NMR data (Tables 1 and 3) of **13** were similar to those of **12**, indicating that **13** was also a bisabolane-type sesquiterpenoid with two acetoxy and two angeloyloxy groups. The only difference between the two compounds was the position of an angeloyloxy group, which was attached at C-8 in **13** instead of C-10 in **12** by the HMBC correlations of H-8/C-1'''. The ^1H NMR coupling constants and NOESY correlations of **13** showed that its relative configuration was identical to that of **12**. Thus, the structure of **13** was constructed as shown.

Pararunine J (**14**), a yellow oil, exhibited a molecular formula of C₂₅H₄₀O₉ as determined by the HRESIMS and ^{13}C NMR data, requiring 6 indices of hydrogen deficiency. Analysis of its 1D and 2D NMR data revealed that **14** was also a bisabolane-type sesquiterpenoid with two angeloyloxy groups, which were attached to C-2 and C-8, respectively, according to the HMBC correlations. The relative configuration of the six-membered ring system in **14** could be proposed by analysis of ^1H NMR coupling constants and NOESY data. H-6 is assumed to be β -oriented, then H-1 should be α -axial, and H-2 should be β -axial based on the large coupling constants of H-1/H-6 ($J_{1,6} = 10.9$ Hz), and H-1/H-2 ($J_{1,2} = 10.2$ Hz). H-4 must be α -equatorial due to the small coupling constants of H-4/H-5 ($J_{4,5} = 1.7, 3.5$ Hz). Therefore, the structure of **14** was defined as shown.

The structures of the known compounds **10** and **11** were respectively confirmed as 1 α -acetoxy-2 α ,8-diangeloyloxy-3 α ,4 α -epoxy-10,11-dihydroxybisabol-7(14)-ene,¹⁷ and 1 α -acetoxy-2 α ,10-diangeloyloxy-3 α ,4 α -epoxy-8,11-dihydroxybisabol-7(14)-ene,¹⁸ by comparing their spectroscopic data with published literature data.

Monoterpenoid coumarins

An examination of the ^1H NMR data (Table 3) in the aromatic region of compounds **15**–**18** revealed a pair of doublets at *ca.* 6.26, 7.63 (each 1H, d, $J = 9.5$ Hz) and an ABX pattern signals at *ca.* 6.79 (1H, d, $J = 2.3$ Hz), 6.82 (1H, dd, $J = 8.5, 2.3$ Hz), and 7.37 (1H, d, $J = 8.5$ Hz) corresponding to 7-substituted coumarin moieties. Accordingly, the maximal absorption of the



UV spectra (*ca.* 258 and 320 nm) of compounds **15–18** also indicated the presences of 7-oxygenated coumarin skeletons. Besides, the combined analysis of 2D NMR of **15–18** revealed the structures of their different side chains to be similar acyclic monoterpenoid moieties. The experimental electronic circular dichroism (ECD) data of **15–18** were collected to determine their absolute configurations, but unfortunately, no valuable calculated ECD data was obtained due to the flexible linear side chain of each compound. In addition, as far as we are aware, no effective method has been established to determine the absolute configurations of this type of monoterpenoid coumarins.

Pararubcoumarin A (**15**) was obtained as a yellow oil, and the molecular formula, $C_{19}H_{22}O_5$, was determined based on its NMR data (Tables 1 and 3) and the HRESIMS analysis, which indicated 11 indices of hydrogen deficiency. Apart from the 7-substituted coumarin moiety, there were ten more carbons for a side chain, which included one ketone carbonyl group [δ_C 205.7 (C-5')], three methyl groups [δ_H 0.95 (3H, d, $J = 4.5$ Hz, H-8'), 0.97 (3H, d, $J = 4.5$ Hz, H-9'), 1.52 (3H, s, H-10'); δ_C 22.8 (C-8'), 22.8 (C-9'), 22.7 (C-10')], three methylene groups [δ_H 4.13 (2H, m, H-1'), 2.02 (2H, s, H-2'), 2.47 (2H, d, $J = 6.9$ Hz, H-6'); δ_C 65.3 (C-1'), 31.5 (C-2'), 50.4 (C-6')], two methine groups [δ_H 3.45 (1H, s, H-4'), 2.20 (1H, s, H-7'); δ_C 65.5 (C-4'), 24.6 (C-7')], and a quaternary carbon [δ_C 62.2 (C-3')]. Analysis of its 1H - 1H COSY spectrum revealed the assignment of two spin systems, $-CH_2(H-1')-CH_2(H-2')-$ and $-CH_2(H-6')-CH(H-7')-CH_3(H-8')-CH_3(H-9')-$ (Fig. 5), which were connected to form an acyclic monoterpenoid moiety by the HMBC correlations (Fig. 6) of H-1', H-4'/C-3'; H-10'/C-2'; H-10'/C-4'; and H-4', H-6'/C-5'. Furthermore, to accommodate 9 indices of hydrogen deficiency (the 7-substituted coumarin moiety and one ketone carbonyl group

accounted for 8 of those), compound **15** was proposed to have an epoxy group [δ_H 3.45 (1H, s, H-4'); δ_C 62.2 (C-3'), 65.5 (C-4')]. Thus, the structure of **15** was determined as 7-[(3',7'-dimethylocta-3',4'-epoxy-5'-one)oxy]coumarin, named pararubcoumarin A.

Pararubcoumarin B (**16**), isolated as a yellow oil, has the molecular formula of $C_{19}H_{20}O_6$ as determined by the HRESIMS and ^{13}C NMR data, suggesting 10 indices of hydrogen deficiency. The 1D NMR data (Tables 1 and 3) of **16** were similar to those of **15**, except for the presence of an extra epoxy group [δ_H 3.52 (1H, s, H-6'); δ_C 65.1 (C-6'), 62.1 (C-7')], the position of which was determined by the HMBC correlations of H-6'/C-5' and H-8'/C-6'. Thus, the structure of **16** was defined as shown, given the name pararubcoumarin B.

Pararubcoumarin C (**17**), a yellow oil, has the molecular formula of $C_{20}H_{24}O_6$ as established by the ^{13}C NMR and HRESIMS data, corresponding to 9 indices of hydrogen deficiency. The NMR data (Tables 1 and 3) of **17** also resembled those of **15**, with the only difference being the presence of an extra methoxy group at C-7' in **17**, which was confirmed by the HMBC correlation of MeO-7'/C-7'. Thus, the structure of **17** was determined as shown, named pararubcoumarin C.

Pararubcoumarin D (**18**) was isolated as a yellow oil. Its molecular formula was established as $C_{19}H_{20}O_5$, with 10 indices of hydrogen deficiency by the HRESIMS and ^{13}C NMR data. Comparison of the NMR data (Tables 1 and 3) of **18** with those of **15** displayed the major difference being the presence of an extra $\Delta^{6',7'}$ double bond in **18**, which was confirmed by the HMBC correlations H-8', H-4'/C-6', given the name pararubcoumarin D.

The known compound **19** was defined as 7-[(3'E)-(3',7'-dimethylocta-3',6'-dien-5'-one)oxy]coumarin,¹⁹ by comparing its spectroscopic data with published literature data.

It was reported that some highly oxygenated bisabolane-type sesquiterpenoids possess potential antimicrobial or cytotoxic activities^{15,20,21} and some highly oxygenated oplopane-type sesquiterpenoids show varying degrees of cytotoxic activity.²² Therefore, all isolates were evaluated for their *in vitro* cytotoxicity against B16 mouse melanoma, HepG2 human hepatocellular carcinoma, and MCF7 human breast adenocarcinoma cell

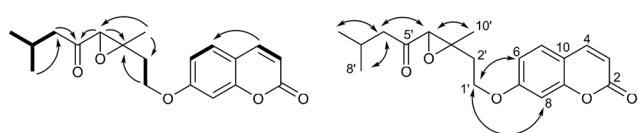


Fig. 6 Key HMBC (\rightarrow), 1H - 1H COSY (thick lines) and NOESY (\leftrightarrow) correlations of compound **15**.

Table 4 Cytotoxicity of compounds **1–19** against B16, HepG2 and MCF7 cell lines^a

Compound (100 μ M)	Cell viability (%)			Compound (100 μ M)	Cell viability (%)		
	B16	HepG2	MCF7		B16	HepG2	MCF7
1	52.11 \pm 2.72	66.34 \pm 3.80	77.53 \pm 1.07	10	78.54 \pm 1.96	69.64 \pm 2.86	59.22 \pm 4.74
2	71.12 \pm 5.66	63.50 \pm 0.66	72.47 \pm 3.43	11	80.54 \pm 0.69	74.20 \pm 1.46	56.60 \pm 3.68
3	70.82 \pm 3.21	58.58 \pm 1.94	61.05 \pm 6.38	12	60.42 \pm 1.78	70.39 \pm 1.06	76.47 \pm 2.52
4	78.04 \pm 5.98	49.20 \pm 1.78	64.83 \pm 5.42	13	52.11 \pm 2.72	66.34 \pm 3.80	77.53 \pm 1.07
5	65.63 \pm 2.17	60.47 \pm 2.29	49.38 \pm 4.41	14	54.59 \pm 1.73	73.24 \pm 7.57	66.42 \pm 4.07
6	64.74 \pm 5.36	55.73 \pm 3.53	73.43 \pm 3.65	15	62.67 \pm 0.74	69.02 \pm 3.58	51.25 \pm 4.16
7	84.41 \pm 1.54	76.21 \pm 0.38	67.67 \pm 5.14	16	57.15 \pm 1.04	79.94 \pm 2.31	67.23 \pm 1.02
8	51.05 \pm 1.42	76.18 \pm 8.69	58.16 \pm 4.81	17	59.45 \pm 0.99	75.27 \pm 1.26	58.87 \pm 3.78
9	86.78 \pm 2.19	73.23 \pm 3.94	62.59 \pm 7.26	18	26.40 \pm 0.73	38.30 \pm 1.07	22.54 \pm 1.46
Doxorubicin	18.89 \pm 2.07	21.33 \pm 0.46	23.53 \pm 0.52	19	77.38 \pm 3.23	87.66 \pm 0.29	87.30 \pm 1.84

^a The data (cell viability, measured by the MTT assay) were normalized and expressed as a percentage of the control group, which is set to 100%. Doxorubicin was used as a positive control.



lines, as well as their antimicrobial effects against *Staphylococcus aureus*, *Escherichia coli*, and *Monilia albicans*. However, none of them showed any appreciable cytotoxic (Table 4) or antimicrobial effect.

Conclusions

Previous phytochemical research on *Parasenecio* demonstrated that eremophilane-type sesquiterpenoids accounted for the characteristic constituents of this genus.^{3–8} However, in our previous¹¹ and present studies on *P. rubescens*, with the identifications of nearly three dozen bisabolane-type sesquiterpenoids, not a single one of eremophilane was found. Also it is noteworthy that compounds isolated from *P. rubescens* tend to be highly oxygenated. Our current research indicates that highly oxygenated bisabolane-type and oplopane-type sesquiterpenoids, together with monoterpenoid coumarins, may play important roles in the chemotaxonomic significance of *P. rubescens* as well as the genus *Parasenecio*.

Experimental section

General experimental procedures

Optical rotations were measured by a PerkinElmer model 341 polarimeter. UV spectra were obtained on a Jasco J-810 spectropolarimeter. IR spectra were recorded on a Bruker VERTEX 70 FT-IR microscopic spectroscopy. NMR experiments were carried out on a Bruker-AM-400 spectrometer. X-ray crystallographic analysis was collected on a Bruker APEX DUO diffractometer using Cu K α radiation. HRESIMS spectra were obtained from a Thermo Scientific LTQ-Orbitrap XL mass spectrometer. Column silica gel (200–300 or 300–400 mesh; Qingdao Marine Chemical Inc.), Sephadex LH-20 gel (GE Healthcare), and MCI gel (CHP20P, 75–150 μ m; Mitsubishi Chemical Industries Ltd.) were used for column chromatography. HPLC was conducted on an Agilent 1260 system using YMC-Pack ODS-A HPLC column (250 mm \times 10 mm i.d.) and YMC-Pack SIL-06 HPLC column (250 mm \times 10 mm i.d.).

Plant material

Whole plants of *P. rubescens* were collected in the Lushan mountains of Jiangxi Province, P. R. China, in September 2012. The plant was identified by research scientist Bei-Li Huang (Lushan Botanical Garden). A voucher specimen (ID 20120901) was deposited in the Herbarium of Materia Medica, Faculty of Pharmacy, Tongji Medical College of Huazhong University of Science and Technology, Wuhan, P. R. China.

Extraction and isolation

The air-dried whole plants of *P. rubescens* (18.4 kg) were extracted with 90% acetone (5 \times 50 L) at room temperature and concentrated *in vacuo* to give 10 L of extract. The solution was sequentially partitioned with PE and EtOAc. The EtOAc-soluble fraction (180.0 g) was subjected to a silica gel column, eluting with PE/acetone (20 : 1–1 : 10 step gradient elution), to obtain five major fractions (A–E). Fraction A (35.0 g) was decolorized on

an MCI gel column (MeOH/H₂O, from 75 to 100%) to afford three fractions, A1–A3, which were chromatographed on a Sephadex LH-20 column (MeOH) separately. Fraction A1.2 was subjected to silica gel column (PE/acetone, from 20 : 1 to 1 : 10) and semipreparative HPLC (MeOH/H₂O, 7 : 3) to yield **3** (5.7 mg, t_R 28.1 min). Fraction A2.2 was subjected to semipreparative HPLC (MeOH/H₂O, 7 : 3) to yield **15** (5.8 mg, t_R 27.2 min) and **19** (6.1 mg, t_R 31.6 min), while A2.3 was also subjected to semipreparative HPLC (MeOH/H₂O, 7 : 3) to yield **16** (1.5 mg, t_R 30.5 min), **17** (2.2 mg, t_R 40.7 min), and **18** (4.3 mg, t_R 35.3 min). Fraction A3.1 was separated on ODS silica gel column (MeOH/H₂O, from 30 to 100%) and semipreparative HPLC (MeOH/H₂O, 7 : 3) to yield **11** (5.7 mg, t_R 41.8 min), and similarly, fraction A3.2 was purified in the same way as A3.1 to yield **1** (5.2 mg, t_R 38.8 min), **2** (3.7 mg, t_R 45.8 min), and **4** (19.0 mg, t_R 31.2 min). Fraction B (62.3 g) was decolorized on an MCI gel column (MeOH/H₂O, from 75% to 100%) to afford three fractions, B1–B3. Fraction B3 was fractionated using a Sephadex LH-20 column (MeOH) to produce four subfractions, B3.1–B3.4. Fraction B3.1 was further resolved by ODS silica gel (MeOH/H₂O, from 30 to 100%), semipreparative HPLC (MeOH/H₂O, 7 : 3), and normal phase HPLC (*n*-hexane/2-propanol, 15 : 1) columns to yield **12** (4.8 mg, t_R 17.3 min), **13** (4.4 mg, t_R 15.4 min), and **14** (2.0 mg, t_R 21.5 min). Using the same methods applied in B3.1, **5** (1.0 mg, t_R 17.8 min), **6** (2.9 mg, t_R 15.8 min), **7** (4.8 mg, t_R 23.6 min), and **9** (4.4 mg, t_R 20.4 min) were afforded from B3.2, and **8** (3.7 mg, t_R 21.7 min), **10** (18.0 mg, t_R 16.3 min) were obtained from B3.3.

Pararunine A (1). Colorless crystals (MeOH); mp 147–149 °C; $[\alpha]_D^{20} +31$ (*c* 0.11, CH₃OH); UV (CH₃OH) λ_{max} (log ϵ) 215 (4.40) nm; IR (KBr) ν_{max} 2969, 2937, 2877, 1738, 1655 cm^{−1}; ¹H and ¹³C NMR data see Tables 1 and 2; HRESIMS m/z 727.3301 [$M + Na$]⁺ (calcd for C₃₇H₅₂O₁₃Na, 727.3306). Crystal data: C₃₇H₅₂O₁₃, M_W = 704.79, orthorhombic, space group *P*2₁2₁2₁, *a* = 12.2069 (2) Å, *b* = 14.3991 (2) Å, *c* = 22.2180 (4) Å, $\alpha = \beta = \gamma = 90.00^\circ$, *V* = 3905.22 (11) Å³, *Z* = 4, ρ (calcd) = 1.199 g cm^{−3}, and μ (Cu K α) = 0.749 mm^{−1}; reflections collected 53 820, independent reflections 6906; final *R* indices [*I* > 2 σ (*I*)] *R*₁ = 0.0520, *wR*₂ = 0.1657; *R* indices (all data) *R* = 0.0532, *wR*₂ = 0.1686; largest diff. peak and hole 0.494 and −0.290 e Å^{−3}; flack parameter 0.15 (18). Reflections collected: 53 820 (CCDC No. 1472359).†

Pararunine B (2). Yellow oil; $[\alpha]_D^{20} +36$ (*c* 0.08, CH₃OH); UV (CH₃OH) λ_{max} (log ϵ) 215 (4.38); IR (KBr) ν_{max} 2974, 2938, 2879, 1744, 1657 cm^{−1}; ¹H and ¹³C NMR data see Tables 1 and 2; HRESIMS m/z 687.2988 [$M + Na$]⁺ (calcd for C₃₄H₄₈O₁₃Na, 687.2993).

Pararunine C (5). Yellow oil; $[\alpha]_D^{20} -31$ (*c* 0.02, CH₃OH); UV (CH₃OH) λ_{max} (log ϵ) 216 (4.21) nm; IR (KBr) ν_{max} 3508, 1743, 1648 cm^{−1}; ¹H and ¹³C NMR data see Tables 1 and 2; HRESIMS m/z 489.2454 [$M + Na$]⁺ (calcd for C₂₅H₃₈O₈Na, 489.2464).

Pararunine D (6). Yellow oil; $[\alpha]_D^{20} -26$ (*c* 0.06, CH₃OH); UV (CH₃OH) λ_{max} (log ϵ) 217 (4.40) nm; IR (KBr) ν_{max} 3476, 1718, 1647 cm^{−1}; ¹H and ¹³C NMR data see Tables 1 and 2; HRESIMS m/z 531.2570 [$M + Na$]⁺ (calcd for C₂₇H₄₀O₉Na, 531.2563).

Pararunine E (7). Yellow oil; $[\alpha]_D^{20} -37$ (*c* 0.31, CH₃OH); UV (CH₃OH) λ_{max} (log ϵ) 216 (4.21) nm; IR (KBr) ν_{max} 3508, 1743, 1648 cm^{−1}; ¹H and ¹³C NMR data see Tables 1 and 2; HRESIMS m/z 533.2722 [$M + Na$]⁺ (calcd for C₂₇H₄₂O₉Na, 533.2727).



Pararunine F (8). Yellow oil; $[\alpha]_D^{20} -23$ (c 0.09, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 217 (4.40) nm; IR (KBr) ν_{max} 3476, 1718, 1647 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 2; HRESIMS m/z 545.2722 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{28}\text{H}_{42}\text{O}_9\text{Na}$, 545.2727).

Pararunine G (9). Yellow oil; $[\alpha]_D^{20} -61$ (c 0.29, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 218 (4.42) nm; IR (KBr) ν_{max} 3501, 1744, 1719, 1648 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 2; HRESIMS m/z 545.2723 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{28}\text{H}_{42}\text{O}_9\text{Na}$, 545.2727).

Pararunine H (12). Yellow oil; $[\alpha]_D^{20} -23$ (c 0.12, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 216 (4.36) nm; IR (KBr) ν_{max} 3459, 1720, 1648 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 607.2724 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{29}\text{H}_{44}\text{O}_{12}\text{Na}$, 607.2730).

Pararunine I (13). Yellow oil; $[\alpha]_D^{20} -29$ (c 0.11, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 216 (4.36) nm; IR (KBr) ν_{max} 3444, 1747, 1721, 1649 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 607.2726 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{29}\text{H}_{44}\text{O}_{12}\text{Na}$, 607.2730).

Pararunine J (14). Yellow oil; $[\alpha]_D^{20} -36$ (c 0.05, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 215 (4.25) nm; IR (KBr) ν_{max} 3408, 1700, 1647 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 507.2562 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{28}\text{H}_{42}\text{O}_9\text{Na}$, 507.2570).

Pararubcoumarin A (15). Yellow oil; $[\alpha]_D^{25} -28$ (c 0.12, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 320 (4.19) nm; IR (KBr) ν_{max} 1732, 1614 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 353.1357 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{22}\text{O}_5\text{Na}$, 353.1365).

Pararubcoumarin B (16). Yellow oil; $[\alpha]_D^{20} -20$ (c 0.03, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 320 (3.88) nm; IR (KBr) ν_{max} 1728, 1614 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 367.1145 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{20}\text{O}_6\text{Na}$, 367.1158).

Pararubcoumarin C (17). Yellow oil; $[\alpha]_D^{20} -20$ (c 0.07, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 320 (4.05) nm; IR (KBr) ν_{max} 1733, 1614 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 383.1460 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{20}\text{H}_{24}\text{O}_6\text{Na}$, 383.1470).

Pararubcoumarin D (18). Yellow oil; $[\alpha]_D^{20} -58$ (c 0.12, CH_3OH); UV (CH_3OH) λ_{max} ($\log \epsilon$) 320 (4.09) nm; IR (KBr) ν_{max} 1734, 1615 cm^{-1} ; ^1H and ^{13}C NMR data see Tables 1 and 3; HRESIMS m/z 351.1201 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{19}\text{H}_{20}\text{O}_5\text{Na}$, 351.1208).

Antimicrobial assays

The microbial strains of *Staphylococcus aureus*, *Escherichia coli* and *Monilia albicans* were purchased from the Basic Medical College of Huazhong University of Science and Technology. The paper disk diffusion method²³ was used as an antimicrobial test for compounds 1–19. About 0.1 mL of each sample (200 $\mu\text{g mL}^{-1}$) was added to a piece of paper (6 mm in diameter). Each disk was dried in air and pasted onto a culture dish at 37 °C for 24 h. Antibacterial activity was measured by the inhibitory zone diameter (IZD). Penicillin G, streptomycin and fluconazole standard antimicrobial susceptibility disks (Hangzhou Microbial Reagent Co., Ltd) were used as positive controls for *Staphylococcus aureus*, *Escherichia coli* and *Monilia albicans*, respectively. Each test was performed in duplicate.

Cell viability assay

The three cancer cell lines including B16 (mouse melanoma cell lines), HepG2 (human hepatocellular carcinoma cell lines) and MCF7 (human breast adenocarcinoma cell lines) were purchased from the cell bank of the Basic Medical College of Huazhong University of Science and Technology. Each of these cell lines were maintained in DMEM containing 10% fetal bovine serum, 100 units per mL penicillin and 100 $\mu\text{g mL}^{-1}$ streptomycin at 37 °C in 5% CO_2 . 90 μL of the cell suspension (1×10^4 cell per mL) was seeded in each well of 96-well cell culture plates for 24 h. 10 μL of tested concentrations of compounds 1–19 were then added to each well. After another 24 h of incubation, each well was treated with 15 μL of MTT solution (5 mg mL^{-1}) for 4 h at 37 °C. For MTT assay, the supernatant was discarded and 100 μL of DMSO was added to each well. The optical density at 570 nm was measured using a microplate reader. Doxorubicin was used as the positive control. All samples were cultured in triplicate. The data (cell viability, measured by MTT assay) were normalized and expressed as a percentage of the control group, which is set to 100%.

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