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Oxygen and carbon dioxide dual gas-responsive homopolymers and diblock copolymers synthesized *via* RAFT polymerization†

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The monomer, 2,2,2-trifluoroethyl 3-(*N*-(2-(diethylamino)ethyl)acrylamido)propanoate (TF-DEAE-AM), which contains both O₂ and CO₂-responsive functionalities, was first synthesized from commercially available *N,N*-diethylethylenediamine, 2,2,2-trifluoroethyl acrylate, and acryloyl chloride. Subsequently, a series of dual-gas responsive polymers, poly(2,2,2-trifluoroethyl 3-(*N*-(2-(diethylamino)ethyl)acrylamido)propanoate) (poly(TF-DEAE-AM)) and poly(ethylene glycol)-*b*-poly(2,2,2-trifluoroethyl 3-(*N*-(2-(diethylamino)ethyl)acrylamido)propanoate) (PEG-*b*-poly(TF-DEAE-AM)), were synthesized by employing reversible addition–fragmentation chain transfer (RAFT) polymerization. Due to the protonation between CO₂ and DEAE groups, and the specific van der Waals interactions between O₂ and C–F bonds, micelles consisting of poly(TF-DEAE-AM) or PEG-*b*-poly(TF-DEAE-AM) display distinct CO₂ and O₂ responsiveness in aqueous media. Pyrene, which is a model hydrophobic drug, is able to be effectively encapsulated in the micelles based on PEG-*b*-poly(TF-DEAE-AM). It is found that the release of pyrene sharply increases after bubbling CO₂ or O₂ compared to N₂, and the release rate of the solution bubbling with CO₂ is the fastest. Since both CO₂ and O₂ are key gases for the human body, the CO₂- and O₂-induced release property of poly(TF-DEAE-AM) may open opportunities for the preparation of functional materials with special CO₂ and O₂ responsiveness for potential applications in biomedicine.

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Introduction

Environmentally responsive polymers have attracted significant attention in the last decade due to their prospective applications in biomedicine, nano-therapeutics, and gene transportation.^{1–6} As a type of “smart” polymer, their solubility, conformation, or chemical structure, *e.g.* breakage or formation of covalent and non-covalent bonds, can be tuned in response to an external trigger, such as temperature,^{7,8} pH,^{9,10} light,^{11,12} ionic strength,¹³ and redox.¹⁴

Recently, intensive attention has been paid to abundant and environmentally benign gas stimulation modes, including CO₂, O₂, and NO. The reversible and dynamic features of gas-responsive polymers make them suitable and desirable for

biomedical applications, including controlled drug release, tissue culture, and peripheral vascular embolization materials.^{15–28} Among them, CO₂-responsive polymers have gained growing interest. On one hand, they can be reversibly protonated by introducing CO₂ with an increase in their solubility or deprotonated by bubbling an inert gas, such as N₂ or Ar, with a decrease in their solubility in an aqueous solution. On the other hand, since CO₂ is a key metabolite for life and has good biocompatibility and membrane permeability, CO₂-responsive polymers have potential applications in sensors, surface adhesives, and drug delivery.^{15–24} For example, Yuan *et al.* developed the new idea of utilizing a specific amidine-containing block copolymer to fabricate CO₂-responsive vesicles with a biomimetic “breathing” feature, and the membrane permeability of the vesicles can be simply tuned by regulating the CO₂ stimulation time.¹⁷ By utilization of the controlled semi-permeability of CO₂-responsive polymers, hierarchical cargo release and selective separation of guest molecules could be easily realized.¹⁸ In addition to the amidine group, some polymers containing tertiary amine groups, such as poly(*N,N*-dimethylamino)ethyl methacrylate) (PDMAEMA) and poly(*N,N*-diethylamino)ethyl methacrylate) (PDEAEMA), were proven to possess CO₂ responsiveness.^{20–24} For instance, Zhao *et al.* designed a series of triblock copolymers composed of an outer

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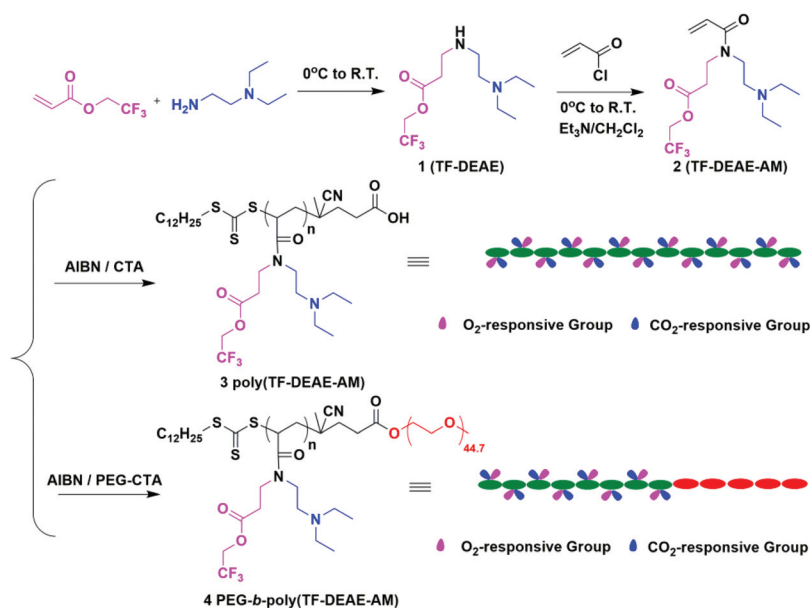


hydrophilic poly(ethylene oxide) segment, middle hydrophobic polystyrene bridging block, and CO₂-responsive interior PDEAEMA flank, which could mimic gas-controllable diversiform deformations of organelles.²⁴ Similar to CO₂, O₂ is another important gas trigger. Both aromatic and aliphatic fluoro-substituted polymers have unique O₂ responsiveness because of the special van der Waals interactions between O₂ molecules and C–F bonds.^{25–29} Zhu *et al.* reported the synthesis of a new type of fluorinated copolymer *via* the atom transfer radical polymerization (ATRP) of commercially available 2,2,2-trifluoroethyl methacrylate (TFMA) and *N,N*-dimethylaminoethyl methacrylate (DMAEMA).²⁶ The obtained copolymer showed a prominent increase in lower critical solution temperature (LCST) from 24.5 °C to 50 °C by the bubbling of O₂, which probably results from its improved solubility due to the interactions between O₂ molecules and 2,2,2-trifluoroethyls.

In contrast to a single-stimulus system, a multi-stimuli system may show a synergistic effect, thereby enhancing the recognition efficiency for distinct drug delivery and release behaviors.^{30–34} Since both CO₂ and O₂ are important to life, the design of dual gas-responsive polymers will significantly expand the scope of stimuli-responsive materials. Although dual CO₂- and O₂-responsive polymers can be prepared by block or random copolymerization of CO₂- and O₂-responsive monomers, the time-consuming steps in the preparation and purification of block copolymers and ill-defined monomer distribution of random copolymer are inherent drawbacks of the block and random copolymer systems, respectively.^{27,28} In order to avoid these drawbacks, installation of CO₂- and O₂-responsive functionalities into the same repeated unit on the basis of the maturity of organic synthesis, that is, the preparation of CO₂- and O₂-responsive homopolymers, has

been considered as an effective and promising alternative. Therefore, it is not surprising that homopolymers with multi-stimuli responsiveness have drawn growing interest in polymer science.^{35–39} However, to the best of our knowledge, the currently available multi-stimuli responsive homopolymers are mainly based on temperature and pH stimuli, and reports on the synthesis of homopolymers with CO₂ and O₂ responsiveness are rare.²⁹

In this study, we develop a new strategy to prepare homopolymers with CO₂ and O₂ responsiveness by the installation of CO₂- and O₂-responsive functional groups in the same repeated unit. Commercially available *N,N*-diethylethylenediamine (DEAE) and 2,2,2-trifluoroethyl acrylate are first connected by the aza-Michael addition reaction (Scheme 1) to give 2,2,2-trifluoroethyl 3-(2-(diethylamino)ethylamino)-propanoate (TF-DEAE). Next, the new monomer, 2,2,2-trifluoroethyl 3-(*N*-(2-(diethylamino)ethyl)acrylamido)propanoate (TF-DEAE-AM), was obtained by the reaction between TF-DEAE and acryloyl chloride. Subsequently, the target poly(TF-DEAE-AM) homopolymers are prepared by reversible addition–fragmentation chain transfer (RAFT) polymerization using 4-cyano-4-(dodecylsulfanylthiocarbonylsulfanyl)pentanoic acid (CDSTSP) as the chain transfer agent with controlled molecular weights and relatively narrow molecular weight distributions. The CO₂ and O₂ responsiveness of the obtained homopolymers are examined *via* TEM, ¹H NMR, ¹⁹F NMR, DLS, and UV-vis spectroscopy. In addition, PEG-*b*-poly(TF-DEAE-AM) diblock polymers are prepared by the RAFT polymerization of TF-DEAE-AM using a PEG-based macromolecular chain transfer agent instead of CDSTSP. It is observed that a variation in both CO₂ and O₂ not only induces a change in the morphology of the aggregates in the aqueous media, but also results in difference in release profiles for hydro-



Scheme 1 Synthetic route for poly(TF-DEAE-AM) and PEG-*b*-poly(TF-DEAE-AM).



phobic dye (pyrene) loading in micelles based on PEG-*b*-poly(TF-DEAE-AM). The CO₂- and O₂-induced release property of poly(TF-DEAE-AM) may open opportunities for the preparation of functional materials with special CO₂ and O₂ responsiveness for potential applications in biomedicine.

Experimental

Materials

2,2'-Azobis(isobutyronitrile) (AIBN, Aldrich, 98%) was recrystallized from anhydrous ethanol. Tetrahydrofuran (THF, Aldrich, 99%) and dichloromethane (CH₂Cl₂, Aldrich, 99%) were dried over CaH₂ and distilled from sodium and benzophenone under N₂, prior to use. Triethylamine (Et₃N, Aldrich, 99.5%) was dried over KOH and distilled over CaH₂ under N₂, prior to use. *N*-Phenyl-1-naphthylamine (PNA, Alfa Aesar, 97%) was purified by recrystallization in ethanol three times. 4-Dimethylaminopyridine (DMAP, Aldrich, 98%) was recrystallized three times from toluene. 2,2,2-Trifluoroethyl acrylate (TCI, 98%), *N,N*-diethylethylenediamine (DEAE, J&K, 99%), *N,N'*-dicyclohexylcarbodiimide (DCC, Aldrich, 99%), acryloyl chloride (Aladdin, 96%), and poly(ethylene glycol) monomethyl ether (PEG-OH, *M_n* = 2000 g mol⁻¹, Aldrich, 99%) were used as received. 4-Cyano-4-(dodecylsulfanyl-thiocarbonylsulfanyl)pentanoic acid (CDSTSP) was synthesized according to previous literature.⁴²

Measurements

All ¹H, ¹³C, and ¹⁹F NMR analyses were performed on a JEOL ECZ 400S spectrometer (400 MHz) in CDCl₃ and D₂O; tetramethylsilane (¹H) and CDCl₃ (¹³C) were used as internal standards, and CF₃CO₂H was used as the external standard for ¹⁹F NMR. FT-IR spectra were recorded on a Nicolet AVATAR-360 spectrophotometer with a 4 cm⁻¹ resolution. Elemental analysis was carried out on a Carlo-Erba 1106 system. Relative molecular weights and molecular weight distributions were measured using a conventional gel permeation chromatography (GPC) system equipped with a Waters 515 Isocratic HPLC pump, Waters 2414 refractive index detector, and a set of Waters Styragel columns (HR3 (500–30 000), HR4 (5000–600 000), and HR5 (50 000–4 000 000), 7.8 × 300 mm, particle size: 5 μm). GPC measurements were carried out at 35 °C using THF as the eluent with a flow rate of 1.0 mL min⁻¹. The system was calibrated with linear poly(methyl methacrylate) (PMMA) standards. The transmittance of polymer solutions and UV-vis spectra were measured using a Hitachi U-2910 spectrophotometer. Steady-state fluorescence spectra were measured at 20 °C on a Hitachi F-2700 fluorescence spectrophotometer with the band width of 5 nm for excitation and emission, and the emission intensity at 418 nm was recorded to determine the critical micelle concentration (cmc), where the excitation wavelength (λ_{ex}) was 340 nm. Hydrodynamic diameter (*D_h*) was measured by dynamic light scattering (DLS) with a Malvern Nano ZS90 Zetasizer after equilibrating for 2 min at 20 °C. The zeta-potential of the solutions

containing polymer aggregates was determined in KCl aqueous media (5.0 mM) at 25 °C on a Malvern Nano ZS90 Zetasizer. Transmission electron spectroscopy (TEM) images were obtained on a JEOL JEM-1230 instrument operated at 80 kV.

Synthesis of 2,2,2-trifluoroethyl 3-(2-(diethylamino)ethylamino)propanoate

N,N-Diethylethylenediamine (8.4 g, 72.0 mmol) was added dropwise to 2,2,2-trifluoroethyl acrylate (9.3 mL, 60.0 mmol) at 0 °C. The reaction mixture was stirred at 0 °C for 30 min and then at room temperature for additional 12 h. The mixture was purified *via* flash chromatography (silica gel column, eluent: ethyl acetate/hexane (v : v = 1 : 4 with Et₃N)) to give 8.5 g (52.4% yield) of amine **1**, 2,2,2-trifluoroethyl 3-(2-(diethylamino)ethylamino)propanoate (TF-DEAE). FT-IR: ν (cm⁻¹): 3313, 2971, 2935, 2816, 1760, 1453, 1412, 1384, 1283, 1168, 1068, 976. ¹H NMR (400 MHz, CDCl₃): δ (ppm): 0.97 (6H, CH₂N(CH₂CH₃)₂), 1.78 (1H, O₂CCH₂CH₂NHCH₂), 2.49 (6H, CH₂N(CH₂CH₃)₂), 2.62 (4H, O₂CCH₂CH₂NHCH₂), 2.91 (2H, O₂CCH₂CH₂NHCH₂), 4.45 (2H, CO₂CH₂CF₃). ¹³C NMR (101 MHz, CDCl₃): δ (ppm): 11.9 (CH₂N(CH₂CH₃)₂), 34.5 (O₂CCH₂CH₂NHCH₂), 45.0 (O₂CCH₂CH₂NHCH₂), 47.2 (CH₂N(CH₂CH₃)₂), 47.5 (O₂CCH₂CH₂NHCH₂), 52.7 (CH₂N(CH₂CH₃)₂), 60.3 (CO₂CH₂CF₃), 123.1 (CO₂CH₂CF₃), 171.2 (CO₂CH₂CF₃). ¹⁹F NMR (376 MHz, CDCl₃): δ (ppm): -73.9 (CO₂CH₂CF₃). Anal. Calcd for C₁₁H₂₁N₂O₂F₃: C, 48.88%; H, 7.83%; N, 10.36%. Found: C, 48.46%; H, 7.83%; N, 10.16%.

Synthesis of 2,2,2-trifluoroethyl 3-(*N*-(2-(diethylamino)ethyl)acrylamido)propanoate

2,2,2-Trifluoroethyl 3-(2-(diethylamino)ethylamino)propanoate (8.1 g, 30.0 mmol) and Et₃N (10.0 mL, 72.0 mmol) were dissolved in 100 mL of anhydrous CH₂Cl₂. The solution was cooled to 0 °C, followed by the addition of acryloyl chloride (3.3 g, 36.0 mmol) dropwise within 15 min. The mixture was slowly warmed to room temperature and stirred for 12 h. The volatiles were evaporated under reduced pressure and the residue was purified using flash chromatography (silica gel column, eluent: ethyl acetate/hexane (v : v = 1 : 4 with Et₃N)) to give 3.0 g (30.9% yield) of the corresponding acrylamide monomer **2**, 2,2,2-trifluoroethyl 3-(*N*-(2-(diethylamino)ethyl)acrylamido)propanoate (TF-DEAE-AM). FT-IR: ν (cm⁻¹): 2970, 2937, 2810, 1758, 1652, 1615, 1448, 1425, 1374, 1281, 1169, 1071, 977, 785. ¹H NMR (400 MHz, CDCl₃): δ (ppm): 0.99 (6H, CH₂N(CH₂CH₃)₂), 2.51 (6H, CH₂N(CH₂CH₃)₂), 2.77 (2H, O₂CCH₂CH₂NCH₂), 3.43 (2H, O₂CCH₂CH₂NCH₂), 3.67 (2H, O₂CCH₂CH₂NCH₂), 4.44 (2H, CO₂CH₂CF₃), 5.68, 6.32 (2H, CH₂=CHCO), 6.54 (1H, CH₂=CHCO). ¹³C NMR (101 MHz, CDCl₃): δ (ppm): 12.0 (CH₂N(CH₂CH₃)₂), 32.4 (O₂CCH₂CH₂NCH₂), 43.8 (CH₂N(CH₂CH₃)₂), 47.6 (CH₂N(CH₂CH₃)₂), 48.1 (O₂CCH₂CH₂NCH₂), 52.7 (O₂CCH₂CH₂NCH₂), 60.5 (CO₂CH₂CF₃), 121.6 (CH₂=CHCO), 124.4 (CH₂=CHCO), 128.0 (CO₂CH₂CF₃), 166.6 (CH₂=CHCO), 170.7 (CO₂CH₂CF₃). ¹⁹F NMR (376 MHz, CDCl₃): δ (ppm): -73.8 (CO₂CH₂CF₃). MS-DART: calculated C₁₄H₂₄N₂O₃F₃ for 325.1734; found



325.1732 $[M + H]^+$. Anal. Calcd for $C_{14}H_{23}N_2O_3F_3$: C, 51.84%; H, 7.15%; N, 8.64%. Found: C, 51.86%; H, 7.15%; N, 8.67%.

RAFT homopolymerization of TF-DEAE-AM

In a typical procedure, AIBN (1.6 mg, 0.01 mmol), CDSTSP (12.4 mg, 0.031 mmol), and TF-DEAE-AM (0.50 g, 1.54 mmol) were first added to a 25 mL Schlenk flask (flame-dried under vacuum prior to use) sealed with a rubber septum for degassing and kept under N_2 . Next, dry THF (0.5 mL) was charged *via* a gastight syringe. The flask was degassed by three cycles of freezing–pumping–thawing, followed by immersing the flask into an oil bath set at 65 °C. The polymerization lasted for 15 h and it was terminated by placing the flask into liquid N_2 . The reaction mixture was precipitated in cold *n*-hexane. The crude product was purified by repeated dissolution in THF and precipitation in *n*-hexane, followed by drying *in vacuo* overnight to give 0.18 g of light yellow solid, poly(TF-DEAE-AM) **3b**. GPC: $M_n = 10\,500\text{ g mol}^{-1}$, $M_w/M_n = 1.36$. FT-IR: ν (cm^{-1}): 2969, 2928, 2812, 1758, 1642, 1452, 1422, 1379, 1280, 1169, 1070, 976, 842. $^1\text{H NMR}$ (400 MHz, CDCl_3): δ (ppm): 1.02 (6H, $\text{CH}_2\text{N}(\text{CH}_2\text{CH}_3)_2$), 1.25 (18H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 1.79 (2H, CH_2CHCON), 2.58 (1H, CH_2CHCON ; 6H, $\text{CH}_2\text{N}(\text{CH}_2\text{CH}_3)_2$; 2H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{NCH}_2$), 3.58 (4H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{NCH}_2$), 4.46 (2H, $\text{CO}_2\text{CH}_2\text{CF}_3$).

Preparation of PEG-CTA

PEG-OH ($M_n = 2000\text{ g mol}^{-1}$, 25.0 g, 12.5 mmol –OH), DMAP (0.3 g, 2.5 mmol), CDSTSP (10.1 g, 25.0 mmol), and dry CH_2Cl_2 (300.0 mL) were first added to a 500 mL flask (flame-dried under vacuum prior to use) sealed with a rubber septum under N_2 . The solution was cooled to 0 °C, followed by the addition of 25 mL of CH_2Cl_2 solution containing DCC (5.2 g, 25.0 mmol) dropwise within 15 min. The mixture was slowly warmed to room temperature and stirred for 12 h. The solid was filtered and the filtrate was concentrated, followed by precipitation in cold diethyl ether. After repeated purification by dissolving in CH_2Cl_2 and precipitation in cold diethyl ether, 16.7 g (56.0% yield) of yellow solid, PEG-CTA, was obtained after drying *in vacuo* overnight. GPC: $M_n = 3800\text{ g mol}^{-1}$, $M_w/M_n = 1.02$. $^1\text{H NMR}$ (400 MHz, CDCl_3): δ (ppm): 0.87 (3H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 1.25 (18H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 1.68 (2H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 1.87 (3H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{C}(\text{CN})\text{CH}_3$), 2.36–2.52 (2H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{C}(\text{CN})\text{CH}_3$), 2.65 (2H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{C}(\text{CN})\text{CH}_3$), 3.31 (2H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 3.37 (3H, OCH_3), 3.45–3.63 (4H, OCH_2CH_2), 3.81 (2H, $\text{CO}_2\text{CH}_2\text{CH}_2\text{O}$), 4.25 (2H, $\text{CO}_2\text{CH}_2\text{CH}_2\text{O}$).

RAFT block copolymerization of TF-DEAE-AM mediated by PEG-CTA

In a typical procedure, AIBN (1.6 mg, 0.01 mmol), PEG-CTA (72.3 mg, 0.030 mmol trithiocarbonate group), and TF-DEAE-AM (0.6 g, 1.85 mmol) were first added to a 25 mL Schlenk flask (flame-dried under vacuum prior to use) sealed with a rubber septum for degassing and kept under N_2 . Next, dry THF (0.6 mL) was charged *via* a gastight syringe. The flask was degassed by three cycles of freezing–pumping–thawing,

followed by immersion of the flask into an oil bath set at 65 °C. The polymerization lasted 48 h and it was terminated by placing the flask into liquid N_2 . The reaction mixture was precipitated in diethyl ether. The collected supernatant was then purified by repeated dissolution in THF and precipitation in diethyl ether followed by drying *in vacuo* overnight to give 0.26 g of light yellow solid, PEG-*b*-poly(TF-DEAE-AM) **4c**. GPC: $M_{n,\text{GPC}} = 8700\text{ g mol}^{-1}$ ($M_{n,\text{NMR}} = 11\,600\text{ g mol}^{-1}$), $M_w/M_n = 1.24$. FT-IR: ν (cm^{-1}): 2970, 2931, 2872, 1757, 1642, 1452, 1422, 1350, 1281, 1147, 976, 845. $^1\text{H NMR}$ (400 MHz, CDCl_3): δ (ppm): 0.88 (3H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 1.00 (6H, $\text{CH}_2\text{N}(\text{CH}_2\text{CH}_3)_2$), 1.26 (18H, $\text{SCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$), 1.84 (3H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{C}(\text{CN})\text{CH}_3$), 1.94 (2H, CH_2CHCON), 2.51 (1H, CH_2CHCON and 6H, $\text{CH}_2\text{N}(\text{CH}_2\text{CH}_3)_2$), 2.69 (2H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{NCH}_2$), 3.30 (4H, $\text{O}_2\text{CCH}_2\text{CH}_2\text{NCH}_2$), 3.38 (3H, OCH_3), 3.64 (OCH_2CH_2), 4.21 (2H, $\text{CO}_2\text{CH}_2\text{CH}_2\text{O}$), 4.46 (2H, $\text{CO}_2\text{CH}_2\text{CF}_3$).

Micellar morphology

A THF solution of poly(TF-DEAE-AM) **3b** homopolymer or PEG-*b*-poly(TF-DEAE-AM) **4b** diblock copolymer (10.0 mg mL^{-1}) was added dropwise to distilled water under vigorous stirring and bubbling with N_2 until the concentration of the polymer reached 0.5 mg mL^{-1} . THF was evaporated by moderate stirring overnight at room temperature. For TEM studies, 50 μL of micelle solution was deposited onto an electron microscopy copper grid coated with carbon film, and the water was evaporated at room temperature.

Titration experiment

A 50 mL three-neck round-bottom flask equipped with a Thermo Scientific™ Orion™ ROSS Ultra™ Refillable pH/ATC Triode™ combination electrode and a Lei-Ci DJS-1C conductance electrode was used to conduct potentiometric titrations. 20 mL of poly(TF-DEAE-AM) **3b** homopolymer (1.0 mg mL^{-1}) in aqueous salt solution ($[\text{NaCl}] = 10\text{ mM}$, pH = 3.0) was titrated using a standard NaOH solution (1.0 M) under constant stirring and bubbling of N_2 .

Transmittance experiment

Poly(TF-DEAE-AM) **3b** homopolymer was uniformly dispersed in distilled water by ultrasonication until the concentration of the homopolymer reached 0.5 mg mL^{-1} . The gas-based transmittance changes of the aqueous solution were measured using a Hitachi U-2910 spectrophotometer at a constant wavelength of 500 nm.

Determination of critical micelle concentration

PNA was used as a fluorescence probe to measure the cmc of the PEG-*b*-poly(TF-DEAE-AM) **4** diblock copolymer in the aqueous media. An acetone solution of PNA ($[\text{PNA}] = 2\text{ mM}$) was added to a large amount of water until the concentration of PNA reached 0.002 mM. The solutions for fluorescence measurement were obtained by adding different amounts of THF solutions of PEG-*b*-poly(TF-DEAE-AM) **4** diblock copolymer (1, 0.1, 0.01, 0.001, or 0.0001 mg mL^{-1}) to water containing PNA ($[\text{PNA}] = 0.002\text{ mM}$). THF was evaporated by constant



stirring and bubbling N₂ overnight at room temperature. The solution was sealed in vials prior to measurement to avoid the influence of CO₂ or O₂ present in air.

Results and discussion

Design and synthesis of TF-DEAE-AM trifunctional acrylamide monomer

Given the well-studied CO₂-responsive PDEAEMA²⁴ and O₂-responsive PTFMA,^{26,27} we attempted to incorporate DEAE and 2,2,2-trifluoroethyl (TF) functional groups into a single monomer, and utilize RAFT polymerization to construct dual-gas-stimulus responsive polymers. The key in the preparation of the target monomer is to incorporate a polymerizable group, DEAE group, and TF group within a monomer. Our group designed a two-step strategy to connect two different temperature- and pH-responsive groups and acrylamide group together.^{40,41} This strategy possesses several advantages, such as facile sample preparation and possibility to accomplish multifunctional materials, and great opportunities to enlarge the applications of this type of polymer. Inspired by the study, we employed a similar strategy in the current study by applying aza-Michael reaction as a key approach to provide the target secondary amine (TF-DEAE) intermediate in a single step, which could be utilized in a further reaction to afford a polymerizable acrylamide monomer (TF-DEAE-AM).

The key intermediate of 2,2,2-trifluoroethyl 3-(2-(diethylamino)ethylamino)propanoate **1** secondary amine was first prepared through aza-Michael addition reaction using 2,2,2-trifluoroethyl acrylate and *N,N*-diethylethylenediamine. ¹H, ¹³C, and ¹⁹F NMR spectra, shown in Fig. S1,† display the expected resonance signals of the target compound, which confirm the chemical structure of the target intermediate, *i.e.* TF-DEAE **1**. Subsequently, TF-DEAE **1** was treated with acryloyl chloride to afford the desired TF-DEAE-AM **2** acrylamide monomer. The chemical structure of TF-DEAE-AM **2** monomer was characterized *via* ¹H NMR, ¹³C NMR, ¹⁹F NMR, HR-MS, FT-IR, and EA in detail. Fig. 1A shows ¹H NMR spectrum of the monomer, which exhibits typical proton resonance signals of a double bond at 6.54, 6.32, and 5.68 ppm (peaks “a” and “b”), 2,2,2-trifluoroethyl unit at 4.44 ppm (peak “c”), and DEAE moiety at 0.99 ppm (peak “i”). The ¹³C NMR spectrum (Fig. 1B) displays the expected resonance signals of carbonyls at 166.6 and 170.7 ppm (peaks “c” and “f”) and double bond at 121.6 and 124.4 ppm (peaks “a” and “b”), respectively. The only resonance signal located at –73.8 ppm in the ¹⁹F NMR spectrum (Fig. 1C) was attributed to fluorine atoms in –CF₃, this verified the existence of TF unit. Furthermore, HR-MS result (325.1732) was also consistent well with the theoretical value (C₁₄H₂₄N₂O₃F₃, 325.1734). All these results clearly confirmed the structure of the acrylamide monomer TF-DEAE-AM **2**.

Synthesis of poly(TF-DEAE-AM) homopolymer

In the present study, RAFT polymerization^{43–47} was chosen for the synthesis of poly(TF-DEAE-AM) **3** homopolymer since

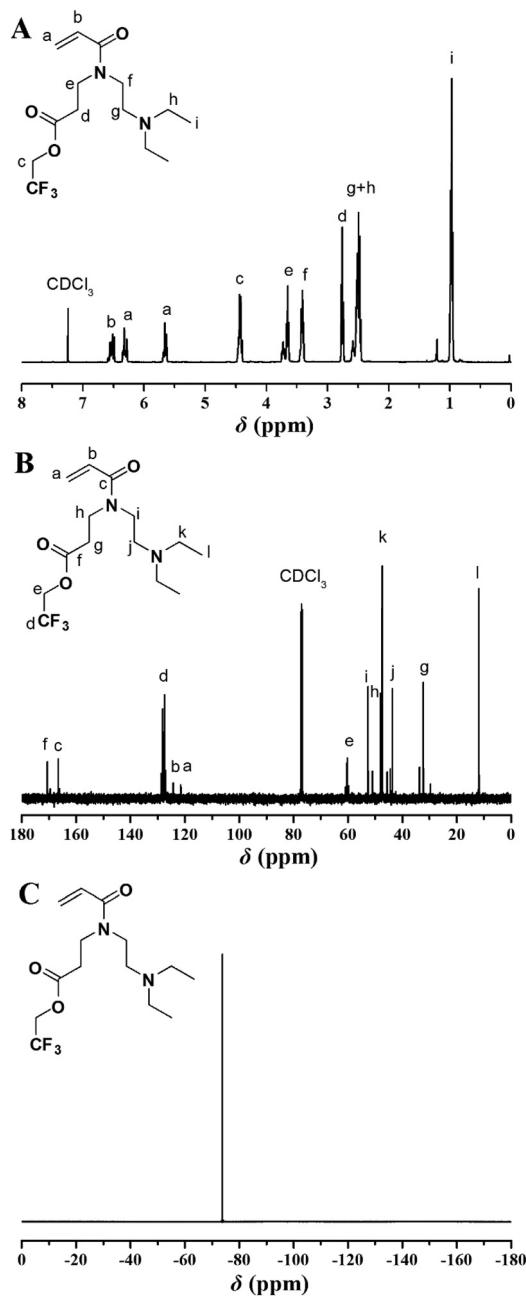


Fig. 1 ¹H (A), ¹³C (B), and ¹⁹F (C) NMR spectra of TF-DEAE-AM **2** in CDCl₃.

RAFT polymerization is one of the most powerful and versatile reversible degenerative radical polymerization (RDRP) processes that ensures precise control over molecular weight and molecular weight distribution in mild reaction conditions. Two poly(TF-DEAE-AM) **3** homopolymers were prepared by varying the feeding ratio of TF-DEAE-AM **2** monomer to CTA of CDSTSP (30 : 1 and 50 : 1) and reaction time, as listed in Table 1.

Both homopolymers show symmetric and unimodal eluent peaks in their GPC curves (Fig. 2A) with relatively narrow molecular weight distributions ($M_w/M_n < 1.40$), which are indicative



Table 1 Synthesis of poly(TF-DEAE-AM) **3** by RAFT homopolymerization^a

Sample	[2]:[CTA]	Time (h)	Conv. ^b (%)	$M_{n,Theo}$ ^c (g mol ⁻¹)	M_n ^d (g mol ⁻¹)	M_w/M_n ^d
3a	30:1	12	80.5	8236	5500	1.29
3b	50:1	15	78.3	13 101	10 500	1.36

^a[AIBN]:[CTA] = 1:3, solvent: THF, temperature: 65 °C. ^bMeasured by ¹H NMR. ^c $M_{n,Theo} = M_{n,2} \times ([2]:[CTA]) \times \text{conversion} + M_{n,CTA}$. ^dMeasured by GPC in THF at 35 °C.

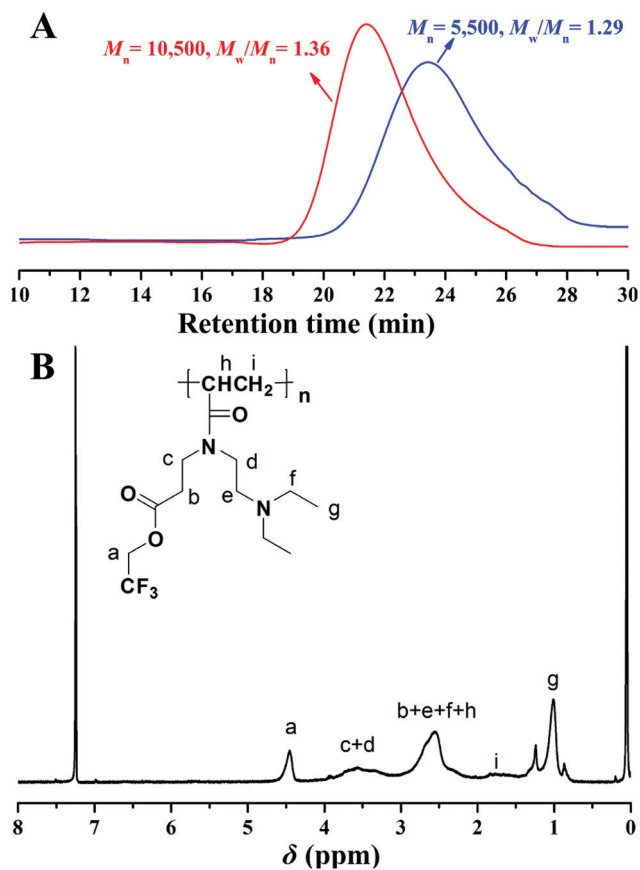


Fig. 2 (A) GPC traces of poly(TF-DEAE-AM) **3** homopolymer in THF; and (B) ¹H NMR spectrum of poly(TF-DEAE-AM) **3** homopolymer in CDCl₃.

of the good choice of CDSTSP as CTA for mediating the polymerization and well-defined structure of the obtained poly(TF-DEAE-AM) **3** homopolymer. The molecular weights of the homopolymers measured by GPC are smaller than theoretical values due to the difference in the structure of poly(TF-DEAE-AM) **3** homopolymer and PMMA standards. A typical ¹H NMR spectrum of poly(TF-DEAE-AM) **3** homopolymer is shown in Fig. 2B, which shows the disappearance of the proton resonance signals of the double bond and the retention of the peaks of TF moiety (peak “a”) and DEAE unit (peak “g”). These observations affirmed the chemical structure of poly(TF-DEAE-AM) **3** homopolymer.

Solution behavior of poly(TF-DEAE-AM) homopolymer

As a type of weak polybase, PDEAEMA homopolymer is soluble in water as a weak cationic polyelectrolyte in acidic surroundings due to the protonation of its tertiary amine group; however, it becomes insoluble at room temperature in a neutral or basic environment. The pK_a of its conjugated acid is about 7.3.^{48–51} Although it is expected that poly(TF-DEAE-AM) **3** homopolymer should have similar pH stimulus responsiveness as PDEAEMA, the difference in their chemical structure might result in a change in pK_a of the conjugated acid of DEAE group. In order to measure the degree of protonation of DEAE group on pH of aqueous solution, pH value and conductivity of an aqueous solution of poly(TF-DEAE-AM) **3b** homopolymer were measured at the same time by adding NaOH solution (1.0 M) to the acidic poly(TF-DEAE-AM) **3b** solution according to the previous report.⁵²

As shown in Fig. 3A, three regions can be divided on the basis of the changes in pH and conductivity, in which the second region between the two transition points (point a, pH ~ 4.7 and point b, pH ~ 8.4) corresponds to the deprotonation process of NH⁺(C₂H₅)₂ group from the fully protonated state to the unionized state. We define that the degree of protonation of DEAE group is 100% when the pH of the solution is 4.7,

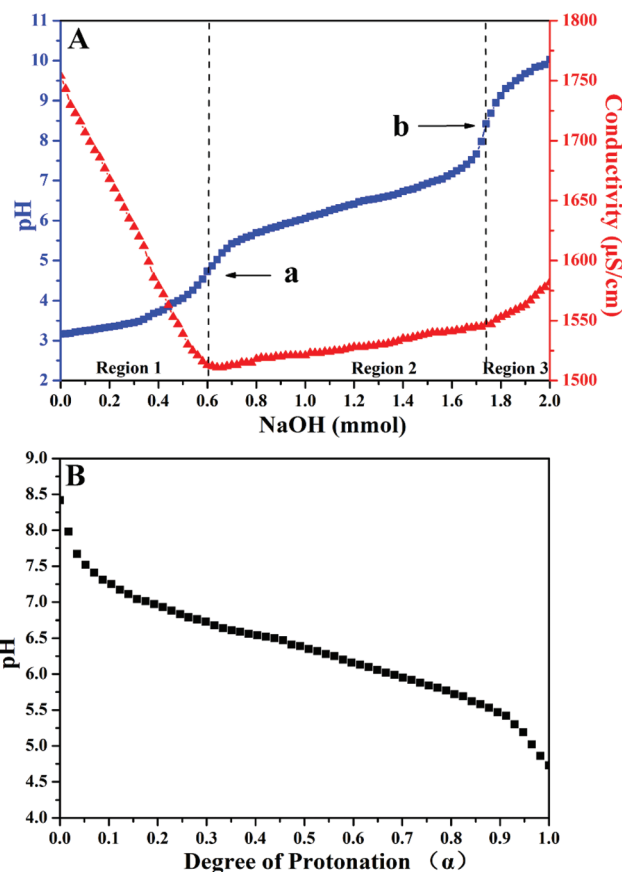


Fig. 3 (A) Titration curves of poly(TF-DEAE-AM) **3b** homopolymer (1.0 mg mL⁻¹) at 25 °C in 10 mM NaCl; and (B) profile for the dependence of protonation on pH.



and the degree of protonation of DEAE group is 0% when pH of the solution is 8.4. Based on this information, we obtained the profile of the dependence of degree of protonation of DEAE group with pH (Fig. 3B). The pH value of a saturated CO₂ aqueous solution is about 3.9 at room temperature and atmosphere pressure so that 100% of tertiary amino of poly(TF-DEAE-AM) **3** homopolymer would be protonated by carbonic acid under this condition.^{20–24} As a result, it can be expected that poly(TF-DEAE-AM) **3** homopolymer will exhibit CO₂-responsive behavior in an aqueous solution.

The transmittance results of poly(TF-DEAE-AM) **3b** aqueous solution at room temperature in response to CO₂ are shown in Fig. 4 and it was found that a turbid solution was well observed at 25 °C (Fig. 4A) before treatment with CO₂ (10 mL min⁻¹). The solution became entirely transparent after the injection of CO₂, while pH decreased from 8.0 to 4.7 due to the complete protonation of the tertiary amine group. The zeta-potential of the solution containing poly(TF-DEAE-AM) **3b** aggregates greatly increased from 15.5 mV to 47.5 mV after the injection of CO₂, which indicates that the protonated DEAE⁺ chains indeed formed a new corona with positive charges.

Subsequently, after passing N₂ (10 mL min⁻¹) through the solution to remove CO₂, a sharp drop in transmittance was observed (Fig. 4B) because of the deprotonation of DEAE group. Repeatable cycles under alternating CO₂/N₂ stimulation (Fig. 4B) indicated the CO₂-responsive reversibility of poly(TF-DEAE-AM) **3** homopolymer. Since the removal of CO₂ by N₂ in aqueous solution represents a physical process, it needs enough time to completely recover. However, considering that a rather long time purging would lead to an evident loss of water, we herein set a consistent time of 1 h to bubble the gas,

and thus, an apparent difference in turbidity was observed during the process of deprotonation.

In order to gain deep insight into the solubility of poly(TF-DEAE-AM) **3** homopolymer in aqueous solution upon bubbling CO₂, ¹H and ¹⁹F NMR spectra of poly(TF-DEAE-AM) **3b** in D₂O before and after bubbling gas were recorded. Before bubbling CO₂, no signal was found in both ¹H and ¹⁹F NMR spectra (green line in Fig. 4C and D). In CO₂-saturated D₂O, however, the characteristic signals of DEAE group attributed to the protons of “a”, “b” and “c” appear at 1.12 and 3.02 ppm, respectively, which is indicative of the good solubility of tertiary amine groups (red line in Fig. 4C). On the other hand, ¹⁹F NMR spectrum exhibits a weak and broad signal centered at -74.1 ppm after treatment with CO₂ (red line in Fig. 4D), which suggests that TF groups are also partly soluble due to the increase in solubility of the close protonated DEAE groups after bubbling CO₂. After the removal of CO₂ by N₂, the two typical signals of DEAE group in ¹H NMR spectrum and characteristic peak originating from TF moiety in ¹⁹F NMR spectrum disappear again because of the deprotonation of DEAE group (blue line in Fig. 4C and D). Moreover, the protonation/deprotonation process could be reproduced over several cycles. It is believed that the interactions between CO₂ and DEAE groups increased the solubility of poly(TF-DEAE-AM). In comparison with the chemical process of protonation by CO₂, van der Waals interactions between O₂ and C–F bonds appeared to be too weak, and there was no significant change in both transmittance and NMR spectrum after O₂ treatment (10 mL min⁻¹).

We then utilized DLS and TEM to examine the size and morphology of the aggregates formed by poly(TF-DEAE-AM) **3b**

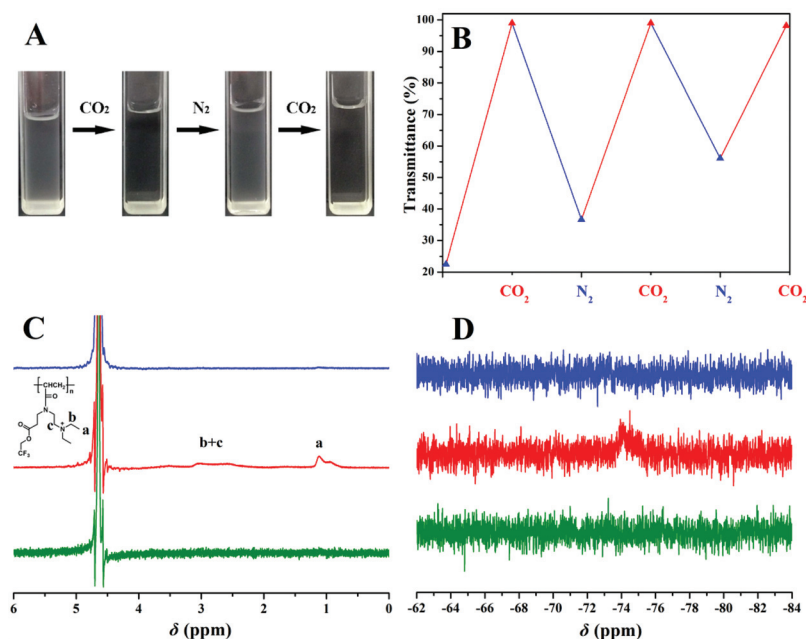


Fig. 4 Optical images (A) and transmittance changes (B) of poly(TF-DEAE-AM) **3b** aqueous solution (0.5 mg mL⁻¹) with CO₂/N₂ gas triggers; and ¹H (C) and ¹⁹F (D) NMR spectra of poly(TF-DEAE-AM) **3b** in D₂O before (green) and after CO₂ (red) or CO₂/N₂ (blue) treatment.



homopolymer upon the treatment of N₂, O₂, and CO₂, respectively. The micellar solution of poly(TF-DEAE-AM) **3b** homopolymer was prepared by adding 0.5 mL of THF solution of poly(TF-DEAE-AM) **3b** homopolymer (10 mg mL⁻¹) in 10 mL of water, followed by removing THF by stirring and bubbling of N₂ at room temperature over 24 h. Two additional aliquots (3.0 mL) taken from the micellar solution were treated with CO₂ and O₂, respectively.

As shown in Fig. 5A, spindle-shaped aggregates with an average hydrodynamic diameter of 379 nm (green line in Fig. 5D) were formed in aqueous media with the bubbling of N₂. After the solution was bubbled with O₂ for 1 h, interestingly, only spherical aggregates (Fig. 5B) with an average hydrodynamic diameter of 234 nm (red line in Fig. 5D), clearly smaller than that without the treatment with O₂, were observed. This phenomenon might indicate that the large spindle-shaped aggregates fell apart and formed spherical aggregates. Although we cannot observe noticeable changes in the solubility, and ¹H and ¹⁹F NMR spectra of poly(TF-DEAE-AM) **3b** homopolymer upon bubbling O₂, this evident change in morphology of aggregates clearly demonstrates the O₂-stimulus responsiveness of poly(TF-DEAE-AM) **3b** homopolymer. Similarly, the spindle-shaped aggregates also changed into ill-defined spherical micelles (Fig. 5C) with an average hydrodynamic diameter of 327 nm (blue line in Fig. 5D) after bubbling CO₂ for 1 h. Different from the spherical micelles observed after the treatment with O₂, there were several isolated darker "dots" in the matrix of the aggregates.

This observation suggests that the aggregates composed of these darker dots are formed by poly(TF-DEAE-AM) **3b** homopolymer.

Synthesis of PEG-*b*-poly(TF-DEAE-AM) diblock copolymer

Although poly(TF-DEAE-AM) **3b** homopolymer has both CO₂ and O₂-stimuli responsiveness, the homopolymer is completely insoluble in physiological environment and as a result, its applications in the biomedical field would be limited. It is necessary to connect poly(TF-DEAE-AM) with a hydrophilic and biocompatible segment for the construction of potential intelligent drug delivery systems, in which the release of hydrophobic drugs loaded in the core of aggregates formed by poly(TF-DEAE-AM) segment could be tuned as a response to CO₂ and O₂. In the current study, we selected PEG as the hydrophilic segment due to its unique biocompatibility and broad application in biomedicine.^{53–56}

A PEG-based macro-CTA (PEG-CTA) was first prepared *via* the attachment of the acidic functional trithiocarbonate of CDSTSP to PEG-OH (*M*_n = 2000 g mol⁻¹) following a commonly-used procedure.^{57–60} ¹H NMR spectrum of PEG-CTA (Fig. S2†) exhibits the attachment of CDSTSP onto the chain end of PEG. Then, PEG-*b*-poly(TF-DEAE-AM) **4** diblock copolymers were obtained by RAFT polymerization of TF-DEAE-AM **2** using PEG-CTA as the chain transfer agent. By varying the polymerization time and feeding ratio of monomer **2** to PEG-CTA, three PEG-*b*-poly(TF-DEAE-AM) **4** diblock copolymers

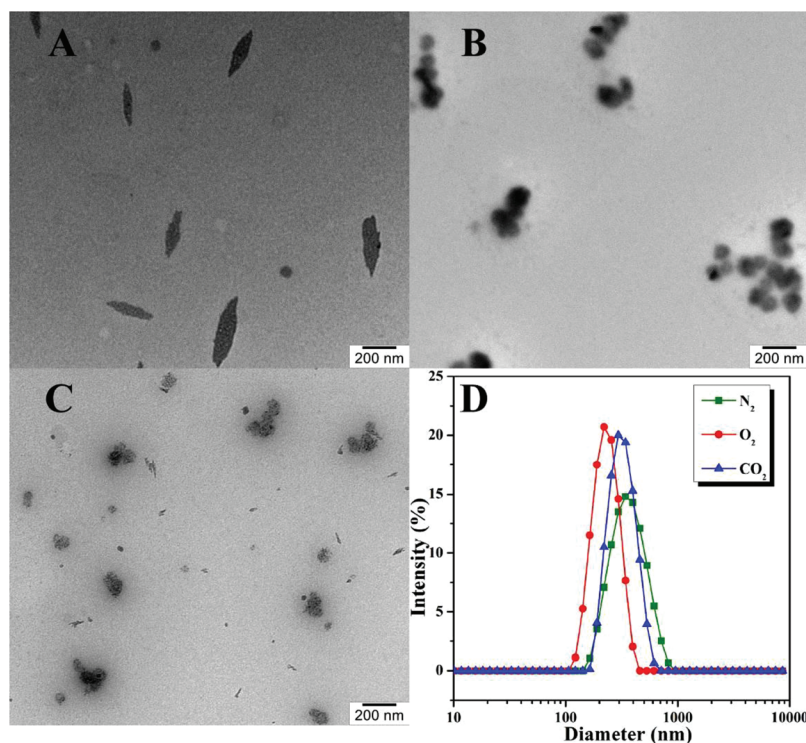


Fig. 5 TEM images of poly(TF-DEAE-AM) **3b** after N₂ (A), O₂ (B), and CO₂ (C) treatment and the corresponding hydrodynamic diameter distributions (D).



Table 2 Synthesis of PEG-*b*-poly(TF-DEAE-AM) 4 by RAFT polymerization^a

Sample	[2]:[CTA]	Time (h)	Conv. ^b (%)	$M_{n,NMR}$ ^c (g mol ⁻¹)	$M_{n,Theo}$ ^d (g mol ⁻¹)	$M_{n,GPC}$ ^e (g mol ⁻¹)	M_w/M_n ^e	f^f	cmc^g (g mL ⁻¹)
4a	30 : 1	12	67.9	4200	8991	5800	1.13	0.43	6.8×10^{-5}
4b	30 : 1	60	98.6	8900	11 978	6700	1.11	0.73	1.8×10^{-5}
4c	60 : 1	48	95.0	11 600	20 872	8700	1.24	0.79	8.3×10^{-6}

^a [AIBN]:[CTA] = 1 : 3, solvent: THF, temperature: 65 °C. ^b Measured by ¹H NMR. ^c Obtained from ¹H NMR. ^d $M_{n,Theo} = M_{n,2} \times ([2]:[CTA]) \times \text{conv.} + M_{n,CTA}$. ^e Measured by GPC in THF at 35 °C. ^f Weight fraction of hydrophobic poly(TF-DEAE-AM) segment in copolymer 4. ^g Determined by fluorescence spectroscopy using PNA as the probe.

with different chain lengths of poly(TF-DEAE-AM) were prepared, as summarized in Table 2.

Fig. 6A shows the GPC traces of the PEG-CTA and PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer. Compared to PEG-CTA, the elution time of the PEG-*b*-poly(TF-DEAE-AM) 4 diblock polymers decreased with an increase in polymerization time and feeding ratio of monomer 2 to PEG-CTA, indicating the extension of the polymer chain of poly(TF-DEAE-AM) from PEG-CTA. In addition, unimodal and symmetrical elution peaks with narrow molecular weight distributions ($M_w/M_n \leq 1.24$)

were observed for PEG-*b*-poly(TF-DEAE-AM) 4 diblock polymers. The chemical structure of PEG-*b*-poly(TF-DEAE-AM) 4 diblock polymer was characterized *via* ¹H NMR observations (Fig. 6B). The typical proton resonance signals originating from PEG block (3.64 ppm) still appear in ¹H NMR spectrum. The peak “a” located at 4.46 ppm is attributed to two protons of CO₂CH₂CF₃ moiety in TF-DEAE-AM repeated unit, whereas the peak “f” located at 4.21 ppm corresponds to two protons of CO₂CH₂CH₂O linkage between poly(TF-DEAE-AM) and PEG in the main chain. On the basis of these results and chain length of PEG, the “absolute” molecular weight of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer could be calculated by ¹H NMR observations according to eqn (1) (S_a and S_f are integral areas of peak “a” and “f” in Fig. 6B, respectively; and 325 and 2385 are the molecular weights of monomer 2 and PEG-CTA, respectively). Since the chemical structure of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer is different from PMMA standards, the molecular weights obtained from ¹H NMR analysis were employed in the current study, which are indeed very different from those obtained from GPC. As listed in Table 2, three different PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymers with 6, 20, and 29 TF-DEAE-AM repeated units were prepared in the current case.

$$M_n = 325(S_a/S_f) + 2385 \quad (1)$$

Solution behavior of PEG-*b*-poly(TF-DEAE-AM) diblock copolymer

The critical micelle concentration (*cmc*) of the amphiphilic PEG-*b*-poly(TF-DEAE-AM) 4 diblock polymer in aqueous solution without external stimuli was firstly determined *via* fluorescence spectroscopy using PNA as the fluorescence probe.⁶¹ The fluorescence intensity of PNA is sensitive to the environment and the polarity of its surroundings and it could be easily quenched by polar solvents such as water.⁶¹ Once micelles are formed in aqueous solution, the hydrophobic PNA could be encapsulated into their hydrophobic core, and thus, the value of I/I_0 would dramatically increase (I and I_0 refer to the fluorescence intensities of PNA at 418 nm with and without PEG-*b*-poly(TF-DEAE-AM) in aqueous solution, respectively).⁶¹ The relationships of I/I_0 of PNA as a function of the concentration of PEG-*b*-poly(TF-DEAE-AM) 4 diblock polymer are plotted in Fig. 7. It is clear that when the concentration of copolymer 4 exceeds a certain value, I/I_0 increases sharply due to the transferring of PNA probe into the hydrophobic core region of the micelles.

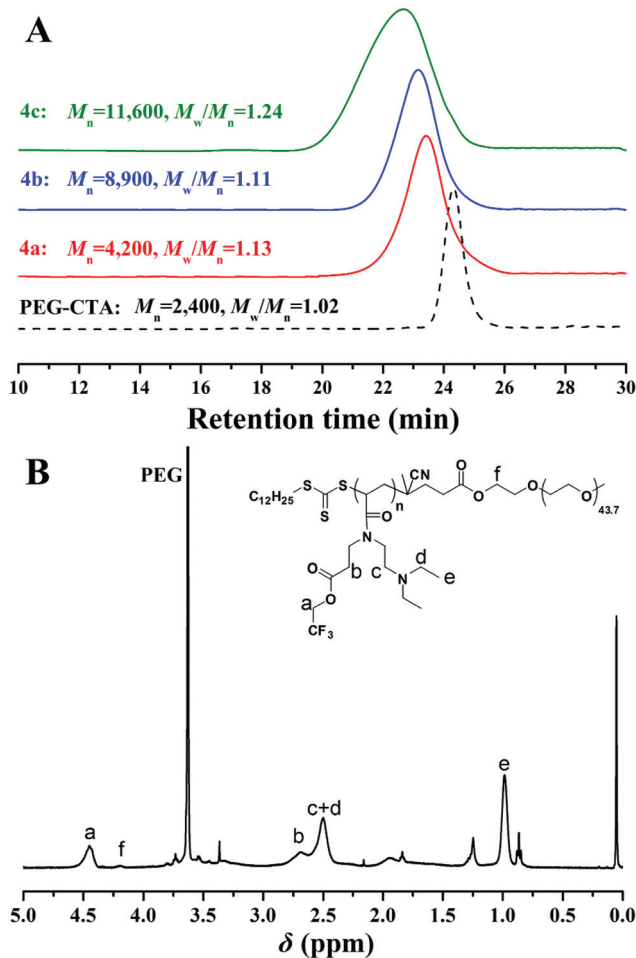


Fig. 6 (A) GPC traces of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer in THF (M_n was obtained from ¹H NMR in CDCl₃); and (B) ¹H NMR spectrum of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer in CDCl₃.



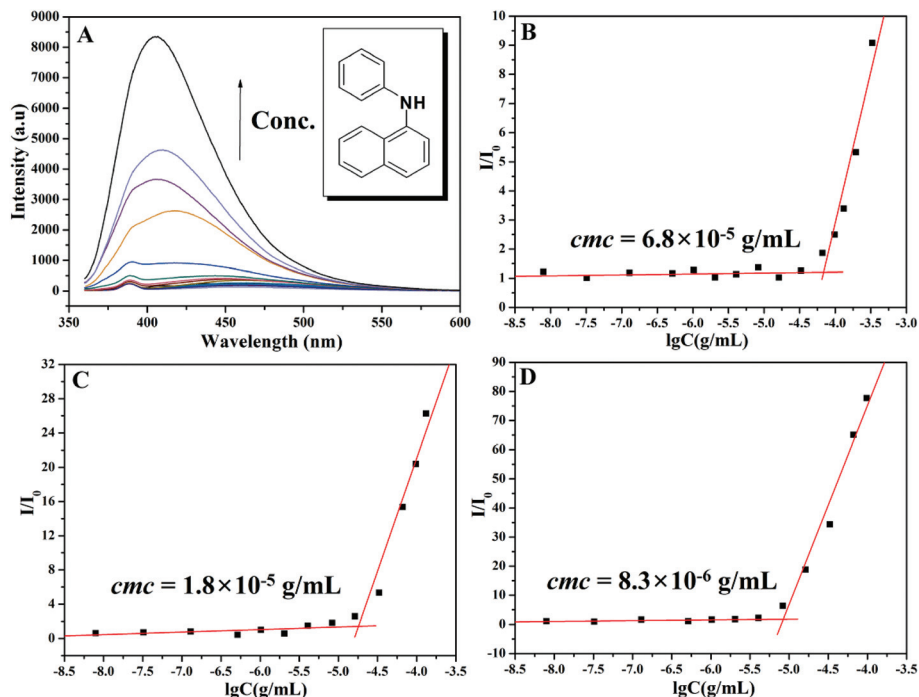


Fig. 7 (A) Fluorescence emission spectra of PNA (0.002 mM) in an aqueous solution of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer. Dependence of the fluorescence intensity (I/I_0) of the PNA emission band at 418 nm on the concentration of PEG-*b*-poly(TF-DEAE-AM) 4a (B), 4b (C), and 4c (D) diblock polymers.

Thus, the *cmc* of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer was determined to be the intersection of two straight lines and the results are summarized in Table 2. The *cmc* of PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer increased from 8.3×10^{-6} g mL⁻¹ to 6.8×10^{-5} g mL⁻¹ with a decrease in the chain length of poly(TF-DEAE-AM) block, which is consistent with a previous report.⁶²

The potentiometric and conductometric titration of PEG-*b*-poly(TF-DEAE-AM) 4b diblock copolymer (Fig. S3†) illustrates that the degree of protonation of DEAE groups was 100% when the pH of the solution was 4.7, whereas the degree of protonation was 0% when the pH of the solution was 8.7. We then examined the CO₂ and O₂-stimuli response behaviors of the micelles formed by PEG-*b*-poly(TF-DEAE-AM) 4 diblock copolymer. 0.5 mL of THF solution of PEG-*b*-poly(TF-DEAE-AM) 4b diblock copolymer (10 mg mL⁻¹) was added to 10 mL of water with stirring and bubbling of N₂ (10 mL min⁻¹). THF was then removed *via* evaporation with stirring and bubbling of N₂ at room temperature over 24 h. Two additional aliquots (3.0 mL) taken from the micellar solution were treated with CO₂ and O₂, separately. PEG-*b*-poly(TF-DEAE-AM) 4b diblock copolymer formed spherical aggregates (Fig. 8B) with a D_h of 140 nm (green line in Fig. 8A). Upon bubbling CO₂ (10 mg mL⁻¹) for 1 h, the aggregates were still spherical (Fig. 8D), although the size of the aggregates increased from 140 nm to 191 nm (blue line in Fig. 8A). The surface of the aggregates became more smooth (Fig. 8D) compared to the aggregates without bubbling CO₂ (Fig. 8B). The zeta potential of the solution containing

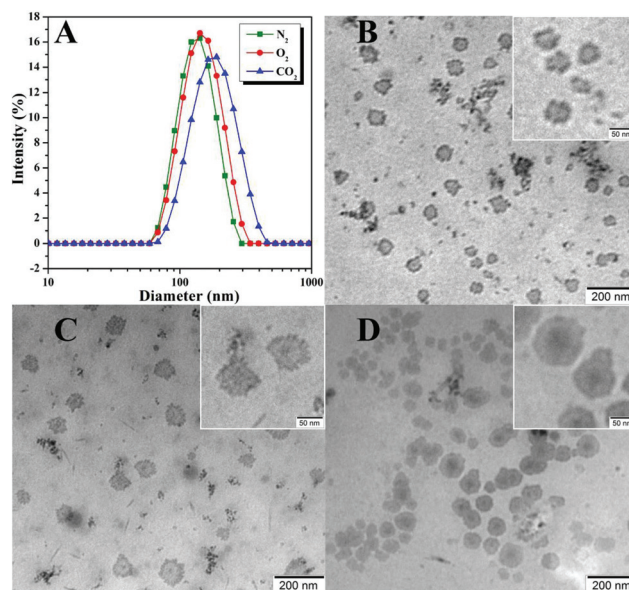


Fig. 8 Hydrodynamic diameter distributions (A) and TEM images of PEG-*b*-poly(TF-DEAE-AM) 4b diblock copolymer in aqueous solution after treatment with N₂ (B), O₂ (C), and CO₂ (D).

PEG-*b*-poly(TF-DEAE-AM) 4b aggregates greatly increased from 9.8 mV to 20.9 mV after CO₂ was injected, which demonstrates that DEAE groups were protonated to DEAE⁺ in order to be extruded out from the core to form a new corona. Fig. 9 shows



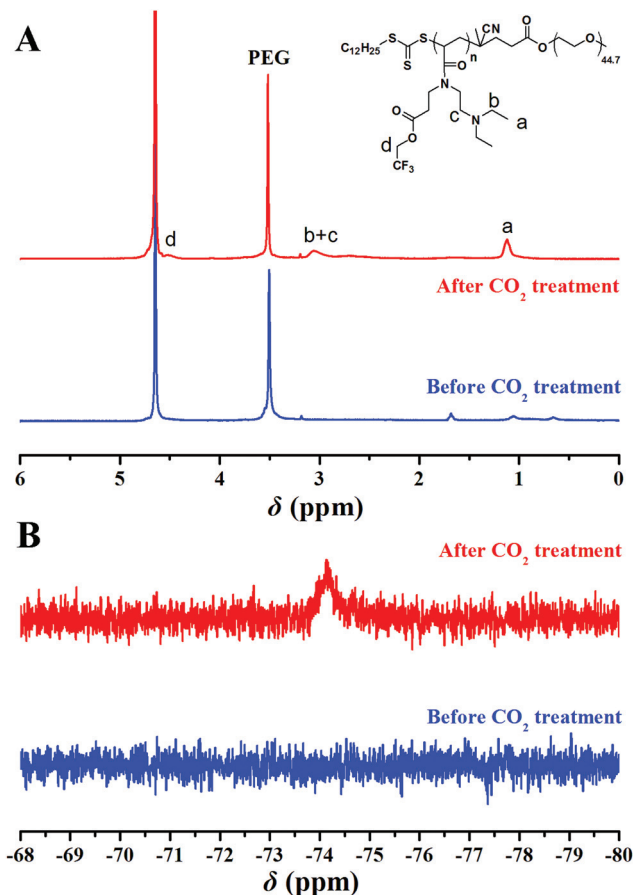


Fig. 9 ^1H (A) and ^{19}F (B) NMR spectra of PEG-*b*-poly(TF-DEAE-AM) **4b** diblock copolymer in D_2O before (blue) and after (red) CO_2 treatment.

^1H and ^{19}F NMR spectra of PEG-*b*-poly(TF-DEAE-AM) **4b** diblock copolymer in D_2O treated with N_2 and CO_2 , respectively. The typical peak at 3.64 ppm corresponding to four protons of the PEG segment could be observed upon the treatment of N_2 , whereas characteristic signals originating from poly(TF-DEAE-AM) block could not be detected both in ^1H (blue line in Fig. 9A) and ^{19}F (blue line in Fig. 9B) NMR

spectra. However, after bubbling CO_2 , the proton resonance signals attributed to DEAE moiety appeared at 1.12 and 3.05 ppm in ^1H NMR spectrum (red line in Fig. 9A), along with the appearance of a signal (-74.2 ppm) coming from TF group in ^{19}F NMR spectrum (red line in Fig. 9B). This observation clearly indicates that DEAE moieties in poly(TF-DEAE-AM) block became soluble, which also improved the solubility of TF units. Thus, it can be concluded that the increase in size of the aggregates upon bubbling CO_2 resulted from the hydration of both DEAE and TF functionalities on the basis of the aforementioned evidence.

Upon bubbling O_2 (10 mg mL^{-1}), although the size of the aggregates only increased from 140 nm to 153 nm (red line in Fig. 8A), an evident change in the outmost layer of aggregates can be noticed (Fig. 8C), where the surface of the aggregates became rougher after bubbling O_2 . We speculate that the weak interaction between TF domains with O_2 improved the solubility of TF domains to some extent, and thus, the size of the aggregates increased because of the partial hydration of TF domains. In addition, the increase in solubility of TF domains might result in micro-phase separation within the core of the aggregates, which leads to a much more rough surface of the aggregates and vague boundary between the aggregates and supporting membrane of TEM grid, although the mobility (solubility) of TF moieties was still not good enough to be detected in ^{19}F NMR spectrum.

In order to further prove the CO_2 - and O_2 -stimuli response behaviors of the aggregates formed by PEG-*b*-poly(TF-DEAE-AM) **4** diblock copolymer and potential application of these aggregates in CO_2 - and O_2 -responsive drug delivery, we examined the release behavior of hydrophobic pyrene, as a model hydrophobic drug loaded in the core of the aggregates upon treatment, with CO_2 and O_2 . The typical UV absorption at around 335 nm attributed to pyrene was observed in an aqueous micellar solution of PEG-*b*-poly(TF-DEAE-AM) **4b** diblock copolymer (blue line in Fig. 10A), which was prepared by adding THF solution containing both pyrene and copolymer **4b** into water, followed by filtration to remove free pyrene and evaporation to remove THF, according to previous literature.⁶³ Since the aqueous solution containing either pyrene (green line in Fig. 10A) or copolymer

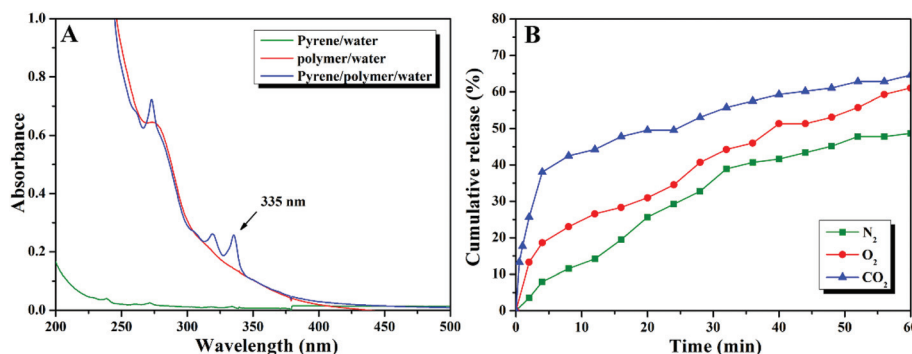


Fig. 10 (A) UV/vis absorption spectra of pyrene/water, PEG-*b*-poly(TF-DEAE-AM) **4b**/water, and pyrene/PEG-*b*-poly(TF-DEAE-AM) **4b**/water; (B) cumulative release profiles of pyrene from the aggregates of PEG-*b*-poly(TF-DEAE-AM) **4b** with the treatment of N_2 , O_2 , and CO_2 .



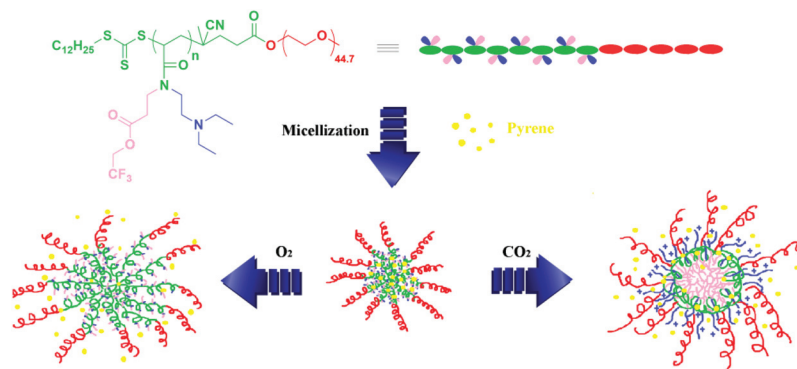


Fig. 11 Fabrication of O_2 - and CO_2 -responsive drug-loaded micelles for the controlled delivery of pyrene.

4b (red line in Fig. 10A) prepared *via* a similar procedure showed almost no UV absorption originating from pyrene, the evident UV absorption in the aqueous solution of copolymer **4b** prepared with the addition of pyrene indicates that pyrene was loaded within the core of aggregates.

Three aliquots of aqueous micellar solution of copolymer **4b** containing pyrene (5 mL) were bubbled with N_2 , O_2 , and CO_2 , separately, and the release of pyrene from the aggregates was traced by monitoring UV absorption of these aqueous solutions varying with time. Fig. 10B shows the cumulative release of pyrene with the stimuli of N_2 , O_2 , and CO_2 . Upon bubbling with O_2 , the release rate increased with an initial burst of about 20% within 5 min and about 55% of pyrene was released from the aggregates after 1 h (red line in Fig. 10B). For the solution treated with CO_2 , almost 40% of the initial pyrene was released within 5 min and the total amount of released pyrene reached about 60% after 1 h (blue line in Fig. 10B). Since the bubbling of gas might also enhance the release of pyrene from the aggregates even for inert gas, we also treated the solution with N_2 as a control experiment. As expected, 7% of the initial pyrene was released from the aggregates after 5 min and about 38% of pyrene escaped from the aggregates after 1 h (green line in Fig. 10B). The release rate of pyrene in both O_2 - and CO_2 -bubbling solutions was much faster than that of the solution bubbling with N_2 , which demonstrates that O_2 and CO_2 make the core of the aggregates more hydrophilic. Furthermore, compared to the solution treated with O_2 , the release rate of the solution bubbling with CO_2 was much faster. This result demonstrates that the core of the aggregates treated with CO_2 was looser than that treated with O_2 , which is consistent with the phenomena observed in DLS and TEM observations (Fig. 11).

Conclusion

In this study, we synthesized a new type of CO_2/O_2 dual-gas-responsive acrylamide monomer by combining CO_2 -responsive DEAE moiety, O_2 -responsive TF group, and polymerizable double bond into a single monomer *via* aza-Michael addition

and amidation reactions. RAFT polymerization was utilized to obtain the target poly(TF-DEAE-AM) homopolymer and PEG-*b*-poly(TF-DEAE-AM) diblock copolymer using CDSTSP and PEG-CTA as the chain transfer agents, respectively. In aqueous solution, the aggregates formed by poly(TF-DEAE-AM) homopolymer and PEG-*b*-poly(TF-DEAE-AM) diblock copolymer show CO_2 and O_2 responsiveness, as demonstrated by the changes in the size and morphologies of the aggregates on bubbling with CO_2 and O_2 . In addition, *in vitro* release measurement indicates that pyrene was remarkably blocked within the micelles in the absence of gas stimuli and the release of pyrene was dramatically increased after bubbling with CO_2 or O_2 compared to N_2 . This study not only explores dual-gas-responsive drug delivery systems, but also provides promising potential applications of poly(TF-DEAE-AM)-containing materials in the biomedical area.

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