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Palladium-catalyzed allylic amination: a powerful tool for the enantioselective synthesis of acyclic nucleoside phosphonates†

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Acyclic nucleoside phosphonates have been prepared in a straightforward manner and in high yields by an enantioselective palladium-catalyzed allylic substitution involving nucleic bases as nucleophiles followed by cross-metathesis reaction with diethyl allylphosphonate.

Introduction

Acyclic nucleoside phosphonates (ANPs)¹ belong to a family of modified nucleoside analogues, in which the sugar moiety has been replaced by a functionalized acyclic chain linking the nucleobase at one end and the phosphonic acid group at the other. Relevant ANPs that have been pursued as antiviral agents are cidofovir (HPMPC), adefovir (PMEA), and tenofovir² (PMPA) (Fig. 1). Because of their limited oral bioavailability, the last two compounds have been converted to their oral prodrug forms, adefovir dipivoxil or bis(pivaloyloxymethyl)-

PMEA [bis(POM)-PMEA] and tenofovir disoproxil or bis(isopropylloxycarbonyloxymethyl)-PMPA [bis(POC)-PMPA], respectively. Many analogues of these compounds have been prepared incorporating modifications in the base, alkyl chain or in the phosphonate moiety. Examples of modification in the alkyl chain are vinyl and allyl ANPs, where a CH₂ group replaces the oxygen of the ether function in the alkyl chain (Fig. 1).³

Acyclic nucleosides are commonly synthesized from nucleic bases and chiral starting materials by alkylation, Mitsunobu reaction, epoxide opening,⁴ and by silver-catalyzed addition of different nucleophiles to 9-allyl-9H-purines.⁵ The reported asymmetric synthesis of acyclic nucleosides involves the transition metal-⁶ or organocatalyzed⁷ aza-Michael reaction of purinic bases to acrylates and asymmetric hydrogenation of purine-substituted acrylates.²

Allylic substitution has also been used in the synthesis of acyclic nucleosides to provide compounds with the nucleic base linked to the terminal carbon.⁸ Hartwig recently reported an enantioselective allylic amination using an iridium catalyst to afford acyclic nucleosides where the nucleic base is bonded to a stereocenter.⁹ Trost described the desymmetrization of *meso* 1,4-cyclopentenyl-dibenzoate derivatives using palladium-DACH catalysts in a straightforward synthesis of carbocyclic nucleosides.¹⁰ He found that the use of nucleobases as nucleophiles has a remarkable effect on the catalytic turnover and the enantioselectivity of desymmetrization reactions.⁹ Furthermore, Pd-catalyzed DYKAT of vinyl epoxide **4** is an efficient procedure for preparing 2-amino-3-buten-1-ol derivatives,¹¹ which are attractive intermediates for synthesizing acyclic nucleosides, although for this purpose it will be necessary to explore the behavior of the different nucleic bases in the reaction. Recently, we have demonstrated that allyl carbonate **5**, substituted with a hydroxymethyl group, is an appropriate substrate for the Pd-catalyzed asymmetric allylic amination reaction (Pd-AAA), affording the branched derivative with high

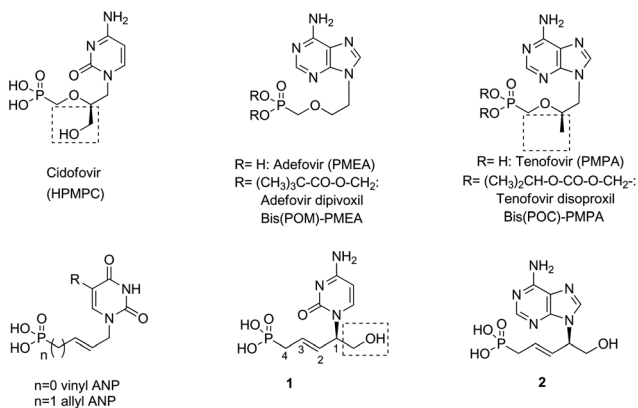


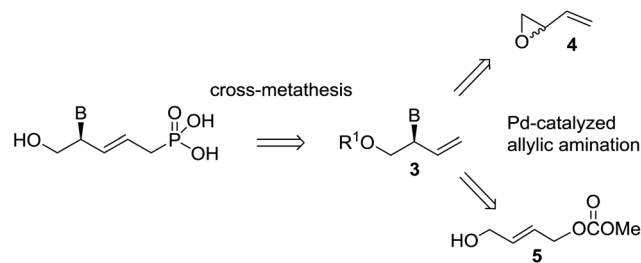
Fig. 1 Relevant reported ANPs and products **1** and **2** described in this paper.

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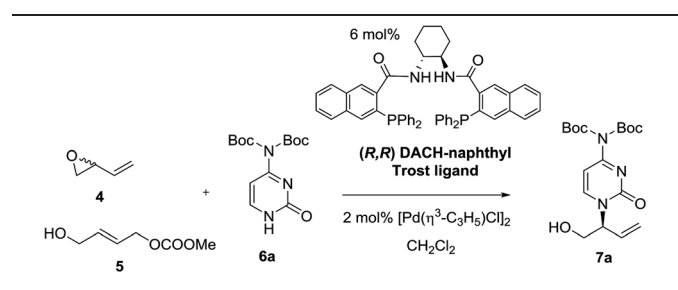
Scheme 1 Retrosynthesis of ANPs 1 and 2.

regio- and enantioselectivities.¹² In this regard, quite a striking different regioselectivity in the reaction of substrates 4 and 5 with amines was observed, with epoxide 4 furnishing equimolar mixtures of linear and branched aminated products. In this work, we report a comparative study of the Pd-catalyzed asymmetric allylic amination reaction starting from vinyl epoxide 4 and carbonate 5 with different nucleic bases, and we show that the resulting derivatives 3 can be transformed into novel ANP nucleosides (1, 2), structurally related to cidofovir, tenofovir and to allyl ANP (Scheme 1). We have also extended this synthesis to the guanine precursor 10 (see Scheme 2).

Results and discussion

Preliminary assays starting from vinyl epoxide 4 and acetylcytosine under reaction conditions optimized in a previous study¹² (6 mol% (*R,R*)-DACH-naphthyl Trost ligand and 2 mol% $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5)\text{Cl}]_2$ as a catalytic system in CH_2Cl_2 , at room temperature), led to the formation of a bis-allylation product containing the 3-buten-1-ol-2-yl units at position 1 and at the exocyclic NH group. This compound was obtained even by reducing the amount of acetylcytosine to 1.1 equivalents. We therefore turned our attention towards di-Boc-cytosine (6a).¹³ When the reaction was conducted under similar conditions using epoxide 4 and di-Boc-cytosine (6a), branched product 7a was obtained in 81% yield and 62% ee (Table 1, entry 1). Decreasing the temperature to 0 °C did not practically improve the enantioselectivity, while conducting the reaction at 35 °C eroded the enantioselectivity significantly (Table 1, entries 2 and 3). The analogous reaction of carbonate 5 with di-Boc-cytosine (6a) using the (*S,S*) ligand (Table 1, entry 4) furnished the enantiomer *ent*-7a in 92% yield and 73% ee. Interesting enough, all reactions from both 4 and 5 were highly regioselective affording exclusively the branched isomer. The use of BSA-KOAc did not improve the results. Lowering the temperature to 0 °C did not improve the enantioselectivity (Table 1, entry 5). Further optimization was focused on the precatalyst loading: a reduction from 2 mol% to 1 mol% produced a drastic decrease in the enantioselectivity (Table 1, entry 6), while an increase in the precatalyst loading to 3 mol% did not provide significant improvement (result not shown in the table). In this context, we selected room tempera-

Table 1 Allylic amination of epoxide 4 or carbonate 5 with cytosine derivative 6a affording 7a using Pd/DACH-naphthyl ligand as catalytic system^a



Entry	Starting material	<i>T</i> (°C)	Conv. ^b (%) (yield) ^c (%)	ee ^d (%)	Product ^e
1 ^f	4	rt	>98 (81)	62	7a
2 ^f	4	0	>98 (85)	65	7a
3 ^f	4	35	>98 (81)	32	7a
4 ^g	5	rt	>98 (92)	73	<i>ent</i> -7a
5 ^g	5	0	>98 (90)	75	<i>ent</i> -7a
6 ^{g,h}	5	rt	>98 (90)	42	<i>ent</i> -7a

^a Conditions: $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5)\text{Cl}]_2$ (2 mol%), naphthyl ligand (6 mol%), substrate (1 equiv.), nucleophile (2 equiv.). Reaction time = 16 h. Concentration = 0.02 M in CH_2Cl_2 . ^b A branched/linear ratio >98 : <2 was observed in all cases. ^c Isolated yield of branched regioisomer. ^d Determined by HPLC on a DAICEL CHIRALCEL OD-H column. ^e The absolute configurations of the products were established by analogy with those obtained by Trost using phthalimide as analogous soft *N*-nucleophiles to nucleic bases (ref. 11a). ^f The (*R,R*)-naphthyl ligand was used. ^g The (*S,S*)-naphthyl ligand was used. ^h 1 mol% $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5)\text{Cl}]_2$ was used.

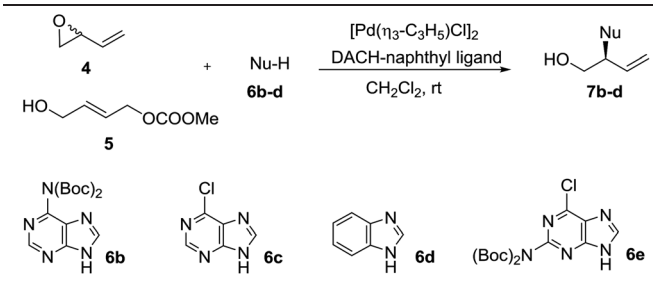
ture and 2 mol% of the palladium precursor as standard reaction conditions.

It can be concluded from Table 1 that, in analogy to reaction from vinyl epoxide 4, Pd-AAA from carbonate 5 with di-Boc-cytosine (6a) using the $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5)\text{Cl}]_2$ /DACH-naphthyl ligand as the catalytic system provides good yields of the branched aminated product 7a/*ent*-7a in complete regioselectivity and moderate enantioselectivity. We therefore decided to extend this comparative study to other purinic bases. The results are collected in Table 2.

Di-Boc-adenosine (6b) reacted with epoxide 4 in the presence of the (*R,R*)-DACH-naphthyl ligand to provide 7b in an excellent yield of 92% and 92% ee (entry 2). When the reaction was conducted with carbonate 5 using the (*S,S*)-DACH-naphthyl ligand, *ent*-7b was obtained in an excellent yield of 92% and 88% ee (entry 2).

6-Chloropurine (6c) is a versatile starting material in nucleoside synthesis since it allows a set of useful and well-known transformations for the synthesis of nucleoside derivatives. When carbonate 5 was treated with 6c in the presence of the (*S,S*)-DACH-naphthyl ligand under the standard conditions, compound *ent*-7c was obtained in very good 91% enantioselectivity, slightly higher than that obtained from 6b (Table 2, entry 2 vs. 4). Pd-AAA of 6c from epoxide 4 furnished the allylated purine *ent*-7c with almost the same enantioselectivity of that obtained from 6b (Table 2, entry 1 vs. 3).



Table 2 Palladium-catalyzed allylic amination reaction of epoxide **4** or carbonate **5** with pyrimidinic and purinic bases **6b–e**^a


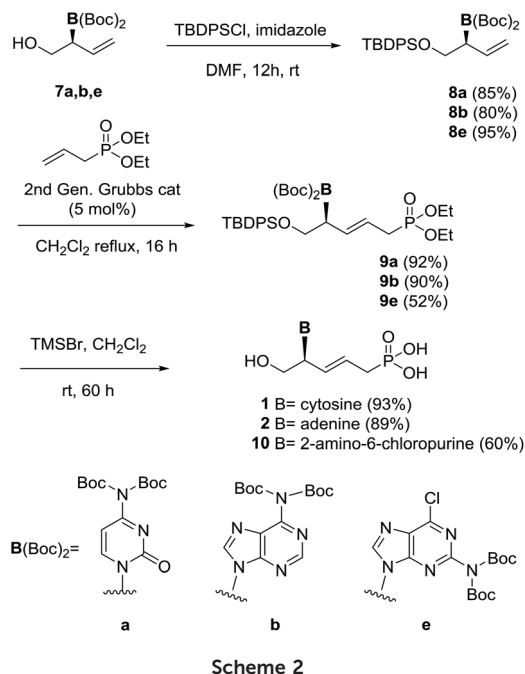
Entry	Starting material	Nucleic base	Product ^b	Conv. ^c (yield) ^d	ee ^e (%)
1 ^f	4	6b	7b	>98 (92)	92
2 ^g	5	6b	<i>ent</i> - 7b	>98 (92)	88
3 ^g	4	6c	<i>ent</i> - 7c	>98 (89)	91.4
4 ^g	5	6c	<i>ent</i> - 7c	>98 (96)	91
5 ^g	4	6d	<i>ent</i> - 7d	>98 (89)	88
6 ^g	5	6d	<i>ent</i> - 7d	>98 (92)	88
7 ^f	4	6e	7e	31 (19)	90
8 ^f	4 ^h	6e	7e	>98 (64)	90
9 ^f	5	6e	7e	72 (43)	92

^a Conditions: [Pd(η^3 -C₃H₅)Cl]₂ (2 mol%), DACH-naphthyl ligand (6 mol%), substrate (1 equiv.), nucleophile (2 equiv.), rt, 16 h. Substrate concentration = 0.02 M. ^b The absolute configurations of the products were established by analogy with those obtained by Trost using phthalimide as analogous soft *N*-nucleophiles to nucleic bases (ref. 11a). ^c A branched/linear ratio >98 : <2 was observed in all cases by ¹H NMR spectroscopy of the crude products. ^d Isolated yield of the branched regioisomer. ^e Determined by HPLC on a DAICEL CHIRALCEL OD-H column. ^f (*R,R*)-DACH-naphthyl ligand was used. ^g (*S,S*)-DACH-naphthyl ligand was used. ^h Using [Pd(η^3 -C₃H₅)Cl]₂ (2 mol%), DACH-naphthyl ligand (6 mol%), substrate (2 equiv.), nucleophile (1 equiv.), rt, 16 h. Substrate concentration = 0.04 M.

Furthermore, the reaction of epoxide **4** and carbonate **5** with benzimidazole (**6d**) as a nucleophile using the (*S,S*)-DACH-naphthyl ligand, afforded compound *ent*-**7d** in good yields and the same enantioselectivity, (88% ee) (Table 2, entries 5 and 6).

Di-Boc-2-amino-6-chloropurine **6e** reacted with epoxide **4** and carbonate **5** under standard conditions to provide **7e** with high enantioselectivity but with low to moderate yields (Table 2, entries 7 and 9). However, on inverting the reaction stoichiometry a substantial improvement in the yield was achieved (64%) with the same enantio-induction (Table 2, entry 8).

Contrary to their complementary behavior in terms of regioselectivity observed with hard amine nucleophiles,¹² both epoxide **4** and carbonate **5** afforded valuable branched products with high degrees of enantioselectivity when reacting with nucleobases. The regioselectivity observed in the reactions of epoxide **4** with nucleobases is comparable to that already observed with other soft nitrogen nucleophiles, such as imides or imidocarboxylates, as a result of a dynamic kinetic asymmetric transformation. As for carbonate **5**, the branched regioisomer is always observed as a major product, regardless of the nitrogen nucleophile used, due to an enantioselective Pd-allylic amination, where hydrogen bonding inter-

**Scheme 2**

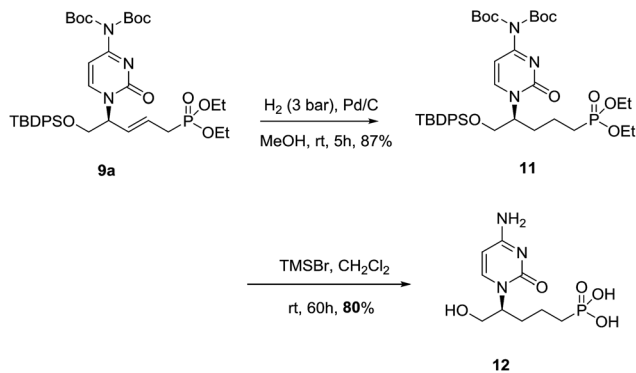
actions play an important role both in the ionization and the nucleophilic attack steps.¹²

Having explored the feasibility of using nucleic bases as nucleophiles for the synthesis of chiral branched heterocycles via a Pd-AAA with the DACH-naphthyl Trost ligand, we selected compounds **7a**, **7b** and **7e** for synthesizing model acyclic pyrimidine and purine nucleosides.

For this purpose, the primary hydroxyl group was initially protected by reaction with *tert*-butyldiphenylsilyl chloride in DMF in the presence of imidazole to afford compounds **8a,b,e** in excellent yields (Scheme 2). According to the retrosynthetic scheme and following previous studies in the field,¹⁴ compounds **8a,b** were then treated with diethyl allylphosphonate^{15,16} in the presence of Grubbs–Nolan's catalyst to afford compounds **9a,b** in excellent 92% and 90% yields, respectively, as a result of a cross-metathesis reaction. The reaction of **8e** under similar conditions afforded **9e** in a moderate 52% yield. Compounds with *E* configuration were exclusively obtained. Removal of the protecting groups¹⁵ in the nucleic base, the phosphonate moiety and the hydroxyl group, was carried out by treatment of compounds **9a,b** with TMSBr¹⁵ in dichloromethane to afford the target acyclic nucleosides **1** and **2** in excellent yields. Additionally, compound **9e** was treated under the deprotection conditions to afford the 2-amino-6-chloro-9*H*-purine derivative **10** in good yield.

The saturated nucleoside **11**, with an increased conformational mobility, was prepared by the reduction of the double bond in **9a** (Scheme 3). Attempts to reduce the double bond under 1 bar of hydrogen pressure and using palladium on charcoal¹⁷ as catalysts led to no conversion. At higher hydrogen pressure (10 bar) the presence of two products was observed by ¹H NMR, corresponding to the expected





Scheme 3

compound **11** and to a secondary product resulting from the reduction of the double bond and partial reduction of the cytosine. Optimal conditions were found driving the reaction at room temperature and using 3 bar of hydrogen pressure. In this way, full conversion and 87% yield of compound **11** was obtained. Removal of protecting groups using TMSBr afforded acyclic nucleoside phosphonate **12** in 80% yield.

In conclusion, acyclic nucleosides **1**, **2**, **10** and **12** were successfully prepared in high yields by palladium-catalyzed allylic amination from compounds **4** and **5**, using pyrimidinic and purinic bases as nucleophiles, followed by ruthenium-catalyzed cross-metathesis with diethyl allylphosphonate and removal of protecting groups. Excellent enantioselectivity was obtained for the purinic derivatives **2** and **10**.

Experimental section

General methods

All chemicals used were reagent grade and used as supplied unless otherwise specified. HPLC grade dichloromethane (CH_2Cl_2), tetrahydrofuran (THF) and dimethylformamide (DMF) were dried using a solvent purification system (Pure-Solv 400). Toluene was purified using standard procedures.¹⁸ ^1H and ^{13}C NMR spectra were recorded on a Varian® Mercury VX 400 (400 MHz and 100.6 MHz respectively) or Varian 400-MR spectrometer in CDCl_3 as a solvent, with chemical shifts (δ) referenced to the internal standard CDCl_3 (7.26 ppm ^1H , 77.23 ppm ^{13}C) or Me_4Si as an internal reference (0.00 ppm). 2D correlation spectra (gCOSY, NOESY, gHSQC, gHMBC) were visualized using a VNMR program (Varian®). ESI MS were run on an Agilent® 1100 Series LC/MSD instrument. Optical rotations were measured at room temperature in a PerkinElmer® 241 MC apparatus with 10 cm cells. IR spectra were recorded on a JASCO FT/IR-600 Plus Fourier transform infrared spectrometer ATR Specac Golden Gate. Optical rotations were measured at 598 nm on a Jasco DIP-370 digital polarimeter using a 100 mm cell. The enantiomeric excess was determined by an HPLC analysis using a DAICEL CHIRALCEL OD-H column. Reactions were monitored by TLC carried out

on 0.25 mm E. Merck® silica gel 60F254 glass or aluminium plates. Developed TLC plates were visualized under a short-wave UV lamp (250 nm) and by heating plates that were dipped in ethanol/ H_2SO_4 (15:1) and a basic solution of potassium permanganate. Flash column chromatography was carried out using forced flow of the indicated solvent on Fluka® or Merck® silica gel 60 (230–400 mesh). Radial chromatography was performed on 1 or 2 mm plates of Kieselgel 60 PF254 silica gel, depending on the amount of the product. Flash column chromatography (FCC) was performed using flash silica gel (32–63 μm) and employed a solvent polarity correlated with TLC mobility.

General procedure 1. Palladium-catalyzed allylic amination from butadiene monoepoxide (4**).** In a Schlenk tube, $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5\text{Cl})_2]$ (2 mol%), the DACH-naphthyl ligand (see tables) (6 mol%), the nucleophile (2.0 eq.) and CH_2Cl_2 ($c = 0.02 \text{ M}$) were introduced under an argon atmosphere. The resulting solution was stirred for 20 minutes. Butadiene monoepoxide (**4**) (1.0 eq.) was then added in one portion. The mixture was stirred at room temperature for 18 h. The reaction mixture was then diluted with H_2O . The phases were separated and the aqueous phase was extracted with CH_2Cl_2 . The combined organic phases were dried over MgSO_4 , filtered, and concentrated under vacuum. The resulting crude was purified by flash chromatography to afford a pure product.

General procedure 2. Palladium-catalyzed allylic amination from (*E*)-4-hydroxybut-2-en-1-yl methyl carbonate (5**).** In a Schlenk tube, $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5\text{Cl})_2]$ (2 mol%), the (*S,S*)-DACH-naphthyl ligand (6 mol%) and CH_2Cl_2 ($c = 0.02 \text{ M}$) were introduced under an argon atmosphere. The resulting mixture was stirred for 20 minutes. Carbonate **5** (1.0 eq.) and nucleophile (1.1 eq.) were then successively introduced. The mixture was stirred at room temperature for 18 h. The reaction mixture was then diluted with H_2O . The phases were separated and the aqueous phase was extracted with CH_2Cl_2 . The combined organic phases were dried over MgSO_4 , filtered, and concentrated under vacuum. The resulting crude was purified by flash chromatography to afford a pure product.

(*R*)-*N,N*-Bis(*tert*-butoxycarbonyl)-1-(1-hydroxybut-3-en-2-yl)-cytosine (ent-7a**).** Following the general procedure 2, compound **ent-7a** was prepared from carbonate **5** (17 mg, 0.116 mmol), compound **6a** (72 mg, 0.23 mmol), $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5\text{Cl})_2]$ (1 mg, 0.002 mmol) and (*S,S*)-DACH-naphthyl ligand (5.5 mg, 0.008 mmol) in dichloromethane (12 mL) at $-10 \text{ }^\circ\text{C}$. Purification by silica chromatography (hexanes/ EtOAc 10:2) provided 41 mg (90%) of compound **ent-7a** as a yellow syrup and 75% ee (determined by chiral HPLC (Daicel Chiralcel OD-H, *n*-hexane- i PrOH, 90:10, flow = 0.5 mL min^{-1} , detection, UV 210 nm; retention times (min), 9.79, 11.42, $t_{\text{R}}(\text{R}) = 9.79 \text{ min}$ and $t_{\text{R}}(\text{S}) = 11.42 \text{ min}$). $[\alpha]_{\text{D}}^{25} +7.13$ (c 1.06, CHCl_3). FTIR-ATR (cm^{-1}): 2918, 2849, 1775, 1741, 1665, 1460, 1369, 1313, 1253, 1154, 1131, 788. ^1H NMR (CDCl_3 , 400 MHz): δ (ppm) 7.67 (d, 1H, $J = 7.5 \text{ Hz}$), 7.05 (d, 1H, $J = 7.5 \text{ Hz}$), 6.00 (ddd, 1H, $J = 17.0, 10.6, 5.9 \text{ Hz}$), 5.42 (dd, 1H, $J = 10.6, 1.5 \text{ Hz}$), 5.30 (dd, 1H, $J = 17.0, 1.5 \text{ Hz}$), 5.28–5.23 (m, 1H), 4.00–3.94 (m, 2H), 3.15 (bs, 1H), 1.55 (s, 18H). ^{13}C NMR (CDCl_3 , 100.6 MHz): δ (ppm)



161.9, 155.8, 149.7, 147.2, 147.0, 132.7, 120.5, 96.2, 85.1, 62.7, 61.2, 27.8. HRMS (ESI-TOF) m/z : $[M + H]^+$ calcd for $C_{18}H_{28}N_3O_6$: 382.1973, found: 382.1964.

(S)-N,N-Bis(tert-butoxycarbonyl)-9-(1-hydroxybut-3-en-2-yl)adenine (7b). Following the general procedure 1, compound **7b** was prepared from butadiene monoepoxide (**4**) (0.02 mL, 0.25 mmol), compound **6b** (100 mg, 0.3 mmol), $[Pd(\eta^3-C_3H_5)Cl]_2$ (2 mg, 0.0054 mmol) and (*R,R*)-DACH-naphthyl ligand (13 mg, 0.016 mmol) in dichloromethane (27 mL). Purification by silica gel chromatography (hexanes/EtOAc 1:1) provided 93 mg (92%) of product **7b** as a yellow syrup and 92% ee (determined by chiral HPLC (Daicel Chiralcel OD-H, *n*-hexane-ⁱPrOH 85:15, flow = 0.5 mL min⁻¹, detection, UV 254 nm; retention times (min), 11.77, 13.47, $t_R(R)$ = 11.77 min and $t_R(S)$ = 13.47 min)). $[\alpha]_D^{25}$ -15.80 (*c* 0.92, CHCl₃). FTIR-ATR (cm⁻¹): 3347, 2979, 2926, 1787, 1600, 1107. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 8.84 (s, 1H), 8.17 (s, 1H), 6.20 (ddd, 1H, *J* = 17.1, 10.5, 6.4 Hz), 5.40 (d, 1H, *J* = 10.5 Hz), 5.30–5.18 (m, 1H), 5.14 (d, 1H, *J* = 17.1 Hz), 4.26–4.10 (m, 2H), 3.80 (bs, 1H), 1.47 (s, 18H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 153.0, 151.7, 150.8, 150.6, 145.1, 132.7, 129.1, 119.7, 84.1, 63.8, 61.3, 27.9. HRMS (ESI-TOF) m/z : $[M + H]^+$ calcd for $C_{18}H_{28}N_3O_5$: 406.2085, found: 406.2073.

(R)-6-Chloro-9-(1-hydroxybut-3-en-2-yl)-9H-purine (ent-7c). Following the general procedure 1, compound **ent-7c** was prepared from butadiene monoepoxide (**4**) (0.024 mL, 0.3 mmol), compound **6d** (50 mg, 0.32 mmol), $[Pd(\eta^3-C_3H_5)Cl]_2$ (2 mg, 0.006 mmol) and (*S,S*)-DACH-naphthyl ligand (14 mg, 0.017 mmol) in dichloromethane (30 mL) at rt. Purification by silica gel chromatography (hexanes/EtOAc 1:1) provided 60 mg (89%) product **ent-7c** as a colourless oil and 91.4% ee (determined by chiral HPLC (Daicel Chiralcel OD-H, *n*-hexane-ⁱPrOH, 90:10, flow = 1 mL min⁻¹, detection, UV 210 nm; retention times (min), 17.12, 18.36, $t_R(R)$ = 18.36 min and $t_R(S)$ = 17.12 min)). $[\alpha]_D^{25}$ +10.8 (*c* 1.23, CHCl₃). FTIR-ATR (cm⁻¹): 3347, 2927, 1591, 1561, 1337. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 8.70 (s, 1H), 8.23 (s, 1H), 6.21 (ddd, 1H, *J* = 17.0, 10.4, 6.4 Hz), 5.43 (dd, 1H, *J* = 10.4, 1.4 Hz), 5.30–5.25 (m, 1H), 5.24 (dd, 1H, *J* = 17.0, 1.4 Hz), 4.28–4.22 (m, 1H), 4.21–4.15 (m, 1H), 3.86 (t, 1H, *J* = 6.3 Hz). ¹³C NMR (CDCl₃, 100.6 MHz): 151.8, 151.5, 151.4, 145.4, 132.2, 131.8, 120.4, 63.7, 61.0. HRMS (ESI-TOF) m/z : $[M + H]^+$ calcd for $C_9H_{10}ClN_4O$: 225.0538, found: 225.0532.

(R)-1-(1-Hydroxybut-3-en-2-yl)-1H-benzo[d]imidazole (ent-7d). Following the general procedure 2, compound **ent-7d** was prepared from carbonate **5** (17 mg, 0.116 mmol), compound **6d** (72 mg, 0.23 mmol), $(\eta^3-C_3H_5)PdCl_2$ (1 mg, 0.002 mmol) and (*S,S*)-DACH-naphthyl ligand (5.5 mg, 0.008 mmol) in dichloromethane (12 mL). Purification by silica gel chromatography (hexanes/EtOAc 1:1) provided 20 mg (92%) of product **ent-7d** as a colourless oil and 88% ee (determined by chiral HPLC (Daicel Chiralcel OD-H, *n*-hexane-ⁱPrOH, 92:08, flow = 1.0 mL min⁻¹, detection, UV 220 nm; retention times (min), 17.84, 25.57, $t_R(R)$ = 17.84 min and $t_R(S)$ = 25.57 min)). $[\alpha]_D^{25}$ +26.0 (*c* 0.80, CHCl₃). FTIR-ATR (cm⁻¹): 3089, 2922, 1492, 1457. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 7.84 (s, 1H), 7.53 (d, 1H,

J = 7.7 Hz), 7.36 (d, 1H, *J* = 7.7 Hz), 7.21 (t, 1H, *J* = 7.7 Hz), 7.14 (t, 1H, *J* = 7.7 Hz), 6.11 (ddd, 1H, *J* = 17.0, 10.8, 5.7 Hz), 5.37 (d, 1H, *J* = 10.8 Hz), 5.20 (d, 1H, *J* = 17.0 Hz), 5.05–4.97 (m, 1H), 4.22–4.10 (m, 2H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 143.2, 142.2, 140.8, 132.9, 123.0, 122.5, 119.8, 119.4, 110.7, 63.3, 60.6. HRMS (ESI-TOF) m/z : $[M + H]^+$ calcd for $C_{11}H_{13}N_2O$: 189.1022, found: 189.1017.

(S)-2-Bis-tert-butoxycarbonyl-amino-6-chloro-9-(1-hydroxybut-3-en-2-yl)-9H-purine (7e). Following the general procedure 2, compound **7e** was prepared from carbonate **5** (17 mg, 0.116 mmol), compound **6e** (85 mg, 0.23 mmol), $[Pd(\eta^3-C_3H_5)Cl]_2$ (1 mg, 0.002 mmol) and (*R,R*)-DACH-naphthyl ligand (5.5 mg, 0.008 mmol) in dichloromethane (6 mL) at room temperature. Purification by silica chromatography (hexanes/EtOAc 4:6) provided 22 mg (43%) of compound **7e** as a yellow syrup and 92% ee (determined by chiral HPLC (Daicel Chiralcel OD-H, *n*-hexane-ⁱPrOH, 90:10, flow = 0.6 mL min⁻¹, detection, UV 230 nm; retention times (min), 13.5, 15.1, $t_R(R)$ = 13.5 min and $t_R(S)$ = 15.1 min)). $[\alpha]_D^{25}$ -34.0 (*c* 0.1, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 8.25 (s, 1H), 6.17 (ddd, *J* = 16.9, 10.5, 6.3 Hz, 1H), 5.40 (d, *J* = 10.4 Hz, 1H), 5.24–5.16 (m, 2H), 4.22 (dd, *J* = 12, 5.9 Hz, 1H), 4.11 (dd, *J* = 12.0, 3.0 Hz, 1H), 3.27 (br s, 1H), 1.44 (s, 18H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 152.2, 151.5, 151.3, 150.4, 146.0, 132.2, 129.9, 120.0, 83.9, 63.4, 60.9, 27.8. HRMS (ESI-TOF) m/z : $[M + Na]^+$ calcd for $C_{19}H_{26}ClN_5O_5Na$: 462.1520, found: 462.1524.

(S)-N,N-Bis(tert-butoxycarbonyl)-1-[1-(tert-butylidiphenylsilyloxy)but-3-en-2-yl]cytosine (8a). Compound **7a** (100 mg, 0.26 mmol) was dissolved in dry DMF (3 mL), treated under argon with *tert*-butylidiphenylchlorosilane (0.08 mL, 0.3 mmol) and imidazole (40 mg, 0.6 mmol), and the reaction mixture was stirred for 16 h at room temperature. The solution was then diluted with water and extracted with Et₂O, and the organic layer was dried and evaporated. The residue was purified by column chromatography on silica gel (hexanes/EtOAc 10:2) to provide 137 mg (85%) of compound **8a**. $[\alpha]_D^{25}$ -13.5 (*c* 1.27, CHCl₃). FTIR-ATR (cm⁻¹): 2931, 2857, 1742, 1671, 1524, 1455, 1370, 1319, 1256, 1137, 1110, 784, 701. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 7.75 (d, 1H, *J* = 7.5 Hz), 7.57 (d, 2H, *J* = 6.4 Hz), 7.48 (d, 2H, *J* = 6.4 Hz), 7.49–7.30 (m, 6H), 7.98 (d, 1H, *J* = 7.5 Hz), 6.01 (ddd, 1H, *J* = 17.6, 10.4, 6.0 Hz), 5.38–5.34 (m, 2H), 5.32 (d, 1H, *J* = 17.6 Hz), 3.97 (d, 2H, *J* = 3.6 Hz), 1.57 (s, 18H), 1.02 (s, 9H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 161.9, 155.0, 149.8, 147.1, 135.7, 135.5, 133, 132.6, 130.1, 128.0, 120.6, 95.6, 84.9, 63.9, 59.6, 27.9, 26.7, 19.3. HRMS (ESI-TOF) m/z : $[M + H]^+$ calcd for $C_{34}H_{46}N_3O_6Si$ 620.3156, found 620.3171.

(S)-N,N-Bis(tert-butoxycarbonyl)-9-[1-(tert-butylidiphenylsilyloxy)but-3-en-2-yl]adenine (8b). Compound **7b** (250 mg, 0.6 mmol) was dissolved in dry DMF (7 mL) and treated under argon with *tert*-butylidiphenylchlorosilane (0.18 mL, 0.66 mmol) and imidazole (90 mg, 1.32 mmol), and the mixture was then stirred for 16 h at room temperature. The solution was then diluted with water and extracted with Et₂O, and the organic layer was dried and evaporated. The residue was purified by column chromatography on silica gel (hexanes/EtOAc 10:2) to provide 319 mg (80%) of compound **8b** as a colourless oil. $[\alpha]_D^{25}$



–10.93 (*c* 1.13, CHCl₃). FTIR-ATR (cm⁻¹): 2987, 2362, 1733, 1716, 1558, 1540, 1507, 1456, 1395, 1259, 1066, 749. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 8.79 (s, 1H), 8.26 (s, 1H), 7.50–7.25 (m, 10H), 6.02 (ddd, 1H, *J* = 17.2, 10.4, 6.4 Hz), 5.34 (dd, 1H, *J* = 11.6, 0.8 Hz), 5.30–5.20 (m, 1H), 5.17 (dd, 1H, *J* = 17.2, 0.8 Hz), 4.16 (dd, 1H, *J* = 11.2, 6.8 Hz), 4.05 (dd, 1H, *J* = 10.8, 4 Hz), 1.43 (s, 18H), 0.94 (s, 9H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 153.3, 151.8, 150.5, 150.3, 144.5, 135.5, 135.4, 132.5, 130.1, 128.9, 127.9, 119.8, 83.7, 64.8, 59.4, 27.9, 26.8, 19.1. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ calcd for C₃₅H₄₅N₅NaO₅Si 666.3088, found 666.3073.

(*S*)-2-Bis-*tert*-butoxycarbonyl-amino-6-chloro-9-[1-(*tert*-butyl diphenylsilyloxy)-but-3-en-2-yl]-9H-purine (**8e**). Compound **7e** (113 mg, 0.257 mmol) was dissolved in dry DMF (3 mL) and treated under argon with *tert*-butyldiphenylchlorosilane (80.1 μ L, 0.308 mmol) and imidazole (40 mg, 0.59 mmol), and the mixture was then stirred for 16 h at room temperature. The solution was then diluted with water and extracted with Et₂O, and the organic layer was dried and evaporated. The residue was purified by column chromatography on silica gel (hexanes/EtOAc 10 : 2) to provide 244 mg (95%) of compound **8e** as a colourless oil. [α]_D²⁵ –11.0 (*c* 0.3, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.33 (s, 1H), 7.55–7.51 (m, 2H), 7.49–7.46 (m, 2H), 7.45–7.30 (m, 6H), 6.16 (ddd, *J* = 17.0, 10.5, 6.3 Hz, 1H), 5.34 (dd, *J* = 10.5, 0.8 Hz, 1H), 5.27–5.19 (m, 1H), 5.18–5.09 (m, 1H), 4.13 (dd, *J* = 11.1, 5.6 Hz, 1H), 4.01 (dd, *J* = 11.1, 3.8 Hz, 1H), 1.39 (s, 18H), 1.01–0.95 (m, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 152.4, 151.7, 150.9, 150.4, 145.7, 135.5, 135.3, 132.3, 132.2, 132.1, 130.1, 130.0, 129.7, 127.9, 119.9, 83.4, 64.7, 59.2, 27.8, 26.7, 19.1. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ calcd for C₃₅H₄₄ClN₅O₅SiNa: 700.2698, found: 700.2700.

Diethyl (2*E*,4*S*)-*N,N*-bis(*tert*-butoxycarbonyl)-1-[1-(*tert*-butyl diphenylsilyloxy)-5-phosphonopent-3-en-2-yl]cytosine (**9a**). To a solution of product **8a** (50 mg, 0.08 mmol) and II generation Grubbs catalyst (4 mg, 0.004 mmol) in dichloromethane (4 mL) was added diethylallylphosphonate (0.06 mL, 0.32 mmol) and the reaction mixture was heated to reflux for 16 h. The solvent was then evaporated and the reaction crude was purified by silica gel chromatography (hexanes/EtOAc 1 : 2) to provide 57 mg (92%) of compound **9a** as a yellow liquid. [α]_D²⁵ –21.4 (*c* 1.2, CHCl₃). FTIR-ATR (cm⁻¹): 2969, 2931, 1741, 1671, 1455, 1370, 1319, 1255, 1111, 1024, 735, 701. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 7.68 (d, 1H, *J* = 7.6 Hz), 7.56 (d, 2H, *J* = 6.4 Hz), 7.45 (d, 2H, *J* = 6.4 Hz), 7.42–7.30 (m, 6H), 6.97 (d, 1H, *J* = 7.6 Hz), 5.87–5.79 (m, 2H), 5.29 (brs, 1H), 4.15–4.02 (m, 4H), 3.96 (d, 2H, *J* = 4.0 Hz), 2.60 (dd, 2H, *J* = 21.0, 6.4 Hz), 1.57 (s, 18H), 1.27 (t, 6H, *J* = 6.8 Hz), 1.03 (s, 9H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 161.7, 154.6, 149.5, 146.9, 135.5, 135.3, 132.5, 132.4, 130, 129.9, 129.5, 129.4, 128.5, 127.8, 126.5, 126.3, 95.4, 84.7, 63.6, 62, 59.4, 30.6 (*J* = 140 Hz) 27.7, 26.8, 19.1, 16.4. HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₃₉H₅₇N₃O₉PSi 770.3602, found 770.3628.

Diethyl (2*E*,4*S*)-*N,N*-bis(*tert*-butoxycarbonyl)-1-[1-(*tert*-butyl diphenylsilyloxy)-5-phosphonopent-3-en-2-yl]adenine (**9b**). To a solution of product **8b** (140 mg, 0.22 mmol) and II generation Grubbs catalyst (9 mg, 0.011 mmol) in dichloromethane

(11 mL) was added diethylallylphosphonate (0.16 mL, 0.88 mmol) and the reaction mixture was heated to reflux for 16 h. The solvent was then evaporated and the reaction crude was purified by silica gel chromatography (hexanes/EtOAc 2 : 1) to provide 157 mg (90%) of product **9b** as a green liquid. [α]_D²⁵ –14.6 (*c* 1.36, CHCl₃). FTIR-ATR (cm⁻¹): 2929, 2856, 1788, 1599, 1452, 1369, 1252, 1139, 1111, 1026, 704. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 8.77 (s, 1H), 8.20 (s, 1H), 7.42–7.29 (m, 10H), 6.15–6.02 (m, 1H), 5.76 (ddd, 1H, *J* = 15.6, 10.4, 7.6 Hz), 5.25 (brs, 1H), 4.16 (dd, 2H, *J* = 10.4, 6.4 Hz), 4.07–3.95 (m, 4H), 2.56 (dd, 1H, *J* = 21.0, 7.6 Hz), 1.43 (s, 18H), 1.22 (t, 6H, *J* = 6.8 Hz), 0.93 (s, 9H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 153.2, 151.9, 150.6, 150.4, 144.5, 135.2, 132.4, 130.1, 129.1, 129.0, 128.0, 126.2, 83.8, 64.8, 62.2, 62.1, 59.2, 30.6 (*J* = 140 Hz), 28.0, 26.9, 19.2, 16.6. HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₄₀H₅₇N₅O₈PSi 794.3714, found 794.3694.

Diethyl (2*E*,4*S*) 2-bis-*tert*-butoxycarbonyl-amino-6-chloro-9-[1-(*tert*-butyl diphenylsilyloxy)-5-phosphonopent-3-en-2-yl]-9H-purine (**9e**). To a solution of product **8e** (134 mg, 0.197 mmol) and II generation Grubbs catalyst (8 mg, 0.009 mmol) in dichloromethane (8 mL) was added diethylallylphosphonate (0.134 mL, 0.788 mmol) and the reaction mixture was heated to reflux for 16 h. The solvent was then evaporated and the reaction crude was purified by silica gel chromatography (hexanes/EtOAc 1 : 2) to provide 86 mg (52%) of compound **9e** as a colourless oil. [α]_D²⁵ –2.4 (*c* 0.4, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.22 (s, 1H), 7.55–7.26 (m, 10H), 6.09–5.98 (m, 1H), 5.75 (dt, *J* = 14.6, 7.0 Hz, 1H), 5.18 (brs, 1H), 4.10 (dd, *J* = 11.0, 5.6 Hz, 2H), 4.03 (ddd, *J* = 14.3, 9.3, 5.3 Hz, 2H), 3.96 (dd, *J* = 11.0, 3.8 Hz, 2H), 2.61 (d, *J* = 7.4 Hz, 1H), 2.55 (d, *J* = 7.4 Hz, 1H), 1.40 (s, 18H), 1.24 (dd, *J* = 12.8, 7.0 Hz, 6H), 0.97 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 152.1, 151.6, 150.9, 150.5, 145.5, 135.4, 135.3, 132.1, 130.1, 130.0, 129.8, 128.6, 128.5, 127.9, 127.8, 126.5, 126.4, 83.5, 64.7, 62.1, 61.9, 58.7, 31.2, 29.8, 27.9, 26.7, 19.0, 16.5, 16.4, 16.3. HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₄₀H₅₆ClN₅O₈PSi: 828.3324, found 828.3364.

(2*E*,4*S*)-1-[5-Phosphono-1-hydroxypent-3-en-2-yl]cytosine (**1**). TMSBr (0.08 mL, 0.6 mmol) was added to a solution of compound **9a** (80 mg, 0.10 mmol) in CH₂Cl₂ (7 mL), and the mixture was stirred for 60 h at room temperature under positive pressure of dry Ar. The reaction was quenched by adding MeOH (3 mL) and the reaction mixture was then evaporated by heating at 60 °C. This process was repeated three times. The resulting residue was extracted with H₂O and CH₂Cl₂, and the aqueous phase was evaporated to dryness to afford compound **1** (25 mg, 93%) as a yellow liquid. [α]_D²⁵ –11.2 (*c* 2.89, MeOH). FTIR-ATR (cm⁻¹): 2969, 1715, 1669, 1540, 1394, 1043, 973, 872, 793, 762, 748, 702. ¹H NMR (D₂O, 400 MHz): δ (ppm) 7.85 (d, 1H, *J* = 7.8 Hz), 6.16 (d, 1H, *J* = 7.8 Hz), 5.75–5.56 (m, 2H), 5.16 (bs, 1H), 3.88 (d, 2H, *J* = 6.0 Hz), 2.45 (d, 2H, *J* = 21.0, 7.0 Hz). ¹³C NMR (D₂O, 100.6 MHz): δ (ppm) 158.8, 149.0, 147.0, 127.5, 127.4, 94.6, 60.9, 59.8, 31.8 (d, *J* = 130 Hz). HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₉H₁₄N₃O₅P 276.0749, found 276.0719.

(2*E*,4*S*)-1-[5-Phosphono-1-hydroxypent-3-en-2-yl]adenine (**2**). TMSBr (0.64 mL, 0.36 mmol) was added to a solution of compound **9b** (50 mg, 0.06 mmol) in CH₂Cl₂ (4 mL), and the



mixture was stirred for 60 h at room temperature under positive pressure of dry Ar. The reaction was quenched by adding MeOH (3 mL) and the reaction mixture was then evaporated by heating at 60 °C. This process was repeated three times. The resulting residue was extracted with H₂O and CH₂Cl₂, and the aqueous phase was evaporated to dryness to afford compound **2** (16 mg, 89%) as a yellow liquid. $[\alpha]_D^{25}$ -12.4 (*c* 1.93, MeOH). FTIR-ATR (cm⁻¹): 3070, 2325, 1691, 1609, 1531, 1496, 1425, 1387, 1224, 1107, 937, 770. ¹H NMR (D₂O, 400 MHz): δ (ppm) 8.44 (s, 1H), 8.40 (d, 1H), 6.07–5.97 (m, 1H), 5.84–5.69 (m, 1H), 5.38–5.31 (m, 1H), 4.12 (dd, 1H, *J* = 12.0, 8.4 Hz), 4.06 (dd, 1H, *J* = 12.0, 5.2 Hz), 2.62 (dd, 2H, *J* = 21.0, 7.0 Hz). ¹³C NMR (D₂O, 100.6 MHz): δ (ppm) 149.6, 148.3, 143.9, 143.6, 127.8, 127.6, 118.1, 62.3, 59.2, 32.4 (d, *J* = 130 Hz). HRMS (ESI-TOF) *m/z*: [M + Na]⁺ calcd for C₁₀H₁₅N₅NaO₄P 323.0759, found 323.0769.

(2E,4S)-2-Amino-6-chloro-9-(5-phosphono-1-hydroxypent-3-en-2-yl)-9H-purine (10). TMSBr (0.75 mL, 0.572 mmol) was added to a solution of compound **9e** (79 mg, 0.095 mmol) in CH₂Cl₂ (7 mL), and the mixture was stirred for 60 h at room temperature under positive pressure of dry Ar. The reaction was quenched by adding MeOH (3 mL) and the reaction mixture was then evaporated by heating at 60 °C. This process was repeated three times. The resulting residue was extracted with H₂O and CH₂Cl₂, and the aqueous phase was evaporated to dryness to afford compound **10** (19 mg, 60%) as a brown oil. $[\alpha]_D^{25}$ +6.22 (*c* 0.9, MeOH). ¹H NMR (500 MHz, D₂O) δ 8.92 (s, 1H), 5.98–5.88 (m, 1H), 5.79 (td, *J* = 14.4, 7.0 Hz, 1H), 5.23 (br s, 1H), 4.03 (dd, *J* = 12.2, 6.5 Hz, 1H), 3.97 (dd, *J* = 12.2, 4.1 Hz, 1H), 2.64 (d, *J* = 7.3 Hz, 1H), 2.59 (d, *J* = 7.4 Hz, 1H). ¹³C NMR (126 MHz, D₂O) δ 155.2, 149.9, 137.5, 136.7, 135.6, 128.0, 107.7, 61.8, 60.9 (m), 31.2 (d, *J* = 132.1 Hz). HRMS (ESI-TOF) *m/z*: [M + Na]⁺ calcd for C₁₀H₁₃ClN₅NaO₄P 356.0291, found 356.0381.

Diethyl (S)-N,N-bis(tert-butoxycarbonyl)-1-[1-(tert-butylidiphenylsilyloxy)-5-phosphonopent-2-yl]cytosine (11). To a solution of **9a** (150 mg, 0.2 mmol) in MeOH (6 mL), 5% palladium/carbon was added. The mixture was stirred under a hydrogen pressure (3 bar) at room temperature for 5 h. The reaction mixture was then filtered through Celite, washed with MeOH and concentrated under reduced pressure. The crude was purified by silica gel column chromatography (EtOAc) to give the compound **11** as a colourless oil (0.134 g, 87%). $[\alpha]_D^{25}$ -16.0 (*c* 2.00, CHCl₃). FTIR-ATR (neat) 2930, 1743, 1669, 1456, 1320, 1264, 1137, 1113, 733, 702 cm⁻¹. ¹H NMR (CDCl₃, 400 MHz): δ (ppm) 7.72 (d, 1H, *J* = 7.2 Hz), 7.55 (d, 2H, *J* = 7.6 Hz), 7.47 (d, 2H, *J* = 7.6 Hz), 7.40–7.30 (m, 6H), 6.99 (d, 1H, *J* = 7.2 Hz), 4.80 (brs, 1H), 4.12–3.93 (m, 4H), 3.84 (dd, 1H, *J* = 3.6, 11.2 Hz), 3.80 (dd, 1H, *J* = 3.2, 11.2 Hz), 1.95–1.86 (m, 2H), 1.77–1.63 (m, 4H), 1.55 (s, 18H), 1.26 (t, 6H, *J* = 6.8 Hz), 1.01 (s, 9H). ¹³C NMR (CDCl₃, 100.6 MHz): δ (ppm) 161.8, 155.3, 149.8, 146.2, 135.7, 132.7, 130.2, 128.0, 95.7, 85, 64.4, 61.8, 61.7, 29.8, 27.9, 26.5 (*J* = 140 Hz), 24.5, 19.3, 16.7, 16.6. HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₃₉H₅₉N₃O₉PSi 772.3758, found: 772.3733.

(S)-1-[5-Phosphono-1-hydroxypent-2-yl]cytosine (12). TMSBr (0.15 ml, 0.9 mmol) was added to a solution of compound **11**

(120 mg, 0.15 mmol) in CH₂Cl₂ (12 mL) and the mixture was stirred for 60 h at room temperature under positive argon pressure. Then MeOH (3 mL) was added and evaporated with heating (*ca.* 60 °C). This process was repeated three times. The residue was extracted with H₂O and CH₂Cl₂, and the aqueous phase was evaporated. The crude product was purified by reverse phase chromatography on Silica C18 using (H₂O, MeOH) as the eluent to give 33 mg (80%) of compound **12** as a white foam. $[\alpha]_D^{25}$ -12.0 (*c* 0.20, MeOH). FTIR-ATR (neat, cm⁻¹): 3327, 2918, 1668, 1540, 1456, 1387, 1148, 1050, 920, 785 cm⁻¹. ¹H NMR (D₂O, 400 MHz): δ (ppm) 7.87 (d, 1H, *J* = 8.0 Hz), 6.19 (d, 1H, *J* = 8.0 Hz), 4.67–4.62 (m, 1H), 3.76–3.70 (m, 2H), 1.81–1.75 (m, 4H), 1.55–1.48 (m, 2H). ¹³C NMR (D₂O, 100.6 MHz): δ (ppm) 158.7, 149.4, 146.2, 94.2, 61.7, 58.5, 28.8, 25.4 (*J* = 130 Hz), 18.3. HRMS (ESI-TOF) *m/z*: [M + Na]⁺ calcd for C₉H₁₆N₃NaO₅P: 300.0725, found: 300.0733.

Conflicts of interest

There are no conflicts to declare.

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