Externally controllable glycan presentation on nanoparticle surfaces to modulate lectin recognition†

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Nature dynamically controls carbohydrate expression on cells rather than static presentation. Here we report synthetic glycosylated nanoparticles that contain polymeric ‘gates’ to enable external control (via temperature changes) of glycan surface expression, as an alternative to enzymatic control in nature. This approach offers a new dynamic multivalent scaffold for glycan recognition.

Glycans (sugars) mediate a diverse range of biological recognition and signal transduction pathways and are implicated in diseases such as cancer (aberrant glycosylation) or as sites for pathogen adhesion. The ‘readers’ of glycosylation state are lectins; carbohydrate binding proteins which are neither antibodies nor enzymes.1 The typical affinity for a glycan to a lectin is rather weak, so Nature presents multiple copies of glycans on cell surfaces to benefit from the cluster glycoside effect – a non-linear enhancement in binding affinity when multiple glycans are present in proximity to each other.2 Inspired by this, multivalent systems such as polymers, peptides, surfaces or nanoparticles functionalized with glycans have been used to generate high avidity binders. Due to their high affinity, glycopolymers3,4 have been explored as anti-adhesive agents against, HIV,5 cholera,6 Shiga toxins7 and also to recruit growth factors to control stem cell fate8 with affinities on the nM scale.

Despite this vast range of structures synthesized, most glycomaterials are static entities with the sugars always accessible for binding. This is in stark contrast to cell-surface glycans which are highly dynamic with the glycans presented changing depending on disease state and for protein folding quality control.9,10 Current synthetic materials do not enable control over glycan expression to be modulated, and hence do not fully mimic the natural environment. Dynamic chemical bonds have been used to generate glycopolymers which reconfigure due to the action of lectin binding (i.e. internal trigger), but not to an external trigger.11–13 In contrast, externally addressable polymers (often termed as ‘smart’ or ‘responsive’) have been extensively studied where an external stimulus, such as light, heat, pH, radiation, metal ions etc., can trigger a (reversible) change in material properties. In particular, thermoresponsive polymers have attracted attention as due to their easy synthesis and diverse range of applications from triggered cellular uptake, trypsin-free cell release14 and drug delivery.15 Polymers which display an LCST (lower critical solution temperature) undergo a chain collapse (soluble–insoluble) upon heating providing a macroscopic effect from the external trigger.16,17 Typical thermo-responsive polymers with an LCST include poly(N-isopropylacrylamide) (pNIPAM) and poly[(oligoethylene glycol) methacrylates] (pOEGMAs) due to their transitions being close to 37 °C. Many other classes have been developed and extensively reviewed.18,19 Immobilization of responsive polymers onto metal or soft nanoparticle enables dynamic control over aggregation state based on an external trigger.20,21 Mastrotto et al. used pNIPAM collapse to expose folate moieties on gold nanoparticle surfaces to enable temperature triggered uptake into cancerous cell lines.22 Temperature gating has also been used to control access to biotin functionality on glass surfaces.23 Gold nanoparticles (AuNPs) are widely used due to their easy functionalization with...
thiols and unique optical properties which make them excellent contrast agents in electron microscopy, or dark field microscopy, but also as colorimetric sensors due to the coupling of their SPR bands when aggregated leading to a red-blue colour shift. Immobilization of glycans onto AuNPs has been used as biosensors. Field et al., used α,β-thio-linked sialic acid to detect human influenza, and Richards et al. have used glycosylated gold nanoparticles libraries as multiplex sensors.

Considering the above, we reasoned that if a nanoparticle surface could be formulated correctly, a responsive polymer could be used as an externally addressable ‘gate’ which upon application of a stimulus, is ‘opened’ (via chain collapse) to enable access to a glycan and hence enable binding. This can be considered as a synthetic alternative to enzyme expression levels, which in vivo control glycan expression based upon biological triggers.

To provide the desired gating mechanism on the nanoparticle surface pNIPAM was selected as the thermo-responsive polymer due to its well characterized switchable behavior and high grafting density onto gold. Well-defined poly(hydroxyethyl acrylamide) (pHEA) was selected as a non-responsive co-coating as we have previously demonstrated it to be an excellent stabilizing polymer for glycans onto gold. 27

Density onto gold. 27 Well-defined poly(hydroxyethyl acrylamide) (pHEA) was selected as a non-responsive co-coating as we have previously demonstrated it to be an excellent stabilizing polymer for biological triggers.

Determined 1H NMR.

<table>
<thead>
<tr>
<th>Polymer</th>
<th>[M]/[CTA]/[I] [mol]</th>
<th>$M_n$ target [g mol$^{-1}$]</th>
<th>Conversion[a] [%]</th>
<th>$M_n$/Thet[a] [g mol$^{-1}$]</th>
<th>$M_n$/SEC[c] [g mol$^{-1}$]</th>
<th>$M_n$/Mn [-]</th>
<th>Cloud point[d] [°C]</th>
</tr>
</thead>
<tbody>
<tr>
<td>pNIPAM$_{25}$</td>
<td>25/1/0.2</td>
<td>3200</td>
<td>87%</td>
<td>2800</td>
<td>2900</td>
<td>1.07</td>
<td>36</td>
</tr>
<tr>
<td>pNIPAM$_{50}$</td>
<td>50/1/0.2</td>
<td>6000</td>
<td>86%</td>
<td>5200</td>
<td>7100</td>
<td>1.10</td>
<td>38</td>
</tr>
<tr>
<td>PFP–pHEA$_{15}$</td>
<td>15/1/0.2</td>
<td>2300</td>
<td>93%</td>
<td>2100</td>
<td>4800</td>
<td>1.10</td>
<td>—</td>
</tr>
</tbody>
</table>

pNIPAM$_{xxx}$/pHEA$_{xxx}$ = poly(N-isopropylacrylamide)/poly(hydroxyethyl acrylamide) where average degree of polymerization indicated by xxx.

[a] Determined 1H NMR. [b] Calculated from the [monomer]/[CTA] ratio and of conversion. [c] Determined by SEC in DMF using PMMA standards. [d] Cloud point was measured in water upon heating from 25 °C to 80 °C, 1.0 mg mL$^{-1}$ polymer concentration.
access to the Gal residues. Upon increasing the temperature to 40 °C, in the presence of SBA, there was a small, but significant shift in the UV-Vis spectra with an increase at 700 nm and decrease at 540 nm indicative of lectin binding and aggregation. This clearly demonstrated that the concept of responsive gating to glycan access could be achieved, but that the surface coating has to be precisely tuned to achieve the balance required (Fig. 2).

Further optimization studies revealed that changing the ratio of pHEA_{15}-Gal : pNIPAM_{50} from 8 : 2 to 9 : 1 provided the optimum balance between glycan affinity (i.e. aggregation) and switchability (ESI†). To ensure the changes seen were due to particle aggregation (indicative of lectin-cross-linking), SBA binding was investigated using dynamic light scattering and transmission electron microscopy (TEM) Fig. 3. The optimized nanoparticle formulation (above) was incubated with SBA at both 20 and 40 °C for 30 minutes and the observed hydrodynamic diameters shown. At 20 °C, in the presence of various concentrations of SBA there was no change in hydrodynamic diameter from the initial 60 nm. At 40 °C, with no SBA added, the particles were stable with the initial diameter of ~80 nm (due to some surface reconfiguration compared to at 40 °C) being retained, in agreement with the UV-Vis data. Following 30 minutes of incubation there was a clear dose-dependent increase in the aggregate size as [SBA] was increased, again supporting the hypothesis that the pNIPAM is gating access to the glycan. TEM analysis was also conducted to provide direct evidence of temperature triggered lectin/particle agglutination. Fig. 3C shows nanoparticles plus SBA at 20 °C, which are clearly well-dispersed and Fig. 3D shows the aggregates which form upon heating to 40 °C only in the presence of SBA (Fig. 4).

As a final test of the system, the optimized nanoparticle formulation (with the 9 : 1 ratio of pHEA_{15}-Gal : pNIPAM_{50}) was interrogated with a panel of lectins, with different binding specificities on both 20 and 40 °C. SBA, WGA (Wheat germ agglutinin), UEA (Ulex europaeus agglutinin) and RCA120 (Ricinus communis agglutinin) were employed. At 20 °C there was no measurable change in UV-Vis spectra upon incubation with SBA, WGA, UEA or RCA120 lectins, indicating that the glycan is stERICALLY shielded against all the lectins (ESI†). Increasing the
temperature to 40 °C, however, lead to clear changes in the UV-Vis spectra for SBA, RCA$_{20}$ and WGA as would be expected with their known affinities for Gal (or GalNAc/GluNAc for WGA). The control lectin UEA, which has specificity for fucose residues did not bind at any temperature, proving the specificity of the interaction and that temperature-induced aggregation is not a factor. A partial isotherm showing the relative changes is included in the ESI$^\dagger$ as a measure of the relative affinity, in the order SBA $>$ WGA $>$ RCA$_{20}$. UEA. Additional control experiments using BSA as a model non-carbohydrate binding protein revealed there were non-specific interactions (see ESI$^\dagger$). The ability to control glycan expression would be a powerful tool for studying the role of multivalency intracellularly, where the glycan is only exposed one trafficked to the desired location, potentially provided spatiotemporal control. They could also be used as new biomolecular logic gates.

In summary, we have demonstrated a new concept in glyco-engineering where responsive polymer surfaces, rather than external enzyme expression levels, control the display of sugars on the surface of a nanoparticle, which could be considered a simple cell mimetic. The ‘gate’ pNIPAM had to be added in a relatively low ratio compared to the glycan-bearing polymer to ensure a binary on/off effect, with significant lectin binding above the pNIPAM LCST observed. The specificity of the GlycoAuNP was confirmed against a panel of lectins, which glycan expression only being induced above the critical temperature. The complex function of this relatively simple system with in-built optical outputs (AuNP colour changes) is highly versatile and presents a new method to dynamically control glycan expression using fully synthetic systems. By controlling presentation on the AuNP surface with an external trigger, we can envisage this being used as a tool to probe glycan function under very controlled environments, including intracellularly and could be considered a molecular ‘AND’ gate. Furthermore, the pNIPAM collapse could be replaced with a range of other stimuli responsive polymers, to enable biochemical rather than temperature trigger to probe more complex cellular environments.

MIG acknowledges the ERC for a Starting Grant, CRYOMAT 638661 and BBSRC (BB/M02878X/1). SW thanks UoW for a chancellors international scholarship. SJR was a UoW IAS Early Career fellow.

Notes and references