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Comparison of the Ca^{2+} complexing properties of isosaccharinate and gluconate – is gluconate a reliable structural and functional model of isosaccharinate?†

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The calcium complexation and acid–base properties of α -D-isosaccharinate (Isa^-) in neutral and in (hyper) alkaline solutions have been investigated via potentiometric titrations, multinuclear NMR, ESI-MS and quantum chemical calculations. Isa^- is the primary alkaline degradation product of cellulose, and may be present in radioactive waste repositories and therefore, it could contribute to the mobilization of radioactive nuclei. Because of its limited availability, D-gluconate (Gluc^-) is commonly used as a structural and functional model of Isa^- . Therefore, the thermodynamic and structural data obtained for Isa^- were compared with those of Gluc^- . The formation constants of the CaIsa^+ and CaGluc^+ complexes present in neutral solutions are practically identical, but the binding sites are in different positions and the CaIsa_2^0 solution species cannot be detected. The stepwise formation constant of the CaIsaH_{-1}^0 complex (forming in alkaline medium) is somewhat larger than that of CaGlucH_{-1}^0 , which is in line with the observation that IsaH_{-1}^{2-} is a stronger base than GlucH_{-1}^{2-} . The most striking difference is that, unlike Gluc^- , Isa^- does not form polynuclear complexes with Ca^{2+} . The structural reason for this is that the alcoholate groups on C2 and C3 adjacent to the carboxylate moiety on Gluc^- are able to simultaneously bind Ca^{2+} , making the formation of polynuclear Ca-complexes possible. On Isa^- , only the alcoholate on C2 is involved, while the other one on C6 is not (supposedly for steric reasons). In conclusion, during the interactions of Gluc^- and Isa^- with Ca^{2+} , differences rather than similarities prevail.

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Introduction

Recently, it has been shown *via* using a variety of thermodynamic and structural experimental means that in (hyper)alka-

line solutions containing Ca^{2+} and various sugar carboxylates ($\text{L}^- = \text{D-gluconate}$, Gluc^- and D-heptagluconate , Hglu^-), bi- and trinuclear Ca^{2+} -sugar-carboxylate complexes with surprisingly high stability were formed.^{1,2} As a consequence, in solutions with $\text{pH} > 12$, Ca^{2+} will be present predominantly in the form of $\text{Ca}_2\text{LH}_{-3}^0$ and $\text{Ca}_3\text{L}_2\text{H}_{-4}^0$. Additionally, the formation of mononuclear CaLH_{-1}^0 with lower stability was also detected and it is noticeable that all of the complexes formed are neutral.

Organic ligands containing $-\text{OH}$ and $-\text{COO}^-$ donor groups (*i.e.*, sugar carboxylates) are known to form stable binary solution complexes with various metal ions (*e.g.*, alkaline earth and transition metals,^{1–11} lanthanides,^{12–15} actinides,^{16–26} *etc.*), and their complexing ability is more pronounced under (hyper)alkaline conditions than under neutral conditions, due to the binding of the metal ion to the alcoholate group(s).^{1,2,5,9,10,13,14,16,19,21,24} In an even more complex system, where polyhydroxy carboxylate-, Ca^{2+} and further three- or four valent cations (*e.g.*, radionuclides) were present, the formation of heteropolynuclear complexes with remarkably high stability was observed in some cases.^{21,23,27–30}

Hyperalkaline conditions prevail in Ca^{2+} -containing cementitious environments foreseen in some repository concepts for

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† Electronic supplementary information (ESI) available: Distribution diagram for the Ca^{2+} – Isa^- neutral system (Fig. S1), the structure of the CaIsa^+ complex in aqueous solution (Fig. S2), the values of J -coupling constants for the Isa^- protons (Table S1), the distribution diagram of $\text{Isa}^-/\text{IsaH}_{-1}^{2-}$ at $\text{pH} = 12$ –14 (Fig. S3), time dependent ^1H NMR spectra of an alkaline solution supersaturated with respect to $\text{Ca}(\text{Isa})_2$ (Fig. S4), the distribution diagram of the Ca^{2+} – Isa^- and Ca^{2+} – Gluc^- alkaline systems (Fig. S5 and S6, respectively), temperature dependent ^1H NMR spectra in 0.2 M NaOH (Fig. S7 and S8) and the structure of the CaIsaH_{-1}^0 complex with O(C6) as a binding site. See DOI: 10.1039/c7dt03120c

disposal of nuclear waste. One of the disposal options widely considered for low- and intermediate level waste (L/ILW) involves the stabilization of waste with cement or cementitious materials. Furthermore, cement-based materials will be widely used as construction materials. The pH of cement pore water (CPW) varies between $8 < \text{pH} < 13.3$ depending on the composition of cement and brine as well as the solid to brine ratio, *etc.*^{31,32} (generally, the pH in a model CPW is usually set to 13.3²³). If CPW is in contact with various calcium silicates, the total concentration of Ca^{2+} ($[\text{Ca}^{2+}]_{\text{T}}$; hereafter $[\text{X}]_{\text{T}}$ denotes the total concentration, while $[\text{X}]$ stands for the equilibrium concentration of species X) changes between $2 \times 10^{-3} \text{ M}$ and $2 \times 10^{-2} \text{ M}$,³¹ while the pH can reach 13 in close-to-saturated NaCl solutions.³¹ In contrast, in MgCl_2 -dominated brines, Ca^{2+} concentrations could be several orders of magnitude higher.³²

Under alkaline conditions, the degradation of organic macromolecules (mainly cellulose) yields various low-molecular-weight carboxylates and hydroxycarboxylates. Among them, α -D-isosaccharinate and its epimeric form, β -D-isosaccharinate are formed in the largest quantities.³³ In the literature, significantly more attention has been paid to the aqueous chemistry of α -D-isosaccharinate (Isa^- hereafter),^{34–36} than to its epimeric form, as the former is claimed to be a stronger complexant^{33,37} than the latter. In the CPW of L/ILW repositories, the maximum concentration of Isa^- can be as high as 0.1 M .³⁸ Another relevant organic compound in cementitious environments is Gluc^- , which is released from the solid matrix itself being one of the most frequently used cement additives (plasticizer) used during the processing of concrete.³⁹ The concentration range for Gluc^- in CPW is considered to be 10^{-5} – 10^{-2} M in numerous studies.²³ Therefore, the complexation of radionuclides with Gluc^- and in particular with Isa^- may increase their solubility and/or decrease their sorption on cement, and can be regarded as a relevant factor contributing to the mobilization of these metal ions from an underground L/ILW repository to the environment.

Gluc^- is often considered^{18,23,39,40} as a readily available functional analogue and model compound of α - Isa^- (see Scheme 1); the latter is not only expensive, when commercially acquired, but also challenging to prepare in sufficiently pure

form.^{5,7,41} Assuming the structural similarity, the formation of the neutral CaLH_{-1}^0 complex (at lower $[\text{Ca}^{2+}]_{\text{T}}$) with a stability constant identical to $\text{L}^- = \text{Gluc}^-$ and Isa^- can be suggested.

In the current publication, the assumed similarity of Isa^- and Gluc^- is scrutinized. For this, we compare the acid–base properties, the Ca^{2+} -binding abilities (both under neutral and alkaline conditions) and the structure of the Ca(II) -complexes of the two sugar carboxylates.

Experimental

Reagents and solutions

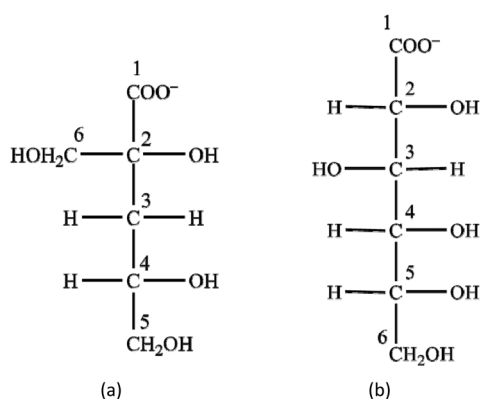
Calcium chloride stock solution was prepared by dissolving $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ (Sigma-Aldrich, $\geq 99\%$ purity) in water, and the exact concentration of Ca^{2+} was determined by titration with EDTA. Sodium hydroxide stock solution was prepared by diluting a carbonate-free concentrated NaOH solution,⁴² and standardized against HCl. The ionic strength was adjusted to 1 M with sodium chloride (Molar Chemicals, a.r. grade) in each sample. All reagents were dissolved in Milli-Q Millipore water.

Calcium α -D-isosaccharinate, Ca(Isa)_2 was prepared according to the method of Whistler and BeMiller.⁴¹ The original procedure was slightly modified as follows: 220 g of lactose monohydrate (Sigma-Aldrich, $\geq 99\%$ purity) and 60 g of Ca(OH)_2 were dissolved in 2 liters of boiled water, and stirred for 3 days at room temperature. After this time, the mixture was boiled for 8 hours. The hot solution was filtered, and the volume of the filtrate was reduced to about 400 mL by evaporating the solvent. The mixture was filtered, and the crude product was recrystallized from water. The success of the synthesis was proven by X-ray diffractometry as well as ^1H and ^{13}C NMR spectroscopy.

Sodium α -D-isosaccharinate, NaIsa stock solution was prepared as follows: 10 g of Ca(Isa)_2 was stirred with an appropriate amount of the hydrogen form of Amberlite® IR120 ion-exchange resin (Sigma-Aldrich) in about 40 mL water for several hours. The mixture was filtered through a $0.45 \mu\text{m}$ hydrophilic PTFE membrane filter (VWR). The α -D-isosaccharinic acid, HIsa solution obtained was standardized against NaOH by adjusting the pH to about 8. The pH of the solution was monitored with a calibrated pH-sensitive glass electrode (Sentix® 62 by WTW).

NMR experiments

^1H and ^{13}C NMR spectra were recorded on a Bruker Avance DRX 500 MHz NMR spectrometer equipped with a 5 mm inverse broadband probe head furnished with z-oriented magnetic field gradient capability. The magnetic field was stabilized by locking it to the ^2D signal of the solvent prior to the measurements. The sample temperature was set to $(25 \pm 1) ^\circ\text{C}$ during the measurements at constant temperature and to 0 – $50 ^\circ\text{C}$ during the temperature-dependent measurement. For the individual samples, 64 and 256 scans were acquired to record ^1H NMR and ^{13}C NMR spectra, respectively. Each highly alkaline solution was placed into a PTFE liner, and the liner



Scheme 1 The structures of α -D-isosaccharinate (a) and D-gluconate (b).



was placed into an external quartz tube containing D₂O. In this way D₂O was not mixed with the H₂O solvent and therefore the results presented here refer to pH values.

For the determination of the deprotonation constant of the alcoholic OH group of Isa[−], two sets of solutions were prepared with [Isa[−]]_T = 0.050 and 0.100 M, where [OH[−]]_T was varied systematically from 0.00 to 1.00 M.

To examine the behavior of Isa[−] in the presence of Ca²⁺ in highly alkaline medium, solutions containing [Isa[−]]_T = 0.20 M, [OH[−]]_T = 0.50 M and [Ca²⁺]_T = 0.00, 0.05, 0.10, 0.25 and 0.50 M were prepared.

To reveal the structure of the complex formed between Isa[−] and Ca²⁺ in alkaline solutions, the temperature dependence of the ¹H NMR spectra was recorded in the temperature range of 0–50 °C for a solution containing [Isa[−]]_T = 0.20 M, [OH[−]]_T = 0.20 M and [Ca²⁺]_T = 0.10 M.

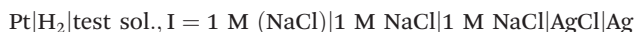
Nonlinear fitting of the ¹³C NMR chemical shifts as a function of [NaOH]_T was performed using the PSEQUAD program.⁴³

Potentiometric titrations

Potentiometric titrations were carried out at constant ionic strength (I = 1 M NaCl) using a Metrohm 794 Titrando instrument.

The complexation between Ca²⁺ and Isa[−] in neutral medium was studied *via* using a Ca-ion selective electrode (Metrohm). All measurements were carried out in a titration cell thermostated to (25.0 ± 0.1) °C, and the samples were stirred continuously during the reactions. For the calibration of the electrode, a solution containing 10^{−4} M CaCl₂ as the initial concentration and 45 ml as the initial volume was titrated with 0.0650 M CaCl₂ up to 120 ml. The response of the Ca-ISE was found to be nonlinear in the −4.0 < log([Ca²⁺]/M) < −1.5 range, thus a nonlinear calibration procedure was used by fitting splines into the calibration curve using the Spline Calculus program.⁴⁴ Several sets of measurements were performed with solutions containing [Ca²⁺]_T = 10^{−4} M and [Isa[−]]_T = 0.011, 0.020, 0.030, 0.059, 0.099, 0.150, 0.194, 0.243 M using the same titrant as in the calibration.

The complex formation between Ca²⁺ and Isa[−] in highly alkaline medium was studied by pH-potentiometric titrations, following the reactions with a H₂/Pt electrode in the pH range of 11.6–12.9. The full electrochemical cell contained a platinized-platinum hydrogen electrode and a thermodynamic Ag/AgCl reference electrode, and was constructed as follows:



Within the employed pH range the response of the electrode was found to be linear with a Nernstian slope (59.0–59.2 mV). The titration cell was externally thermostated to (25.0 ± 0.1) °C. Potentiometric titrations of systems containing both Ca²⁺ and Isa[−] were performed with the initial NaOH concentration of 0.0102 M and V₀ = 20 cm³. The titrant was 1.0136 M NaOH in each case. The initial [Isa[−]]_T concentration was 0.100 M, while the initial [Ca²⁺]_T concentration was set to 0.024 and 0.051 M.

During the data processing, the log([Ca²⁺]/M) or E/mV values for the measurements performed in neutral and alkaline media, respectively, were fitted as a function of added volume of the titrant using PSEQUAD software.

ESI-MS measurements

ESI-MS measurements in positive ion mode were performed using a Micromass Q-TOF Premier (Waters MS Technologies, Manchester, UK) mass spectrometer equipped with an electrospray ion source. Samples were introduced into the MS by using the direct injection method: the built-in syringe pump of the instrument with a Hamilton syringe was used. The electrospray needle was adjusted to 3 kV and N₂ was used as the nebulizer gas.

Quantum chemical structure optimizations

The geometry optimizations of the CaIsa⁺ and CaIsaH_{−1}⁰ complexes were performed using the Gaussian 09 software package⁴⁵ at the B3LYP/6-311++g(d,p) level. The structures were first optimized *in vacuo*, and then further optimization was performed on the thus obtained structures considering implicit solvation. For these calculations, the Conductor-like Polarizable Continuum Model (CPCM)⁴⁶ was utilized at the same theoretical level and with the same basis set (using water as the solvent). During computations, different coordination isomers for CaIsa⁺ and CaIsaH_{−1}⁰ were considered in order to find the most probable structure.

Results and discussion

Ca²⁺ complexation of Isa[−] in neutral solution

The Ca-ISE potentiometric titration curves of Isa[−] containing solutions are shown in Fig. 1. The observed cell potential values are systematically shifted towards smaller [Ca²⁺] values with increasing [Isa[−]]_T indicating complexation. The

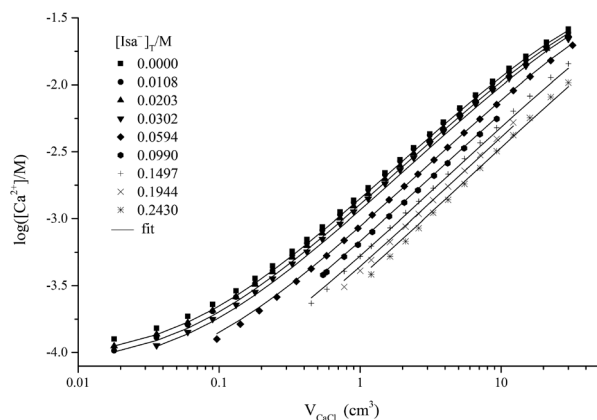


Fig. 1 Ca-ISE potentiometric titration curves of solutions containing Isa[−] and Ca²⁺, in neutral medium, in terms of added titrant volume. V₀ = 45 cm³, [Ca²⁺]_{T,0} = 10^{−4} M, titrant: [CaCl₂]_T = 0.2 M, initial [Isa[−]]_T values are shown in the figure, I = 1 M (NaCl), T = 25.0 ± 0.1 °C. The symbols and solid lines correspond to the measured and fitted data, respectively.



maximum difference is about 10 mV between the cell potentials obtained for solutions containing 0 and 0.24 M Isa^- , respectively. The titration curves were fitted assuming solely the formation of the 1:1 complex (represented by the solid lines in Fig. 1). The optimized value for $\log \beta_{1,1,0}$ (conditional stability constant relating to $I = 1.0$ M (const.) ionic strength) with its \pm SD value was found to be 1.12 ± 0.02 . In general, $\beta_{p,q,r}$ can be defined as

$$\beta_{p,q,r} = \frac{[\text{Ca}_p\text{L}_q\text{OH}_r]^{(2p-q-r)+}}{[\text{Ca}^{2+}]^p[\text{L}^-]^q[\text{OH}^-]^r\text{M}^{1-p-q-r}} \quad (1)$$

where $\text{L}^- = \text{Isa}^-$ or Gluc^- and $\text{M} = 1 \text{ mol dm}^{-3}$. Accordingly, $\beta_{1,1,0}$ is defined as

$$\beta_{1,1,0} = \frac{[\text{CaIsa}^+]}{[\text{Ca}^{2+}][\text{Isa}^-]\text{M}^{-1}} \quad (2)$$

This stability constant corresponds to the significant (up to 60%) formation of the complex CaIsa^+ (Fig. S1†). At $T = 22^\circ\text{C}$ and $I = 0.2$ M, $\log \beta_{1,1,0}$ was found to be 1.25 from potentiometry and 1.29 ± 0.02 from ion-exchange,⁶ while a value of 1.44 ± 0.07 was suggested from solubility measurements.¹⁸

Under the experimental conditions identical to ours ($T = 25^\circ\text{C}$, $I = 1.0$ M NaCl), Ca^{2+} and Gluc^- were found to form both 1:1 ($\log \beta_{1,1,0} = 1.08 \pm 0.01$) and 1:2 ($\log \beta_{1,2,0} = 1.65 \pm 0.03$) complexes,⁴⁷ where

$$\beta_{1,2,0} = \frac{[\text{CaGluc}_2^0]}{[\text{Ca}^{2+}][\text{Gluc}^-]^2\text{M}^{-2}} \quad (3)$$

In the case of Isa^- , a surprisingly high formation constant ($\log \beta_{1,2,0} = 5.40$) was proposed for the CaIsa_2^0 complex,⁴⁸ however, it was further neglected from the comprehensive model describing the $\text{Ca}^{2+}/\text{Isa}^-$ system in the 2–14 pH range.¹⁸ Additionally, its formation was neither observed by us nor reported in other literature sources as well.^{5–7}

The difference in the composition of the solution species in the $\text{Ca}^{2+}/\text{Isa}^-$ and $\text{Ca}^{2+}/\text{Gluc}^-$ systems is most probably associated with the differences between the structures of the two ligands. The optimum geometry of the CaIsa^+ complex obtained from DFT quantum chemical calculations is shown in Fig. 2. The Ca^{2+} ion sits in the nest of three O atoms (one carboxylate oxygen on C1, and the OH oxygens on C6 and C4). From single crystal X-ray diffraction measurements⁴⁹ similar binding mode is present in $\text{Ca}(\text{Isa})_2$. Analogous calculations for the CaGluc^+ complex⁴³ suggested binding of Ca^{2+} to O atoms on C1, C2, C3 and C6. This binding mode seems to be the governing coordination motif for both ligands resulting in an almost identical association constant for the 1:1 species.

In contrast to CaGluc^+ , there is no binding to the alcoholic OH closest to the carboxylate (the latter two rather establish an intramolecular hydrogen bond) in the CaIsa^+ species. Moreover, only one freely rotating C–C bond is present in this rigid structure. Consequently, the binding of a second anion and the simultaneous dehydration of Ca^{2+} are difficult compared to the more flexible Gluc^- . This may be the reason why

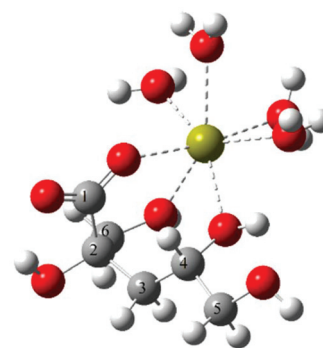


Fig. 2 The optimum geometry calculated for CaIsa^+ at the B3LYP/6-311++G(d,p) level using the PCM model and explicit water molecules.

the CaIsa_2^0 complex has lower stability than CaGluc_2^0 and why the former was not possible to detect experimentally.

Acid–base properties of Isa^- in hyperalkaline solutions

The proton dissociation constant of Isa^- (corresponding to the deprotonation of one or more alcoholic OH groups) taking place in hyperalkaline solutions can be defined as

$$\beta_{0,1,-1} = \frac{[\text{IsaH}_{-1}^{2-}][\text{H}^+]}{[\text{Isa}^-]\text{M}} \quad (4)$$

This deprotonation process causes a significant downfield shift in each ^{13}C NMR signal of Isa^- (see Fig. 3, in which the peak assignments shown in Scheme 1 are used; they were constructed on the basis of the average proton coupling constants of Isa^- shown in Table S1† and from the two-dimensional ^1H – ^{13}C HSQC NMR spectrum of Isa^-). The extent of this variation is 0.08–0.55 ppm and it can be described using saturation curves with the increasing NaOH concentration. The $\log \beta_{0,1,-1}$ dissociation constant extracted from the data points obtained for a solution series with $I = 1$ M NaCl (const.) was found to be -14.5 ± 0.1 (using -13.76 as $\log K_w$, the ionic

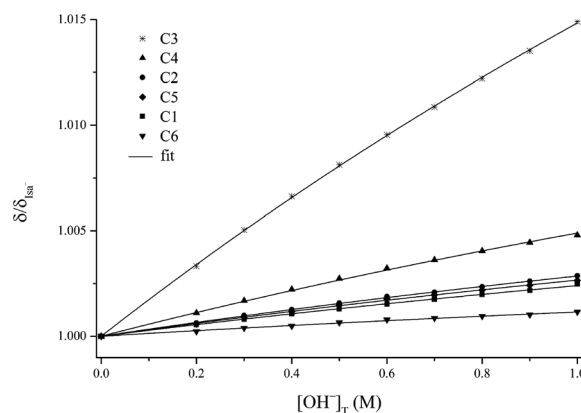


Fig. 3 ^{13}C NMR chemical shifts of each carbon atom (normalized to those of the neat Isa^- ion) as a function of $[\text{OH}^-]_T$ at $[\text{Isa}^-]_T = 0.100$ M, $I = 1$ M (NaCl) and $T = (25 \pm 1)^\circ\text{C}$. Symbols: observed data; solid lines: calculated data.



product of water, for 1 M NaCl⁵⁰). From this dissociation constant, the degree of deprotonation is *ca.* 20% at pH = 14 (Fig. S2†).

The ¹H NMR signals of Isa[−] exhibited an upfield shift with increasing NaOH concentration. This small but detectable variation ($\Delta\delta < 0.1$ ppm) supports the deprotonation of the anion, but the magnitude of these changes makes the ¹H NMR spectra unsuitable for extracting $\log\beta_{0,1,-1}$. The almost uniform variation in the chemical shifts on each proton signal upon increasing NaOH concentration indicates the deprotonation of the alcoholic OH groups without a favored position for the deprotonation. In this respect, ¹H NMR observations analogous to those found for Isa[−] were made for Gluc[−].¹

Ca²⁺ complexation of Isa[−] in alkaline to hyperalkaline solutions

The ¹H NMR spectra of aqueous solutions containing [Isa[−]]_T = 0.20 M, [OH[−]]_T = 0.50 M and varying amounts of added Ca²⁺ (0–0.10 M) are shown in Fig. 4(a). For comparison, the spectra of solutions with identical compositions, but with Gluc[−] in place of Isa[−] are also shown (Fig. 4(b)).

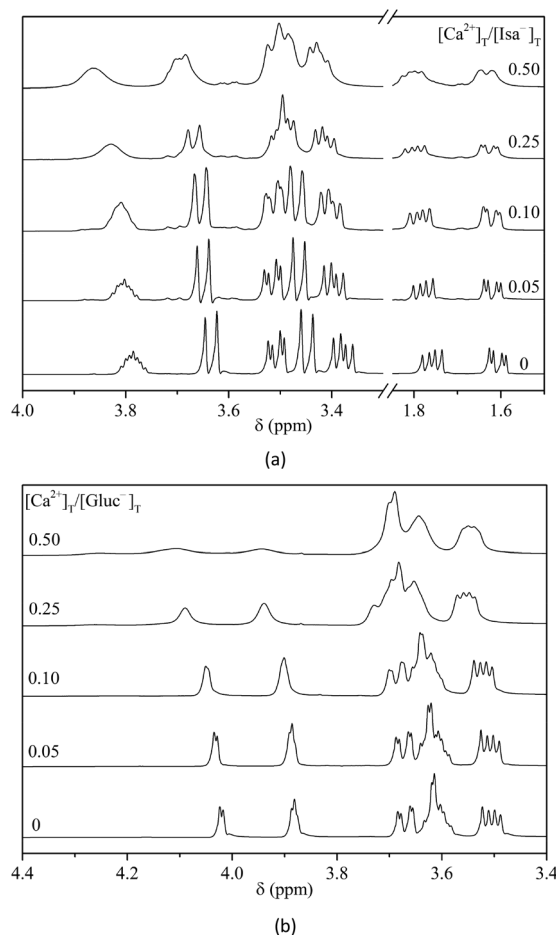


Fig. 4 ¹H NMR spectra of Isa[−] (a) or Gluc[−] (b) solutions (0.20 M) in the presence of [Ca²⁺]_T = 0–0.10 M and at [OH[−]]_T = 0.50 M. I = 1 M (NaCl), T = (25 ± 1) °C.

Addition of Ca(II) ions to the system causes minor but detectable effects in the ¹H NMR spectrum of Isa[−]: the peaks are shifted downfield and broadened; the highest change occurred in the signal of H4. Both experimental observations indicate the interaction between the metal ion and Isa[−]. Many more dramatic effects are seen for identical solutions containing Gluc[−], the displacement of the NMR peaks is significantly larger and the extent of line broadening is so extensive that at a 1 : 2 metal-to-ligand ratio, some peaks (H2 and H3) vanish in the baseline. The profound difference between the two spectral series in Fig. 3 strongly suggests that under these experimental conditions, calcium ions form complexes more readily with Gluc[−] than with Isa[−]. To further enhance the experimental effect seen in Fig. 4(a), the metal-to-ligand ratio should be increased; in this concentration range Ca(Isa)_{2(s)} precipitation occurred in the system right after solution preparation. (The precipitate formation was also observed at lower total concentrations too, as it is exemplified in Fig. S3,† however, the spectra exhibited variation only after the onset of the precipitation. Before that, the spectra are independent of the time elapsed since solution preparation. This indicates that the simultaneous solution equilibria are very rapidly established between the solution species even in supersaturated solutions.)

Although the experimental effects observed in the ¹H NMR spectra undoubtedly proved the complex formation between Ca²⁺ and Isa[−], they were not sufficiently large for the quantitative determination of the composition and structure of the complexes formed. Accordingly, the interaction between Ca²⁺ and Isa[−] in alkaline solutions was studied *via* potentiometric titrations using the H₂/Pt electrode (Fig. 5).

The observed cell potentials are shifted towards the less negative values for solutions containing both calcium and Isa[−]. This corresponds to some OH[−] consuming complexation process resulting in a decrease in [OH[−]]. However, the experi-

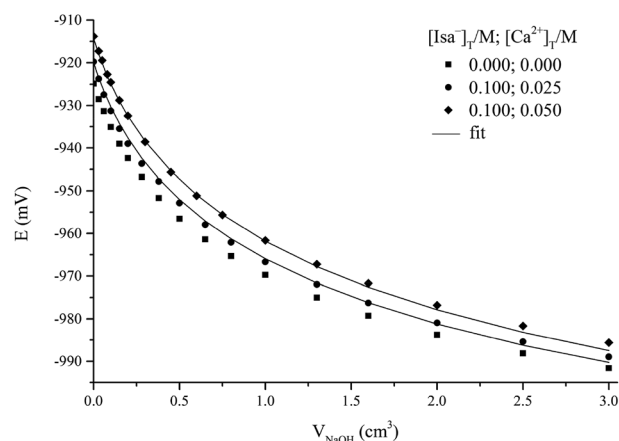


Fig. 5 H₂/Pt electrode potentiometric titration curves of alkaline solutions containing Isa[−] and Ca²⁺ as a function of the added titrant volume. V₀ = 20 cm³, [OH[−]]_{T,0} = 0.010 M, titrant: [NaOH]_T = 1.00 M, initial [Isa[−]]_T and [Ca²⁺]_T values are shown in the figure, I = 1 M (NaCl), T = 25.0 ± 0.1 °C. Symbols: observed potentials; solid lines: fitted data.



mental effect (*i.e.*, the potential difference between the calibration and the titration in question at the same added base volume) is at most ~12 mV. A much larger effect (up to 50 mV) and a unique curvature were seen on the titration curves analogous to these ones but with Gluc[−] as the complexing ligand (see Fig. 1 in ref. 1). This confirms the semi-quantitative statement derived from the ¹H NMR measurements (Fig. 4) on the formation of more stable Ca²⁺ complexes with Gluc[−] than with Isa[−].

During the data processing, the formation constants of IsaH_{−1}^{2−} and CaIsa⁺ (see previous sections), the ionic product of water, pK_w (13.76, taken from the literature⁵⁰) and the formation constants of the CaOH⁺ and Ca(OH)₂⁰ solution species⁵¹ (with log β_{1,0,1} = 0.18 ± 0.02 and log β_{1,0,2} = 0.67 ± 0.02, respectively) were kept constant. Assumption of only one mononuclear complex, CaIsaH_{−1}⁰ with log β_{1,1,−1} = −11.36 ± 0.02, where

$$\beta_{1,1,-1} = \frac{[\text{CaIsaH}_{-1}^0][\text{H}^+]}{[\text{Ca}^{2+}][\text{Isa}^-]} \quad (5)$$

was sufficient to fit the experimentally observed cell potentials (see the solid curves in Fig. 5), and employing polynuclear complexes in the model (like for the Ca²⁺/Gluc[−]/OH[−] systems with similar compositions¹) was not necessary. The formation of this CaIsaH_{−1}⁰ complex is significant (25%) and the degree of formation increases with the increasing pH (Fig. S4†). Under the conditions identical to those shown in Fig. S4†, gluconate-containing solutions are predominated by polynuclear complexes (Fig. S5†), and only a small fraction (maximum 20%) of the calcium ions may be present as CaGlucH_{−1}⁰.

ESI-MS measurements were performed to obtain further information on the composition of the solution species present in the Ca²⁺/Isa[−]/OH[−] systems. The spectrum of the sample containing Ca²⁺ (0.025 M), Isa[−] (0.050 M) and NaOH (0.100 M) was recorded in positive ion mode (Fig. 6). As uncharged complexes are invisible in ESI-MS, the peaks referring to the solution species appeared in the spectrum gaining

positive charge(s) *via* “binding” one (or more) proton(s). Therefore, the peaks referring to CaIsaH_{−1}⁰ are expected to appear at 219.03 *m/z*. Polynuclear complexes with compositions similar to those found for the Ca²⁺/Gluc[−]/OH[−] systems are expected to appear at around 257 and 475 *m/z* values. For the Ca²⁺/Isa[−]/OH[−] system, the experimentally observed mass spectrum was practically empty in this range.

It is interesting to note that the stability constant of the direct association of Ca²⁺ and IsaH_{−1}^{2−} is defined as

$$K_{1,1,-1} = \frac{[\text{CaIsaH}_{-1}^0]}{[\text{Ca}^{2+}][\text{IsaH}_{-1}^{2-}]\text{M}^{-1}} = \frac{\beta_{1,1,-1}}{\beta_{0,1,-1}} \quad (6)$$

the value of which was found to be log K_{1,1,−1} = 3.13. This is considerably larger than that of the CaGlucH_{−1}⁰ complex (2.74¹); this difference in the formation constants is most probably due to the stronger basicity of IsaH_{−1}^{2−} than GlucH_{−1}^{2−}.

The binding sites of Isa[−] in the CaIsaH_{−1}⁰ complex

To identify the Ca²⁺-binding sites of Isa[−] in the CaIsaH_{−1}⁰ complex, ¹H and ¹³C NMR measurements were performed. From the temperature dependence of the ¹H NMR spectrum of an alkaline solution containing Ca²⁺ and Isa[−] (Fig. S6†), the chemical exchange between the various forms of the ligand at low temperatures (0–5 °C) is slow on the NMR time scale, while by increasing the temperature to 30–35 °C, the chemical exchange becomes fast. Below 5 °C, the signals of free Isa[−] and those of its Ca²⁺ complex(es) appear separately (Fig. S7†). The largest change (splitting at low temperatures, displacement and broadening at higher ones) is seen in the ¹H NMR peak of H4 indicating that the alcoholic OH group on C4 (most probably in the form of alcoholate) participates in calcium binding. Because of the complexity of the ¹H NMR spectra and the overlap of various peaks, it is not possible to unambiguously locate further binding sites in the molecule.

The ¹³C NMR spectra of alkaline Isa[−] solutions containing various amounts of added calcium are shown in Fig. 7. With increasing Ca²⁺ concentration, the signals of C1, C2, and C3 significantly broaden to an extent that those of C1 and C2 vanish in the baseline at the highest calcium concentration ([Ca²⁺]_T = 0.05 M, corresponding to a 1:4 metal-to-ligand ratio). Less spectacular but still significant broadening is seen on the peak corresponding to C4, and those of C5 and C6 are the least affected by calcium binding.

These observations indicate that in alkaline solutions Isa[−] acts as a multidentate ligand in the CaIsaH_{−1}⁰ complex; this feature is similar to that of Gluc[−], as it is shown in ref. 1. Furthermore, the peak broadening and loss of intensity imply that the chemical exchange is slower than at neutral pH, since for the 1:1 complex of the Gluc[−] and Hpgl[−] ions^{1,8} the chemical exchange was proven to be fast. This implies stronger interaction between the metal ion and the ligand and can be explained by the binding of an alcoholate group. This reaction takes place most probably on C2–OH since this is the closest to the carboxylate anchor and the ionization of the O–H group

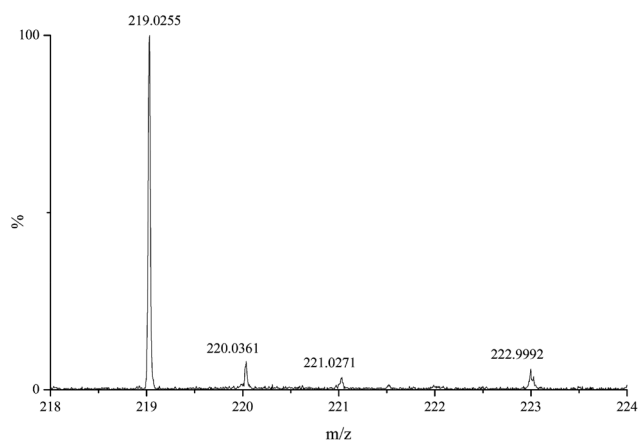


Fig. 6 ESI-MS spectrum of a solution containing 0.05 M Isa[−] and 0.025 M Ca²⁺ in 0.100 M NaOH medium. Spectra were recorded in positive ion mode. The peak at 219.0255 corresponds to the Calsa⁺ species (formed by the protonation of the CalsaH_{−1}⁰ species, predominant in this solution).



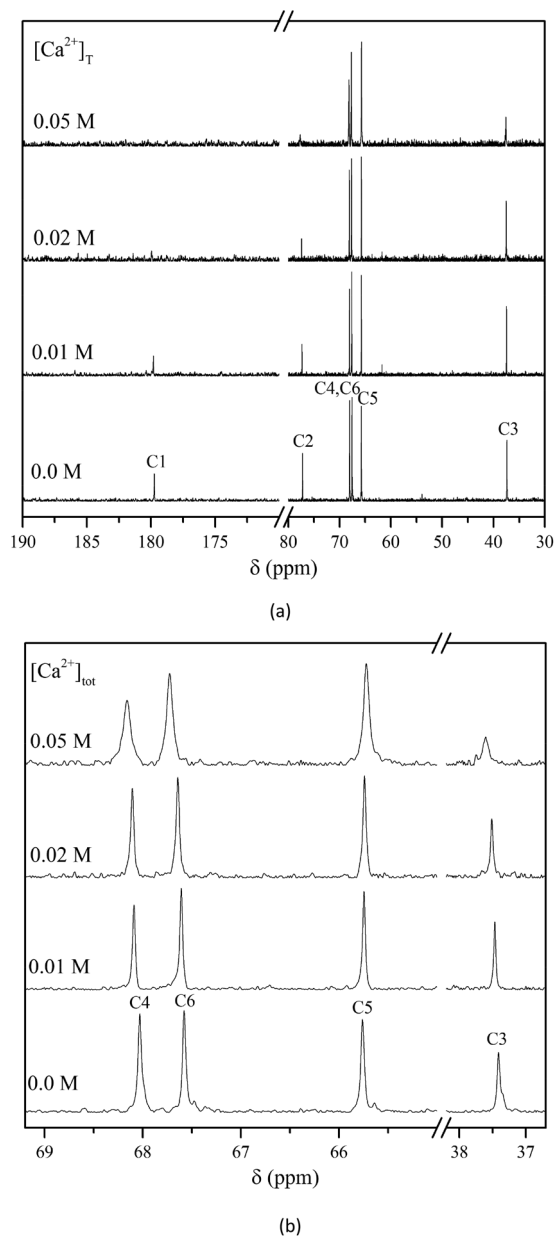


Fig. 7 Normalized ^{13}C NMR spectra of solutions containing $[\text{Isa}^-]_{\text{T}} = 0.2 \text{ M}$, $[\text{OH}^-]_{\text{T}} = 0.2 \text{ M}$ and $[\text{Ca}^{2+}]_{\text{T}} = 0\text{--}0.05 \text{ M}$, $T = (25 \pm 1)^\circ\text{C}$.

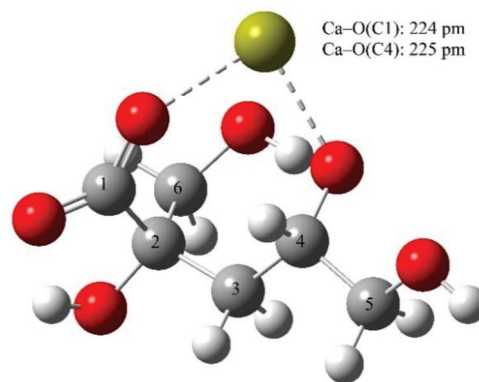
may be facilitated by an intramolecular hydrogen bond (Fig. 2).

Profound differences, however, are seen in the binding arrangement around the calcium ion in CaIsaH_{-1}^0 compared to CaGlucH_{-1}^0 . In the latter, beside the carboxylate oxygens on C1, the alcoholate groups on both the C2 and C3 carbons adjacent to the carboxylate participate in Ca^{2+} binding. This results in the formation of two bonding isomers containing five- or six-membered chelate rings.¹ These arrangements are similar to those observed in analogous systems with neutral and slightly alkaline pH (that is, in the CaGluc^+ solution species).⁸

In CaIsaH_{-1}^0 , unlike in CaGlucH_{-1}^0 , the donor group(s) further away from the carboxylate moiety are most probably

involved in the calcium binding. This is strongly supported by the broadening of the C3 signal upon calcium addition (Fig. 7). Of the two alcoholic OH groups of Isa^- adjacent to the carboxylate (on C2 and C6) only the one on C2 participates in the metal ion binding, while that on C6 does not. In the bonding isomers of CaIsaH_{-1}^0 a carboxylate oxygen on C1 and the alcoholate groups on C2 or C4 coordinate. It is expected that the chelate ring with C1 and C2 oxygens is energetically more favored (5 membered) than that with C1 and C4, which is a 7 membered chelate. The OH on C6 is a non-coordinating group. This is supported not only by the ^{13}C NMR spectra shown in Fig. 7 (the carbon signal barely changes with the increasing calcium concentration), but also by the ^1H NMR spectra shown in Fig. S7† (the proton signal corresponding to H6 and H6' does not change with temperature).

Structural models for the CaIsaH_{-1}^0 complex were also built (Fig. 8 and S8†). From the calculations, the formation of four binding isomers can be proposed. In the two lowest energy bidentate arrangements, Ca^{2+} is bound to the oxygen on C1 and the alcoholate on C2 (Fig. 8a) or C2 and C4 (Fig. 8b); the latter is only 0.7 kJ mol^{-1} higher in energy. In the third structure, oxygens on C1, C4 (alcoholate) and C6 (alcohol) bind Ca^{2+} (Fig. S8a†), while the fourth isomer (Fig. S8b†) with binding oxygens on C1, C6 (alcoholate) and C4



(a)

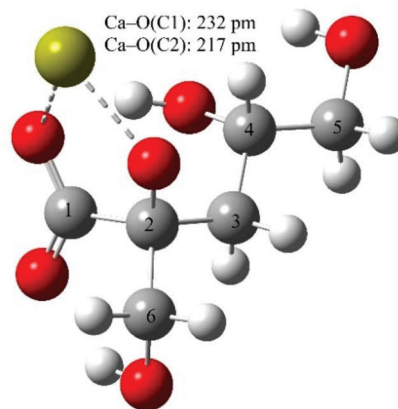


Fig. 8 The optimum geometries calculated for CaIsaH_{-1}^0 at the B3LYP/6-311++G(d,p) level using the PCM model and explicit water molecules.



(alcohol) does not. The formation of these complexes is less probable, since their energies are 2.2 and 2.4 kJ mol⁻¹ higher. From these calculations, the bidentate arrangement with Ca²⁺ is bound to the oxygen on C1 and the alcoholate on C6 is one of the least energetically favored (14.9 kJ mol⁻¹ higher in energy).

Conclusions

The structure of the two ligands (Scheme 1) is similar in the sense that both contain one carboxylate group and several (4 or 5) alcoholic OH groups. On Isa⁻ next to the -COO⁻ group, there are two -OH moieties on C2 and C6; these motifs are practically the same on Gluc⁻ (*i.e.*, the -OH groups on C2 and C3). The structure of Isa⁻ is branched (contains one tertiary C atom), while that of Gluc⁻ can be regarded more like linear (only primary and secondary C atoms are present). Accordingly, Gluc⁻ is expected to be much more flexible than Isa⁻. In the middle of an Isa⁻ anion, there is a methylene group (C3) forming a hydrophobic “bridge” between the two hydrophilic sides of the molecule (comprising of C1, C2 and C6 on one end and C4 and C5 on the other). This hydrophobic motif is completely missing in Gluc⁻, and it is likely to be associated with the difference between the solubility of their calcium salts (pK_{sp} values reported for Ca(Gluc)₂ and Ca(Isa)₂ are 4.19 ± 0.05 and 6.53 ± 0.02, respectively⁴⁰).

In neutral solutions weak complexes are formed with Ca²⁺ ($\log \beta_{1,1,0} = 1.12$ and 1.08 for CaIsa⁺ and CaGluc⁺ at I = 1 M NaCl and at T = 25 °C, respectively). The almost identical formation constants imply the involvement of similar coordinating groups (*i.e.*, carboxylate O and one or two OH). Isa⁻ does not form a 1 : 2 complex with calcium, while at appreciable excess ligand excess, there is unambiguous experimental proof for the formation of the CaGluc_{2(aq.)} solution species. There are indications that the positions of binding sites in CaIsa⁺ and CaGluc⁺ are different: in CaIsa⁺, the OH on C2 does not take part in the metal ion binding, while in CaGluc⁺, both the OH and C2 and that on C3 are capable of coordinating (based on calculations). Either of these features is most probably connected to the rigidity of Isa⁻ compared to Gluc⁻.

The proton dissociation constants of Isa⁻ and Gluc⁻ are also somewhat different ($\log \beta_{0,1,-1} = -14.5 \pm 0.1$ and -13.85 ± 0.02 for Isa⁻ and Gluc⁻, respectively). Isa⁻ is a significantly weaker acid than Gluc⁻, which is due to the presence of an electron withdrawing group (*i.e.*, OH) on C3 of Gluc⁻. Based on the statistical effect of the number of OH groups, the pK of Isa⁻ would only be 0.3 unit higher⁵² which points out the importance of the steric factor as well.

On the basis of the difference between the $\log \beta_{0,1,-1}$ overall proton dissociation constants, IsaH₋₁²⁻ should form more stable Ca²⁺-complex, than GlucH₋₁²⁻, which is reflected in the difference of the stepwise formation constants ($\log \beta_{1,1,-1} = 3.15$ and 2.74 for Isa⁻ and Gluc⁻, respectively). The higher acidity of Gluc⁻ indicates that the CaGluc⁺ complex undergoes deprotonation at a lower pH (the pK of the complex

is $\log \beta_{1,1,0} - \log \beta_{1,1,-1} = 12.02$) than does CaIsa⁺ ($pK = 12.48$). This remarkable difference was found previously for Th(OH)₄GlucH₋₁²⁻; an analogous complex with Isa⁻ in place of Gluc⁻ could not be observed in the same pH range.¹⁹

A more striking difference is seen between the composition and structure of the Ca²⁺-complexes forming in strong alkaline solutions. In Isa⁻-containing solutions, no polynuclear complexes were detected, and under hyperalkaline conditions, the only species is the 1 : 1 : -1 complex. Under identical conditions, solutions with Gluc⁻ are predominated by 2 : 1 : -3 and 3 : 2 : -4 polynuclear complexes with a 1 : 1 : -1 mononuclear complex as a minor species. In the polynuclear complexes, the flexibility of Gluc⁻ makes the simultaneous participation of the alcoholate groups on C2 and C3 in the calcium ion binding possible, and this feature is a prerequisite for the formation of the multinuclear species (see Fig. 6 in ref. 1). As we have shown *via* using multinuclear NMR and molecular modelling calculations, this condition does not hold for Isa⁻; it binds Ca²⁺ only with one alcoholate adjacent to the carboxylate (*i.e.*, on C2). The participation of the other adjacent oxygen donor on C6 is hindered, due to the unfavorable arrangement of the COO⁻, C2-OH and C6-OH groups (Fig. S8†).

Because of this structural difference, it is not expected that Isa⁻ and Gluc⁻ behave identically (or even similarly) in hyperalkaline solutions containing Ca²⁺ ions and furthermore, higher valent cations, such as various actinides and lanthanides, *i.e.*, under conditions relevant to radioactive waste repositories. It is also expected that the thermodynamics and structure of ternary complexes comprising Ca²⁺, L⁻ and various radionuclides depend strongly on whether L⁻ = Isa⁻ or Gluc⁻. Taking this into account, a detailed study, similar to the present one, on the hyperalkaline aqueous chemistry of β-D-isosaccharinate would also be warranted. Birjkumar *et al.* stated that “caution should be exercised when using gluconate as a thermodynamic model for isosaccharinate in uranyl(vi) chemistry”.⁵³ In that sense, our investigations led to similar conclusions with regard to certain aspects of the chemistry of aqueous solutions containing Isa⁻ and calcium.

Conflicts of interest

There are no conflicts to declare.

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