



Cite this: *Phys. Chem. Chem. Phys.*,  
2017, 19, 28512

Received 14th July 2017,  
Accepted 12th September 2017

DOI: 10.1039/c7cp04748g

rsc.li/pccp

# Interplay between the folding mechanism and binding modes in folding coupled to binding processes†

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**Proteins that fold upon binding to their partners exhibit complex binding behavior such as induced-fit. But the connections between the folding mechanism and the binding mode remain unknown. Here we focus on the high affinity complex between the physiologically and marginally unstable, fast folder PSBD and the E1 subunit of pyruvate dehydrogenase. Using coarse-grained simulations we investigate the binding to E1 of a partially disordered PSBD under two folding scenarios: two-state and downhill. Our simulations show that induced-fit binding requires that PSBD folds—unfolds in the downhill folding regime. In contrast, a two-state folding PSBD must fold completely before it binds. The reason is that effective coupling between folding and binding involves partially folded conformations, which are only sufficiently populated under the downhill folding regime. Our results establish a direct mechanistic link between complex binding and downhill folding, supporting the idea that PSBD operates functionally as a conformational rheostat.**

Recent developments are unveiling a plethora of exceptions to the classical paradigm by which proteins fold into unique 3D structures in a single stroke (two-state folding) and bind to their partners in a lock-and-key fashion. The experimental identification of proteins that fold downhill<sup>1</sup> confirmed a key prediction of the energy landscape theory<sup>2</sup> and expanded the palette of folding behaviors. Downhill proteins lose/gain the native structure gradually, eventually leading to one-state folding when the barrier disappears altogether.<sup>3</sup> Such gradual disorder has been characterized at atomic resolution by NMR,<sup>4,5</sup> kinetically using ultrafast methods,<sup>6</sup> and in individual molecules.<sup>7</sup> On the binding front, we now know that intrinsically disordered proteins (IDPs) can fold upon binding<sup>8</sup> in a process that may

provide kinetic advantages (“fly-casting”)<sup>9</sup> and result in complex binding phenomena such as induced-fit and conformational selection.<sup>10</sup> Some IDPs morph upon binding to structurally diverse partners,<sup>11–13</sup> and others exhibit sophisticated allosteric behavior.<sup>14</sup> Interestingly, there is a direct connection between fast, non-cooperative folding and the marginal stability of the native fold that leads to intrinsic disorder.<sup>15</sup> Downhill folding and IDPs thus seem to be two sides of the same coin. It has been in fact proposed that the structural malleability and complex binding modes found on IDPs may require that the protein folds downhill, in which case IDPs could be operating as conformational rheostats rather than conventional switches.<sup>16</sup>

Here we focus on the case of the pyruvate dehydrogenase complex (PDC),<sup>17</sup> a large macromolecular assembly composed of multiple copies of three types of subunits. In this complex a small domain from the central flexible region of E2 (PSBD) binds to both E1 and E3, and it is key to the swinging arm mechanism that channels substrates.<sup>17</sup> Folding studies have shown that PSBDs are marginally stable downhill folders that have been tuned by natural selection to be partially unfolded (disordered) at the living temperature of the host organism.<sup>18,19</sup> In parallel, the binding processes of E1 and E3 to PSBD occur with a 1 : 1 stoichiometry, are mutually exclusive, and are of very high affinity (sub-nanomolar  $K_D$ ).<sup>20</sup> Yet, the structures of PSBD bound to E1 and E3 reveal strikingly superficial binding interfaces (Fig. 1A and B) with few intermolecular contacts (Fig. 1C). In these complexes PSBD exhibits very high *B*-factors that hint at the presence of conformational heterogeneity in the bound state.<sup>21,22</sup> Here we are interested in the role that downhill folding may play in the binding properties of PSBD. In particular, we focus on the E1–PSBD complex (Fig. 1).

To investigate the coupling between PSBD folding and binding to E1, we used coarse-grained simulations, which allow us to control the folding scenario of PSBD and thus investigate the link between folding properties and binding modes. We model the PSBD folding using the Karanicolas and Brooks (KB) model, which is Gō-like, represents the protein geometry using the C $^{\alpha}$  trace (with harmonic potentials for angles and bonds),

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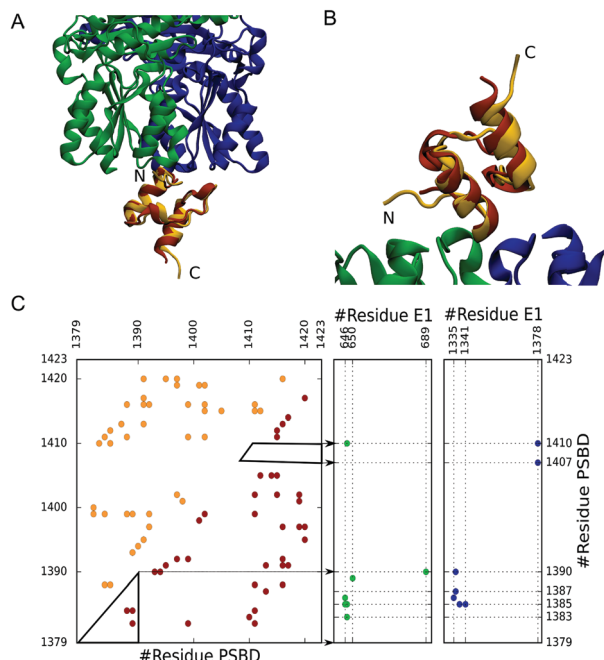
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† Electronic supplementary information (ESI) available: Details on simulation methods and additional examples of transition paths. See DOI: 10.1039/c7cp04748g





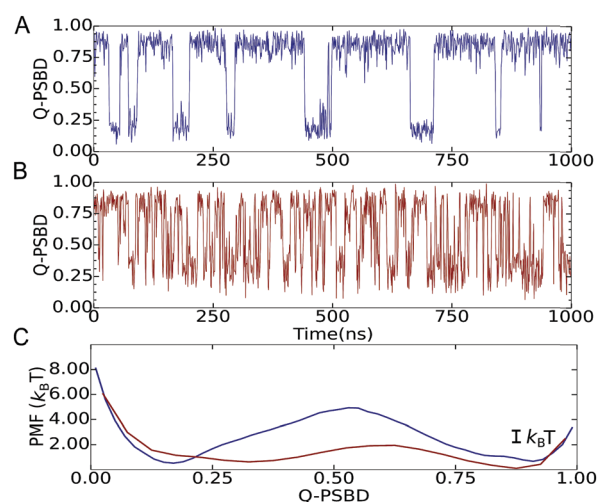
**Fig. 1** Experimental 3D structure of the E1-PSBD complex (1w85) in top-down (A) and bottom-up (B) views. E1 subunits are shown in green and blue. The bound PSBD is shown in dark red. We overlay the NMR structure of monomeric PSBD (1w3d, gold) used in our simulations. (C) Contact map of the PSBD and its intermolecular interactions with E1. For PSBD (left), the upper left triangle (gold) corresponds to the NMR contact map, whereas the lower right triangle (dark red) corresponds to the X-ray structure. The intermolecular contacts with E1 are shown in the right panels in blue and green as the corresponding subunits.

and contains statistical energy terms for torsions as well as a native-centric pairwise contact potential<sup>23</sup> (see the ESI† for details). We parameterized the KB model from the contacts derived from a high-quality NMR structure of PSBD obtained at 293 K (1w3d,<sup>24</sup> gold in Fig. 1) using a 4.5 Å cutoff distance for contacts between heavy atoms, as originally prescribed.<sup>23</sup> The X-ray structure does not include the N- and C-terminal residues of PSBD because they were unstructured in the crystal. To define the interactions in the complex, we added the missing residues without incorporating additional contacts to avoid non-physiological protein end effects (total length of 45 residues). In Fig. 1C we show the contact map that forms the basis of the Gō potential for PSBD folding together with the intermolecular contacts with E1 derived from the complex structure (1w85).<sup>22</sup> Because the binding to E1 also involves multiple charged residues, we added a description of electrostatic interactions using a simple Debye-Hückel term calibrated against experiments<sup>25</sup> and used before in the context of IDP binding.<sup>26</sup>

We adjusted the intramolecular PSBD interaction energy to the folding temperature to mimic the physiological conditions of the host organism (*B. stearotherophilus* lives around 333–343 K, and the experimental denaturation temperature of PSBD is  $\sim 340$  K).<sup>19</sup> Under these conditions PSBD is partially disordered, favoring the coupling of folding and binding. To investigate the role played by PSBD's folding scenario on binding, we fine-tuned the model to modify the free energy

barrier at the folding temperature. This procedure involved extensive equilibrium simulations at multiple temperatures for every version of the model using Langevin dynamics within the Gromacs 4.0.5 package,<sup>27</sup> and analyzing the trajectories with histogram methods.<sup>28</sup> From this exercise we found that the PSBD folding free energy barrier could be modulated by: (1) making the repulsive radii of the C $\alpha$  beads all the same, in contrast to the original KB implementation in which the repulsive radius for each bead is set to the distance to the closest residue in the native structure that is not in a direct contact<sup>23</sup> and (2) scaling down the magnitude of the angular and dihedral contributions to the force-field (see technical details in the ESI†). In contrast, changes in the contact interaction energies had negligible effects on the folding free energy barrier.

We then focused on two folding scenarios. In the first one, PSBD folding-unfolding is two-state-like, as manifested by trajectories projected as a function of the fraction of native PSBD contacts ( $Q_{\text{PSBD}}$ ; see the ESI† for definitions) showing a binary switching pattern (Fig. 2A). The corresponding potential of mean force (PMF) for this model shows two defined basins at  $Q_{\text{PSBD}} \approx 0.2$  and  $0.85$  separated by a  $\sim 4.7 k_{\text{B}}T$  free energy barrier (blue in Fig. 2C). For the downhill folding scenario, the modified model reproduced the marginal thermodynamic folding barrier ( $\sim 1 k_{\text{B}}T$ ) obtained experimentally from differential scanning calorimetry<sup>29</sup> and multi-probe thermodynamic analysis.<sup>19</sup> The downhill version of the model resulted in faster dynamics and more complex folding-unfolding trajectories projected as a function of  $Q_{\text{PSBD}}$  (Fig. 2B), and a PMF with a free energy barrier of  $\sim 1.2 k_{\text{B}}T$  (red in Fig. 2C). Here we note that the behavior shown in Fig. 2B and C lies well within the downhill scenario, but it does not reach the one-state (global downhill) limit.<sup>3</sup> It is also worth noting that lowering the barrier is accompanied by displacement of the unfolded well to higher  $Q_{\text{PSBD}}$ , consistent with the observation of the residual structure in the unfolded states of fast folding proteins.<sup>30</sup>



**Fig. 2** Projection of the Langevin dynamics trajectories on the folding order parameter  $Q$  (fraction of native contacts) for the two-state (A, blue) and downhill (B, red) models of PSBD. (C) PMF for the projection on  $Q$  at the midpoint temperature for both scenarios.

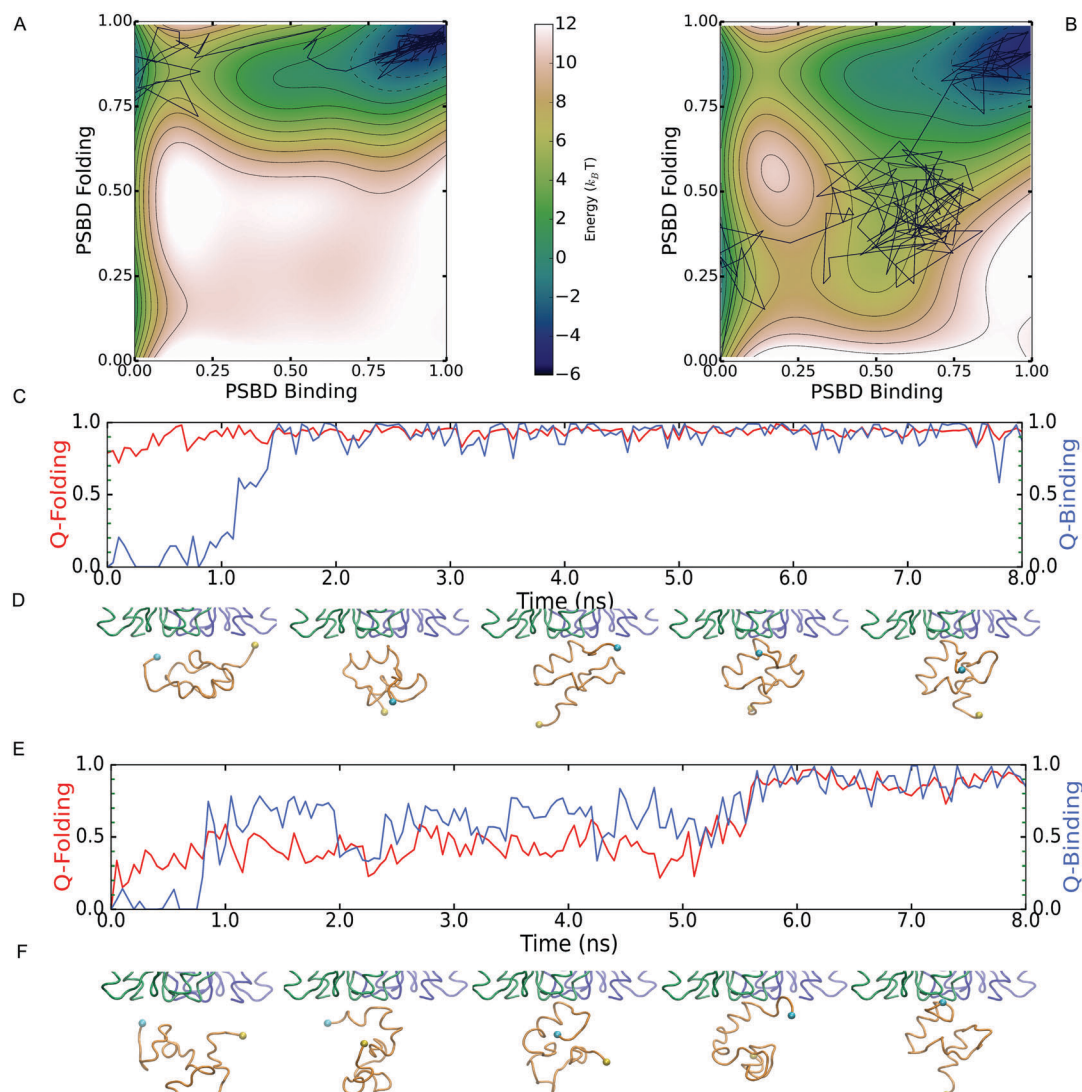


To investigate the coupling between PSBD folding and binding to E1, we parameterized the binding component of the Gō-like force-field using intermolecular contacts derived from the original X-ray structure of the E1-PSBD complex (1w85, Fig. 1C). In this structure, the coordinates for the PSBD are not well resolved (low electron density and very high *B* factors) and several E1 residues and/or atoms are missing.<sup>22</sup> We added all the missing parts using the Modeller software package<sup>31</sup> with a limited number of optimization steps to minimize deviations from the experimental structure. In a second step we grafted the PSBD structure from NMR (1w3d) onto the E1 docking site and fitted its exact position to the crystallographic PSBD model by minimizing the RMSD of select C $\alpha$  atoms (*i.e.* final RMSD = 0.38 Å, see Fig. 1A and B). This exercise, which was necessary to build a complete model for binding, did not change the intermolecular contacts that define the binding interface in the X-ray structure. The final intra-PSBD

and inter-molecular interactions used for all the calculations are shown in Fig. 1C.

We then calibrated the strength of intermolecular interactions on both folding scenarios so that binding simulations reproduced the experimental dissociation constant of the complex,  $K_D = 3.2 \times 10^{-10} \text{ M}^{32}$  (we used a 600 Å per side cubic simulation box, resulting in 7.7  $\mu\text{M}$  protein concentration). Binding coupled to folding was then investigated by performing long replica exchange simulations keeping the E1 coordinates frozen. The simulations sampled well both the bound and unbound states, resulting in an aggregate simulation time of 96 microseconds (see the ESI† for additional details).

Although the two scenarios are identical in terms of the degree of PSBD unfolding and the binding affinity to E1, the simulations revealed important differences in binding behavior. Such differences are readily apparent in Fig. 3A and B, which show the 2D PMFs



**Fig. 3** (A and B) Potentials of mean force as a function of folding (Q-PSBD) and binding (Q-PSBD-E1) order parameters determined for the two-state (A) and downhill folding (B) scenarios. The black lines represent the transition paths shown in the following panels. (C and D) Examples of folding (red) and binding (blue) transition path on the two-state scenario (C) with selected snapshots of the binding event (D). (E and F) Examples of folding (red) and binding (blue) transition path on the downhill folding scenario (E) with selected snapshots of the binding event (F).





calculated as a function of folding and binding order parameters. The PMF for the two-state scenario (Fig. 3A) shows that in the unbound state (low  $Q_{\text{PSBD-E1}}$ ) the protein fluctuates between the equally populated folded and unfolded basins (midpoint conditions). However, the binding to E1 only takes place when PSBD is fully folded ( $Q_{\text{PSBD}} > 0.7$ ) and thus folding and binding are decoupled. The entire process can be described with the simple linear three state model:  $\text{UU} \rightleftharpoons \text{FU} \rightleftharpoons \text{FB}$ , in which the first letter indicates the PSBD folding status (U-unfolded and F-folded) and the second its binding status (U-unbound and B-bound). E1 thus selects the folded conformation of PSBD from an ensemble with equal populations of folded and unfolded species, and hence the binding process can be described as following a conformational selection mechanism.<sup>33</sup> In contrast, the PMF for the downhill folding scenario (Fig. 3B) demonstrates an effective coupling between folding and binding that results in richer behavior. Fig. 3B shows two alternative mechanisms. The first one is the  $\text{UU} \rightleftharpoons \text{FU} \rightleftharpoons \text{FB}$  found in the two-state scenario. The second one involves binding of unfolded PSBD (UB), manifested by a significant population of microstates with low to intermediate values of  $Q_{\text{PSBD}}$  (0.2–0.5) and intermediate to high values of  $Q_{\text{PSBD-E1}}$  (0.5–0.6). This alternative mechanism can be described with the scheme:  $\text{UU} \rightleftharpoons \text{UB} \rightleftharpoons \text{FB}$ , where a partially unfolded PSBD molecule first binds and then folds while being bound, thus following an induced fit mechanism.

Analysis of the binding transition paths provides additional information. Transition paths are the reactive segments of a trajectory, which comprise all the mechanistic information about the binding process. Examples of binding transition paths are provided in Fig. 3C–F. The binding transition path for the two-state scenario (Fig. 3C and D) shows how PSBD is already folded before approaching its docking site in E1. Upon contact with E1, folded PSBD binds *via* a lock-and-key mechanism that results in a sharp transition and the subsequent freezing of PSBD's conformational ensemble in the bound state (*i.e.* FB experiences minimal  $Q_{\text{PSBD}}$  fluctuations). All the observed transition paths for the two-state scenario adhere to this behavior. Therefore, there are no structural correlations between binding and folding in the two-state scenario.

The transition path shown for the downhill folding scenario (Fig. 3E and F) corresponds to the folding–binding diagonal path in the 2D PMF (Fig. 3B). Binding takes place from the unfolded state resulting in concerted folding and binding *via* an induced-fit mechanism. The transition path is accordingly much longer (over an order of magnitude difference) and permits the reconfiguration of PSBD while it remains loosely bound to E1 ( $Q_{\text{PSBD-E1}} \sim 0.50$ ). Binding and folding exhibit correlated fluctuations during which PSBD populates partially unfolded conformations for significant fractions of the time. These conformations are in fact part of the PSBD folding barrier ( $0.55 < Q_{\text{PSBD}} < 0.75$ ) and become effectively coupled to binding because their basal population is significant (marginal barrier). Additional examples of induced-fit binding transition paths are provided in the ESI.† Overall, roughly 50% of the binding transition paths observed under the downhill scenario were induced-fit, whereas the remainder involved complete folding followed by binding (*i.e.* conformational selection).

## Conclusions

In this work we have used a coarse grained computational model to explore how the folding mechanism of PSBD (*i.e.* two-state *vs.* downhill) influences its binding to E1. We find that the two-state version of PSBD must fold before binding to E1. Binding occurs *via* a conformational selection mechanism where PSBD remains structurally rigid while bound. In contrast, the downhill PSBD version populates myriads of partially folded conformations that are able to bind to E1, resulting in induced-fit binding. We can thus conclude that effective coupling between folding of PSBD and binding to E1 requires: (1) a downhill folding scenario and (2) a structurally localized binding interface that facilitates binding of partially folded conformations. Therefore, the presence of intrinsic disorder is not sufficient *per se* to produce complex binding behavior such as induced-fit.

Our results provide a practical demonstration of a conformational rheostat, in which the conformational flexibility of a downhill folding ensemble makes it possible to implement complex functions in single-domain proteins.<sup>1</sup> The possibility that PSBD operates as a conformational rheostat sheds some light onto the role that this small domain plays in the swinging arm of PDC. Particularly, it hints at how a partially disordered, downhill folding PSBD could adapt its folding ensemble to bind to the structurally distinct E1 and E3 and result in the effective coupling of both binding events. Such coupling could result in the alternating binding cycles that are required for efficiently channeling substrates during PDC catalysis.<sup>17</sup> Moreover, the changes in the microenvironment (reactants, products, and cofactors) that take place during this complex reaction could further tune the PSBD folding ensemble to trigger binding to (or dissociation from) E1 or E3, thus synchronizing catalysis and binding throughout the entire reaction.

More generally, the connection that we observe between the folding scenario of PSBD and its binding behavior originates from fundamental physical principles and is not specific of the molecular system studied. Hence, the same connection between the folding mechanism and binding should apply to the binding modes of many other proteins that fold in the downhill regime. Downhill folding proteins share certain features, including a small size, a primarily helical secondary structure and a loosely knit network of contacts in the hydrophobic core,<sup>30</sup> which are also often associated with some IDPs. These commonalities lead us to hypothesize that the complex binding modes and allosteric behavior that are being reported in the folding upon binding of IDPs<sup>16</sup> do not simply emerge from the protein being unstructured, but require the IDP to fold downhill. In other words, we hypothesize that a conformational rheostat (*i.e.* a partly disordered downhill folder) is the molecular mechanism behind the functioning of IDPs.<sup>16</sup>

## Conflicts of interest

There are no conflicts to declare.



## Acknowledgements

RS acknowledges an FPI fellowship from the Spanish Ministry of Economy and Competitiveness. DDS was supported by an Ikerbasque Research Fellowship and acknowledges Robert B. Best for useful discussions and the kind donation of computational tools. This research was supported by grants BIO 2011-28092 (Spanish Ministry of Economy and Competitiveness), ERC-2012-ADG-323059 (European Research Council) and MCB-1616759 (National Science Foundation) to VM, and grant CTQ2015-65320-R (MINECO/FEDER) to DDS.

## References

- 1 M. M. Garcia-Mira, M. Sadqi, N. Fischer, J. M. Sanchez-Ruiz and V. Munoz, *Science*, 2002, **298**, 2191–2195.
- 2 J. D. Bryngelson, J. N. Onuchic, N. D. Socci and P. G. Wolynes, *Proteins*, 1995, **21**, 167–195.
- 3 V. Muñoz, *Annu. Rev. Biophys. Biomol. Struct.*, 2007, **36**, 395–412.
- 4 M. Sadqi, D. Fushman and V. Munoz, *Nature*, 2006, **442**, 317–321.
- 5 L. Sborgi, A. Verma, S. Piana, K. Lindorff-Larsen, M. Cerminara, C. M. Santiveri, D. E. Shaw, E. de Alba and V. Muñoz, *J. Am. Chem. Soc.*, 2015, **137**, 6506–6516.
- 6 P. Li, F. Y. Oliva, A. N. Naganathan and V. Muñoz, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 103–108.
- 7 J. Liu, L. A. Campos, M. Cerminara, X. Wang, R. Ramanathan, D. S. English and V. Muñoz, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 179–184.
- 8 V. N. Uversky, *Chem. Rev.*, 2014, **114**, 6557–6560.
- 9 B. A. Shoemaker, J. Portman and P. G. Wolynes, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 8868–8873.
- 10 R. B. Berlow, H. J. Dyson and P. E. Wright, *FEBS Lett.*, 2015, **589**, 2433–2440.
- 11 B. Y. Qin, C. Liu, H. Srinath, S. S. Lam, J. J. Correia, R. Derynck and K. Lin, *Structure*, 2005, **13**, 1269–1277.
- 12 J. C. Ferreón, C. W. Lee, M. Arai, M. A. Martinez-Yamout, H. J. Dyson and P. E. Wright, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 6591–6596.
- 13 M. Kjaergaard, K. Teilum and F. M. Poulsen, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 12535–12540.
- 14 A. C. Ferreón, J. C. Ferreón, P. E. Wright and A. A. Deniz, *Nature*, 2013, **498**, 390–394.
- 15 A. N. Naganathan, U. Doshi and V. Muñoz, *J. Am. Chem. Soc.*, 2007, **129**, 5673–5682.
- 16 V. Muñoz, L. A. Campos and M. Sadqi, *Curr. Opin. Struct. Biol.*, 2016, **36**, 58–66.
- 17 R. N. Perham, *Annu. Rev. Biochem.*, 2000, **69**, 961–1004.
- 18 A. N. Naganathan, P. Li, R. Perez-Jimenez, J. M. Sanchez-Ruiz and V. Muñoz, *J. Am. Chem. Soc.*, 2010, **132**, 11183–11190.
- 19 A. N. Naganathan and V. Munoz, *J. Phys. Chem. B*, 2014, **118**, 8982–8994.
- 20 I. A. Lessard and R. N. Perham, *Biochem. J.*, 1995, **306**(Pt 3), 727–733.
- 21 S. S. Mande, S. Sarfaty, M. D. Allen, R. N. Perham and W. G. Hol, *Structure*, 1996, **4**, 277–286.
- 22 R. A. Frank, J. V. Pratap, X. Y. Pei, R. N. Perham and B. F. Luisi, *Structure*, 2005, **13**, 1119–1130.
- 23 J. Karanicolas and C. L. Brooks, *Protein Sci.*, 2002, **11**, 2351–2361.
- 24 M. D. Allen, R. W. Broadhurst, R. G. Solomon and R. N. Perham, *FEBS J.*, 2005, **272**, 259–268.
- 25 Y. C. Kim and G. Hummer, *J. Mol. Biol.*, 2008, **375**, 1416–1433.
- 26 D. De Sancho and R. B. Best, *Mol. Biosyst.*, 2012, **8**, 256–267.
- 27 B. Hess, C. Kutzner, D. van der Spoel and E. Lindahl, *J. Chem. Theory Comput.*, 2008, **4**, 435–447.
- 28 S. Kumar, J. M. Rosenberg, D. Bouzida, R. H. Swendsen and P. A. Kollman, *J. Comput. Chem.*, 1992, **13**, 1011–1021.
- 29 A. N. Naganathan, R. Perez-Jimenez, V. Muñoz and J. M. Sanchez-Ruiz, *Phys. Chem. Chem. Phys.*, 2011, **13**, 17064–17076.
- 30 D. De Sancho, U. Doshi and V. Munoz, *J. Am. Chem. Soc.*, 2009, **131**, 2074–2075.
- 31 N. Eswar, B. Webb, M. A. Marti-Renom, M. S. Madhusudhan, D. Eramian, M.-y. Shen, U. Pieper and A. Sali, *Current protocols in bioinformatics*, ed. Andreas D. Baxevanis *et al.*, 2006, ch. 5, unit-5.6.
- 32 H.-I. Jung, S. J. Bowden, A. Cooper and R. N. Perham, *Protein Sci.*, 2002, **11**, 1091–1100.
- 33 D. D. Boehr, R. Nussinov and P. E. Wright, *Nat. Chem. Biol.*, 2009, **5**, 789–796.

