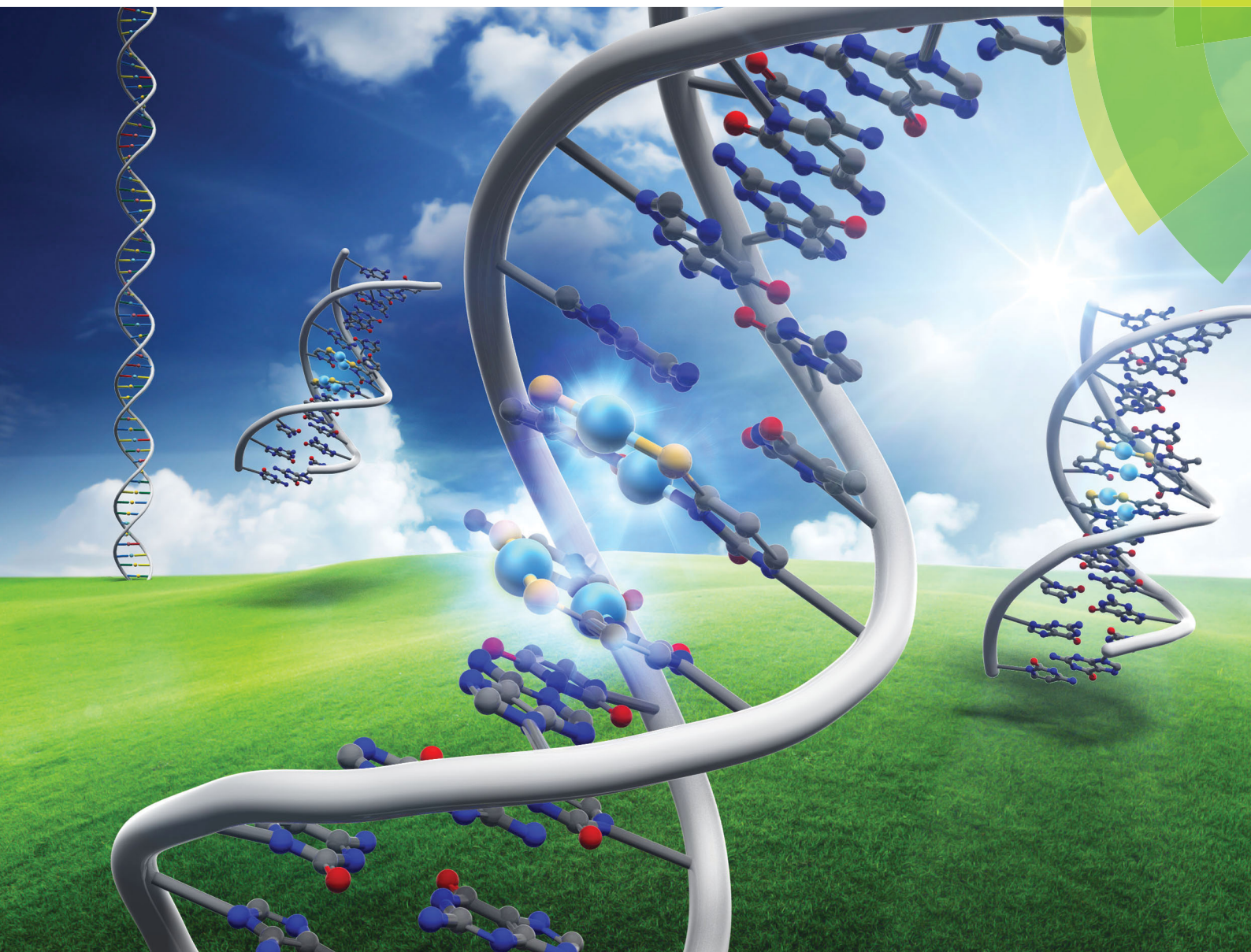


ChemComm

Chemical Communications

rsc.li/chemcomm



ISSN 1359-7345



COMMUNICATION

Jiro Kondo, Akira Ono *et al.*

Crystal structure of a DNA duplex containing four Ag(I) ions in consecutive dinuclear Ag(I)-mediated base pairs: 4-thiothymine–2Ag(I)–4-thiothymine



Cite this: *Chem. Commun.*, 2017, 53, 11747

Received 7th August 2017,
Accepted 29th August 2017

DOI: 10.1039/c7cc06153f

rsc.li/chemcomm

Herein, we determined a high-resolution crystal structure of a B-form DNA duplex containing consecutive dinuclear metal ion-mediated base pairs, namely, 4-thiothymine–2Ag(I)–4-thiothymine (S–2Ag(I)–S), and four Ag(I) ions form a rectangular network and the distances between the Ag(I) ions are 2.8–3.2 Å, which may indicate the existence of metallophilic attractions.

Strategies to use DNA as templates to generate functional molecules, precisely designed molecular architecture, and nano-scale devices have been widely investigated since nucleic acids can be assembled in a sequence-directed manner to form multi-dimensional structures.¹ Recently, duplexes containing metal-mediated base pairs (metallo-base pairs), in which Watson–Crick base pairs are replaced by metal–base coordination complexes, have been identified. In addition, extensive studies on metal-mediated pairs of natural and artificial bases have expanded the design possibilities of functional DNA molecules.² Metallo-base pairs have been applied to DNA molecular devices, such as ion-sensors,³ electric transport nano-wires,⁴ and DNA magnets.⁵ Many metallo-base pairs consist of two bases (heterocycles) and a metal ion placed between the bases to connect them.² For example, in the presence of Hg(II) ions, a thymine–thymine (T–T) pair in a duplex forms a metallo-base pair, T–Hg(II)–T (Fig. 1a),⁶ and in the presence of Ag(I) ions, a cytosine–cytosine (C–C) pair forms C–Ag(I)–C (Fig. 1b).⁷ Recently, metallo-base pairs containing two metal ions between bases have been identified. A 4-thiothymine–4-thiothymine (S–S) in a duplex captures two Ag(I) ions to form a dinuclear Ag(I)-mediated base pair, S–2Ag(I)–S; thus, the duplex was thermally stabilized in the presence of Ag(I) ions.⁸ The predicted structure of S–2Ag(I)–S is shown in Fig. 1c. Metal ion binding to the pyrimidine base

Crystal structure of a DNA duplex containing four Ag(I) ions in consecutive dinuclear Ag(I)-mediated base pairs: 4-thiothymine–2Ag(I)–4-thiothymine†

Jiro Kondo,^a Toru Sugawara,^b Hisao Saneyoshi^b and Akira Ono^b

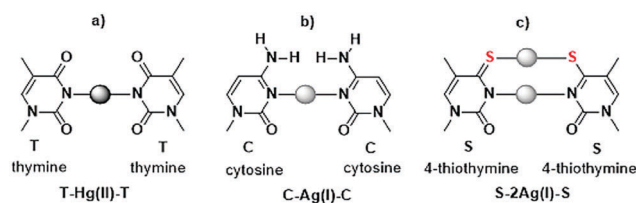


Fig. 1 Metallo-base pairs consisting of pyrimidine bases and metal ions.

pairs is highly selective; thus, Ag(I) ions do not bind to T–T pairs and Hg(II) ions do not bind to C–C pairs under ordinary circumstances.⁹ Therefore, a minor modification of the pyrimidine base strongly affects the metal ion binding properties, such as the 4-carbonyl group being converted to a thiocarbonyl group⁸ and substitutions at the 5-position of uracil residues.¹⁰

To date, several dinuclear metal ion-mediated base pairs have been reported.¹¹ Müller and colleagues used an artificial base to form metallo-base pairs containing two Ag(I) ions¹² and two Hg(II) ions¹³ in antiparallel- and parallel-oriented duplexes. In addition, Seela and colleagues used ring-expanded pyrimidine bases to form extremely stable dinuclear Ag(I)-mediated base pairs in antiparallel- and parallel-oriented duplexes.¹⁴

Only a few studies have provided structural information on nucleic acid duplexes containing metallo-base pairs.^{2e} Recently, we solved the solution structure of a DNA duplex containing two consecutive T–Hg(II)–T base pairs using NMR spectroscopy^{6f} and X-ray crystallography.^{6g} In the present study, we performed X-ray analysis of a DNA duplex containing consecutive dinuclear Ag(I)-mediated base pairs, S–2Ag(I)–S (Fig. 2). Four Ag(I) ions metallophilically interacted in a B-form duplex. Data acquired in crystalline and solution states provide a reliable structural basis for advanced design and further development of metallo-DNAs.

The DNA dodecamer d(CGCGASSTCGCG) (S = 4-thiothymine), hereafter referred to as 12SS, was designed to fold as a self-complementary duplex containing tandem S–S mismatches at the centre (Fig. 2a). These DNA fragments are commonly used as models in crystallographic studies of B-form DNAs containing natural bases, artificial bases, and metallo-base pairs. Recently,

^a Department of Materials and Life Sciences, Faculty of Science and Technology, Sophia University, 7-1 Kioi-cho, Chiyoda-ku, Tokyo 102-8554, Japan.

E-mail: j.kondo@sophia.ac.jp

^b Department of Materials & Life Chemistry, Faculty of Engineering, Kanagawa University, 3-27-1 Rokkakubashi, Kanagawa-ku, Yokohama, 221-8686 Kanagawa, Japan. E-mail: akiraono@kanagawa-u.ac.jp

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7cc06153f



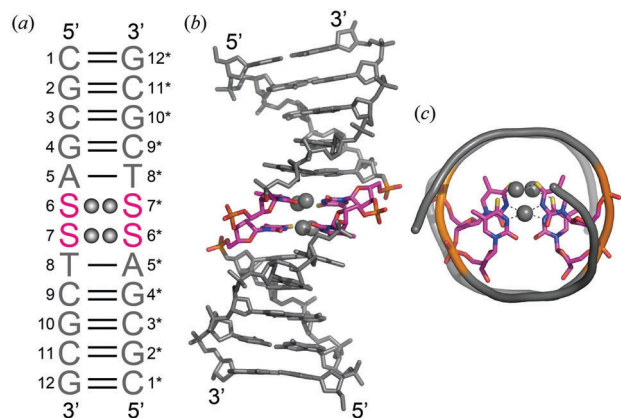


Fig. 2 Secondary structure (a) and crystal structure (side view (b) and top view (c)) of **12SS/Ag(I)**. In these figures, Ag(I) ions are shown as gray spheres. In (c), only two S–2Ag(I)–S base pairs are shown, and the covalent bonds between Ag(I) and base residues are represented by dashed lines.

the crystal structure of a DNA duplex containing consecutive T–Hg(II)–T base pairs has been solved using d(CGCGATTTCGCG), **12TT**.^{6g}

The DNA dodecamer **12SS** was synthesized on a DNA synthesizer using commercially available phosphoramidite units, after which the dodecamer was deprotected using protocols recommended by the provider. The deprotected dodecamer was purified using HPLC with a reverse-phase silica gel column.⁸ The **12SS** was co-crystallized with Ag(I) ions. The crystal structures of **12SS/Ag(I)** were deposited in the Protein Data Bank (PDB) under ID codes 5XUV. A detailed description of the Materials and methods is included in the ESI.†

In the presence of Ag(I) ions, **12SS** formed an antiparallel right-handed double helix (Fig. 2a and b). At both ends of the double helix, canonical Watson–Crick G–C and A–T pairs were formed. In the center of the double helix, two Ag(I) ions were placed in each S–S mispair and formed S–2Ag(I)–S metallo-base pairs (Fig. 1c); thus, the four Ag(I) ions were placed in proximity. Additionally, a K⁺ ion bridges two O₂ atoms of the S–S mispair (Fig. 3a). The local helical parameters, intra-base pair parameters, and pseudorotation phase angles clearly indicate that the duplex adopts a standard B-conformation (ESI,† Tables S2 and S3). Furthermore, the overall duplex structure was similar to that of a duplex containing consecutive T–Hg(II)–T metallo-base pairs, **12TT/Hg**.^{6g} The formation of S–2Ag(I)–S base pairs did not significantly affect the overall B-form conformation.

The electron density maps and geometries of the S–2Ag(I)–S base pairs are shown in Fig. 3a. The $2|F_o| - |F_c|$ maps for S–2Ag(I)–S pairs are broadened at the center, where the Ag(I) ions are found. Based on the omit map calculated after removing the four Ag(I) ions, the metal ion occupies the center between the two S residues. N3...Ag(I) and S4...Ag(I) distances are around 2.1 Å and 2.4 Å, respectively; thus, N3...N3 and S4...S4 distances in each metallo-base pair are around 4.2 Å and 4.8 Å, respectively, and the numerical values are indicative of the disappearance of hydrogens (imino protons). Overall, the imino protons dissociated to generate anionic pyrimidine bases, between which

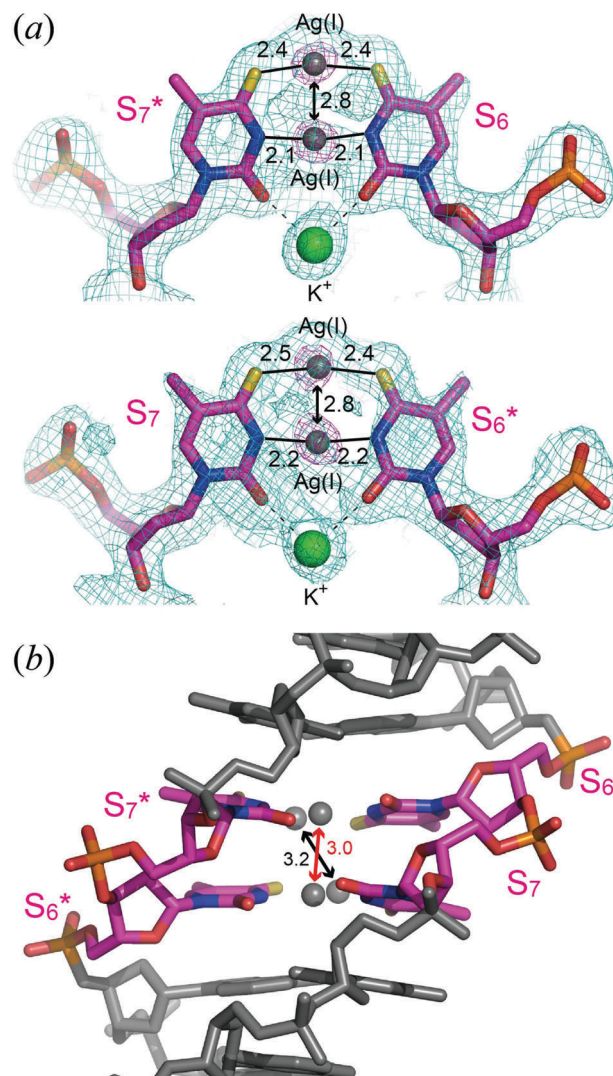


Fig. 3 Structures of S–2Ag(I)–S base pairs. (a) Local $2|F_o| - |F_c|$ (blue: 1 σ contour level) and omit (magenta: 10 σ contour level) maps for S–2Ag(I)–S base pairs. Ag(I) ion arrangements in metallo-base pair planes are shown. Ag(I) ions and K⁺ ions are shown as gray and green spheres, respectively. Distances (Å) between N3 and Ag(I), thiocarbonyl and Ag(I) are represented by lines, and distances between Ag(I) ions are represented by double headed arrows. (b) Ag(I) ion arrangements between the metallo-base pairs are shown. For better understanding, K⁺ ions are not shown in this figure. The inter-base pair Ag(I)...Ag(I) distance of N3–Ag(I)–N3 is shown in red and that of S–2Ag(I)–S in black.

Ag(I) ions are incorporated to form the metallo-base pair, S–2Ag(I)–S.

The propeller twist angles of the S–2Ag(I)–S base pairs (–19° and –21°) are larger than that of the canonical Watson–Crick base pairs in the B-form DNA (–1°) (Fig. 3b and ESI,† Table S2). This property is similar to that of T–Hg(II)–T^{6g} and C–Ag(I)–C^{7b} pairs, in which propeller twist occurs since there is no extra bond (excluding the N3–Hg(II)–N3 and N3–Ag(I)–N3 bonds between two pyrimidine residues) and due to the repulsion of carbonyl groups and amino groups, respectively (Fig. 1). Although 4-thiathymine residues are connected by two silver-mediated bonds, N3–Ag(I)–N3 and S4–Ag(I)–S4, and a potassium-mediated



bond, O2–K⁺–O2, the propeller twist angles of the S–2Ag(I)–S pairs are as large as those of T–Hg(II)–T and C–Ag(I)–C pairs. The C1'···C1' distances of the S–2Ag(I)–S base pairs (9.2 Å and 9.4 Å) are more than 1 Å shorter than those in the canonical Watson–Crick base pairs (~10.7 Å), but the S–2Ag(I)–S base pairs fit into the B-form conformation (ESI,† Tables S2 and S3).

The intra-base pair Ag(I)···Ag(I) distance in each S–2Ag(I)–S is 2.8 Å (Fig. 3a), much less than the sum of their van der Waals radii (3.44 Å).¹⁵ The numerical value, 2.8 Å, is suggestive of a metallophilic interaction between Ag(I) ions.¹⁶ Similar values for Ag(I)···Ag(I) distances in metallo-base pairs were estimated based on DFT calculations.^{12a,c,e}

In the B-form DNA, the helical axis runs through the center of the base pairs, and the Ag(I) ions in the S–2Ag(I)–S base pairs are aligned along the helical axis (Fig. 2c). The inter-base pair Ag(I)···Ag(I) distances are 3.0 Å and 3.2 Å (Fig. 3b). The distance between the two Ag(I) ions between the N3 positions (shown in red, in the center of the helix) is slightly shorter than that of the two Ag(I) ions between the 4-thiocarbonyl groups (shown in black, slightly strayed off the center of the axis). The numerical values, 3.0 Å and 3.2 Å, are shorter than the distances between Watson–Crick base pairs in B-form DNA (3.4 Å), which is indicative of a metallophilic attraction between Ag(I) inter-base pairs. A similar metallophilic attraction has been observed in the above-mentioned crystals of the DNA duplex containing consecutive T–Hg(II)–T base pairs.^{6g} A Hg(II)···Hg(II) distance of 3.3 Å was indicative of a metallophilic attraction for Hg(II). The NMR solution structure of a B-form DNA duplex containing three consecutive imidazole–Ag(I)–imidazole base pairs¹⁷ showed an average Ag(I)···Ag(I) distance of 3.45 Å. Thus, it is important to compare the Hg(II)···Hg(II) distance^{6g} and Ag(I)···Ag(I) distances in this study since the DNA sequences and whole duplex structures in the two crystals are similar. The Ag(I)···Ag(I) distances (3.0 Å and 3.2 Å) are smaller than the Hg(II)···Hg(II) distance (3.3 Å), which may indicate that S–2Ag(I)–S pairs are more tightly fixed into the duplex structure. The short distances and the relatively large propeller twist angles of the S–2Ag(I)–S base pairs might be explained by metallophilic attractions.

A schematic representation of the steric arrangement of four Ag(I) ions is shown in Fig. 4(i). The four Ag(I) ions were arranged in a rectangle (approximately square)-shaped network and the distances between the Ag(I) ions were 2.8–3.2 Å, which may indicate the existence of metallophilic attractions. In addition, four Ag(I) ion clusters are regularly arranged in a three-dimensional space in the crystal (Fig. 4(ii)). The distances between the four Ag(I) ion clusters are 25–30 Å.

Recently, a strategy to knit DNA strands and obtain nanoscale architecture (known as “DNA origami”) has been developed.¹⁸ Crystals consisting of functional oligonucleotides may be another strategy to construct DNA-based architecture, in which the functional groups are precisely arranged in a three-dimensional space.^{9b,19} For instance, crystals consisting of metal–DNA nanowires with uninterrupted one-dimensional silver arrays have been identified.^{9b} In the crystals, silver atom arrays are precisely arranged to form the three-dimensional nanoscale architecture.

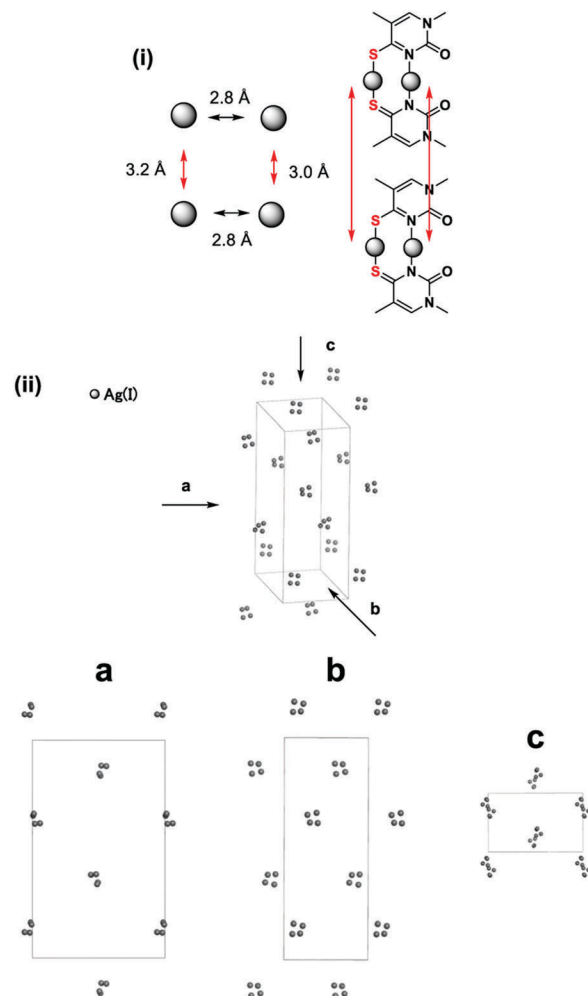


Fig. 4 (i) A schematic representation of a steric arrangement of four Ag(I) ions. (ii) A three-dimensional representation of Ag(I) in the crystal. Ag(I) is extracted and oligonucleotides are removed. The views are down the crystallographic *a*-, *b*- and *c*-axes, respectively.

In conclusion, crystal structures of DNA duplexes revealed the following: (i) Ag(I) ions stabilize the B-form conformation by forming S–2Ag(I)–S base pairs, (ii) the dinuclear Ag(I)-mediated base pair is structurally replaceable with normal Watson–Crick base pairs based on the size and position within the B-form DNA duplex, (iii) four Ag(I) ions form a rectangular network and the relatively short Ag(I)···Ag(I) distances observed inside DNA are suggestive of the stabilizing metallophilic attraction between Ag(I) ions in consecutive S–2Ag(I)–S base pairs. The crystal structure of the metallo–DNA duplex containing the consecutive dinuclear Ag(I)-mediated base pairs provides the basis for the structure-based design of metal-conjugated nucleic acid nanomaterials.

This work was supported by a Grant in Aid for Scientific Research (B) (No. 17H03033), a Strategic Development of Research Infrastructure for Private Universities from the Ministry of Education, Culture, Sports, Science and Technology, Japan (MEXT). We thank the Photon Factory for provision of synchrotron radiation facilities (No. 2015G533) and acknowledge the staff of the BL-1A beamline.



Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 *DNA in Supramolecular Chemistry and Nanotechnology*, ed. E. Stulz, G. H. Clever, Wiley, Chichester, 2015.
- 2 (a) G. H. Clever, C. Kaul and T. Carell, *Angew. Chem.*, 2007, **119**, 6340–6350 (*Angew. Chem., Int. Ed.*, 2007, **46**, 6226–6236); (b) A. Ono, H. Torigoe, Y. Tanaka and I. Okamoto, *Chem. Soc. Rev.*, 2011, **40**, 5855–5866; (c) Y. Takezawa and M. Shionoya, *Acc. Chem. Res.*, 2012, **45**, 2066–2076; (d) P. Scharf and J. Müller, *ChemPlusChem*, 2013, **78**, 20–34; (e) Y. Tanaka, J. Kondo, V. Sychrovský, J. Šebera, T. Dairaku, H. Saneyoshi, H. Urata, H. Torigoe and A. Ono, *Chem. Commun.*, 2015, **51**, 17343–17360; (f) Y. Takezawa, J. Müller and M. Shionoya, *Chem. Lett.*, 2017, **46**, 622–633.
- 3 (a) A. Ono and H. Togashi, *Angew. Chem.*, 2004, **116**, 4400–4402 (*Angew. Chem., Int. Ed.*, 2004, **43**, 4300–4302); (b) E. M. Nolan and S. J. Lippard, *Chem. Rev.*, 2008, **108**, 3443–3480; (c) D. L. Ma, D. S. Chan, B. Y. Man and C. H. Leung, *Chem. – Asian J.*, 2011, **6**, 986–1003; (d) Y. Song, W. Wei and X. Qu, *Adv. Mater.*, 2011, **23**, 4215–4236.
- 4 (a) T. Carell, C. Behrens and J. Gierlich, *Org. Biomol. Chem.*, 2003, **1**, 2221–2228; (b) T. Ito, G. Nikaïdo and S. Nishimoto, *J. Inorg. Biochem.*, 2007, **101**, 1090–1093; (c) J. Joseph and G. B. Schuster, *Org. Lett.*, 2007, **9**, 1843–1846; (d) L. Guo, N. Yin and G. Chen, *J. Phys. Chem. C*, 2011, **115**, 4837–4842; (e) H. Isobe, N. Yamazaki, A. Asano, T. Fujino, W. Nakanishi and S. Seki, *Chem. Lett.*, 2011, **40**, 318–319; (f) S. Liu, G. H. Clever, Y. Takezawa, M. Kaneko, K. Tanaka, X. Guo and M. Shionoya, *Angew. Chem.*, 2011, **123**, 9048–9052 (*Angew. Chem., Int. Ed.*, 2011, **50**, 8886–8890).
- 5 (a) K. Tanaka, A. Tengeiji, T. Kato, N. Toyama and M. Shionoya, *Science*, 2003, **299**, 1212–1213; (b) S. S. Mallajosyula and S. K. Pati, *Angew. Chem.*, 2009, **121**, 5077–5081 (*Angew. Chem., Int. Ed.*, 2009, **48**, 4977–4981); (c) G. H. Clever, S. J. Reitmeier, T. Carell and O. Schiemann, *Angew. Chem.*, 2010, **122**, 5047–5049 (*Angew. Chem., Int. Ed.*, 2010, **49**, 4927–4929).
- 6 (a) S. Katz, *Nature*, 1962, **195**, 997–998; (b) S. Katz, *Biochim. Biophys. Acta*, 1963, **68**, 240–253; (c) Z. Kuklenyik and L. G. Marzilli, *Inorg. Chem.*, 1996, **35**, 5654–5662; (d) Y. Miyake, H. Togashi, M. Tashiro, H. Yamaguchi, S. Oda, M. Kudo, Y. Tanaka, Y. Kondo, R. Sawa, T. Fujimoto, T. Machinami and A. Ono, *J. Am. Chem. Soc.*, 2006, **128**, 2172–2173; (e) Y. Tanaka, S. Oda, H. Yamaguchi, Y. Kondo, C. Kojima and A. Ono, *J. Am. Chem. Soc.*, 2007, **129**, 244–245; (f) H. Yamaguchi, J. Šebera, J. Kondo, S. Oda, T. Komuro, T. Kawamura, T. Dairaku, Y. Kondo, I. Okamoto, A. Ono, J. V. Burda, C. Kojima, V. Sychrovský and Y. Tanaka, *Nucleic Acids Res.*, 2014, **42**, 4094–4099; (g) J. Kondo, T. Yamada, C. Hirose, I. Okamoto, Y. Tanaka and A. Ono, *Angew. Chem.*, 2014, **126**, 2417–2420 (*Angew. Chem., Int. Ed.*, 2014, **53**, 2385–2388).
- 7 (a) A. Ono, S. Cao, H. Togashi, M. Tashiro, T. Fujimoto, T. Machinami, S. Oda, Y. Miyake, I. Okamoto and Y. Tanaka, *Chem. Commun.*, 2008, 4825–4827; (b) J. Kondo, Y. Tada, T. Dairaku, H. Saneyoshi, I. Okamoto, Y. Tanaka and A. Ono, *Angew. Chem.*, 2015, **127**, 13521–23524 (*Angew. Chem., Int. Ed.*, 2015, **54**, 13323–13326); (c) T. Dairaku, K. Furuita, H. Sato, J. Šebera, K. Nakashima, J. Kondo, D. Yamanaka, Y. Kondo, I. Okamoto, A. Ono, V. Sychrovský, C. Kojima and Y. Tanaka, *Chem. – Eur. J.*, 2016, **22**, 13028–13031; (d) H. Liu, C. Cai, P. Haruehanroengra, Q. Yao, Y. Chen, C. Yang, Q. Luo, B. Wu, J. Li, J. Ma, J. Sheng and J. Gan, *Nucleic Acids Res.*, 2017, **45**, 2910–2918.
- 8 I. Okamoto, T. Ono, R. Sameshima and A. Ono, *Chem. Commun.*, 2012, **48**, 4347–4349.
- 9 For the first time, we have observed the formation of thymine–Ag(I)–thymine pairs in crystals consisting of all metallo-base pairs. (a) J. Kondo, Y. Tada, T. Dairaku, H. Saneyoshi, Y. Tanaka and A. Ono, Crystal Structure of Silver–DNA Hybrid Nanowire, The 43th International Symposium on Nucleic Acids Chemistry (ISNAC2016), 100th Anniversary Hall, Kumamoto University, Sept. 27–29, 2016, pp. 108–109; (b) J. Kondo, Y. Tada, T. Dairaku, Y. Hattori, H. Saneyoshi, A. Ono and Y. Tanaka, *Nat. Chem.*, 2017, DOI: 10.1038/nchem.2808, published online.
- 10 (a) I. Okamoto, K. Iwamoto, Y. Watanabe, Y. Miyake and A. Ono, *Angew. Chem.*, 2009, **121**, 1676–1679 (*Angew. Chem., Int. Ed.*, 2009, **48**, 1648–1651); (b) X. Guo, S. A. Ingale, H. Yang, Y. He and F. Seela, *Org. Biomol. Chem.*, 2017, **15**, 870–883.
- 11 S. Mandal and J. J. Müller, *Curr. Opin. Chem. Biol.*, 2017, **37**, 71–79.
- 12 (a) D. A. Megger, C. F. Guerra, J. Hoffmann, B. Brutschy, F. Matthias Bickelhaupt and J. Müller, *Chem. – Eur. J.*, 2011, **17**, 6533–6544; (b) S. Mandal, A. Hepp and J. Müller, *Dalton Trans.*, 2015, **44**, 3540–3543; (c) S. Litau and J. Müller, *Z. Anorg. Allg. Chem.*, 2015, **641**, 2169–2173; (d) S. Mandal, C. Wang, R. K. Prajapati, J. Kösters, S. Verma, L. Chi and J. J. Müller, *Inorg. Chem.*, 2016, **55**, 7041–7050; (e) S. Mandal, M. Hebenbrock and J. Müller, *Chem. – Eur. J.*, 2017, **23**, 5962–5965.
- 13 S. Mandal, M. Hebenbrock and J. Müller, *Angew. Chem.*, 2016, **128**, 15747–15750 (*Angew. Chem., Int. Ed.*, 2016, **55**, 15520–15523).
- 14 (a) H. Mei, I. Röhl and F. Seela, *J. Org. Chem.*, 2013, **78**, 9457–9463; (b) H. Mei, S. A. Ingale and F. Seela, *Chem. – Eur. J.*, 2014, **20**, 16248–16257; (c) H. Mei, H. Yang, I. Röhl and F. Seela, *ChemPlusChem*, 2014, **79**, 914–918; (d) H. Yang, H. Mei and F. Seela, *Chem. – Eur. J.*, 2015, **21**, 10207–10219; (e) S. K. Jana, X. Guo and H. Mei, F. Seela, *Chem. Commun.*, 2015, **51**, 17301–17304.
- 15 A. Bondi, *J. Phys. Chem.*, 1964, **68**, 441–451.
- 16 H. Schmidbaur and A. Schier, *Angew. Chem.*, 2015, **127**, 756–797 (*Angew. Chem., Int. Ed.*, 2015, **54**, 746–784).
- 17 (a) S. Johannsen, N. Megger, D. Böhme, R. K. O. Sigel and J. Müller, *Nat. Chem.*, 2010, **2**, 229–234; (b) S. Kumbhar, S. Johannsen, R. K. O. Sigel, M. P. Waller and J. Müller, *J. Inorg. Biochem.*, 2013, **127**, 203–210.
- 18 (a) N. Seeman, *Nature*, 2003, **421**, 427–431; (b) A. R. Chandrasekaran and O. Levchenko, *Chem. Mater.*, 2016, **28**, 5569–5581.
- 19 H. Liu, F. Shen, P. Haruehanroengra, Q. Yao, Y. Cheng, Y. Chen, C. Yang, J. Zhang, B. Wu, Q. Luo, R. Cui, J. Li, J. Ma and J. Sheng, *Angew. Chem.*, 2017, **129**, 9558–9562 (*Angew. Chem., Int. Ed.*, 2017, **56**, 9430–9434).

