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**Molecular determinants of FlA1 fluorinase specificity were probed using 5'-chloro-5'-deoxyadenosine (5'-ClDA) analogs as substrates and FlA1 active site mutants. Modifications at F213 or A279 residues are beneficial towards these modified substrates, including 5'-chloro-5'-deoxy-2-ethynyladenosine, ClDEA (>10-fold activity improvement), and conferred novel activity towards substrates not readily accepted by wild-type FlA1.**

Identification and characterization of a growing number of naturally occurring fluorinase enzymes (FlA,<sup>1,2</sup> FlA1,<sup>3</sup> FlA3,<sup>3</sup> NobA<sup>3,4</sup> and FlA4<sup>5,6</sup>) offer an exciting prospect of producing fluorinated compounds under mild conditions.<sup>7</sup> These fluorinases catalyze the conversion of *S*-adenosyl-L-methionine (SAM) and fluoride ion to generate 5'-fluoro-5'-deoxyadenosine (5'-FDA) *via* a nucleophilic substitution (S<sub>N</sub>2) mechanism.<sup>8–10</sup> In order to expand the scope of application, substrate promiscuity was explored on non-native substrates. However, successful examples were only limited to 5'-chloro-5'-deoxyadenosine (5'-ClDA),<sup>11</sup> 2'-deoxyadenosine,<sup>12</sup> L-methionine (L-Met) analogs,<sup>13</sup> methylaza-analog of SAM,<sup>14</sup> and C-2 modified substrates (5'-chloro-5'-deoxy-2-ethynyladenosine, ClDEA,<sup>15,16</sup> and di- and tetra-cyclic peptide constructs of ClDEA<sup>17</sup>). In addition, the catalytic efficiency of wild-type fluorinases on these non-native substrates is often reduced.<sup>13–16</sup> There are only a handful of reports demonstrating limited success at enhancing

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## Probing the molecular determinants of fluorinase specificity†

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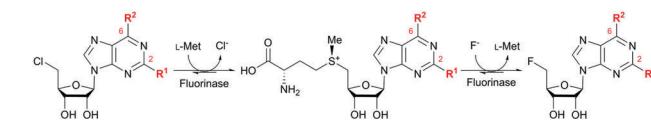
substrate specificity through enzyme modification.<sup>4,6,10,13,18</sup> The challenge here is the lack of understanding of the fluorinase–substrate interactions to enable a better engineered enzyme for fluorination.

Herein, we probed the molecular determinants of FlA1 fluorinase specificity towards non-native substrates which are modified at the adenine ring, as varying this moiety can give rise to a variety of useful fluorinated nucleosides.<sup>15,16,19–24</sup> We have recently demonstrated that the catalytic efficiency of FlA1 fluorinase on a non-native substrate 5'-ClDA can be improved *via* directed evolution.<sup>18</sup> With the improved variants as a starting point, we probed their promiscuity against substrates that are modified at the C-2 (R<sup>1</sup> substitution) and the unexplored C-6 (R<sup>2</sup> substitution) positions of the adenine ring (Table 1).

O'Hagan's group reported that the reaction of ClDEA 2 to 5'-fluoro-5'-deoxy-2-ethynyladenosine (FDEA 2a) using L-selenomethionine progressed at about 60% of the rate of conversion of 5'-ClDA 1 to 5'-FDA 1a which demonstrated that the FlA fluorinase from *Streptomyces cattleya* has a specificity weakness

**Table 1** Substrates (**1–11**) tested for the two-step fluorinase reaction and their corresponding products (**1a–11a**)

Substrates	Products	R <sup>1</sup>	R <sup>2</sup>
<b>1</b>	<b>1a</b>	H	NH <sub>2</sub>
<b>2</b>	<b>2a</b>	C≡CH	NH <sub>2</sub>
<b>3</b>	<b>3a</b>	Cl	NH <sub>2</sub>
<b>4</b>	<b>4a</b>	I	NH <sub>2</sub>
<b>5</b>	<b>5a</b>	NH <sub>2</sub>	NH <sub>2</sub>
<b>6</b>	<b>6a</b>	NH <sub>2</sub>	Cl
<b>7</b>	<b>7a</b>	H	Cl
<b>8</b>	<b>8a</b>	NH <sub>2</sub>	H
<b>9</b>	<b>9a</b>	H	H
<b>10</b>	<b>10a</b>	NH <sub>2</sub>	OMe
<b>11</b>	<b>11a</b>	NH <sub>2</sub>	O



at the C-2 position.<sup>15</sup> Similarly, FlA1 fluorinase from *Streptomyces* sp. MA37 also showed a reduction in activity when 5'-ClIDA **1** was substituted with ClDEA **2** in the L-Met reaction (Table 2A). However, using our previously identified variants of fah2047 (A279L) and fah2081 (A279Y), we found that their activities for CIDEA **2** were either maintained or improved, respectively as compared to 5'-ClIDA **1**. In particular, both variants also demonstrated >9-fold improvement in activity over FlA1 for ClDEA **2**. These results suggest that position 279 could play a role in facilitating binding to the ethynyl group of the substrate at the C-2 position. When R<sup>1</sup> substituents are bulkier groups (**3** and **4**), the % yield dropped further for all the enzyme variants, suggesting that the C-2 position is sensitive to steric bulk. Nonetheless, most of our enzyme variants showed improved activity over FlA1 for substrates **1–4**, except for frh2066 (Y77W) which only showed improved activity for substrate **1**. Thus, mutations F213Y and/or A279LY have led to improved activity against a wider range of R<sup>1</sup> substituted substrates. When the R<sup>1</sup> substituent is an amino group (**5**), the % yield for all the fluorinases increased as compared to that of 5'-ClIDA **1**, with all our enzyme variants still outperforming the fluorination efficiency of FlA1. This is an unprecedented improvement in the fluorination efficiency on 5'-ClIDA analogs. Our results showed that having a hydrogen donor such as an amino

group at the C-2 position of the adenine group enables the modified substrate to interact better with the fluorinase enzyme, thus achieving higher % yield of the corresponding fluorinated product.

With the improvement in activity observed due to the amino group at the C-2 position (substrate **5**), we went on to explore the changes at the C-6 position of the adenine group (R<sup>2</sup> substitution). When R<sup>2</sup> is either a chloro or hydrogen substituent (**6** and **8** respectively), the % yield of all the fluorinases dropped significantly compared to substrate **5** after a 4-h reaction (Table S1, ESI<sup>†</sup>). To enable better comparison between the C-6 modified substrates, the reaction time was lengthened to 24 h (Table 2B). Three of our enzyme variants (fah2019, fah2047 and fah2114) showed improved activity (up to 27-fold) over FlA1 for substrates **6** and **8**. However, no activity was seen for any of the fluorinases for substrates **10** or **11** even with an amino group at the C-2 position. These results suggested that F213Y and A279L mutations are beneficial for selected R<sup>2</sup> substituents such as chlorine or hydrogen. When R<sup>1</sup> was changed from an amino group to a hydrogen atom for substrates **6** and **8** (to give **7** and **9** respectively), there was a decrease in % yield similar to what was observed between **5** and **1**. For substrate **7**, there was no detectable activity for wild-type FlA1 whereas activity could be observed with fah2114 (F213Y, A279L). For substrate **9**, marginal activity was observed for FlA1 and

**Table 2** % product yield and fold improvement (FI) of fluorinases over FlA1 at 37 °C. Reaction conditions: Synthesized substrate (0.2 mM), L-Met (0.1 mM), NaF (80 mM), and fluorinase (50 μM) at a single time point of 4 h or 24 h. % yield (mean of triplicates) = (concentration of product generated/concentration of substrate) × 100%. FI = % yield of evolved variants/% yield of FlA1. A. 4 h reactions using substrates **1–5**. B. 24 h reactions using substrates **6–11**

A

Fluorinase	% yield		FI		% yield		FI		% yield		FI		% yield		FI	
	FlA1	6.6 ± 0.1	—	—	2.3 ± 0 <sup>a</sup>	—	—	3.5 ± 0.1	—	0	—	—	28.8 ± 0.3	—	—	—
fah2019	12.5 ± 0.2	1.9	10.3 ± 0.1	4.5	7.8 ± 0 <sup>a</sup>	2.2	0.2 ± 0 <sup>a</sup>	—	35.4 ± 0.5	1.2	—	—	35.4 ± 0.5	1.2	—	—
fah2047	19.9 ± 0.3	3.0	20.5 ± 0.5	9.0	8.2 ± 0.1	2.3	0.2 ± 0 <sup>a</sup>	—	38.0 ± 0.2	1.3	—	—	38.0 ± 0.2	1.3	—	—
frh2066	13.9 ± 0.2	2.1	0.3 ± 0 <sup>a</sup>	0.1	0.1 ± 0 <sup>a</sup>	0	0	—	31.6 ± 0.4	1.1	—	—	31.6 ± 0.4	1.1	—	—
fah2081	15.3 ± 0.1	2.3	23.9 ± 0.6	10.5	8.5 ± 0.2	2.4	2.2 ± 0 <sup>a</sup>	—	37.7 ± 0.2	1.3	—	—	37.7 ± 0.2	1.3	—	—
fah2114	18.1 ± 0.3	2.7	13.5 ± 0.3	6.0	9.4 ± 0.1	2.7	0.1 ± 0 <sup>a</sup>	—	37.6 ± 0.5	1.3	—	—	37.6 ± 0.5	1.3	—	—

B

Fluorinase	% yield		FI		% yield		FI		% yield		FI		% yield		FI	
	FlA1	0.2 ± 0 <sup>a</sup>	—	—	0	—	15.2 ± 0.3	—	0.1 ± 0 <sup>a</sup>	—	0	—	0	—	0	—
fah2019	1.4 ± 0 <sup>a</sup>	6.6	0	—	33.2 ± 0.3	2.2	0.5 ± 0 <sup>a</sup>	6.5	0	—	0	—	0	—	0	—
fah2047	2.0 ± 0.1	9.8	0	—	37.4 ± 0.7	2.5	0.1 ± 0 <sup>a</sup>	0.9	0	—	0	—	0	—	0	—
frh2066	0	0	0	—	1.2 ± 0.1	0.1	0	0	0	—	0	—	0	—	0	—
fah2081	0.4 ± 0 <sup>a</sup>	2.0	0	—	7.5 ± 0.1	0.5	0	0	0	—	0	—	0	—	0	—
fah2114	5.8 ± 0.1	27.7	0.1 ± 0 <sup>a</sup>	—	30.2 ± 0.4	2.0	0.4 ± 0 <sup>a</sup>	4.7	0	—	0	—	0	—	0	—

<sup>a</sup> Standard deviation less than 0.1.



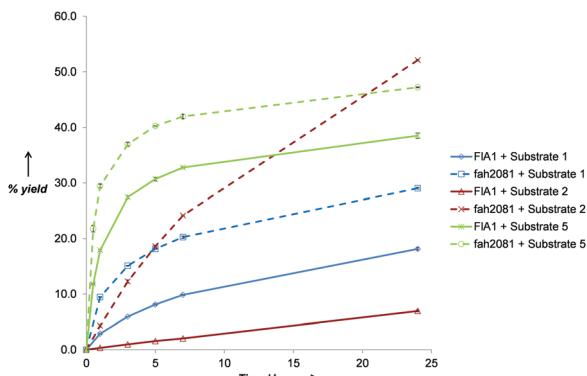


Fig. 1 Comparison of reaction rates for FIA1 and fah2081 with substrates **1**, **2** and **5** at 37 °C over 24 h. Reaction conditions: Synthesized substrate (0.2 mM), L-Met (0.1 mM), NaF (80 mM), and fluorinase (50  $\mu$ M) at different time points. % yield (mean of triplicates) = (concentration of product generated/concentration of substrate)  $\times$  100%, error bars = standard deviation of triplicates.

fah2047 (A279L) whereas improved activity (4–6-fold) was observed for fah2019 (F213Y) and fah2114 (F213Y, A279L). These observations suggest the benefits of a single F213Y mutation for improved activity on substrate **9**.

To elucidate the rates of the best performing substrates, time-course reactions were conducted for FIA1 and fah2081 with substrates **1**, **2** and **5** (Fig. 1). The initial rates for the substrates tested were significantly disparate, with FIA1 showing rates in the order **5**  $\gg$  **1**  $\gg$  **2** and fah2081 showing in the order **5**  $\gg$  **1**  $\approx$  **2**. An interesting anomaly was substrate **2**, which gave the highest % yield with fah2081 at 24 h, whereas the rates of the rest showed trends of considerable slowdown after  $\sim$ 7 h. This suggests that tyrosine at the 279 position has imparted additional benefit towards substrate **2** even though it has a slower initial rate. As for FIA1, substrate **5** still gave the highest % yield after 24 h although the rate was plateaued after  $\sim$ 7 h. The much lower activity seen for R<sup>2</sup> substitution as compared to R<sup>1</sup> substitution suggests that the C-6 position, which is largely unexplored, plays a more important role than the C-2 position in the binding of the substrate to the enzyme. Control experiments had ruled out the possibility that the inactivity observed was due to substrate or product degradation under the reaction conditions. It is also interesting to note that frh2066 (Y77W) showed improved activity over FIA1 for only two of the substrates (5'-ClDA **1** and **5**). Dong *et al.* reported that the Y77 residue is involved in facilitating halide binding for FIA.<sup>9</sup> This implies that the Y77 residue is critical for the fluorination reaction and should not be modified.<sup>6</sup> The Y77 residue is conserved in FIA1, which showed 88% identity to FIA.<sup>3</sup> For our previous work of directed evolution of FIA1 with 5'-ClDA **1** and L-Met substrates, saturation mutagenesis of the Y77 residue showed that Y77F (unpublished) and Y77W mutations displayed improvement over FIA1. This suggests that the Y77 residue may play an additional role in the FIA1 enzymatic reaction, although the model complexes for Y77W do not show any obvious reason for the improved activity towards substrates **1** and **5**.

To understand how the mutants impact the binding of substrates, structural models of the FIA1 mutants were generated

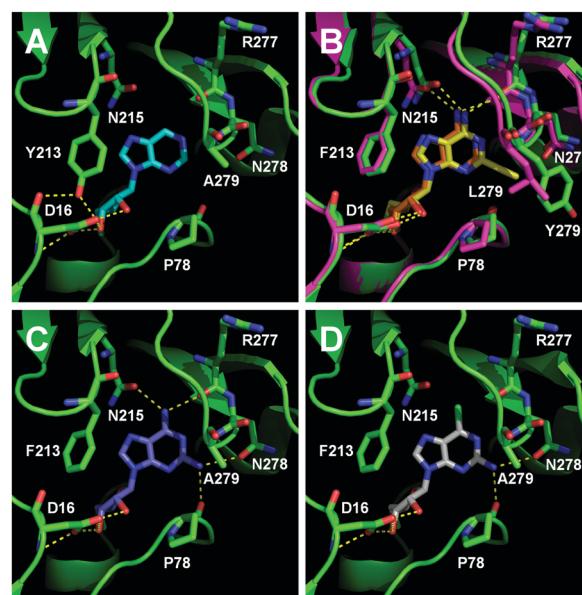


Fig. 2 Novel substrate binding to wild-type FIA1 or FIA1 variants. (A) Substrate **9** (in light blue) bound to fah2019, showing the interaction between Y213 and D16 residues. (B) Substrate **2** (in orange and yellow) bound to fah2047 (in magenta) and fah2081 (in green), showing the interaction of the R<sup>1</sup> substituent of the ethynyl group with L279 and Y279 residues respectively. (C) Substrate **5** (in blue) bound to wild-type FIA1, with hydrogen bonds shown between the substrate and enzyme. (D) Substrate **6** (in white) bound to FIA1 showing the loss of hydrogen bond interaction with the removal of the R<sup>2</sup> substituent of the amino group.

using ROSETTA,<sup>25</sup> with the recently solved crystal structure of FIA1 (PDB code 5B6I)<sup>18</sup> as the template. Substrate coordinates were derived from the crystal structure of the *Streptomyces cattleya* fluorinase trimer complexed with substrate **2** (PDB code 4CQJ)<sup>15</sup> and docked into the FIA1 mutant active site using PyMOL.<sup>26</sup> All models were then subjected to energy minimisation using AMBER.<sup>27</sup> The models identify key interactions that may have led to improved binding affinity translating to increased activity. The F213Y mutation is located in the binding pocket at a dimer interface, and is probably hydrogen bonded to D16 in the opposing monomer. This is likely to increase the stability of the enzyme, and may improve activity by increasing affinity towards the substrate. This is particularly noticeable with substrate **9** (Fig. 2A). A279Y also bridges the two monomers through a hydrogen bond between the tyrosine side-chain and the peptide carbonyl of W50 on an adjacent chain. These interactions translate into an overall improvement in activity for these mutations in their variants for most of the substrates.

The improved activity observed for our enzyme variants for R<sup>1</sup> substitutions on the adenine ring (2–5) is favourable for several reasons. Firstly, this C-2 position points into the bulk solvent, and therefore should have minimal effect on the binding to the FIA1 enzyme, similar to what was observed in the FIA enzyme.<sup>15</sup> Secondly, the A279L mutation, where the leucine side-chain directly interacts with the substituent, is likely to form favourable van der Waals contacts with the substituent atoms (Fig. 2B). As for the A279Y mutation, the tyrosine side-chain brings the enzyme in closer contact to the ethynyl substituent of substrate **2**,

hence forming favourable van der Waals interactions. Thirdly, the amino group substituent is able to form favourable hydrogen bonds with the enzyme through the peptide carbonyl of P78 and the amide side-chain of N278 (Fig. 2C). The iodinated substrate **4** has much lowered activity for all mutants, and this might be because the P78 carbonyl and the N278 side-chain are unlikely to form any polar interactions or hydrogen bonds with the bulky iodine substituent.

The FlA1-5'-ClDA enzyme complex model when compared with the bound models for **6–11** identifies the main reason for diminished or loss of activity. The R<sup>2</sup> substituent of the amino group is hydrogen-bonded to the enzyme through the amide side-chain of N215, and the peptide carbonyl of R277 (Fig. 2C). The loss of this group removes this interaction, which would lower the affinity of the enzyme for the substrate, and substituting with chlorine (**6** and **7**), hydrogen (**8** and **9**) or oxygen (**11**) will not form any stabilizing hydrogen bonds with N215 (Fig. 2D and Fig. S1A and B, ESI<sup>†</sup>). In the case of **10**, the substituted methoxy group could be too bulky to fit into this pocket, and thus cannot form any stabilizing hydrogen bonds (Fig. S1C, ESI<sup>†</sup>). We tried to mutate N215 in order to explore potential alterations in specificity at the C-6 position, but so far the mutations we tested (N215LQ) have rendered the protein insoluble (data not shown). The addition of an amino group at the C-2 position helps in compensating for the loss of key interactions at the C-6 position and improving the binding of some substrates (e.g. **6** and **8**) due to the additional hydrogen bonding (*vide supra*), as well as the favourable van der Waals interactions from A279L and Y.

In conclusion, we found that substitutions at the C-2 position of the adenine group are sensitive to steric bulk whereas an amino group at the C-2 position led to enhanced fluorination yield. Additionally, our evolved FlA1 fluorinase variants are active against substrates that are modified at the C-2 and/or C-6 positions of the adenine ring despite being evolved against 5'-ClDA **1** and L-Met substrates. Modifications at F213 and/or A279 residues showed improved activity over FlA1 on the eight new substrates tested, including novel activity for two of the substrates, **4** and **7**. Based on our previous kinetic studies,<sup>18</sup> it is likely that the improvement in activity shown by our enzyme variants over FlA1 for these substrates is due to the enhancement of the first step, *i.e.* conversion of the substrate and L-Met to a SAM-analog in this two-step fluorination process. This work sheds light on the specificity of the FlA1 fluorinase with respect to modifications at the adenine moiety of the 5'-ClDA substrate and the mutations that can improve the promiscuity of the enzyme against non-native substrates. We believe that our work can serve as a starting point for engineering of enzymes to perform fluorination on more structurally diverse small molecules.

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