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## Silver-induced reconstruction of an adeninate-based metal–organic framework for encapsulation of luminescent adenine-stabilized silver clusters†

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Bright luminescent silver-adenine species were successfully stabilized in the pores of the MOF-69A (zinc biphenyldicarboxylate) metal–organic framework, starting from the intrinsically blue luminescent bio-MOF-1 (zinc adeninate 4,4'-biphenyldicarboxylate). Bio-MOF-1 is transformed to the MOF-69A framework by selectively leaching structural adenine linkers from the original framework using silver nitrate solutions in aqueous ethanol. Simultaneously, bright blue-green luminescent silver-adenine clusters are formed inside the pores of the recrystallized MOF-69A matrix in high local concentrations. The structural transition and concurrent changes in optical properties were characterized using a range of structural, physicochemical and spectroscopic techniques (steady-state and time-resolved luminescence, quantum yield determination, fluorescence microscopy). The presented results open new avenues for exploring the use of MOFs containing luminescent silver clusters for solid-state lighting and sensor applications.

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### Introduction

The electronic and optical properties of silver are heavily influenced by the number of silver atoms in the aggregate, ranging from single ions, over clusters of a few atoms and larger nanoparticles, to the bulk material.<sup>1</sup> The typical band structure of bulk silver with freely moving electrons results in good electrical conduction and ensures reflective optical properties. Reducing the physical size of silver to nanoparticles considerably alters its catalytic, chemical, electrical, optical and magnetic properties, which are all size and shape dependent.<sup>2</sup> For instance, the collective light-responsive oscillation of the conduction band electrons, known as surface plasmon resonance (SPR), endows silver nanoparticles with bright colours.<sup>3</sup> Sub-nanometre sized silver clusters exhibit an electronic structure

fragmented into discrete energy levels. Clusters of a few to up to ten silver atoms have a size in the range of the Fermi wavelength of electrons, which is approximately 0.5 nm for silver. These discrete energy levels allow molecular-like electronic transitions stimulated by light, generally in the UV-vis region with subsequent luminescence.<sup>1</sup> Because of these discrete energy levels and luminescence, small metal clusters are of considerable importance as their electronic behaviour and properties bridge ionic and nanoparticle/metallic behaviour.<sup>1,3–6</sup> Their luminescent properties enable interesting applications, *e.g.* as fluorescent markers for biological imaging<sup>2</sup> or secondary light sources in fluorescent lamps.<sup>7</sup> Beside their luminescent properties, these silver nanoclusters also feature unique catalytic properties.<sup>8,9</sup> The catalytic properties of such low-atomicity metal clusters are enhanced compared to metal nanoparticles because more atoms are exposed and available for adsorption and catalysis.<sup>10</sup>

The inherent tendency of small metal clusters to aggregate to non-luminescent metallic silver nanostructures and nanoparticles has led to the development of strategies to stabilize them. The two most popular strategies are (i) to exploit the large affinity of silver for nitrogen-containing DNA bases in short single stranded oligonucleotides (ssDNA) and to subsequently reduce the silver ions with NaBH<sub>4</sub>,<sup>11–17</sup> and (ii) the use of microporous host materials allowing confinement of the clusters, *e.g.* in the sodalite cages of zeolites. In the latter case, the reduction is performed either thermally or by UV or X-ray irradiation.<sup>7,18–25</sup> Other, less frequently used stabilizing materials employed for

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luminescent silver cluster entrapment are cryogenic noble gas matrices,<sup>26,27</sup> inorganic glasses,<sup>28,29</sup> silver oxide films<sup>13</sup> and dendrimers.<sup>30</sup>

Another class of porous materials called metal-organic frameworks (MOFs) has emerged recently as suitable hosts for the stabilization of luminescent silver clusters.<sup>31</sup> The confinement approach in microporous materials (MOFs and zeolites) limits the cluster size to the pore/cage size of the material, *i.e.* one nanometer or less. MOFs consist of metal ion nodes and multitopic organic linkers which assemble into porous, three-dimensional architectures. Because of their high surface area and highly functionalisable porous structure,<sup>32–34</sup> research on MOFs has strongly expanded<sup>35</sup> with the discovery of many new structures and more recently the search for innovative applications.<sup>32,34,36</sup> These comprise gas storage<sup>37</sup> and separations,<sup>38</sup> liquid phase separations of hydrocarbons and key molecules<sup>39–41</sup> for (bio-)chemical industry, catalysis,<sup>42–46</sup> inherent framework luminescence for sensing<sup>47–49</sup> and drug delivery,<sup>50</sup> among others.

MOFs are also known to stabilize luminescent species, like CdSe quantum dots.<sup>51</sup> However, so far, the luminescence of oligo-atomic silver clusters confined in metal-organic frameworks has not yet been studied. In this paper, the large affinity of ionic silver for adenine and the rigid confining scaffold of a MOF framework are exploited to stabilize luminescent silver-adenine clusters in MOF pores. Our approach differs from previous studies in which only non-luminescent metal nanoparticles<sup>31,52–56</sup> and metallic microstructures<sup>57</sup> have been reported.

## Results and discussion

### Exploration of water-stable MOFs and selecting nucleobase MOFs

Aqueous conditions are preferred for loading silver ions onto porous materials like MOFs, since water is an excellent solvent for most ionic silver sources like silver nitrate. Among the large number of MOF structures, only few are truly water stable. The best known water-stable MOFs are zirconium-based (*e.g.* UiO-66, MIL-140C and MOF-808) with strong hydrolysis-resistant Zr-carboxylate bonds. Other water-stable MOFs are aluminium-based (*e.g.* MIL-53, MIL-68 and MIL-96) and the zinc-imidazolate, zeolitic ZIF-8 framework.

Typical reducing treatments for generating silver clusters are chemical reduction with NaBH<sub>4</sub> or H<sub>2</sub>, thermal activation or reduction by electromagnetic radiation (gamma rays, X-rays and UV light). Thermal reduction as performed in zeolites is not suitable for MOFs, since in general the thermal stability of MOFs is limited to 300–400 °C, with only few exceptions (*e.g.* UiO-66, MIL-140). Therefore, we attempted to create luminescent silver clusters by introducing silver ions from mixtures of water and ethanol to generate a mildly reducing environment. However, during the preliminary screening, these conditions did not result in the formation of luminescent silver clusters on any of the tested materials. Even thermal treatments at 300–450 °C, as for zeolites,<sup>19</sup> of silver-loaded and thermally stable materials

(*e.g.* UiO-66 or MIL-140C) yielded no luminescent silver clusters. Therefore, our attention turned towards another group of potentially water stable MOFs, namely the bio-MOFs. These materials contain the DNA nucleobase adenine as one of the linkers in the framework, in combination with dicarboxylates<sup>50,58–64</sup> or tricarboxylates,<sup>65,66</sup> and various metal ions (Zn<sup>2+</sup>, Co<sup>2+</sup> and Cu<sup>2+</sup>). Of the wide range of adenine-containing MOFs,<sup>67–72</sup> only bio-MOF-14 and bio-MOF-1 have confirmed water stability.<sup>50,59</sup>

Exploratory experiments with the nucleobase-containing framework bio-MOF-1 resulted in immediate formation of a luminescent silver phase when the material was loaded with silver using AgNO<sub>3</sub> solutions with intermediate ethanol-water ratios. This result sharply contrasts to all other water-stable MOFs and the related ZJU-48 (a Zn adeninate 4,4'-*trans*-stilbenedicarboxylate), tested under the same circumstances. A detailed description of the structure of the two tested adeninate MOF structures, bio-MOF-1<sup>50</sup> and ZJU-48,<sup>61</sup> is given in Fig. S1 (ESI†). Basically, in these structures adenine occupies the major part of the coordination sphere of Zn<sup>2+</sup>, while the carboxylates are used to space the zinc-adeninate building units. This results in adenine moieties at well-defined positions at the corners of large square, one-dimensional pores with a diameter of about 1 nm. The pore walls consist of the organic carboxylate linkers. An important property of bio-MOF-1 is cation exchange capacity.<sup>50,61</sup> Typically, *N,N*-dimethyl ammonium (DMA<sup>+</sup>), a by-product of the DMF synthesis resides in the pores of bio-MOF-1 for charge compensation. This DMA<sup>+</sup> can be exchanged for many other organic<sup>50,62,73,74</sup> or inorganic<sup>75–77</sup> cations. In principle, this ion exchange should also be possible with silver ions. Stabilization of formed cationic or metallic luminescent silver clusters in this case would therefore be enabled by two possible mechanisms: (i) interaction of silver, either in its ionic or in partially reduced state with adenine, or (ii) ionic interactions stabilize these cationic silver clusters in the anionic framework, as in the case of silver-containing zeolites.

### Structural transformation of bio-MOF-1 to MOF-69A

Even though bio-MOF-1 is stable in pure water, its structure fully degrades when exposed to aqueous solutions of silver nitrate (40 mM) at a 1–1 ratio of silver to adenine. Bio-MOF-1 is transformed into another crystalline material, as evidenced by the X-ray diffraction pattern in Fig. 1a. In contrast, ZJU-48 undergoes a transformation to a nonporous, poorly crystalline silver adeninate phase upon contact with silver ions as shown in Fig. S2 (ESI†). This instability of both adeninate MOFs is most likely due to the strong interactions between silver ions and the nitrogen atoms of the adeninate linkers. PXRD patterns of bio-MOF-1 samples with silver-adenine ratios lower than 1 contain the original reflections of bio-MOF-1 but also new emerging reflections, originating from the transformed phase, as shown in Fig. S3 (ESI†). These strong interactions can be understood in the light of the Pearson hard/soft acid/base (HSAB) theory,<sup>78</sup> which states that a soft Lewis acid (*e.g.* Ag<sup>+</sup>) will preferably interact with bases of an intermediately soft nature (*e.g.* adeninate) rather than with hard bases (*e.g.* carboxylates). The new crystalline phase that remains after the transformation



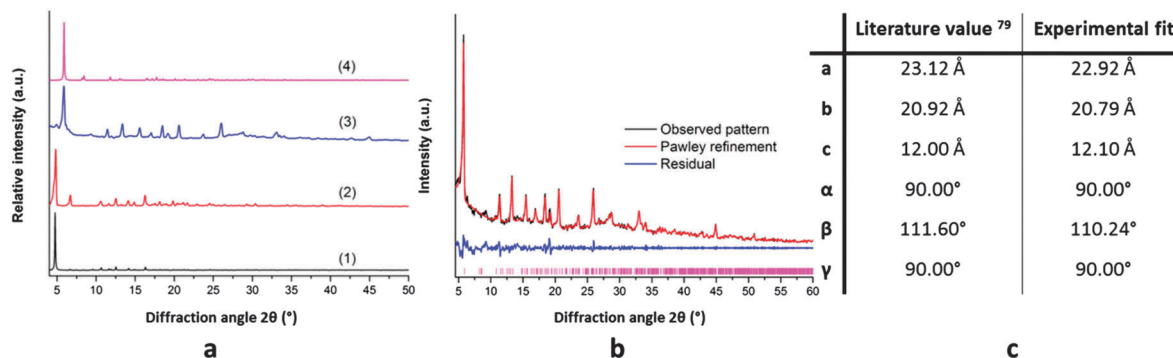


Fig. 1 (a) Powder X-ray diffractograms: (1) theoretical pattern of bio-MOF-1, (2) as synthesized bio-MOF-1, (3) bio-MOF-1 after Ag-induced structural transformation, and (4) theoretical MOF-69A pattern; (b) Pawley fit of the MOF-69A material obtained by Ag-induced transformation of bio-MOF-1; (c) calculated cell parameters and literature values for MOF-69A.

of bio-MOF-1 could either be a crystalline framework built from silver and adenine, or from the two other building blocks of bio-MOF-1, *viz.* Zn<sup>2+</sup> and 4,4'-BPDA.

It seems unlikely that the newly formed crystalline phase is a silver-adeninate, since the gel formed from an equimolar aqueous mixture of silver nitrate and adenine has PXRD reflections above 10° 2θ after drying. The resulting pattern is very different from the diffractogram obtained for the silver-transformed bio-MOF-1. Therefore, more plausible candidate structures for the new crystalline phase are zinc carboxylates like IRMOF-10, the 4,4'-BPDA isostructural form of MOF-5, or MOF-69A,<sup>79</sup> the 4,4'-BPDA isostructural form of MOF-69C. The latter material contains rods of tetrahedral and octahedral Zn<sup>2+</sup> ions interconnected with a dicarboxylic acid to form unidimensional diamond-shaped channels. Since it is known that MOF-5 converts to MOF-69C in excess of water,<sup>80,81</sup> MOF-69A seems to be the most likely structure of these Zn<sup>2+</sup>-carboxylate frameworks. To investigate this, a Pawley refinement of the experimental diffraction pattern of the transformed phase was undertaken in the monoclinic space group, with cell parameters corresponding to those of MOF-69A.<sup>79</sup> Overall, a good fit to the data is obtained ( $R_{wp} = 5.83\%$ ;  $GoF = 3.67$ ), as shown in Fig. 1b. The deviation of non-matching intensities compared to literature values can be explained by the presence of residual silver-adenine guests in the silver-transformed bio-MOF-1 material, which distorts the MOF-69A host framework. For the sake of comparison, attempts were made to directly prepare MOF-69A and load this material with such silver clusters. However no phase-pure MOF-69A powder could be synthesized when following literature procedures.<sup>79,82,83</sup>

This silver-induced MOF-to-MOF transformation was also performed using different ethanol to water ratios. These solvent ratios had only a minor influence on the transformation or on the crystallinity of the formed MOF-69A product, as shown by the XRD patterns (Fig. S4, ESI<sup>†</sup>). Nevertheless, as will be demonstrated below, an appropriate solvent ratio is essential to generate luminescent silver clusters.

For the silver-transformed bio-MOF-1, no reflections of Ag<sup>0</sup> nanoparticles were observed in PXRD at the typical diffraction angles of 38.1° and 44.3° 2θ.<sup>19</sup> This was further confirmed by

Auger spectra of silver obtained by XPS (Fig. S5, ESI<sup>†</sup>) which do not show any evidence of the typical peaks associated with Ag<sup>0</sup> nanoparticles or metal. Furthermore the so-called modified Auger parameters<sup>84</sup> calculated from the XPS data are characteristic of small silver clusters.<sup>85,86</sup> Photoelectron spectroscopy in air (PESA) showed ionisation energies of the silver-transformed MOFs of  $5.62 \pm 0.05$  eV and  $5.52 \pm 0.11$  eV, with these values probably coming from the MOF framework itself ( $5.30 \pm 0.04$  eV before transformation) or possibly the silver clusters, but certainly not from metallic silver whose work function is about 4.3 eV.<sup>87</sup>

While Zn<sup>2+</sup> and 4,4'-BPDA both end up in the MOF-69A framework, the fate of the adenine needs additional clarification. As mentioned above, mixing equimolar solutions of adenine and silver nitrate in aqueous conditions results in precipitation of a white gel, which indicates that adenine complexes with silver nitrate are poorly soluble. Therefore, it is unlikely that silver extracts adenine to the aqueous solution during the silver-induced transformation of bio-MOF-1. Rather all adenine is expected to remain occluded in the pores during the MOF-to-MOF transformation. *Via* <sup>1</sup>H-NMR analysis of (transformed) bio-MOF-1 samples digested with HF (Fig. 2), it was confirmed that there was no significant adenine loss from the solid material during the silver-induced transformation of bio-MOF-1. The spectrum obtained after digestion of as-synthesized bio-MOF-1 (Fig. 2a) shows the two doublets of 4,4'-BPDA (8.05, 7.85 ppm; 8 H), while the signals of the two ring protons of adenine coincide at 8.32 ppm. As expected, based on the structural formula of bio-MOF-1 ( $[Zn_8(ad)_4(BPDC)_6O_2Me_2NH_2^+]$ ), the ratio of adenine to 4,4'-BPDA is 2 to 3. After silver-induced transformation to MOF-69A (Fig. 2b) this ratio remains identical, proving that no adenine has been leached to the aqueous solution containing silver. However, the adeninate signals have shifted significantly downfield and are split (8.36 and 8.38 ppm), possibly pointing to a stable complex of adenine with silver, even after digestion with HF.

When starting from the analogous 4,4'-*trans*-SBDA containing ZJU-48, there is no crystal structure transformation. Instead, a poorly crystalline product is observed, as shown in Fig. S2 (ESI<sup>†</sup>). This contrast with the case of bio-MOF-1 can be explained by the fact that there are no IRMOF or MOF-69 structures known in



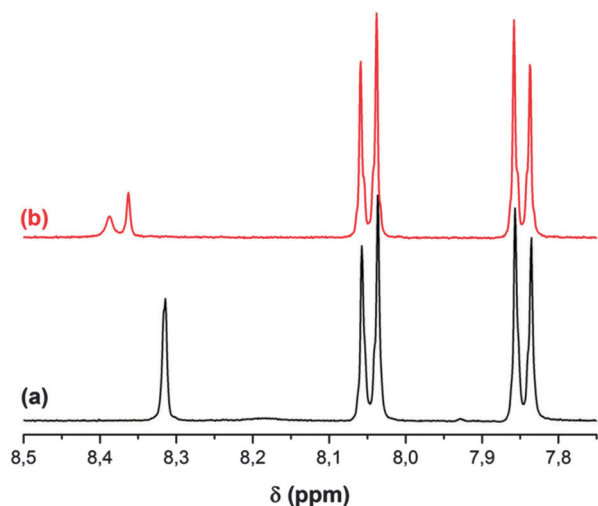


Fig. 2  $^1\text{H-NMR}$  spectra (aromatic region) for (a) as-synthesized bio-MOF-1 and (b) the same material after silver-induced transformation in 4–1 EtOH–H<sub>2</sub>O.

literature with the 4,4'-*trans*-SBDA linker, since this dicarboxylate linker has a slightly different geometry compared to uniaxial linkers like 4,4'-BPDA. Hence, recrystallization to a MOF-69 type material with 4,4'-*trans*-SBDA seems less plausible from crystallographic point of view.

### Physicochemical characterisation of silver-transformed bio-MOF-1

The metal content of the silver-transformed bio-MOF-1 materials was determined using both ICP for the bulk composition and XPS for the surface composition of selected samples. Table 1 shows bulk and surface metal contents for pristine bio-MOF-1 and silver-transformed MOF-69A samples from various EtOH–H<sub>2</sub>O ratios. Comparing the bulk metal and surface metal content

demonstrates that silver is slightly enriched in the outer layers of the crystals. The drastically lowered BET surface areas after silver-induced transformation, also indicated in Table 1, are likely due to filling of the pores of the newly crystallized MOF-69A with poorly soluble silver-adenine species, which strongly reduce available pore volume.

FT-IR measurements were performed to monitor the interactions between silver and adenine after the structural MOF-to-MOF transformation. These spectra reveal that adenine in bio-MOF-1 has a free amine function, as demonstrated by the distinct N–H stretching vibration signals (3330 and 3185 cm<sup>-1</sup>); therefore, the zinc must be bound to the nitrogen atoms in the heterocycle, as also shown in Fig. S1a (ESI<sup>†</sup>). The N–H stretching signals are also retained after the silver-induced bio-MOF-1 transformation, giving a qualitative indication that adenine is still present and is not bound to silver through its free –NH<sub>2</sub> group (Fig. 3). Moreover, an additional, sharp O–H stretching vibration band of the hydroxyl groups on Zn<sup>2+</sup> in the formed MOF-69A<sup>79</sup> is visible at 3544 cm<sup>-1</sup> (Fig. 3). On the other hand, after the silver-induced transformation of ZJU-48, the FT-IR signals are more similar to those of free adenine, independently of the used solvent ratios for silver loading (Fig. S6, ESI<sup>†</sup>). In contrast to the transformation of bio-MOF-1, there is no sharp O–H stretch for the silver-transformed ZJU-48. It is therefore likely that the well-defined O–H stretching vibration at 3544 cm<sup>-1</sup> in the silver-transformed bio-MOF-1 is due to the vibrations of the structural (Zn)–OH groups in MOF-69A, rather than to amorphous zinc hydroxide. Differential thermogravimetric analysis (TGA) (Fig. S7, ESI<sup>†</sup>) shows that the two organic linkers in the as-synthesized bio-MOF-1 degrade separately with maxima around 400 °C and 500 °C. In the following analysis, we only consider weight losses above 200 °C, at which temperature solvent molecules like DMF and water have been removed from the structure.

Based on the structural formula of bio-MOF-1, [Zn<sub>8</sub>(ad)<sub>4</sub>-O(BPDA)<sub>6</sub>·(DMA<sup>+</sup>)<sub>2</sub>], the weight losses in TGA can be assigned to

Table 1 Metal content (bulk and surface), BET surface areas and quantum yields (QY) of bio-MOF-1 powders before and after silver-induced transformation to MOF-69A using various EtOH–H<sub>2</sub>O mixtures

|                                 | Ag/Zn (ICP)<br>(wt%/wt%) | Ag/Zn (XPS)<br>(wt%/wt%) | BET surface area<br>(m <sup>2</sup> g <sup>-1</sup> ) | QY (%)<br>λ <sub>exc</sub> = 330 nm | QY (%)<br>λ <sub>exc</sub> = 366 nm |
|---------------------------------|--------------------------|--------------------------|---|-------------------------------------|-------------------------------------|
| Bio-MOF-1                       | 0/14.8                   | 0/25.6                   | 815.4   | 0                                   | 8.3                                 |
| Adenine                         |                          |                          |   | 0                                   | 2.0                                 |
| 4,4'-BPDA                       |                          |                          |   | 20.6                                | 21.4                                |
| After Ag-induced transformation |                          |                          |   |                                     |                                     |
| From H <sub>2</sub> O           | 9.6/11.9                 |                          | 37.2  | 4.1                                 | 7.2                                 |
| From 1–7 EtOH–H <sub>2</sub> O  | 9.2/11.9                 |                          | 98.1  | 3.0                                 | 12.3                                |
| From 1–4 EtOH–H <sub>2</sub> O  | 9.4/11.9                 |                          | 87.1  | 2.8                                 | 9.2                                 |
| From 1–2 EtOH–H <sub>2</sub> O  | 8.9/12.0                 |                          | 88.1  | 0                                   | 10.3                                |
| From 1–1 EtOH–H <sub>2</sub> O  | 8.8/11.9                 |                          | 101.6   | 0                                   | 18.8                                |
| From 2–1 EtOH–H <sub>2</sub> O  | 9.2/11.7                 |                          | 62.9  | 0                                   | 17.2                                |
| From 4–1 EtOH–H <sub>2</sub> O  | 12.2/11.3                |                          | 45.7  | <1                                  | 15.0                                |
| From 5–1 EtOH–H <sub>2</sub> O  | 8.2/13.9                 | 19.2/14.5 <sup>a</sup>   | 23.5  | 2.1                                 | 14.5                                |
| From 7–1 EtOH–H <sub>2</sub> O  | 9.9/11.6                 |                          | 33.0  | 4.8                                 | 18.0                                |
| From 9–1 EtOH–H <sub>2</sub> O  | 9.9/14.0                 | 22.2/11.3 <sup>a</sup>   | 124.0   | 11.1                                | 24.7                                |
| From EtOH                       | 12.4/11.8                |                          | 171.5   | 20.1                                | 33.0                                |

<sup>a</sup> These weight percentages do not include hydrogen in the calculations, as hydrogen cannot be detected by XPS. This will cause the presented values to be slightly lower, but retaining the observation of increased silver contents at the surface.



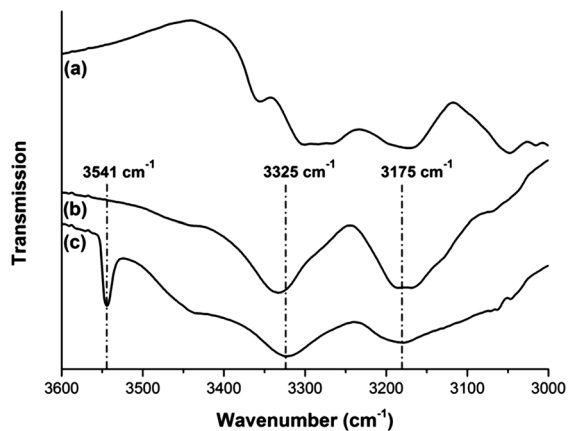


Fig. 3 FT-IR spectra of (a) adenine, (b) bio-MOF-1 and (c) MOF-69A obtained by silver-induced transformation of bio-MOF-1. The bands at 3175 and 3325  $\text{cm}^{-1}$  in (c) prove that the adenine remains present in the transformed material; the O–H stretching vibration at 3544  $\text{cm}^{-1}$  originates from the structural hydroxyl groups (Zn(O–H)) in MOF-69A.

4,4'-BPDA (400 °C; 53.4 wt% in TGA) and to adenine (500 °C; 17.7 wt% in TGA); the total loss of linker (71.1 wt%) corresponds well with the theoretically expected weight loss upon conversion of the desolvated MOF to ZnO (71.7 wt%). Note that in contrast to the free ligands, adenine in bio-MOF-1 degrades at a higher temperature than 4,4'-BPDA. The residue of 25.2 wt% of ZnO is in agreement with the theoretical  $\text{Zn}^{2+}$  content of the framework (theoretical: 24.9 wt%). After the silver-induced transformation, the weight loss of the sample is reduced to a single step, at a temperature corresponding to 4,4'-BPDA loss in the original material. It is likely that both structural 4,4'-BPDA and intraporous residual adenine are lost simultaneously in this single step. This may indicate that the adenine is less strongly encapsulated in MOF-69A than in the original bio-MOF-1 framework; alternatively, the contained silver may act as an oxidation catalyst for combustion of both linkers during analysis. Obviously, the inorganic residue after silver-loading is increased, in line with the ICP data of Table 1. Taking into account the ZnO content of the inorganic residue for the respective samples, the organic linker content with respect to ZnO can again be determined after Ag-induced transformation. Analysis of the data in Fig. S7 (ESI<sup>†</sup>) shows that there is no perceptible loss of organic linkers caused by the exposure to silver, which confirms the <sup>1</sup>H-NMR data of Fig. 2. Remarkably, the small weight loss of exchanged  $\text{DMA}^+$ , observed at 240 °C in the starting bio-MOF-1, is absent in the silver-loaded materials. This shows that  $\text{DMA}^+$  can initially have been exchanged for  $\text{Ag}^+$ . Also note that after the structural transition, the MOF-69A no longer possesses such cation exchange capacity.

In the adopted synthesis conditions, bio-MOF-1 crystallizes as 20–100  $\mu\text{m}$  sized tetragonal prisms capped with tetragonal pyramids on both sides. These crystals are often large single crystals (Fig. S8, ESI<sup>†</sup>). Scanning electron microscopy (SEM) studies show that the morphology is largely retained after the silver-induced MOF-to-MOF transformation, but large fractures are visible perpendicular to the *c* axis of the crystals. This can be

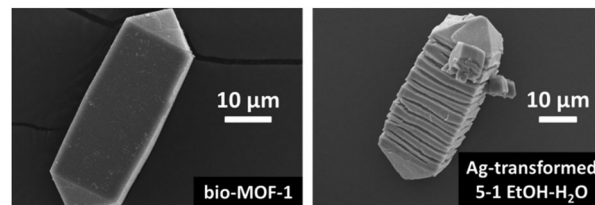


Fig. 4 SEM pictures of pristine bio-MOF-1 and silver-transformed material from 5–1 EtOH–H<sub>2</sub>O.

the result of the severe stress that arises due to silver-induced adenine extraction from the original bio-MOF-1 (Fig. 4). Again, the influence of the used solvent ratios was very drastic: the crystal habitus is generally best preserved in intermediate EtOH–water mixtures (Fig. S8, ESI<sup>†</sup>), mostly in the range of 4–1 and 5–1 EtOH–H<sub>2</sub>O.

### Optical characterisation

For metal cations with closed shell electron configurations, like alkali(ne earth) and  $d^0$  or  $d^{10}$  transition metals (e.g.  $\text{Zn}^{2+}$  in bio-MOF-1), the linker-centred luminescence is often hardly altered after incorporation of luminescent linkers in crystalline MOF structures.<sup>47–49</sup> Since two different organic linkers are present in the structure of bio-MOF-1, they can both contribute to the absorption and emission properties. This is indeed the case: the UV-vis absorption spectrum of the parent bio-MOF-1 contains the spectral signatures of both linkers. The DRS spectrum (Fig. S10, ESI<sup>†</sup>) is dominated by the UV absorption of adenine between 250 and 300 nm and further shows a shoulder between 300 and 330 nm from 4,4'-BPDA. Based on the spectral similarity, the emission profile of bio-MOF-1 can largely be attributed to adenine luminescence (Fig. S11, ESI<sup>†</sup>). The strong 4,4'-BPDA luminescence observed in the solid state (see Table 1) possibly becomes quenched when 4,4'-BPDA is integrated into the bio-MOF-1 structure, as quenching and emission shifts for similar dicarboxylates are known from literature.<sup>88,89</sup> However, after the structural transformation to MOF-69A the main luminescence in many cases still stems from 4,4'-BPDA (ESI,† Fig. S11 and S12).

After the transformation of bio-MOF-1 to MOF-69A the optical properties change drastically. The composition (EtOH–H<sub>2</sub>O ratio) of the silver nitrate solution has an important impact on the reduced Ag species formed (Fig. S12, ESI<sup>†</sup>). At either low or high EtOH–H<sub>2</sub>O ratios, only the luminescence of 4,4'-BPDA in the formed MOF-69A is discerned, indicating the transformation of bio-MOF-1 without formation of luminescent silver clusters. Only at intermediate EtOH–H<sub>2</sub>O ratios (1–1 to 4–1), a new, very strong blue-green emission emerges ( $\lambda_{\text{em}}$  455–530 nm,  $\lambda_{\text{max}}$  at 485 nm) using excitation in the UV between 340 and 390 nm (Fig. 5). This emission cannot be attributed to the original linkers (Fig. S11 and S12, ESI<sup>†</sup>), nor to the zinc oxide nodes or possibly formed ZnO nanoparticles.<sup>90</sup> The latter typically have a maximum, usually narrow emission at 530 nm. The spectral properties, including the larger Stokes shift, show a large similarity to those observed for luminescent



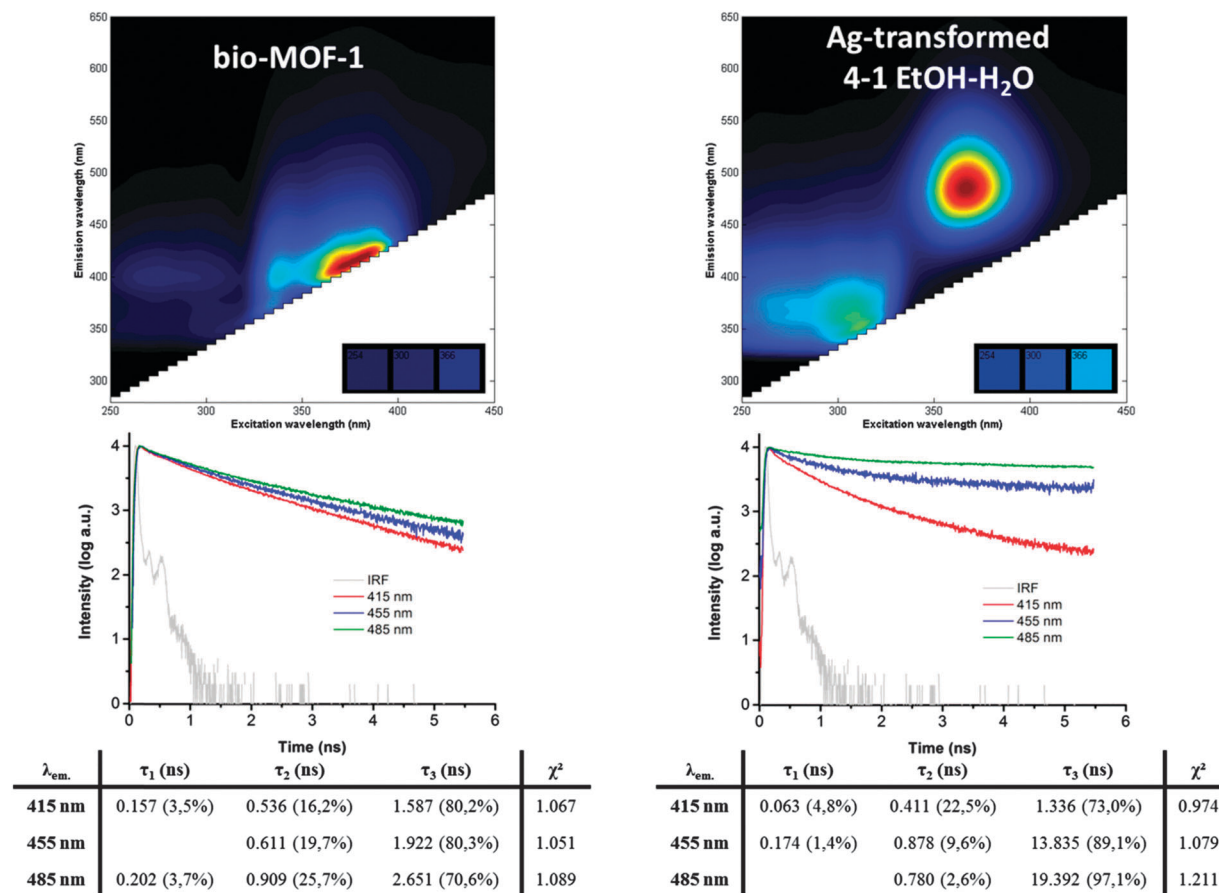


Fig. 5 Spectroscopic study of bio-MOF-1 (left) and of the same material after Ag-induced transformation to MOF-69A (right): steady-state (top) and time-resolved (bottom) luminescence characterization at  $\lambda_{exc} = 375$  nm show the appearance of a strong and long-lived luminescent feature upon silver loading.

silver clusters stabilized by ssDNA<sup>14,17</sup> but even more similarities compared to luminescent Ag-zeolites, as previously reported by our group.<sup>24</sup> See also Fig. S13 (ESI<sup>†</sup>) for a comparison of the Ag-transformed MOF from 4-1 EtOH-H<sub>2</sub>O with Ag-zeolites. Clearly, during the transformation to MOF-69A, these blue-green luminescent silver-cluster species can only be formed under well-defined, moderately reducing conditions. The formed silver clusters are successfully stabilized by the adenine released from the original bio-MOF-1 structure during the MOF-to-MOF transformation. In contrast, the absorption-emission spectra of ssDNA-stabilized silver clusters in solution<sup>12</sup> continuously change over time due to non-specific formation of different emissive silver clusters,<sup>91</sup> gradual additional reduction and cluster aggregation<sup>12</sup> because the DNA strands are too flexible. Our proposed luminescent system is more stable over time, as the silver clusters are contained by adenine in the pores of the MOF-69A framework, much like silver clusters in zeolites.<sup>20,22,23</sup> In zeolites A and Y, the smallest reported emissive clusters, most probably Ag<sub>3</sub><sup>+</sup>, also display the most blue-shifted emission. Based on these analogies, we propose that the blue-green luminescent adenine-stabilized clusters formed in MOF-69A are no larger than four silver atoms. However, based on theoretical studies of silver cluster in zeolites,<sup>25</sup> it was found that the optical

properties of the clusters is highly dependent on nuclearity, oxidation state and hydration level, which all determine the electronic properties of the metal clusters. For the Ag-loaded MOF samples, similar processes can be expected, but nevertheless this is subject to further research beyond the scope of this work.

Based on the excitation-emission profiles of our samples, the quantum yields were determined at two different wavelengths to discriminate between luminescence of the adenine-stabilized silver cluster species ( $\lambda_{exc} = 366$  nm) and of the 4,4'-BPDA linker ( $\lambda_{exc} = 330$  nm), as shown in Table 1. The parent bio-MOF-1 structure has a quantum yield (QY) of 8.3% ( $\lambda_{exc} = 366$  nm), which is much lower than the quantum yield of up to 18.8% for the new blue-green luminescence ( $\lambda_{exc} = 366$  nm) that appears after the MOF-to-MOF transformation from 1-1 EtOH-H<sub>2</sub>O. For samples prepared in ethanol-rich conditions, this green-blue luminescent species is overpowered by the blue luminescence of the 4,4'-BPDA linker, which causes the general quantum yield to rise for both tested excitation wavelengths. This is also clear from the excitation-emission profiles in Fig. S12 (ESI<sup>†</sup>). The UV-vis absorption properties of all silver-transformed bio-MOF-1 samples are still mainly determined by the organic linkers, originally present in bio-MOF-1 (Fig. S10, ESI<sup>†</sup>).



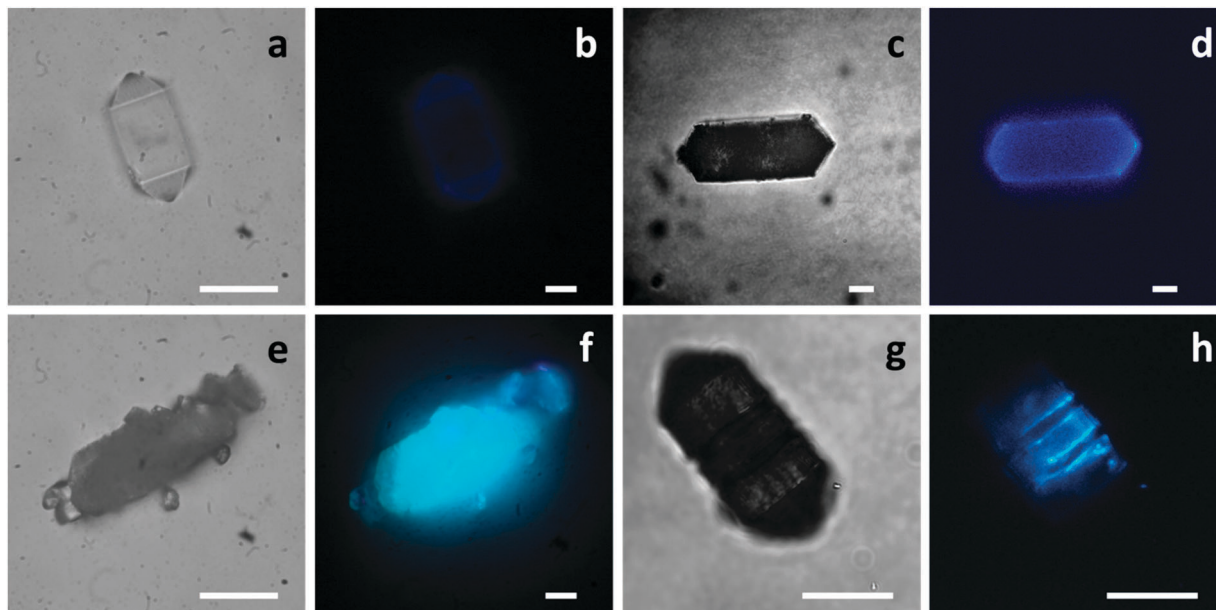


Fig. 6 Microscopy images of bio-MOF-1: (a and b) transmission and wide-field fluorescence, (c and d) transmission and confocal fluorescence microscopy (detection channel 430–470 nm for d); after silver-induced transformation from 4–1 EtOH–H<sub>2</sub>O: (e and f) transmission and wide-field fluorescence, (g and h) transmission and confocal fluorescence microscopy (detection channel 505–540 nm for h).

The emission differences between the starting bio-MOF-1 samples and the silver-transformed samples (4–1 ethanol–H<sub>2</sub>O) are clearly visible by eye under a standard laboratory UV lamp (Fig. S14, ESI<sup>†</sup>). Using wide-field and confocal fluorescence microscopy, not only the spectroscopic changes of single MOF crystals but also the structural transformation can be followed (Fig. 6). When scanning across the entire crystal depth of pristine bio-MOF-1 crystals, the blue luminescence appears homogeneously throughout the crystals, using the 430–470 nm emission detection channel (Fig. 6d). In contrast, for bio-MOF-1 crystals after the silver-induced transformation from 4–1 ethanol–water there appears to be an enrichment of the luminescent silver cluster species ( $\lambda_{\text{em}} = 485$  nm) towards the outer rim of the crystals, but some luminescence is also clearly detected inside the crystals using a 505–540 nm emission detection window (Fig. 6h). This detection window was applied to properly discriminate between the blue-green luminescent Ag species and the background luminescence of 4,4'-BPDA. This observation also lines up with the differences in outer-surface and bulk silver contents shown in Table 1.

From time-resolved fluorescence measurements ( $\lambda_{\text{ex}} = 375$  nm) for bio-MOF-1 and the Ag-transformed sample (4–1 EtOH–H<sub>2</sub>O), it is clear that the fluorescence lifetimes in both materials are different (Fig. 5). The fluorescence decay curves were fitted with a multi-exponential function; see Fig. S15 (ESI<sup>†</sup>) for residual signals. The decays in both samples at  $\lambda_{\text{em}} = 415$  nm are very similar, which possibly means they originate from the same molecule. Presumably, this molecule is adenine, based on the spectral information in Fig. 5 and Fig. S11 (ESI<sup>†</sup>). When specifically looking at the emission wavelengths of the adenine-stabilized silver clusters ( $\lambda_{\text{em}} = 485$  nm), a much slower fluorescence decay of about 20 ns is recorded for the MOF after transformation, while for the original bio-MOF-1 the

lifetime is only a few nanoseconds. This long lifetime component involves up to 97% of emitted photons' lifetime, as illustrated in Fig. 5. However, this 20 ns lifetime is much longer than the sub-10 ns lifetimes usually reported for most Ag-ssDNA materials.<sup>17</sup> This long lifetime is possibly caused by excited state processes that delay the fluorescence, such as charge transfers and electron recombination, as described in MOFs<sup>92</sup> and related silver cluster materials.<sup>93,94</sup>

## Conclusions

In this work, the adeninate framework bio-MOF-1 was investigated for its ability to stabilize luminescent silver clusters. A novel method for generating such clusters was developed by applying a well-adjusted reducing power using various EtOH–H<sub>2</sub>O ratios in the silver-containing solution. Among the large number of tested water-stable MOFs, only bio-MOF-1 appeared capable of producing new luminescent silver species. During the silver loading of bio-MOF-1, the framework reorganizes to MOF-69A due to the strong affinity of the silver ions for the adenine linkers. In this way, silver-adenine cluster species were generated in very high local concentrations in the pores of the formed MOF-69A. Adenine does not leave the system during this transformation, as confirmed by <sup>1</sup>H-NMR, TGA and DRS. These formed silver clusters possess a peculiar blue-green luminescence ( $\lambda_{\text{em max}} = 485$  nm) with long lifetimes and high quantum yield compared to the parent bio-MOF-1 material and silver clusters stabilized in ssDNA. This work is the proof-of-principle that metal–organic frameworks are potential hosts, under the correct conditions, for the stabilization of luminescent silver clusters, by using the versatility of these materials and



a confinement scaffold approach similar to that used to stabilize silver clusters in zeolites. Probing this new type of host materials can yield additional, deeper insights in the photophysical processes occurring for luminescent silver clusters in the large variety of researched host materials. This proof-of-principle study opens opportunities for application of luminescent MOFs in lighting applications and sensor devices.

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## References

- J. Zheng, P. R. Nicovich and R. M. Dickson, *Annu. Rev. Phys. Chem.*, 2007, **58**, 409–431.
- I. Díez and R. A. Ras, in *Advanced Fluorescence Reporters in Chemistry and Biology II*, ed. A. P. Demchenko, Springer, Berlin, Heidelberg, 2010, vol. 9, ch. 10, pp. 307–332.
- I. Díez and R. H. A. Ras, *Nanoscale*, 2011, **3**, 1963–1970.
- H. Xu and K. S. Suslick, *Adv. Mater.*, 2010, **22**, 1078–1082.
- J. P. Wilcoxon and B. L. Abrams, *Chem. Soc. Rev.*, 2006, **35**, 1162–1194.
- Y. Lu and W. Chen, *Chem. Soc. Rev.*, 2012, **41**, 3594–3623.
- G. De Cremer, Y. Antoku, M. B. J. Roeflaers, M. Sliwa, J. Van Noyen, S. Smout, J. Hofkens, D. E. De Vos, B. F. Sels and T. Vosch, *Angew. Chem., Int. Ed.*, 2008, **47**, 2813–2816.
- T. Baba, Y. Iwase, K. Inazu, D. Masih and A. Matsumoto, *Microporous Mesoporous Mater.*, 2007, **101**, 142–147.
- J. A. Martens, A. Cauvel, A. Francis, C. Hermans, F. Jayat, M. Remy, M. Keung, J. Lievens and P. A. Jacobs, *Angew. Chem., Int. Ed.*, 1998, **37**, 1901–1903.
- A. Corma, P. Concepción, M. Boronat, M. J. Sabater, J. Navas, M. J. Yacaman, E. Larios, A. Posadas, M. A. López-Quintela, D. Buceta, E. Mendoza, G. Guilera and A. Mayoral, *Nat. Chem.*, 2013, **5**, 775–781.
- J. T. Petty, C. Fan, S. P. Story, B. Sengupta, A. St. John Iyer, Z. Prudowsky and R. M. Dickson, *J. Phys. Chem. Lett.*, 2010, **1**, 2524–2529.
- J. T. Petty, J. Zheng, N. V. Hud and R. M. Dickson, *J. Am. Chem. Soc.*, 2004, **126**, 5207–5212.
- L. A. Peyser, A. E. Vinson, A. P. Bartko and R. M. Dickson, *Science*, 2001, **291**, 103–106.
- C. I. Richards, S. Choi, J.-C. Hsiang, Y. Antoku, T. Vosch, A. Bongiorno, Y.-L. Tzeng and R. M. Dickson, *J. Am. Chem. Soc.*, 2008, **130**, 5038–5039.
- T. Vosch, Y. Antoku, J.-C. Hsiang, C. I. Richards, J. I. Gonzalez and R. M. Dickson, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 12616–12621.
- S. M. Copp, D. Schultz, S. Swasey, J. Pavlovich, M. Debord, A. Chiu, K. Olsson and E. Gwinn, *J. Phys. Chem. Lett.*, 2014, **5**, 959–963.
- S. Choi, R. M. Dickson and J. Yu, *Chem. Soc. Rev.*, 2012, **41**, 1867–1891.
- T. Sun and K. Seff, *Chem. Rev.*, 1994, **94**, 857–870.
- G. De Cremer, E. Coutiño-Gonzalez, M. B. Roeflaers, B. Moens, J. Ollevier, M. Van der Auweraer, R. Schoonheydt, P. A. Jacobs, F. C. De Schryver, J. Hofkens, D. E. De Vos, B. F. Sels and T. Vosch, *J. Am. Chem. Soc.*, 2009, **131**, 3049–3056.
- G. De Cremer, E. Coutiño-Gonzalez, M. B. J. Roeflaers, D. E. De Vos, J. Hofkens, T. Vosch and B. F. Sels, *ChemPhysChem*, 2010, **11**, 1627–1631.
- G. De Cremer, B. F. Sels, J.-I. Hotta, M. B. J. Roeflaers, E. Bartholomeeusen, E. Coutiño-Gonzalez, V. Valtchev, D. E. De Vos, T. Vosch and J. Hofkens, *Adv. Mater.*, 2010, **22**, 957–960.
- E. Coutiño-Gonzalez, M. B. J. Roeflaers, B. Dieu, G. De Cremer, S. Leyre, P. Hanselaer, W. Fyen, B. Sels and J. Hofkens, *J. Phys. Chem. C*, 2013, **117**, 6998–7004.
- E. Coutiño-Gonzalez, D. Grandjean, M. Roeflaers, K. Kvashnina, E. Fron, B. Dieu, G. De Cremer, P. Lievens, B. Sels and J. Hofkens, *Chem. Commun.*, 2014, **50**, 1350–1352.
- E. Coutiño-Gonzalez, W. Baekelant, D. Grandjean, M. B. J. Roeflaers, E. Fron, M. S. Aghakhani, N. Bovet, M. Van der Auweraer, P. Lievens, T. Vosch, B. Sels and J. Hofkens, *J. Mater. Chem. C*, 2015, **3**, 11857–11867.
- N. T. Cuong, H. M. T. Nguyen and M. T. Nguyen, *Phys. Chem. Chem. Phys.*, 2013, **15**, 15404–15415.
- G. A. Ozin and H. Huber, *Inorg. Chem.*, 1978, **17**, 155–163.
- L. König, I. Rabin, W. Schulze and G. Ertl, *Science*, 1996, **274**, 1353–1354.
- E. Maik, R. Klaus, H. Armin, M. T. Dragomir, W. Wilfried, S. E. Reinhard and P. Gianfranco, *Nanotechnology*, 2008, **19**, 135701.
- M. V. Shestakov, L. F. Chibotaru, V. K. Tikhomirov, V. D. Rodriguez, J. J. Velazquez and V. V. Moshchalkov, *Phys. Chem. Chem. Phys.*, 2013, **15**, 15949–15953.
- J. Zheng and R. M. Dickson, *J. Am. Chem. Soc.*, 2002, **124**, 13982–13983.
- R. J. T. Houk, B. W. Jacobs, F. E. Gabaly, N. N. Chang, A. A. Talin, D. D. Graham, S. D. House, I. M. Robertson and M. D. Allendorf, *Nano Lett.*, 2009, **9**, 3413–3418.
- G. Ferey, *Chem. Soc. Rev.*, 2008, **37**, 191–214.
- C. Janiak, *Dalton Trans.*, 2003, 2781–2804, DOI: 10.1039/b305705b.
- C. Janiak and J. K. Vieth, *New J. Chem.*, 2010, **34**, 2366–2388.



- 35 J. R. Long and O. M. Yaghi, *Chem. Soc. Rev.*, 2009, **38**, 1213–1214.
- 36 S. Kitagawa, R. Kitaura and S. Noro, *Angew. Chem., Int. Ed.*, 2004, **43**, 2334–2375.
- 37 L. J. Murray, M. Dinca and J. R. Long, *Chem. Soc. Rev.*, 2009, **38**, 1294–1314.
- 38 D. Britt, H. Furukawa, B. Wang, T. G. Glover and O. M. Yaghi, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 20637–20640.
- 39 B. Van de Voorde, B. Bueken, J. Denayer and D. De Vos, *Chem. Soc. Rev.*, 2014, **43**, 5766–5788.
- 40 J.-R. Li, J. Sculley and H.-C. Zhou, *Chem. Rev.*, 2011, **112**, 869–932.
- 41 B. Van de Voorde, D. Damasceno Borges, F. Vermoortele, R. Wouters, B. Bozbiyik, J. Denayer, F. Taulelle, C. Martineau, C. Serre, G. Maurin and D. De Vos, *ChemSusChem*, 2015, **8**, 3159–3166.
- 42 A. Corma, H. Garcia and F. X. L. Xamena, *Chem. Rev.*, 2010, **110**, 4606–4655.
- 43 F. Vermoortele, R. Ameloot, A. Vimont, C. Serre and D. De Vos, *Chem. Commun.*, 2011, **47**, 1521–1523.
- 44 P. Valvekens, F. Vermoortele and D. De Vos, *Catal. Sci. Technol.*, 2013, **3**, 1435–1445.
- 45 P. Valvekens, M. Vandichel, M. Waroquier, V. Van Speybroeck and D. De Vos, *J. Catal.*, 2014, **317**, 1–10.
- 46 P. Valvekens, D. Jonckheere, T. De Baerdemaeker, A. V. Kubarev, M. Vandichel, K. Hemelsoet, M. Waroquier, V. Van Speybroeck, E. Smolders, D. Depla, M. B. J. Roefiaers and D. De Vos, *Chem. Sci.*, 2014, **5**, 4517–4524.
- 47 M. D. Allendorf, C. A. Bauer, R. K. Bhakta and R. J. T. Houk, *Chem. Soc. Rev.*, 2009, **38**, 1330–1352.
- 48 Y. Cui, Y. Yue, G. Qian and B. Chen, *Chem. Rev.*, 2011, **112**, 1126–1162.
- 49 J. Heine and K. Muller-Buschbaum, *Chem. Soc. Rev.*, 2013, **42**, 9232–9242.
- 50 J. An, S. J. Geib and N. L. Rosi, *J. Am. Chem. Soc.*, 2009, **131**, 8376–8377.
- 51 T. Wakaoka, K. Hirai, K. Murayama, Y. Takano, H. Takagi, S. Furukawa and S. Kitagawa, *J. Mater. Chem. C*, 2014, **2**, 7173–7175.
- 52 C. M. Doherty, D. Buso, A. J. Hill, S. Furukawa, S. Kitagawa and P. Falcaro, *Acc. Chem. Res.*, 2014, **47**, 396–405.
- 53 P. Falcaro, R. Ricco, A. Yazdi, I. Imaz, S. Furukawa, D. MasPOCH, R. Ameloot, J. D. Evans and C. J. Doonan, *Coord. Chem. Rev.*, 2016, **307**(part 2), 237–254.
- 54 B. W. Jacobs, R. J. T. Houk, M. R. Anstey, S. D. House, I. M. Robertson, A. A. Talin and M. D. Allendorf, *Chem. Sci.*, 2011, **2**, 411–416.
- 55 X. Cheng, M. Liu, A. Zhang, S. Hu, C. Song, G. Zhang and X. Guo, *Nanoscale*, 2015, **7**, 9738–9745.
- 56 L. He, L. F. Dumeé, D. Liu, L. Velleman, F. She, C. Banos, J. B. Davies and L. Kong, *RSC Adv.*, 2015, **5**, 10707–10715.
- 57 R. Ameloot, M. B. J. Roefiaers, G. De Cremer, F. Vermoortele, J. Hofkens, B. F. Sels and D. E. De Vos, *Adv. Mater.*, 2011, **23**, 1788–1791.
- 58 J. An, S. J. Geib and N. L. Rosi, *J. Am. Chem. Soc.*, 2009, **132**, 38–39.
- 59 T. Li, D.-L. Chen, J. E. Sullivan, M. T. Kozlowski, J. K. Johnson and N. L. Rosi, *Chem. Sci.*, 2013, **4**, 1746–1755.
- 60 S. Pérez-Yáñez, G. Beobide, O. Castillo, J. Cepeda, A. Luque, A. T. Aguayo and P. Román, *Inorg. Chem.*, 2011, **50**, 5330–5332.
- 61 H. Xu, J. Cai, S. Xiang, Z. Zhang, C. Wu, X. Rao, Y. Cui, Y. Yang, R. Krishna, B. Chen and G. Qian, *J. Mater. Chem.*, 2013, **1**, 9916–9921.
- 62 H. Oh, T. Li and J. An, *Chem. – Eur. J.*, 2015, **21**, 17010–17015.
- 63 J. An, O. K. Farha, J. T. Hupp, E. Pohl, J. I. Yeh and N. L. Rosi, *Nat. Commun.*, 2012, **3**, 1–6.
- 64 T. Li, M. T. Kozlowski, E. A. Doud, M. N. Blakely and N. L. Rosi, *J. Am. Chem. Soc.*, 2013, **135**, 11688–11691.
- 65 H. Cai, M. Li, X.-R. Lin, W. Chen, G.-H. Chen, X.-C. Huang and D. Li, *Angew. Chem., Int. Ed.*, 2015, **54**, 10454–10459.
- 66 J. An, S. Geib, M.-G. Kim, S. Choi and W. Lim, *J. Porous Mater.*, 2015, **22**, 867–875.
- 67 Y. Song, X. Yin, B. Tu, Q. Pang, H. Li, X. Ren, B. Wang and Q. Li, *CrystEngComm*, 2014, **16**, 3082–3085.
- 68 J. P. García-Terán, O. Castillo, A. Luque, U. García-Couceiro, P. Román and L. Lezama, *Inorg. Chem.*, 2004, **43**, 4549–4551.
- 69 F. Wang and Y. Kang, *Inorg. Chem. Commun.*, 2012, **20**, 266–268.
- 70 K. C. Stylianou, J. E. Warren, S. Y. Chong, J. Rabone, J. Bacsá, D. Bradshaw and M. J. Rosseinsky, *Chem. Commun.*, 2011, **47**, 3389–3391.
- 71 S. Pérez-Yáñez, G. Beobide, O. Castillo, J. Cepeda, A. Luque and P. Román, *Cryst. Growth Des.*, 2012, **12**, 3324–3334.
- 72 P. S. Nugent, V. L. Rhodus, T. Pham, K. Forrest, L. Wojtas, B. Space and M. J. Zaworotko, *J. Am. Chem. Soc.*, 2013, **135**, 10950–10953.
- 73 J. Yu, Y. Cui, H. Xu, Y. Yang, Z. Wang, B. Chen and G. Qian, *Nat. Commun.*, 2013, **4**, 1–7.
- 74 J. An and N. L. Rosi, *J. Am. Chem. Soc.*, 2010, **132**, 5578–5579.
- 75 J. An, C. M. Shade, D. A. Chengelis-Czegan, S. Petoud and N. L. Rosi, *J. Am. Chem. Soc.*, 2011, **133**, 1220–1223.
- 76 X. Shen and B. Yan, *J. Mater. Chem. C*, 2015, **3**, 7038–7044.
- 77 B. Li, Y. Zhang, D. Ma, T. Ma, Z. Shi and S. Ma, *J. Am. Chem. Soc.*, 2014, **136**, 1202–1205.
- 78 R. G. Pearson, *J. Am. Chem. Soc.*, 1963, **85**, 3533–3539.
- 79 N. L. Rosi, M. Eddaoudi, J. Kim, M. O’Keeffe and O. M. Yaghi, *Angew. Chem., Int. Ed.*, 2002, **41**, 284–287.
- 80 S. S. Kaye, A. Dailly, O. M. Yaghi and J. R. Long, *J. Am. Chem. Soc.*, 2007, **129**, 14176–14177.
- 81 S. Hausdorf, J. Wagler, R. Moßig and F. O. R. L. Mertens, *J. Phys. Chem. A*, 2008, **112**, 7567–7576.
- 82 N. L. Rosi, J. Kim, M. Eddaoudi, B. Chen, M. O’Keeffe and O. M. Yaghi, *J. Am. Chem. Soc.*, 2005, **127**, 1504–1518.
- 83 U. Ravon, M. Savonnet, S. Aguado, M. E. Domine, E. Janneau and D. Farrusseng, *Microporous Mesoporous Mater.*, 2010, **129**, 319–329.
- 84 S. W. Gaarenstroom and N. Winograd, *J. Chem. Phys.*, 1977, **67**, 3500–3506.
- 85 A. M. Fonseca and I. C. Neves, *Microporous Mesoporous Mater.*, 2013, **181**, 83–87.
- 86 O. Fenwick, E. Coutino-Gonzalez, W. Baekelant, F. Richard, S. Bonacchi, M. B. J. Roefiaers, J. Hofkens and P. Samori, under review.
- 87 G. W. C. Kaye and T. H. Laby, *Tables of Physical & Chemical Constants*, 16th edn, 1995.



- 88 D. Rankine, T. D. Keene, C. J. Sumby and C. J. Doonan, *CrystEngComm*, 2013, **15**, 9722–9728.
- 89 C. A. Bauer, T. V. Timofeeva, T. B. Settersten, B. D. Patterson, V. H. Liu, B. A. Simmons and M. D. Allendorf, *J. Am. Chem. Soc.*, 2007, **129**, 7136–7144.
- 90 N. Kiomarsipour and R. Shoja Razavi, *Superlattices Microstruct.*, 2012, **52**, 704–710.
- 91 I. Diez, R. H. A. Ras, M. I. Kanyuk and A. P. Demchenko, *Phys. Chem. Chem. Phys.*, 2013, **15**, 979–985.
- 92 K. G. M. Laurier, E. Fron, P. Atienzar, K. Kennes, H. Garcia, M. Van der Auweraer, D. E. De Vos, J. Hofkens and M. B. J. Roeffaers, *Phys. Chem. Chem. Phys.*, 2014, **16**, 5044–5047.
- 93 V. Soto-Verdugo, H. Metiu and E. Gwinn, *J. Chem. Phys.*, 2010, **132**, 195102.
- 94 S. A. Patel, M. Cozzuol, J. M. Hales, C. I. Richards, M. Sartin, J.-C. Hsiang, T. Vosch, J. W. Perry and R. M. Dickson, *J. Phys. Chem. C*, 2009, **113**, 20264–20270.

