

Cite this: *Chem. Sci.*, 2016, **7**, 6846

Cubic three-dimensional hybrid silica solids for nuclear hyperpolarization[†]

D. Baudouin,^{*a} H. A. van Kalkeren,^a A. Bornet,^b B. Vuichoud,^b L. Veyre,^a M. Cavaillès,^a M. Schwarzwälder,^c W.-C. Liao,^c D. Gajan,^d G. Bodenhausen,^{befg} L. Emsley,^b A. Lesage,^d S. Jannin,^b C. Copéret^{*c} and C. Thieuleux^{*a}

Hyperpolarization of metabolites by dissolution dynamic nuclear polarization (D-DNP) for MRI applications often requires fast and efficient removal of the radicals (polarizing agents). Ordered mesoporous SBA-15 silica materials containing homogeneously dispersed radicals, referred to as HYperPolarizing SOLids (HYPSOs), enable high polarization – $P(^1\text{H}) = 50\%$ at 1.2 K – and straightforward separation of the polarizing HYPSO material from the hyperpolarized solution by filtration. However, the one-dimensional tubular pores of SBA-15 type materials are not ideal for nuclear spin diffusion, which may limit efficient polarization. Here, we develop a generation of hyperpolarizing solids based on a SBA-16 structure with a network of pores interconnected in three dimensions, which allows a significant increase of polarization, *i.e.* $P(^1\text{H}) = 63\%$ at 1.2 K. This result illustrates how one can improve materials by combining a control of the incorporation of radicals with a better design of the porous network structures.

Received 11th May 2016

Accepted 15th July 2016

DOI: 10.1039/c6sc02055k

www.rsc.org/chemicalscience

One of the major limitations of nuclear magnetic resonance spectroscopy (NMR) and magnetic resonance imaging (MRI) is their intrinsic low sensitivity, which arises from low thermal equilibrium (Boltzmann) polarization of the nuclear spins at room temperature, even with the highest field instruments. This low sensitivity is particularly challenging for low-gamma nuclear spins. For carbon-13, polarization is as low as $P(^{13}\text{C}) \approx 2 \times 10^{-5}$ at $T = 300$ K and $B_0 = 23.5$ T. This weakness can be circumvented by dissolution dynamic nuclear polarization (D-DNP).^{1,2} D-DNP takes advantage of the high electron spin polarization at low temperatures to enhance the nuclear spin polarization well beyond thermodynamic equilibrium (>10 000 times) *via* microwave irradiation. Such huge gains in sensitivity allow metabolic imaging and have for example enabled the detection of anomalous metabolic rates in prostate tumors in living patients.³

^aUniversité de Lyon, Institut de Chimie de Lyon, LC2P2, UMR 5265 CNRS-CPE Lyon-UCBL, CPE Lyon, 43 Bd du 11 Novembre 1918, 69100 Villeurbanne, France. E-mail: david.baudouin@univ-lyon1.fr; chloé.thieuleux@univ-lyon1.fr

^bInstitut des Sciences et Ingénierie Chimiques, Ecole Polytechnique Fédérale de Lausanne (EPFL), CH-1015 Lausanne, Switzerland

^cETH Zürich, Department of Chemistry and Applied Biosciences, Vladimir-Prelog-Weg 1-5/10, 8093 Zürich, Switzerland. E-mail: ccoperet@inorg.chem.ethz.ch

^dUniversité de Lyon, Institut des Sciences Analytiques, UMR 5280, CNRS, Université Lyon 1, ENS Lyon 5 rue de la Doua, F-69100 Villeurbanne, France

^eDépartement de Chimie, Ecole Normale Supérieure, 24 Rue Lhomond, 75231 Paris Cedex 05, France

^fUniversité Pierre-et-Marie Curie, Paris, France

^gUMR 7203, CNRS/UPMC/ENS, Paris, France

[†] Electronic supplementary information (ESI) available. See DOI: 10.1039/c6sc02055k

Hyperpolarization by D-DNP involves microwave irradiation at low temperatures in moderate magnetic fields (typically $T = 1.2$ K and $B_0 = 3.35$ or 6.7 T) of frozen glassy solutions doped with stable free radicals and molecules of interest (*e.g.* metabolites or tracers). The preparation of a glassy frozen sample is important because it ensures that the radicals are statistically distributed, without the formation of ice crystals, which leads to optimal DNP efficiency. For that purpose, glass-forming agents such as glycerol, DMSO or ethanol are usually included in high concentrations. In a typical D-DNP experiment, following polarization, the polarized solution is rapidly brought to room temperature using superheated water and quickly transferred to the MRI or NMR machine for further studies. Once the solution is hyperpolarized, both radicals and glass-forming agents are unwanted and should obviously not be injected into patients. Furthermore, radicals act as paramagnetic relaxing agents, inducing faster depolarization.⁴ Therefore, rapid removal of radicals after polarization and before use is essential. In the case of trityl and BDPA radicals, the removal can be achieved by precipitation followed by filtration or by ion exchange.^{5,6} However both methods are limited to specific sample formulations. Alternatively, nitroxide based radicals can be scavenged by ascorbate (vitamin C),⁷ which attenuates paramagnetic relaxation but leads to contamination of the samples by hydroxylamines. Radicals can be incorporated into polymers such as polystyrene particles⁸ or hydrogels,⁹ allowing physical separation of the polarizing agent from the solution, but the efficiency for D-DNP system is limited and the filtration not straightforward.



In this context, we have recently developed a family of solid polarizing matrices based on hybrid materials containing covalently bound radicals, coined HYperPolarizing SOLids (HYPSo). These materials provide in principle a universal solution to the above-mentioned issues: *i.e.* fast and easy removal of radicals by filtration, and the absence of glass-forming agents. HYPSo are porous and robust silica-based solids on which any radicals (*i.e.* TEMPO, trityl...) can be covalently and homogeneously attached to the surface of their pores.^{10,11} We showed an efficient direct polarization approaching $P(^{13}\text{C}) = 15\%$ with a build-up time of 2 h in a 3 M $1\text{-}^{13}\text{C}$ sodium pyruvate aqueous solution impregnated with trityl-based HYPSo.¹¹ The first generation TEMPO-based materials, HYPSo-1, allowed reaching a ^{13}C polarization as high as $P(^{13}\text{C}) = 33\%$ in only 20 min using a state-of-the-art polarizer including microwave frequency modulation¹² and ^1H - ^{13}C cross-polarization.¹³ Despite significant research efforts to improve the DNP performances of the first generations of HYPSo, it was not possible to enhance the proton polarization beyond $P(^1\text{H}) = 50\%$, well below the $P(^1\text{H}) = 90\%$ that can be obtained under similar conditions in glassy water/glycerol TEMPOL solutions.¹³

The two first generations of HYPSo were based on ordered mesoporous SBA-15 type structures, with a skeleton consisting of 8–10 nm diameter 1D-pore channels stacked in a 2D-hexagonal arrangement (Fig. 1 – top right). Two generations of nitroxide-based materials, HYPSo-1 and HYPSo-2, were hence prepared and differ from the linker used to anchor the radical to the solid surface, a propylamido¹⁰ and a 1,2,3-triazole-propyl tethers, respectively.¹¹ Using a direct synthesis, the radicals were homogeneously incorporated onto the pore surface of HYPSo-1 and HYPSo-2 by peptide coupling or click chemistry, respectively, avoiding radical aggregation, which is important for

D-DNP. However, in such a structure, the pores do not communicate with each other; we thus hypothesized that this could be a limiting factor for both nuclear spin diffusion and for the three-dimensional distribution of the radicals, in comparison to frozen glassy solutions. We therefore reasoned that a silica architecture with a 3D cubic porous network (for example using SBA-16 like structures) could improve the DNP performance. Here we describe the development of materials with cubic network arrangement (HYPSo-3) and show that they lead to greater polarization with respect to the one-dimensional porous HYPSo-1/2 materials, yielding proton polarization up to $P(^1\text{H}) = 63\%$. We show how the 3D cubic material can also be efficiently used under Magic Angle Spinning (MAS) DNP conditions.

Results and discussion

Preparation and characterization of materials

First, propylazido functionalized SBA-16 materials were prepared by co-condensation of (3-azidopropyl)triethoxysilane and tetraethyl orthosilicate in an HCl/NaCl aqueous medium in the presence of pluronic F127 as a structure-directing agent (SDA).¹⁴ The ratios between (3-azidopropyl)triethoxysilane and tetraethyl orthosilicate varied in the range 1/34, 1/60, 1/100, 1/140, 1/320, corresponding to a loading of organic functionalities of 472, 272, 164, 118 and 52 $\mu\text{mol}_{\text{SiR}}\text{ g}^{-1}$. The materials obtained were filtered, and the SDA was extracted with a pyridine/HCl solution for 24 h and finally washed and dried under 10^{-5} mbar vacuum at 135 °C.¹⁵ The different materials were analyzed by N_2 adsorption, which showed a type IV isotherm characteristic of mesoporous materials. The BET model indicated surface areas ranging from *ca.* 910 to 1080 $\text{m}^2\text{ g}^{-1}$ (see Table 1) with total pore volumes ($P/P_0 < 0.99$) of 0.62–0.82 $\text{cm}^3\text{ g}^{-1}$, of which 37–46% are micropores according to the α -plot model. The pore sizes have narrow bimodal distributions at 1.4–1.7 (inter-connecting windows, MP model) and 6.2–7.0 nm (cavities, BJH model). The sharp drop in the desorption branch of the isotherm observed at *ca.* 0.45 P/P_0 is typical of nitrogen desorption from mesoporous materials through smaller pores (ink bottle effect), in line with the formation of 6–7 nm cages interconnected by 1–2 nm connecting micropores. Small Angle X-Ray Diffraction (SA-XRD, see Table SI-1†) of 1/34_N₃_SBA-16 presented a strong peak at *ca.* $2\theta = 0.75^\circ$ assigned to the X-ray diffraction of the {110} family of planes, indicative of well-structured cubic-centered body SBA-16 materials ($Im\bar{3}m$ space group). SA-XRD and N_2 adsorption/desorption data allow one to calculate a mean micropore length of *ca.* 10 nm (Table 1 and SI-1†). Transmission Electron Micrographs (TEM) of 1/140_N₃_SBA-16 confirm the formation of a cubic ordered mesoporous material (see representative pictures in Fig. 1 and S1†).

For comparison, several polarizing matrices with 2D hexagonal arrangements of mesopore tube-like pores (SBA-15 type materials), hereafter named HYPSo-2, were also prepared.¹⁰ These materials are highly porous with a BET surface area, a total pore volume, BJH and MP pore diameters of 770–870 $\text{m}^2\text{ g}^{-1}$, 1.1–1.2 $\text{cm}^3\text{ g}^{-1}$ and 8.0–9.2 nm, respectively (Table S2†).

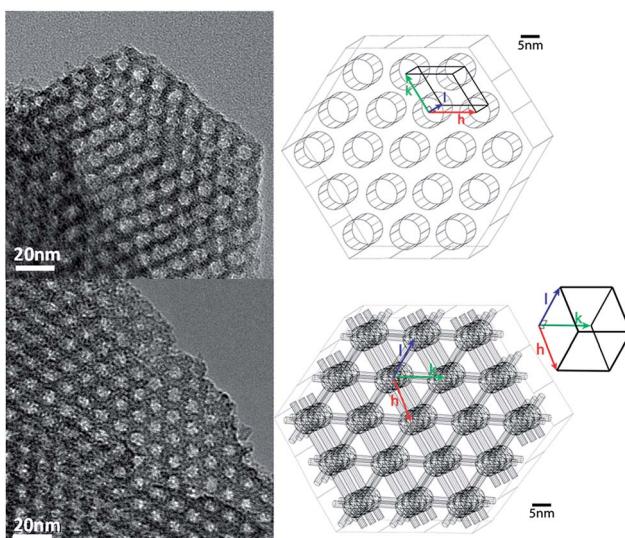


Fig. 1 Left: TEM pictures of hexagonal 1/100_N₃_SBA-15 in the [001] axis (top) and of cubic 1/140_N₃_SBA-16 in the [111] axis (bottom). Right: Schematic representations of the 2D pore structure in the [001] axis of SBA-15/HYPSo-2 (top) and the 3D pore structure in the [111] axis of SBA-16/HYPSo-3 (axis). The radicals are distributed uniformly over the surface of the pores. See Fig. SI-1† for SBA-16 in [100] axis.



Table 1 Organic function loadings and textural characteristics of the materials obtained from SA-XRD and N_2 -adsorption desorption at 77 K

Materials	$[\equiv SiR]/\mu mol_{SiR} g^{-1}$	$S_{BET}/m^2 g^{-1}$	$V_p^a (tot.)/m^2 g^{-1}$	$V_p^b (\mu)/m^2 g^{-1}$	D_p^c/nm	$L_{\mu pore}^d /nm$
1/34_N ₃ _SBA-16	472	1012	0.68	0.31	1.7/6.2	9.6
1/34_HYPSO-3	472	729	0.50	0.19	1.6/5.4	10.4
1/60_N ₃ _SBA-16	272	1010	0.66	0.29	1.6/6.3	n.d.
1/60_HYPSO-3	272	752	0.52	0.22	1.6/5.4	10.2
1/100_N ₃ _SBA-16	164	913	0.62	0.11	1.7/6.2	n.d.
1/100_HYPSO-3	164	893	0.63	0.26	1.3/6.3	8.6
1/140_N ₃ _SBA-16	118	1184	0.82	0.33	1.4/7.0	n.d.
1/140_HYPSO-3	118	983	0.69	0.26	1.7/7.1	9.3
1/320_N ₃ _SBA-16	52	1068	0.75	0.48	1.7/7.0	n.d.
1/320_HYPSO-3	52	714	0.48	0.27	1.6/5.4	12.3

^a Total pore volume corresponding to the quantity of N_2 adsorbed at $P/P_0 = 0.99$. ^b Micropore volume, calculated from the α_S plot model.

^c Micropore mean diameter calculated using MP model/mesopore mean diameter calculated using the BJH model (adsorption branch).

^d Micropore mean length, calculated using $L_{\mu pore} = (d_{(110)}/\cos(\pi/4) - D_{meso})$ using the mesoporous diameter D_p and the d -spacing $d_{(110)}$ obtained from Small Angle XRD analysis.

Importantly, these SBA-15 type materials exhibit non-interconnected 1.8 nm micropores (5–8% of pore volume). In contrast, HYPSO-3 (SBA-16 materials) exhibits spherical mesopores interconnected by micro-channels in all three dimensions. This pore structure leads to different textures: a lower pore volume and a different intra-grain pore volume distribution (Fig. 1).

Post-functionalization of N_3 -SBA-16 to obtain HYPSO-3 materials was performed using copper-catalyzed azide-alkyne cycloaddition (Cu-AAC)¹⁶ in the presence of *o*-propargyl TEMPO, CuI, dry DMF and Et_3N (see details in ESI and Fig. S2 and S3†). Diffuse Reflectance Infrared Fourier Transform Spectroscopy (DRIFTS) analysis of the powder allowed evaluation of the efficiency of the cycloaddition (referred to as Cu-AAC yield) on HYPSO-3.

As shown in Table 2, 88–64% of the starting azido $-N_3$ reacted with the *o*-propargyl TEMPO reactant, the yield decreasing slowly when decreasing the molar concentration of radicals (quoted 1/xx ratio that stands for 1 mol of radical per xx mol of SiO_2). The concentration of radical incorporated in HYPSO-3 was quantified by recording X-band CW EPR spectra at room temperature. Nitroxyl radical loadings of 246, 135, 79, 50 and 33 $\mu mol_{NO} g^{-1}$ were measured for ratios of 1/34, 1/60, 1/100, 1/140 and 1/320 respectively, corresponding to radical concentrations of 491, 260, 125, 72 and 67 $\mu mol_{NO} cm^{-3}$ within the

total volume of the pores. The radical concentrations show that the yields of post-functionalization are in the range 41–63%, similar to 2D-hexagonal materials characterized by the same method (42–57%). No significant difference between the EPR profiles of HYPSO-2 and HYPSO-3 could be observed at room temperature (Fig. S4†).

EPR spectroscopy

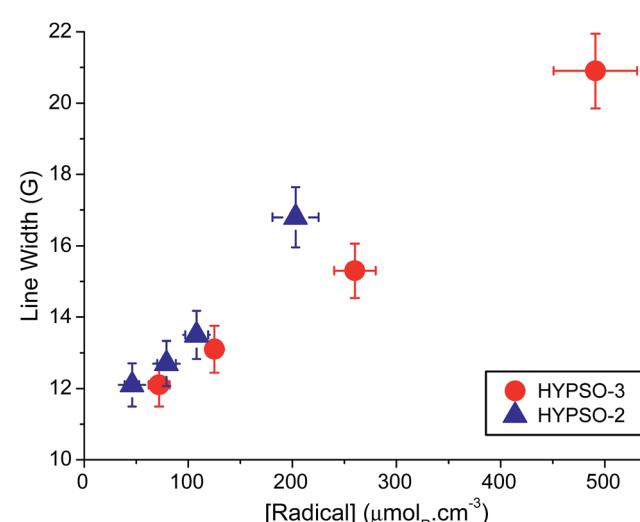
The average inter-radical distances in HYPSO-3 were evaluated from the line width of the central EPR signal at 110 K, which is known to be broadened by electron–electron dipolar couplings and by spin exchange.¹⁷ Such an analysis is limited to an average inter-radical distance $r_{RR} < 2$ nm. For larger distances, the dipolar line width is masked by inhomogeneous broadening (≈ 12 G). The EPR line-widths measured are 20.9, 15.3, 13.1 and 12.1 G for HYPSO-3 with ratios $N_3/TEOS = 1/34, 1/60, 1/100$ and 1/140 respectively (Fig. 2 and Table S3†). The dipolar broadening is almost proportional to the radical concentrations for

Table 2 Characteristics of HYPSO-3 materials

Ratio	$[R]/\mu mol g^{-1}$	$[NO^\cdot]/\mu mol g^{-1}$	Cu-AAC yield ^a (%)	EPR yield ^b (%)	$[NO^\cdot]^c / \mu mol cm^{-3}$
1/34	472	246	88	52	491
1/60	272	135	81	50	260
1/100	164	79	77	48	125
1/140	118	50	74	41	72
1/320	52	33	64	63	67

^a Percentage of N_3 reacted after Cu-AAC (obtained by DRIFTS).

^b Percentage of NO^\cdot compared to initial N_3 . ^c Concentration per total pore volume ($P/P_0 = 0.99$).

Fig. 2 EPR linewidths of HYPSO-2 and -3 as a function of the molar radical concentration (in $\mu mol_{NO} cm^{-3}$).

HYPSO-2 and -3, although the slopes are different. Indeed, the line width of HYPSO-3 (SBA-16) is narrower than that of HYPSO-2 (SBA-15, data from ref. 11) for the same volumetric radical concentration. This narrower line width can be attributed to a more uniform 3D distribution of the radicals. We believe that this improvement likely originates from the difference in pore shape (tubular *vs.* interconnected cages) and structuration between HYPSO-2 and HYPSO-3.

DNP performances

The DNP efficiency of HYPSO-3 was first evaluated for samples spinning at the magic angle (MAS) near 100 K and gave results similar to HYPSO-2^{10,18,19} (see ESI†). The DNP performance of HYPSO-2 and -3 was then determined at 4.2 and 1.2 K. Both materials were impregnated by filling *ca.* 95% of the pore volume with D₂O : H₂O (8 : 2). The results obtained at 4.2 K with microwave frequency modulation¹² are presented in Fig. 3.

One can observe a maximum polarization in the vicinity of $[R] = 50 \mu\text{mol cm}^{-3}$. At this concentration, HYPSO-3 yields a polarization $P(^1\text{H}) = 12.5\%$, significantly higher than the 7.5% obtained with HYPSO-2. The use of microwave frequency

modulation improves the DNP performances of both HYPSO-2 and -3 but only for $[R] < 100 \mu\text{mol cm}^{-3}$ (see Fig. S6†).

For comparison, an isotropic “glassy” H₂O : D₂O : glycerol-d₈ (10 : 40 : 50) matrix without HYPSO doped with 40 mM TEMPOL ($40 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$) gave rise to $P(^1\text{H}) = 21.5\%$. Note that a $80 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$ solution gives comparable results. When impregnating HYPSO-3 type matrices containing surface azido-groups instead of TEMPO units (1/140_N₃_SBA-16) with a 40 mM TEMPO solution, a polarization $P(^1\text{H}) = 19\%$ was obtained. This polarization value is close to that of the isotropic glassy DNP solution ($P(^1\text{H}) = 21.5\%$). On the contrary, when impregnated in a HYPSO-2 matrix without any radicals (1/140_N₃_SBA-15), the ¹H polarization dropped to $P(^1\text{H}) = 9\%$. We take this as a strong indication that the cubic 3D porous network is advantageous for efficient DNP as compared to a one-dimensional network.

The build-up time constant (Fig. S6†) was found to be $\tau_{\text{DNP}} = 74 \text{ s}$ for the reference DNP solution without HYPSO at 4.2 K. The same solution impregnated in 1/140_N₃_SBA-16 (same as HYPSO-3) gave $\tau_{\text{DNP}} = 145 \text{ s}$ for a similar polarization, and $\tau_{\text{DNP}} = 77 \text{ s}$ when impregnated in 1/140_N₃_SBA-15 (same as HYPSO 2) (Fig. S6†). EPR studies confirmed that this lengthening of the DNP build-up in HYPSO-3 was not due to a radical quenching effect.

At 1.2 K, HYPSO-3 yielded a polarization $P(^1\text{H}) > 40\%$ over a broad range of radical concentrations $50 < [R] < 160 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$, reaching $P(^1\text{H}) = 63\%$ for $[R] = 67 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$ (cf. Fig. 3, bottom).

By comparison, HYPSO-2 only yielded a maximum polarization $P(^1\text{H}) = 50\%$ for an optimal $[R] = 79 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$. The polarization in the H₂O : D₂O : glycerol-d₈ (10 : 40 : 50) mixture containing 40 mM TEMPOL can reach $P(^1\text{H}) = 90\%$ under the same conditions.¹³ At 1.2 K, frequency modulation was found to have a positive effect on DNP for HYPSO-2 and -3 when $[R] < 75 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$. When the optimal DNP solution was impregnated in 1/140_N₃_SBA-16 (radical-free HYPSO-3) and 1/140_N₃_SBA-15 (radical-free HYPSO-2), we observed $P(^1\text{H}) = 63\%$ for HYPSO-3 and 50% for HYPSO-2 at 1.2 K. Note that higher polarization might be reached using HYPSO-3 with lower radical concentration, but build up times (τ_{DNP}) would become

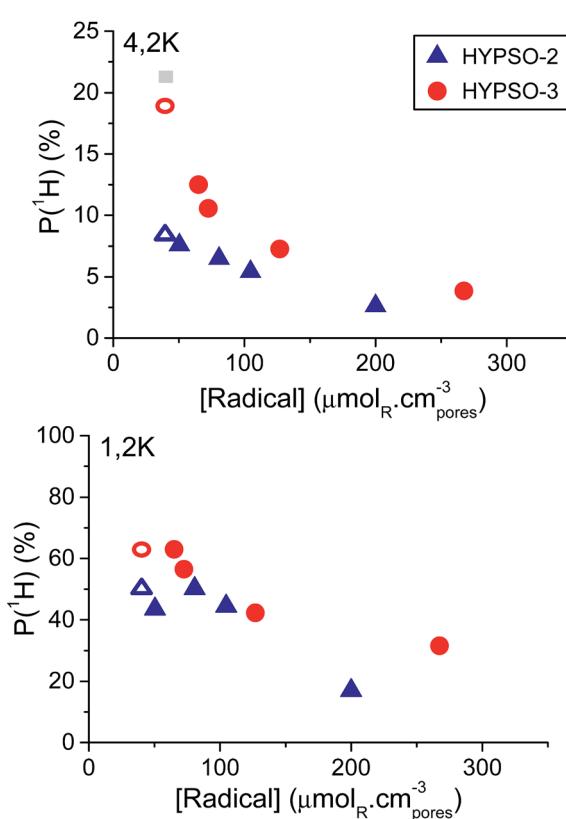


Fig. 3 Polarization $P(^1\text{H})$ measured using microwave frequency modulation at 4.2 K (top) and 1.2 K (bottom) and 6.7 T as a function of radical concentration (in $\mu\text{mol}_{\text{NO}} \text{cm}^{-3}$) for: HYPSO-2 (▲) and HYPSO-3 (●) impregnated with D₂O : H₂O (8 : 2). The open circles ○ and triangles △ correspond to a solution of 40 mM TEMPOL in H₂O : D₂O : glycerol-d₈ (10 : 40 : 50) impregnated in 1/140_N₃_SBA-16 (radical-free HYPSO-3) and 1/140_N₃_SBA-15 (radical-free HYPSO-2), respectively. The grey square ■ corresponds to a TEMPOL reference DNP solution, without HYPSO.

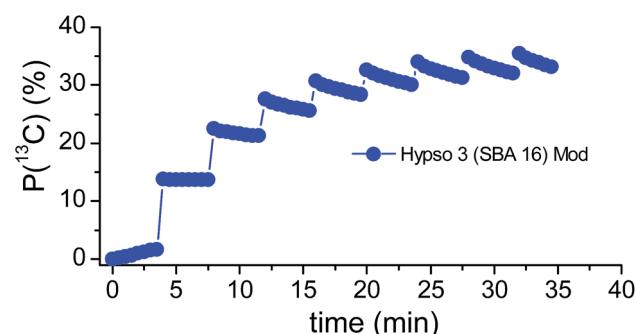


Fig. 4 Left: $^1\text{H} \rightarrow ^{13}\text{C}$ CP-DNP performed on HYPSO-3 material ($67 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$) impregnated with a 3 M solution of [1-¹³C]-acetate in D₂O. $P(^{13}\text{C}) = 36\%$ is reached in 32.5 min with $^1\text{H} \rightarrow ^{13}\text{C}$ CP applied at 4 min intervals.



very long (>300 s) (see Fig. S6†). Finally, we prepared a 3 M solution of sodium [$1\text{-}^{13}\text{C}$]-acetate in $\text{H}_2\text{O} : \text{D}_2\text{O}$ (1 : 9) to impregnate HYPSO-3 (with $[\text{R}] = 67 \mu\text{mol}_{\text{NO}} \text{cm}^{-3}$) and we obtained $P(^1\text{H}) \approx 50\%$. Cross polarization²⁰ was performed with 8 contacts every 4 min, yielding $P(^1\text{H} \rightarrow ^{13}\text{C}) = 36\%$ after *ca.* 30 minutes (Fig. 4). After dissolution, the aqueous acetate solution was recovered by filtration and centrifugation and subjected to ESR analysis which confirmed the presence of a negligible quantity of radicals in the liquid (*ca.* 1 $\mu\text{mol L}^{-1}$ *i.e.* $<0.3\%$ of HYPSO) probably arising from the presence of very small grains of HYPSO-3 in the solution.

Conclusions

In conclusion, we have shown that the architecture of the porous network affects the hyperpolarization properties of HYPSO materials for DNP. Here, a material with 1D tubular pores (HYPSO-2) allows one to reach $P(^1\text{H}) = 50\%$ at 1.2 K, while a material with 3D interconnected cage-like pores in cubic symmetry (HYPSO-3) leads to a polarization of $P(^1\text{H}) = 63\%$. We propose that this is due to (i) a more homogeneous 3D distribution of nitroxyl radicals in HYPSO-3, as demonstrated by EPR, and (ii) the interconnection of the pores, which allows for nuclear spin diffusion in all three directions, as in ideal solutions. We are currently working on designing optimal materials based on these underlying principles.

Acknowledgements

This work was supported by the CNRS, the SATT Lyon (Pulsalys), the European Research Council (ERC) Advanced Grant No. 320860, the Swiss National Science Foundation (SNSF), the Ecole Polytechnique Fédérale de Lausanne (EPFL), ETH Zürich, and the Swiss Commission for Technology & Innovation (CTI). Mathieu Baudin is kindly acknowledged for his contribution to dissolution DNP experiments.

References

- 1 J. H. Ardenkjær-Larsen, B. Fridlund, A. Gram, G. Hansson, L. Hansson, M. H. Lerche, R. Servin, M. Thaning and K. Golman, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 10158–10163.
- 2 J. H. Ardenkjær-Larsen, *J. Magn. Reson.*, 2016, **264**, 3–12.
- 3 S. J. Nelson, J. Kurhanewicz, D. B. Vigneron, P. E. Z. Larson, A. L. Harzstark, M. Ferrone, M. van Criekinge, J. W. Chang, R. Bok, I. Park, G. Reed, L. Carvajal, E. J. Small, P. Munster, V. K. Weinberg, J. H. Ardenkjær-Larsen, A. P. Chen, R. E. Hurd, L.-I. Odegardstuen, F. J. Robb, J. Tropp and J. A. Murray, *Sci. Transl. Med.*, 2013, **5**, 198ra108.
- 4 P. Miéville, S. Jannin and G. Bodenhausen, *J. Magn. Reson.*, 2011, **210**, 137–140.
- 5 J. A. Urbahn, J. Ardenkjær-Larsen, A. M. Leach, E. Telfeyan, D. K. Dietrich, D. B. Whitt, P. Miller and E. W. Stautner, *US Pat.*, US 2008/0240998 A1, 2007.
- 6 J. H. Ardenkjær-Larsen, A. M. Leach, N. Clarke, J. Urbahn, D. Anderson and T. W. Skloss, *NMR Biomed.*, 2011, **24**, 927–932.
- 7 P. Miéville, P. Ahuja, R. Sarkar, S. Jannin, P. R. Vasos, S. Gerber-Lemaire, M. Mishkovsky, A. Comment, R. Gruetter, O. Ouari, P. Tordo and G. Bodenhausen, *Angew. Chem., Int. Ed.*, 2010, **49**, 6182–6185.
- 8 Y. Zhang, P. J. Baker and L. B. Casabianca, *J. Phys. Chem. B*, 2016, **120**, 18–24.
- 9 B. C. Dollmann, M. J. N. Junk, M. Drechsler, H. W. Spiess, D. Hinderberger and K. Munnemann, *Phys. Chem. Chem. Phys.*, 2010, **12**, 5879–5882; T. Cheng, M. Mishkovsky, M. J. N. Junk, K. Munnemann and A. Comment, *Macromol. Rapid Commun.*, 2016, **37**, 1074–1078.
- 10 D. Gajan, M. Schwarzwälder, M. P. Conley, W. R. Grüning, A. J. Rossini, A. Zagdoun, M. Lelli, M. Yulikov, G. Jeschke, C. Sauvée, O. Ouari, P. Tordo, L. Veyre, A. Lesage, C. Thieuleux, L. Emsley and C. Copéret, *J. Am. Chem. Soc.*, 2013, **135**, 15459–15466.
- 11 D. Gajan, A. Bornet, B. Vuichoud, J. Milani, R. Melzi, H. A. van Kalkeren, L. Veyre, C. Thieuleux, M. P. Conley, W. R. Grüning, M. Schwarzwälder, A. Lesage, C. Copéret, G. Bodenhausen, L. Emsley and S. Jannin, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 14693–14697.
- 12 A. Bornet, J. Milani, B. Vuichoud, A. J. Perez Linde, G. Bodenhausen and S. Jannin, *Chem. Phys. Lett.*, 2014, **602**, 63–67.
- 13 S. Jannin, A. Bornet, R. Melzi and G. Bodenhausen, *Chem. Phys. Lett.*, 2012, **549**, 99–102.
- 14 A. Boullanger, J. Alauzun, A. Mehdi, C. Reye and R. J. P. Corriu, *New J. Chem.*, 2010, **34**, 738–743.
- 15 R. J. P. Corriu, A. Mehdi, C. Reye and C. Thieuleux, *New J. Chem.*, 2003, **27**, 905–908.
- 16 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**, 2004.
- 17 G. Jeschke, A. Koch, U. Jonas and A. Godt, *J. Magn. Reson.*, 2002, **155**, 72–82.
- 18 A. J. Rossini, A. Zagdoun, M. Lelli, A. Lesage, C. Copéret and L. Emsley, *Acc. Chem. Res.*, 2013, **46**, 1942–1951.
- 19 A. Lesage, M. Lelli, D. Gajan, M. A. Caporini, V. Vitzthum, P. Miéville, J. Alauzun, A. Roussey, C. Thieuleux, A. Mehdi, G. Bodenhausen, C. Copéret and L. Emsley, *J. Am. Chem. Soc.*, 2010, **132**, 15459–15461.
- 20 A. Bornet, R. Melzi, A. J. P. Linde, P. Hautle, B. van den Brandt, S. Jannin and G. Bodenhausen, *J. Phys. Chem. Lett.*, 2013, **4**, 111–114.

