A reactivity-based $^{18}$FFDG probe for in vivo formaldehyde imaging using positron emission tomography†

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Formaldehyde (FA) is a reactive carbonyl species (RCS) that plays a broad spectrum of roles in epigenetics, toxicology, and progression of diseases ranging from cancer to diabetes to neurodegeneration, motivating the development of translatable technologies for FA imaging. Here we report formaldehyde-caged-$^{18}$F fluorodeoxyglucose-1 ([$^{18}$F]FAC-FDG-1), an aza-Cope-based reactivity probe for in vivo FA imaging using positron emission tomography (PET). [$^{18}$F]FAC-FDG-1 reacts selectively with FA over potentially competing analytes to generate [$^{18}$F]FDG, allowing its FA-dependent uptake and retention in cell culture as well as in animal models. The relative uptake of [$^{18}$F]FAC-FDG-1 was evaluated using FA-treated PC3 prostate cancer and U87-MG glioblastoma cells demonstrating a dose-dependent response to exogenously added FA. Moreover, [$^{18}$F]FAC-FDG-1 is capable of FA detection in vivo using a PC3 tumor xenograft model. In addition to providing a unique tool for monitoring FA in living animals, these data establish a general approach for translatable detection of FA and other reactive biological analytes in vivo by exploiting the widely-available clinical [$^{18}$F]FDG tracer as a masked aldehyde that can be caged by analyte-responsive triggers.

Introduction

Reaction-based chemical probes for selective and non-invasive molecular imaging of biologically important species have attracted significant attention. By utilizing biocompatible chemical transformations, a variety of small-molecule reagents have been developed to detect a diverse range of analytes in living systems.1–5 Among the many non-invasive molecular imaging techniques, fluorescence is currently the most well studied modality, particularly at the cellular level, owing to its high spatiotemporal resolution, high sensitivity, relative simplicity and the widespread use of confocal and other light microscopy. However, in part because of relatively poor tissue penetration, in vivo imaging with the fluorescence modality has had limited clinical translation compared to positron emission tomography (PET), which has been widely applied to oncology, neurology, cardiology and pharmacokinetic studies.6 As such, new chemical strategies for designing functional PET imaging agents for in vivo use are of interest, and in this context, reaction-based PET probes remain largely undeveloped compared to radiolabeled ligands for receptors and other biomolecular targets.

One design strategy for bioanalyte sensing using PET relies on caging a clinically utilized PET tracer, as an analogy to reaction-based fluorescent probes that un cage useful dyes for light microscopy. In the presence of a specific bioanalyte, the caged species is degraded to the parent tracer, which can subsequently be trapped and accumulated in adjacent cells. We have recently employed this approach with success for PET-based monitoring of hydrogen peroxide7 and acidic pH.8 In view of the synthetic ease and wide availability of [$^{18}$F]fluorodeoxyglucose ($^{18}$F-FDG), the most commonly used PET tracer, we decided to pursue $^{18}$F-FDG as a general platform for developing reaction-based PET probes. In particular, we recognized that $^{18}$F-FDG could be thought of as a latent masked aldehyde and reasoned that the aldehyde group of this open-chain form of $^{18}$F-FDG could be converted to a reactive trigger through suitable chemical modification, which can selectively respond to the bioanalytes of interest and release parent $^{18}$F-FDG. Thus, the engineered $^{18}$F-FDG could be used as a reaction-based PET probe (Scheme 1).

To illustrate this concept with a representative example, we targeted the detection of formaldehyde (FA), a reactive carbonyl species (RCS) involved in a diverse array of processes related to...
in cells upon cleavage of an FA-sensitive moiety and can be used to image FA levels within tumor xenografts in living animals.

**Result and discussion**

**Design and synthesis of [18F]FAC-FDG-1 and [18F]Ctrl-FAC-FDG-1**

We envisioned that the acyclic aldehyde form of [18F]FDG could be masked by a homoallylic amine, which would release the parent [18F]FDG tracer upon condensation with FA (Fig. 1). This glucose analogue is then transported into cells via the glucose transporter (GLUT) and subsequently phosphorylated by hexokinase (HK) resulting in its metabolic trapping. Indeed, the widespread availability of [18F]FDG has led to its use in a variety of FDG derivatives bearing stable linkages. Based on these considerations we prepared [18F]FAC-FDG-1 (Fig. 1), noting that accumulation of intracellular [18F]FDG could result from either extracellular reaction-immolation of [18F]FAC-FDG-1 into [18F]FDG followed by GLUT transport or via passive diffusion of [18F]FAC-FDG-1 into cells and subsequent intracellular reaction with FA to generate [18F]FDG. In both cases, [18F]FDG would undergo phosphorylation by HK, resulting in trapped radiotracer and an accumulation in signal within cells with elevated levels of extracellular and/or intracellular FA.

Scheme 2 outlines the synthesis of FAC-FDG-1 via aminolysis of [18F]FADG with adamantaneamethyl amine and pinacol allylboration. We reasoned that an adamantyl functionality would increase cell permeability. We also designed and synthesized the control probe Ctrl-FAC-FDG-1, which is identical to FAC-FDG-1 except for an ethyl group on the amine, rendering Ctrl-FAC-FDG-1 unable to condense with FA. Ctrl-FAC-FDG-1 was synthesized via reductive ethylation of FAC-FDG-1 with acetaldheyde. [18F]FAC-FDG-1 and [18F]Ctrl-FAC-FDG-1 were obtained in a 45 ± 13% (n = 6) and 14 ± 6% (n = 3) decay corrected radiochemical yields, respectively.

**Fig. 1** [18F]FAC-FDG-1, a PET tracer designed to exhibit FA-dependent cellular accumulation of [18F]FDG via aza-Cope uncaging of its masked aldehyde functionality.
Response and selectivity

With these probes in hand, we then evaluated the reactivity of $[^{18}\text{F}]$FAC-FDG-1 with FA and a variety of reactive carbonyl species (RCS) by monitoring its conversion to $[^{18}\text{F}]$FDG using radio-HPLC (Fig. 2). In the presence of 1 mM FA under simulated physiological conditions, (20 mM, pH = 7.4 PBS), consumption of $[^{18}\text{F}]$FAC-FDG-1 with concomitant formation of $[^{18}\text{F}]$FDG was observed (Fig. S1†), leading to 43% and 76% conversions to product within 1 and 2 hours, respectively. In control experiments, no $[^{18}\text{F}]$FDG formation was observed in the absence of FA or upon treatment of $[^{18}\text{F}]$Ctrl-FAC-FDG-1 with FA under the same conditions. Moreover, $[^{18}\text{F}]$FAC-FDG-1 shows high selectivity for FA over other potentially competing species, including acetaldehyde, glucose, sodium pyruvate, benzaldehyde, methylglyoxal, dehydroascorbic acid, glucosone, and hydrogen peroxide (Fig. 2). $[^{18}\text{F}]$FAC-FDG-1 shows a small change in FA levels using PC3 prostate cancer and U87-MG glioblastoma cells, as these cell lines exhibit high FDG avidity. $[^{18}\text{F}]$FAC-FDG-1 responses to added FA concentrations ranging from 0–1000 μM showed a FA dose-dependent (Fig. 3a) and time-dependent accumulation in cells (Fig. 3b), with a 4.4 fold increase in signal from 1.3 ± 0.2% cell associated activity at 0 μM FA to 5.7 ± 0.4% cell associated activity at 1000 μM FA at 1 h. Similarly, in U87-MG cancer cells, a 5.5-fold increase in signal was observed (Fig. S2 and S3†). The ctrl experiments showed that uptake of FDG in the same cell lines was not affected by varying FA concentrations (Fig. S5†). Moreover, $[^{18}\text{F}]$Ctrl-FAC-FDG-1 did not exhibit a significant change in accumulation at 1 h ± 1 1000 μM FA (Fig. S4†). Also, at 1 h with 1000 μM FA, cell uptake of $[^{18}\text{F}]$FAC-FDG-1 is effectively blocked by the addition of cytochalasin, a known GLUT inhibitor, suggesting that $[^{18}\text{F}]$ accumulation occurs by GLUT-dependent transport. These data suggest that $[^{18}\text{F}]$FAC-FDG-1 reacts with FA mainly via an extracellular process and the resulting $[^{18}\text{F}]$FDG is transported intracellularly by GLUT and is then trapped by hexokinase.

Cellular FA detection with $[^{18}\text{F}]$FAC-FDG-1

We next tested whether $[^{18}\text{F}]$FAC-FDG-1 could respond to changes in FA levels using PC3 prostate cancer and U87-MG glioblastoma cells, as these cell lines exhibit high FDG avidity. $[^{18}\text{F}]$FAC-FDG-1 responses to added FA concentrations ranging from 0–1000 μM showed a FA dose-dependent (Fig. 3a) and time-dependent accumulation in cells (Fig. 3b), with a 4.4 fold increase in signal from 1.3 ± 0.2% cell associated activity at 0 μM FA to 5.7 ± 0.4% cell associated activity at 1000 μM FA at 1 h. Similarly, in U87-MG cancer cells, a 5.5-fold increase in signal was observed (Fig. S2 and S3†). The ctrl experiments showed that uptake of FDG in the same cell lines was not affected by varying FA concentrations (Fig. S5†). Moreover, $[^{18}\text{F}]$Ctrl-FAC-FDG-1 did not exhibit a significant change in accumulation at 1 h ± 1 1000 μM FA (Fig. S4†). Also, at 1 h with 1000 μM FA, cell uptake of $[^{18}\text{F}]$FAC-FDG-1 is effectively blocked by the addition of cytochalasin, a known GLUT inhibitor, suggesting that $[^{18}\text{F}]$ accumulation occurs by GLUT-dependent transport. These data suggest that $[^{18}\text{F}]$FAC-FDG-1 reacts with FA mainly via an extracellular process and the resulting $[^{18}\text{F}]$FDG is transported intracellularly by GLUT and is then trapped by hexokinase.

In vivo imaging of FA

Finally, we evaluated the ability of $[^{18}\text{F}]$FAC-FDG-1 to image changes in FA levels in vivo using a murine cancer model. Specifically, $[^{18}\text{F}]$ PET imaging was performed 7–8 weeks following implantation of PC3-derived xenograft tumors on the flanks of nu/nu mice. $[^{18}\text{F}]$FAC-FDG-1 shows detectable uptake within the PC3-derived tumor as revealed by $[^{18}\text{F}]$ imaging in living mice (Fig. 4a), and the signal increases upon intratumoral injection of FA (Fig. 4b). As anticipated, the control probe $[^{18}\text{F}]$Ctrl-FAC-FDG-1 does not exhibit significant uptake within tumor, with only hepatobiliary and renal clearance observed (Fig. 4c) and $[^{18}\text{F}]$FDG providing a positive control (Fig. 4d and S7† for biodistribution). Biodistribution analysis of mice treated with $[^{18}\text{F}]$FAC-FDG-1 and imaged establish that $[^{18}\text{F}]$ uptake in the tumor increased from 1.8 ± 0.26 ID% per g to 2.6 ± 0.24 ID% per g after the intratumoral injection of FA ($n = 3$, $p < 0.05$, data were analyzed using unpaired two-tailed Student's t-test; Fig. 4e and S6 for biodistribution in other organs). Taken together, these results demonstrate that $[^{18}\text{F}]$FAC-FDG-1 is a new class of imaging tool for studying biological formaldehyde in vivo.
underway to apply $^{18}$F-FAC-FDG-1 and related reactivity-based imaging probes to various preclinical models, with particular interest in the epigenetic modifications seen in cancer and neurodegeneration. The use of aldehyde-caged $^{18}$F-FDG tracers provides a general synthetic platform for the potential design of a wide variety of responsive molecular imaging probes.

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Notes and references


Conclusions

To close, we have presented the design, synthesis, and cellular and in vivo properties of $^{18}$F-FAC-FDG-1, a unique reactivity-based PET probe for selective imaging of FA in living animals. $^{18}$F-FAC-FDG-1 reacts with FA via a 2-aza-Cope rearrangement to uncage the clinically-used PET tracer $^{18}$F-FDG in a FA-dependent manner, allowing for detection of changes of FA in living cells and animals most likely via an extracellular pathway. While we are encouraged by these proof-of-principle results, potential limitations may include the short $^{18}$F lifetime vs. FA uncaging as well as the short circulation time of the probe, and, therefore, future improvements will seek to improve probe kinetics by tuning the reactive trigger and optimize the pharmacokinetic properties of the probe. Current efforts are...