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Polydopamine coated manganese oxide nanoparticles with ultrahigh relaxivity as nanotheranostic agents for magnetic resonance imaging guided synergistic chemo-/photothermal therapy†

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Mn-based nanoparticles have been regarded as a new class of probes for magnetic resonance imaging (MRI), but their low relaxivity is the major obstacle for applications *in vivo*. Herein, we designed and constructed a multifunctional nanotheranostic (FA-Mn₃O₄@PDA@PEG) for MRI guided combinatorial chemo-/photothermal therapy (PTT) for cancer. The ultrahigh relaxivity of 14.47 mM⁻¹ s⁻¹ makes the nanotheranostic an excellent contrast agent for MRI *in vitro* and *in vivo*, and provides comprehensive information for tumor diagnosis. When irradiated with an 808 nm NIR laser, FA-Mn₃O₄@PDA@PEG exhibits a remarkably improved and synergistic therapeutic effect compared to PTT or chemotherapy alone, providing high therapeutic efficiency and low side effects of drugs. These findings are of great interest and will inspire us to develop highly effective MRI guided synergistic chemo-/photothermal therapy for cancer treatment.

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Introduction

Recent developments in the field of biomedicine have triggered significant research efforts to exploit multifunctional nanocomposites (so-called “theranostics”) that integrate imaging and therapy into a single system for imaging-guided cancer therapy since they play an important role in overcoming the limitations of traditional cancer therapy.¹ Chemotherapy, as a general therapeutic approach for cancer, suffers greatly from limited therapeutic efficacy and severe side effects in patients. Designing multifunctional nanocomposites, which are responsive to external stimuli for on-demand chemotherapy, and are able to perform other therapeutic approaches for combined cancer therapy, is a promising strategy for addressing these problems.² Lately, photothermal therapy (PTT) has received significant interest owing to its high efficiency and selectivity, minimal invasiveness, and favorable biosafety in normal

tissues.³ Importantly, near-infrared (NIR) light could trigger the release of drugs loaded on nanocomposites, and improve the cellular uptake of the drugs. However, commonly used photothermal agents are currently inorganic nanomaterials, such as various gold nanostructures,⁴ copper sulfide nanoparticles,⁵ and carbon nanomaterials,⁶ which may cause long-term toxicity concerns in their further clinical implementation. Polydopamine (PDA), a type of melanin, has been proposed as a new generation of PTT agents with great potential for *in vivo* applications owing to its advantages of good biodegradability, no long-term toxicity, and high photothermal conversion efficiency (~40%).⁷ Along this line, constructing theranostics based on PDA, which not only exhibits a strong photothermal effect, but also provides an active surface for loading aromatic chemotherapy drugs, could effectively improve therapeutic effectiveness and alleviate side effects.

For effective guidance for cancer therapy, an appropriate medical imaging modality is significant for identifying the tumor location and size, monitoring the biodistribution of nanocomposites and assessing the therapeutic efficacy. Magnetic resonance imaging (MRI), as a routine diagnostic tool in clinical medicine, is particularly attractive owing to its advantages of noninvasiveness, high spatial resolution and soft tissue contrast, and three-dimensional imaging.⁸ To compensate for the innate low sensitivity, a variety of contrast agents, especially positive or *T*₁ contrast agents, are explored to increase contrast for obtaining an accurate diagnosis.⁹ Most *T*₁ MRI

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contrast agents adopted currently are based on gadolinium (Gd^{3+}) in the form of paramagnetic chelates or colloidal nanoparticles.¹⁰ Unfortunately, their potential renal toxicity, which is occasionally associated with nephrogenic system fibrosis, has aroused widespread concerns and promoted the search to find alternatives. Manganese (Mn)-based nanoparticles, especially Mn_3O_4 nanoparticles, have been regarded as promising alternatives owing to their lower intrinsic toxicity than Gd^{3+} ions, relatively high electronic spin, and fast water exchange rates. Nevertheless, although different size and shape Mn_3O_4 nanoparticles have been reported by several groups as potential T_1 contrast agents,¹¹ their relaxivities are usually lower than those of commercial Gd-based agents. Therefore, it is highly desirable to design and construct a theranostics nanoplatform based on Mn_3O_4 nanoparticles with high relaxivity for MRI guided synergistic chemo-/photothermal therapy, but it remains a great challenge.¹²

Herein, we designed and synthesized novel multifunctional Mn_3O_4 @PDA core/shell nanocomposites using a water-in-oil microemulsion method, then applied them as MR imaging contrast agents, photothermal agents and drug carriers for combined anticancer therapy (PTT and chemotherapy), which has not been reported until now to our best knowledge (Fig. 1). Biamino polyethylene glycol ($\text{NH}_2\text{--PEG--NH}_2$) was modified on the surface of Mn_3O_4 @PDA for further conjugation with folic acid (FA), improving the ability to target tumors and the stability in physiological conditions. The obtained $\text{FA-Mn}_3\text{O}_4$ @PDA@PEG exhibited ultrahigh longitudinal relaxivity ($14.47 \text{ mM}^{-1} \text{ s}^{-1}$), which is nearly three times higher than that of the commercial Magnevist ($4.96 \text{ mM}^{-1} \text{ s}^{-1}$) and the highest value reported to date over all Mn-based nanoparticles. MRI experiments were employed and the results demonstrated that $\text{FA-Mn}_3\text{O}_4$ @PDA@PEG can serve well as a contrast agent *in vitro* and *in vivo*. Doxorubicin (DOX) as a model anticancer drug was loaded on the surface of PDA *via* $\pi\text{--}\pi$ stacking and hydrogen bonding interactions, and the release could be triggered by NIR light, reducing side effects owing to the on-demand drug release. Moreover, a synergistic effect of combined photothermal therapy and chemotherapy is expected to improve the therapeutic efficiency in both *in vitro* and *in vivo* experiments.

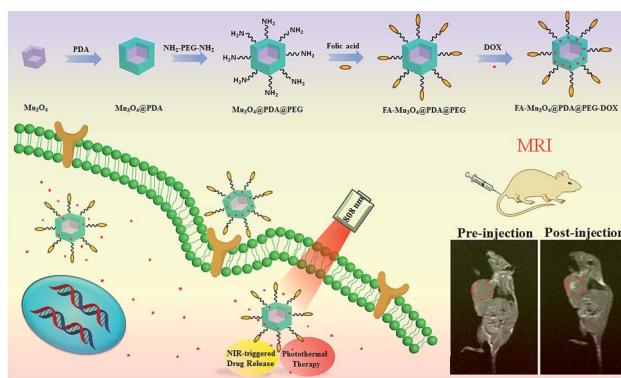


Fig. 1 Schematic illustration of the design and synthesis of core/shell nanotheranostic Mn_3O_4 @PDA for MRI guided synergistic chemo-/photothermal therapy.

Results and discussion

The overall synthetic procedure for $\text{FA-Mn}_3\text{O}_4$ @PDA@PEG is illustrated in Fig. 1. Briefly, Mn_3O_4 nanocrystals were firstly synthesized using a low-temperature method, and the average diameter was determined to be 16.8 nm using transmission electron microscopy (TEM), as presented in Fig. 2a. The crystalline nature of the as-prepared nanocrystals is further confirmed by the corresponding X-ray diffraction (XRD) pattern (Fig. S1, see ESI†), which can be indexed well to the hausmannite Mn_3O_4 (JCPDS 24-0734) phase.¹³ X-ray photoelectron spectroscopy (XPS) was further employed to analyze the Mn oxidation states. As shown in Fig. S2a,† in the survey spectrum, the detected peaks of Mn2p and O1s confirm the presence of Mn and O elements in the sample. The high-resolution Mn2p3/2 spectrum (Fig. S2b, see ESI†) can be resolved into two peaks at 641.2 and 642.9 eV, corresponding to the data reported for Mn^{2+} and Mn^{4+} , respectively.¹⁴ Afterwards, Mn_3O_4 nanocrystals were coated with uniform PDA by a water-in-oil microemulsion method. The TEM image in Fig. 2b reveals that the PDA wrapping process on Mn_3O_4 was successful and the thickness of the PDA shell is approximately 4 nm. The XPS spectrum clearly shows the peaks of N1s, confirming the presence of PDA on the Mn_3O_4 (Fig. S3, see ESI†). With the aim of improving the accumulation of nanoparticles in tumors, $\text{NH}_2\text{--PEG--NH}_2$ was modified on the surface of the Mn_3O_4 @PDA through a reaction between the amine group of PEG and PDA, and then the retained amine groups were available for reacting with FA *via* a carbodiimide conjugation reaction and the ether bond of the PEG chains. The amount of PEG conjugated on the surface of the Mn_3O_4 @PDA was approximately 4%, which was assessed using thermogravimetric analysis (Fig. S4, see ESI†). The successful PEG and FA targeted functionalization is evidenced by Fourier-transform infrared spectroscopy (FT-IR) and zeta potential (Fig. S5 and S6, see ESI†). The as-obtained $\text{FA-Mn}_3\text{O}_4$ @

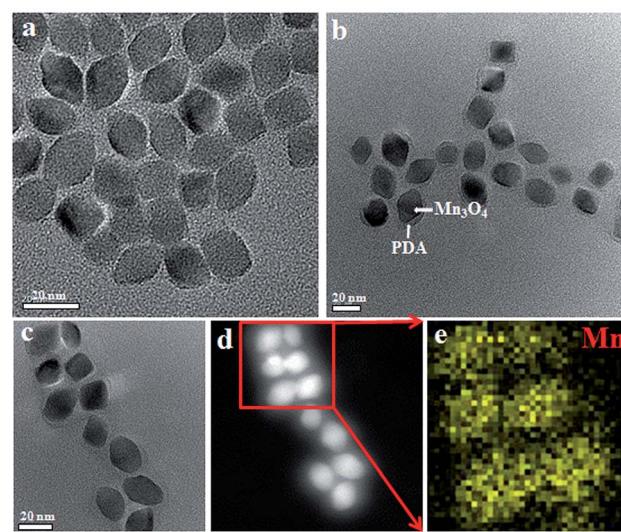


Fig. 2 (a-c) TEM images of Mn_3O_4 , Mn_3O_4 @PDA, and $\text{FA-Mn}_3\text{O}_4$ @PDA@PEG, (d) HAADF-STEM image of $\text{FA-Mn}_3\text{O}_4$ @PDA@PEG, and (e) EDS mapping of Mn according to the image in (d).



PDA@PEG is still uniform in size, and HAADF-STEM and elemental mapping analysis further confirm its core-shell structure (Fig. 2c–e).

The FA-Mn₃O₄@PDA@PEG exhibits excellent colloidal stability in different dispersants including H₂O, PBS, and 0.9% NaCl solution (Fig. S7, see ESI†). Importantly, such FA-Mn₃O₄@PDA@PEG shows strong NIR absorption owing to the oxidation of dopamine and the following self-polymerization process (Fig. S8, see ESI†).^{7a} Then, to investigate the photothermal performance, FA-Mn₃O₄@PDA@PEG aqueous solutions with different concentrations were exposed to an 808 nm NIR laser at 2 W cm⁻² for 420 s. In marked contrast to pure water, FA-Mn₃O₄@PDA@PEG displays effective photothermal heating of the solutions (Fig. 3a). The temperatures of the solutions increase with increasing FA-Mn₃O₄@PDA@PEG concentration or irradiation time. After irradiation for 420 s, a rapid temperature increase of the FA-Mn₃O₄@PDA@PEG solution (Mn content: 200 ppm) is noted from 25 °C to 50 °C (the temperature of efficient killing of cancerous cells). Furthermore, there is no colour change or absorption decrease of the FA-Mn₃O₄@PDA@PEG after irradiation by an 808 nm NIR laser for 1 h (Fig. 3b and S9, see ESI†), indicating that FA-Mn₃O₄@PDA@PEG has high photostability. Such high photothermal efficiency and photostability makes FA-Mn₃O₄@PDA@PEG superior as a promising PTT agent.

Along with the capability of acting as a photothermal agent for PTT of cancer, FA-Mn₃O₄@PDA@PEG can load therapeutic molecules for chemotherapy. We chose an extensively used clinical chemotherapy drug DOX as a model to evaluate the carrying and releasing properties of FA-Mn₃O₄@PDA@PEG. After incubating with FA-Mn₃O₄@PDA@PEG, DOX was loaded

on the surface of FA-Mn₃O₄@PDA@PEG at a content of 0.8014 mg mg⁻¹. Such high loading capacity could be attributed to the strong π–π stacking and hydrogen bond interactions of the PDA shell with DOX since PDA has abundant phenyl, amino, and hydroxyl groups on its surface.¹⁵ The subsequent test of sustained release of the drug under different pH values in Fig. 3c shows that a slow drug release from the obtained FA-Mn₃O₄@PDA@PEG-DOX is detected at pH 7.4. By comparison, the carrier shows a relatively faster release process at pH 5, which contributes to the protonation of the amino group in the DOX molecule that offers DOX a positive charge, thus facilitating drug release under acidic conditions. Importantly, after irradiation under an 808 nm NIR laser (2 W cm⁻², 10 min for each pulse), the release of DOX sharply increases from the FA-Mn₃O₄@PDA@PEG-DOX at pH 5.0 (as shown in Fig. 3d). However, there is only limited DOX release from FA-Mn₃O₄@PDA@PEG-DOX at pH 7.4 under the same conditions. Such pH-dependent NIR-responsive release properties show that FA-Mn₃O₄@PDA@PEG-DOX could be applied as a better on-demand drug delivery system which can enhance antitumor efficacy and minimize side effects of the drug.

Prior to the theranostic application of FA-Mn₃O₄@PDA@PEG, investigation of the cytotoxicity is an important prerequisite, so the cytotoxicity of FA-Mn₃O₄@PDA@PEG was assessed using a traditional MTT assay. Fig. S10† shows the *in vitro* cell viability of human breast cancer cells (MCF-7) incubated with FA-Mn₃O₄@PDA@PEG with different concentrations ranging from 25 to 800 µg mL⁻¹ for 24 h. It can be seen that the cell viabilities are still higher than 88%, even at the highest concentration of 800 µg mL⁻¹. Such findings demonstrate the near non-cytotoxicity *in vitro* for all dosages of FA-Mn₃O₄@PDA@PEG, which implies that FA-Mn₃O₄@PDA@PEG can potentially serve as a theranostic probe for simultaneous MRI and combinatorial PTT and chemotherapy of cancer.

The cellular uptake process and targeting recognition capability of FA-Mn₃O₄@PDA@PEG-DOX were investigated using a confocal laser scanning microscope (CLSM). Fig. 4 shows CLSM images of MCF-7 cells incubated with Mn₃O₄@PDA@PEG-DOX and FA-Mn₃O₄@PDA@PEG-DOX for 1 h and 3 h at 37 °C. Obviously, the stronger red fluorescence from the cells incubated with FA-Mn₃O₄@PDA@PEG-DOX than that from Mn₃O₄@PDA@PEG-DOX can be seen, which indicates that more folate-modified nanocomposites were taken up specifically by MCF-7 cells. These results demonstrate that the as-prepared FA-Mn₃O₄@PDA@PEG-DOX is a promising candidate for targeted imaging and combinatorial PTT and chemotherapy of cancer.

Owing to their intrinsic paramagnetic properties, Mn₃O₄ nanoparticles are potential *T*₁-MRI contrast agents by accelerating the longitudinal relaxation of water protons. To evaluate the capacity of FA-Mn₃O₄@PDA@PEG as a *T*₁-MRI contrast agent for diagnostics, we investigated the magnetic resonance signals of FA-Mn₃O₄@PDA@PEG and Magnevist (a traditional MRI contrast agent currently used in clinic) using a 1.2 T MRI system. Fig. 5a displays the *T*₁-weighted relaxivity of FA-Mn₃O₄@PDA@PEG and shows that the MRI contrast is enhanced by increasing the concentration of Mn. The longitudinal relaxivity

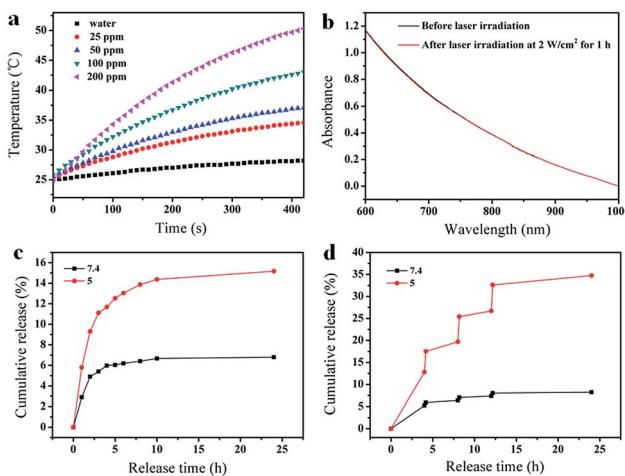


Fig. 3 (a) Temperature elevation of water and FA-Mn₃O₄@PDA@PEG aqueous solutions with different Mn²⁺ concentrations as a function of time under irradiation (808 nm, 2 W cm⁻²), (b) UV-vis absorption spectra of an FA-Mn₃O₄@PDA@PEG dispersion in water before and after laser irradiation for 1 h (808 nm, 2 W cm⁻²), (c) DOX release from FA-Mn₃O₄@PDA@PEG-DOX at pH 7.4 and 5.0 at 37 °C, and (d) NIR-triggered release of DOX from FA-Mn₃O₄@PDA@PEG-DOX at pH 7.4 and 5.0, which were irradiated with an 808 nm NIR laser (2 W cm⁻²) for 10 min at different time points.



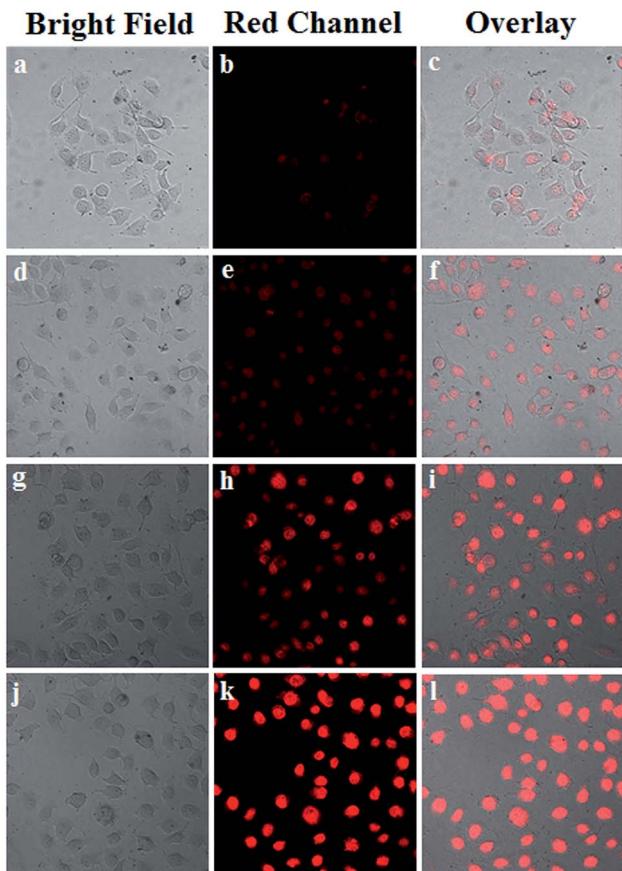


Fig. 4 (a) CLSM images of MCF-7 cells incubated with $\text{Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$ for 1 h (a–c), and 3 h (d–f), and $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$ for 1 h (g–i), and 3 h (j–l) at 37°C .

value (r_1) is calculated to be $14.47 \text{ mM}^{-1} \text{ s}^{-1}$ from the slope of the concentration-dependent relaxation $1/T_1$ (Fig. 5b). This value is nearly three times higher than that of Magnevist ($4.96 \text{ mM}^{-1} \text{ s}^{-1}$) under the same testing conditions, which may be ascribed to the greater number of free sites of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ for water ligation. Most importantly, it is the highest value reported to date for Mn-based NPs (Table S1, see ESI†), which reduces the concentration required for contrast agent detection by MRI. This result further confirms that $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ could operate as an excellent positive MRI contrast agent.

To assess the *in vivo* MR imaging, we intratumorally injected $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ into a tumor-bearing mouse, and used a 3.0 T human MRI scanner. As shown in Fig. 5c, the significantly enhanced MRI signal can be clearly observed in the injection area. Then, $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ was intravenously administered into a tumor-bearing mouse to further evaluate its targeting MRI capacity *in vivo*. As shown in Fig. 5d, different from the homogeneous and slightly dark image before injection, the whole tumor area becomes brighter after 36 h injection. Such results demonstrate that a large amount of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ accumulates in the tumor owing to the active targeting ability of the FA and the enhanced permeability and retention (EPR) effect occurring in the vessels of the cancer

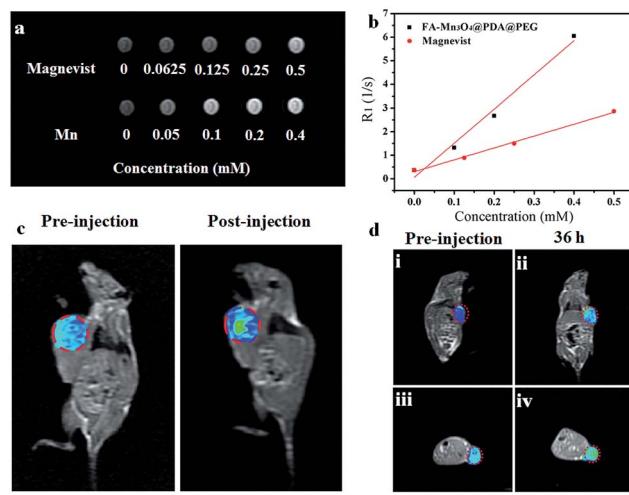


Fig. 5 (a) *In vitro* T_1 -weighted MRI maps of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ and commercial Magnevist with varied concentrations of $\text{Mn}^{2+}/\text{Gd}^{3+}$, (b) the relaxivity (r_1) of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ and commercial Magnevist, (c) T_1 -weighted MR images of a tumor-bearing Balb/c mouse: pre-injection and after injection *in situ*, and (d) *in vivo* T_1 -weighted MR images of a tumor-bearing Balb/c mouse before and after intravenous injection of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$. The red circles indicate the tumor sites.

tissues. Importantly, the enhancement of the MRI signals in the tumor is capable of continuing for 36 hours, which provides adequate time for guiding the following treatment. The active targeting and high MRI contrast ability make $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$ a promising candidate for accurate cancer diagnosis and locating the tumor site to guide the external NIR laser irradiation for photothermal therapy and controlling the drug release on-demand.

Inspired by the properties of high photothermal efficiency and NIR triggered controllable drug release, we further investigated the antitumor effects of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$ *in vitro* and *in vivo*. MCF-7 cells were treated with free DOX, $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}}$, and $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$ for 10 min with or without NIR laser irradiation. As shown in Fig. 6a, after exposure to the NIR laser for 10 min, $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$ exhibits the highest statistically cytotoxic effect at all the tested concentrations compared to the chemotherapy and photothermal therapy alone. For example, when DOX concentration is $20 \text{ }\mu\text{g mL}^{-1}$, the cell viability for $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}} + \text{NIR}$ is remarkably reduced to 11%, which is evidently lower than that of free DOX, $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$ alone, and $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG}} + \text{NIR}$. The remarkably improved therapeutic effect may be attributed to the photothermal effect, which can not only kill the cancer cells but also effectively enhance the sensitivity of the delivery and release of DOX into cells for improved chemotherapy for cancer.

To shed more light on the combined therapeutic effects of $\text{FA-Mn}_3\text{O}_4@\text{PDA}@{\text{PEG-DOX}}$, we further investigated the effectiveness of tumor inhibition *in vivo* using H22 tumor-bearing mice. The H22 (murine hepatocarcinoma) xenograft model was established by injecting H22 cancer cells into the right axilla of

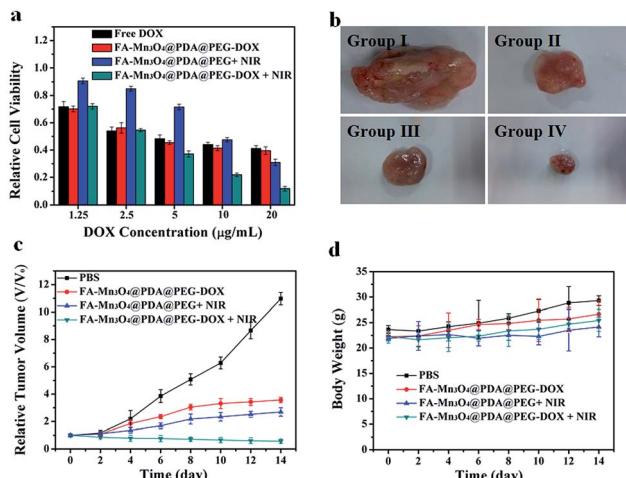


Fig. 6 (a) Cell viability of MCF-7 cells incubated with free DOX, FA- Mn_3O_4 @PDA@PEG-DOX, FA- Mn_3O_4 @PDA@PEG + NIR, and FA- Mn_3O_4 @PDA@PEG-DOX + NIR. (b) Photographs of excised tumors from representative euthanized mice. (c) Relative tumor volume and (d) body weight of mice after treatment with a PBS solution as control, FA- Mn_3O_4 @PDA@PEG-DOX, FA- Mn_3O_4 @PDA@PEG + NIR, and FA- Mn_3O_4 @PDA@PEG-DOX + NIR.

female Balb/c mice. The mice bearing tumors were randomly divided into four groups (group I to IV) and treated with PBS (as control), FA- Mn_3O_4 @PDA@PEG-DOX, FA- Mn_3O_4 @PDA@PEG + NIR irradiation, and FA- Mn_3O_4 @PDA@PEG-DOX + NIR irradiation, respectively. The tumor dimensions were tracked every 2 days with a caliper for 14 days. After treatment, the tumors were isolated from the different groups of mice and weighed. As shown in Fig. 6b and c, the tumor growth of group II and group III was only slightly delayed compared to group I, indicating that either chemotherapy or photothermal therapy by itself exhibited only partial inhibitory effects. In contrast, when the mice were injected with FA- Mn_3O_4 @PDA@PEG-DOX and then exposed to 808 nm NIR laser triggered photothermal therapy and drug release, the tumor growth was remarkably inhibited with tumor growth inhibition (TGI) of 94.81%, demonstrating that combination chemotherapy and PTT exhibited the optimal therapeutic efficacy. Moreover, the body weight of all groups did not decrease with the time prolonged, indicating no significant acute toxicity of our theranostic agent. Such results clearly confirm that FA- Mn_3O_4 @PDA@PEG-DOX has potential applications for *in vivo* combinatorial chemotherapy and PTT for cancer.

Conclusions

In summary, a multifunctional core/shell FA- Mn_3O_4 @PDA@PEG nanotheranostic has been successfully constructed for MRI guided combinatorial chemo-/photothermal therapy for cancer. The ultrahigh relaxivity of $14.47 \text{ mM}^{-1} \text{ s}^{-1}$ makes the nanotheranostic an excellent contrast agent for MRI *in vitro* and *in vivo*, and provides comprehensive information for tumor diagnosis. The PDA shell not only is employed as a high photothermal conversion agent for PTT and aromatic anticancer drug carrier, but also endows the nanotheranostic with robust

biocompatibility owing to its natural characteristics. Moreover, the PTT and on-demand drug release are simultaneously triggered by 808 nm NIR laser irradiation, which effectively enhances the anticancer effect and reduces adverse side effects of drugs. Compared with chemotherapy or photothermal treatment alone, the combined treatment shows a synergistic effect, resulting in higher therapeutic efficacy for *in vitro* and *in vivo* cancer therapy. Therefore, this multifunctional nanocomposite is expected to be employed as a powerful theranostics platform for MRI guided therapeutics allied with significant therapeutic effectiveness but low side effects in future oncotherapy.

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Notes and references

- (a) F. Greco and M. Vicent, *Adv. Drug Delivery Rev.*, 2009, **61**, 1203; (b) D. Lane, *Nat. Biotechnol.*, 2006, **24**, 163.
- (a) J. Liu, C. Wang, X. Wang, X. Wang, L. Cheng, Y. Li and Z. Liu, *Adv. Funct. Mater.*, 2015, **25**, 384; (b) H. Gong, L. Cheng, J. Xiang, H. Xu, L. Feng, X. Shi and Z. Liu, *Adv. Funct. Mater.*, 2013, **23**, 6059; (c) H. Y. Liu, D. Chen, L. L. Li, T. L. Liu, L. F. Tan, X. L. Wu and F. Q. Tang, *Angew. Chem., Int. Ed.*, 2011, **50**, 891.
- (a) S. Lal, S. E. Clare and N. J. Halas, *Acc. Chem. Res.*, 2008, **41**, 1842; (b) J. T. Robinson, S. M. Tabakman, Y. Liang, H. Wang, H. S. Casalongue, D. Vinh and H. Dai, *J. Am. Chem. Soc.*, 2011, **133**, 6825.
- (a) Y. Xia, W. Li, C. M. Cobley, J. Chen, X. Xia, Q. Zhang, M. Yang, E. C. Cho and P. K. Brown, *Acc. Chem. Res.*, 2011, **44**, 914; (b) L. Dykman and N. Khlebtsov, *Chem. Soc. Rev.*, 2012, **41**, 2256; (c) E. Boisselier and D. Astruc, *Chem. Soc. Rev.*, 2009, **38**, 1759.
- (a) Q. Tian, M. Tang, Y. Sun, R. Zou, Z. Chen, M. Zhu, S. Yang, J. Wang and J. Hu, *Adv. Mater.*, 2011, **23**, 3542; (b) Q. Tian, F. Jiang, R. Zou, Q. Liu, Z. Chen, M. Zhu, S. Yang, J. Wang, J. Wang and J. Hu, *ACS Nano*, 2011, **5**, 9761.
- (a) K. Yang, S. Zhang, G. Zhang, X. Sun, S.-T. Lee and Z. Liu, *Nano Lett.*, 2010, **10**, 3318; (b) K. Yang, L. Feng, X. Shi and Z. Liu, *Chem. Soc. Rev.*, 2013, **42**, 530; (c) C. Li, S. Bolisetty, K. Chaitanya, J. Adamek and R. Mezzenga, *Adv. Mater.*, 2013, **25**, 1010.

7 (a) Y. Liu, K. Ai, J. Liu, M. Deng, Y. He and L. Lu, *Adv. Mater.*, 2013, **25**, 1353; (b) J. Stritzker, L. Kirscher, M. Scadeng, N. C. Deliolanis, S. Morscher, P. Symvoulidis, K. Schaefer, Q. Zhang, L. Buckel, M. Hess, U. Donat, W. G. Bradley, V. Ntziachristos and A. A. Szalay, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 3316; (c) Y. Gao, X. Wu, L. Zhou, Y. Su and C. M. Dong, *Macromol. Rapid Commun.*, 2015, **36**, 916.

8 (a) J. A. Feshitan, F. Vlachos, S. R. Sirsi, E. E. Konofagou and M. A. Borden, *Biomaterials*, 2012, **33**, 247; (b) X. Q. Yang, J. J. Grailer, I. J. Rowland, A. Javadi, S. A. Hurley, V. Z. Matson, D. A. Steeber and S. Q. Gong, *ACS Nano*, 2010, **4**, 6805; (c) C. Sanson, O. Diou, J. Thevenot, E. Ibarboure, A. Soum, A. Brulet, S. Miraux, E. Thiaudiere, S. Tan, A. Brisson, V. Dupuis, O. Sandre and S. Lecommandoux, *ACS Nano*, 2011, **5**, 1122; (d) J. S. Basuki, L. Esser, H. T. T. Duong, Q. Zhang, P. Wilson, M. R. Whittaker, D. M. Haddleton, C. Boyer and T. P. Davis, *Chem. Sci.*, 2014, **5**, 715.

9 (a) S. H. Wang, X. Shi, M. Van Antwerp, Z. Cao, S. D. Swanson, X. Bi and J. R. Baker, *Adv. Funct. Mater.*, 2007, **17**, 3043; (b) X. Shi, S. H. Wang, S. D. Swanson, S. Ge, Z. Cao, M. E. Van Antwerp, K. J. Landmark and J. R. Baker, *Adv. Mater.*, 2008, **20**, 1671; (c) H. Cai, K. Li, M. Shen, S. Wen, Y. Luo, C. Peng, G. Zhang and X. Shi, *J. Mater. Chem.*, 2012, **22**, 15110.

10 (a) P. Caravan, *Chem. Soc. Rev.*, 2006, **35**, 512; (b) P. Caravan, J. J. Ellison, T. J. McMurry and R. B. Lauffer, *Chem. Rev.*, 1999, **99**, 2293; (c) R. E. Lauffer, *Chem. Rev.*, 1987, **87**, 901; (d) L. M. Randolph, C. L. M. LeGuyader, M. E. Hahn, C. M. Andolina, J. P. Patterson, R. F. Mattrey, J. E. Millstone, M. Botta, M. Scadeng and N. C. Gianneschi, *Chem. Sci.*, 2016, **7**, 4230.

11 (a) C. C. Huang, N. H. Khu and C. S. Yeh, *Biomaterials*, 2010, **31**, 4073; (b) J. Xiao, X. M. Tian, C. Yang, P. Liu, N. Q. Luo, Y. Liang, H. B. Li, D. H. Chen, C. X. Wang, L. Li and G. W. Yang, *Sci. Rep.*, 2013, **3**, 3424; (c) K. An, M. Park, J. H. Yu, H. B. Na, N. Lee, J. Park, S. H. Choi, I. C. Song, W. K. Moon and T. Hyeon, *Eur. J. Inorg. Chem.*, 2012, 2148; (d) T. Yu, J. Moon, J. Park, Y. Park II, H. B. Na, B. H. Kim, I. C. Song, W. K. Moon and T. Hyeon, *Chem. Mater.*, 2009, **21**, 2272; (e) J. Shin, R. M. Anisur, M. K. Ko, G. H. Im, J. H. Lee and I. S. Lee, *Angew. Chem., Int. Ed.*, 2009, **48**, 321; (f) T. L. Ha, H. J. Kim, J. Shin, G. H. Im, J. W. Lee, H. Heo, J. Yang, C. M. Kang, Y. S. Choe, J. H. Lee and I. S. Lee, *Chem. Commun.*, 2011, **47**, 9176.

12 (a) M. Nafiujiyaman, M. Nurunnabi, S. Kang, G. R. Reeck, H. A. Khand and Y. Lee, *J. Mater. Chem. B*, 2015, **3**, 5815; (b) H. Hu, A. Dai, J. Sun, X. Li, F. Gao, L. Wu, Y. Fang, H. Yang, L. An, H. Wu and S. Yang, *Nanoscale*, 2013, **5**, 10447; (c) R. Savla, O. B. Garbuzenko, S. Chen, L. Rodriguez-Rodriguez and T. Minko, *Pharm. Res.*, 2014, **31**, 3487.

13 Y. Tan, C. Xu, G. Chen, X. Fang, N. Zheng and Q. Xie, *Adv. Funct. Mater.*, 2012, **22**, 4584.

14 (a) V. Di Castro and G. Polzonetti, *J. Electron Spectrosc. Relat. Phenom.*, 1989, **48**, 117; (b) S. Ardizzone, C. L. Bianchi and D. Tirelli, *Colloids Surf. A*, 1998, **134**, 305; (c) K. J. Kim, M. S. Park, J. H. Kim, U. Hwang, N. J. Lee, G. Jeong and Y. J. Kim, *Chem. Commun.*, 2012, **48**, 5455.

15 F. Liu, X. He, Z. Lei, L. Liu, J. Zhang, H. You, H. Zhang and Z. Wang, *Adv. Healthcare Mater.*, 2015, **4**, 559.

