

CrossMark  
click for updatesCite this: *Chem. Sci.*, 2016, 7, 430

# Deacetylation of mycothiol-derived 'waste product' triggers the last biosynthetic steps of lincosamide antibiotics†

Zdenek Kamenik,<sup>‡a</sup> Stanislav Kadlcik,<sup>a</sup> Bojana Radojevic,<sup>‡a</sup> Petra Jiraskova,<sup>a</sup> Marek Kuzma,<sup>a</sup> Radek Gazak,<sup>a</sup> Lucie Najmanova,<sup>a</sup> Jan Kopecky<sup>b</sup> and Jiri Janata<sup>\*a</sup>

The immediate post-condensation steps in lincomycin biosynthesis are reminiscent of the mycothiol-dependent detoxification system of actinomycetes. This machinery provides the last proven lincomycin intermediate, a mercapturic acid derivative, which formally represents the 'waste product' of the detoxification process. We identified and purified new lincomycin intermediates from the culture broth of deletion mutant strains of *Streptomyces lincolnensis* and tested these compounds as substrates for proteins putatively involved in lincomycin biosynthesis. The results, based on LC-MS, in-source collision-induced dissociation mass spectrometry and NMR analysis, revealed the final steps of lincomycin biosynthesis, *i.e.* conversion of the mercapturic acid derivative to lincomycin. Most importantly, we show that deacetylation of the *N'*-acetyl-*S*-cysteine residue of the mercapturic acid derivative is required to 'escape' the detoxification-like system and proceed towards completion of the biosynthetic pathway. Additionally, our results, supported by L-cysteine-<sup>13</sup>C<sub>3</sub>, <sup>15</sup>N incorporation experiments, give evidence that a different type of reaction catalysed by the homologous pair of pyridoxal-5'-phosphate-dependent enzymes, LmbF and CcbF, forms the branch point in the biosynthesis of lincomycin and celesticetin, two related lincosamides.

Received 3rd September 2015  
Accepted 30th September 2015

DOI: 10.1039/c5sc03327f

www.rsc.org/chemicalscience

## Introduction

Zhao *et al.* recently discovered the intriguing participation of two low-molecular-weight thiols, mycothiol (MSH) and ergothioneine, in the biosynthesis of the lincosamide antibiotic, lincomycin A (hereinafter lincomycin or LIN; Fig. 1).<sup>1</sup> Such unusual involvement of these actinomycete thiols (Fig. 2B) in biosynthesis had remained undiscovered for a long time,

mainly because the sulfur atom of MSH remains the only footprint of the thiols in the lincomycin structure. Ergothioneine is, however, required for condensation of the amino acid and amino octose unit to obtain the scaffold of lincomycin, whereas MSH plays its role immediately after the condensation. In the condensation process, the stand-alone adenylation domain LmbC catalyses 4-propyl-L-proline activation and transfer on the *holo*-form of the carrier protein domain of LmbN,<sup>2,3</sup> whereas LmbT catalyses the formation of the octose conjugate with ergothioneine.<sup>1</sup> This conjugate is then condensed with the activated 4-propyl-L-proline *via* an amide bond in a reaction catalysed by LmbD (Fig. 2A–C).<sup>1</sup> Interestingly, the following post-condensation steps are at least formally reminiscent of the MSH-dependent detoxification system present in actinomycetes, which is generally used for the elimination of electrophilic toxins, including various antibiotics and their metabolites, from the cell.<sup>4</sup> Following this parallel, ergothioneine is from the *S*-conjugated condensation product replaced for MSH by nucleophilic substitution in a reaction catalysed by LmbV.<sup>1</sup> The resulting product corresponds to a 'toxin'–MSH conjugate, where the lincomycin scaffold formally represents the 'toxin'.

Subsequently, LmbE cleaves off the pseudo-disaccharide 1-*O*-glucosamine-*D*-myo-inositol, resulting in compound **507**, a mercapturic acid derivative characterized by the *N'*-acetyl-*S*-cysteine residue, the only remainder of MSH in its structure

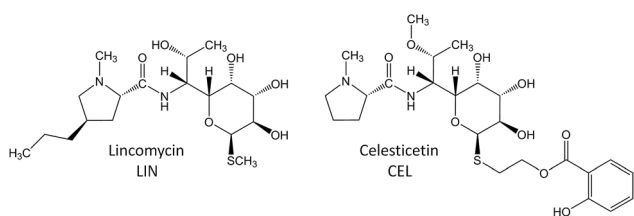


Fig. 1 Structures of the main natural lincosamide antibiotics.

<sup>a</sup>Institute of Microbiology ASCR, Videnska 1083, Prague 4, Czech Republic. E-mail: janata@biomed.cas.cz

<sup>b</sup>Crop Research Institute, Drnovska 507, Prague 6, Czech Republic

† Electronic supplementary information (ESI) available: Experimental procedures, additional figures, chromatograms and NMR data. See DOI: 10.1039/c5sc03327f

‡ These authors contributed equally.

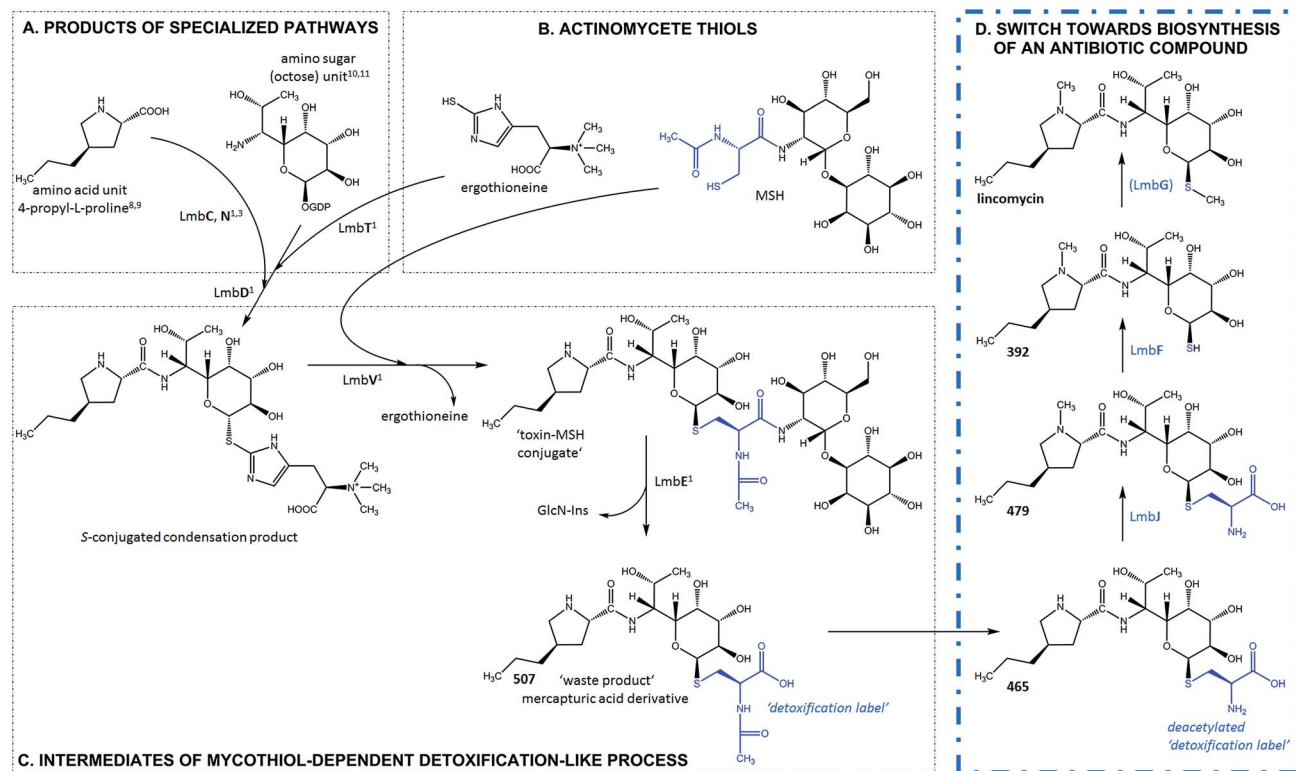


Fig. 2 Lincomycin biosynthesis. Condensation and MSH-dependent detoxification-like process ((A–C), grey frame – evidenced previously<sup>1,3,8–11</sup>); switch towards biosynthesis of an antibiotic compound ((D), blue frame – evidenced in this paper). The blue structural features highlight the *N'*-acetyl-*S*-cysteine residue of MSH and its processing during lincomycin maturation. GlcN-Ins – 1-*O*-glucosamine-*D*-myo-inositol; MSH – mycothiol; (LmbG) – the assignment of the catalysing enzyme based on bioinformatic analysis.

(Fig. 2C).<sup>1</sup> The structure of **507** corresponds to the 'waste product' of the MSH-dependent detoxification system, where mercapturic acid derivatives are generally predetermined for transport out of the cell.<sup>4,5</sup> Thus, the *N'*-acetyl-*S*-cysteinyl of the mercapturic acid derivative may be considered as a 'detoxification label'. Interestingly, the seeming 'waste product' **507** is further matured to give the antibiotic product lincomycin. When comparing the structures of **507** and lincomycin, it is obvious that the *N'*-acetyl-*S*-cysteinyl 'detoxification label' of **507** has to be removed and that two methylations at the nitrogen and sulfur atoms have to occur to give rise to lincomycin. The *N*-methylation has been demonstrated to be catalysed by LmbJ in the biosynthesis of lincomycin and its homologue CcbJ in the biosynthesis of the related lincosamide, celesticetin (Fig. 1).<sup>6</sup> Moreover, the crystal structure of the homohexamer CcbJ has been solved, and the detailed reaction mechanism has been proposed.<sup>7</sup> However, the mechanism that switches the machinery from the 'detoxification' mode to the biosynthetic mode and how the final biosynthetic steps proceed remains elusive.

Here, we demonstrate that the acetyl group of the *N'*-acetyl-*S*-cysteinyl 'detoxification label' represents a locked door for further maturation of the 'waste product' **507** into the antibiotic lincomycin. Furthermore, we present the complete processing of the 'detoxification label' so that only its sulfur atom remains in the structure of lincomycin. In addition, we discuss apparent

differences in the final steps of lincomycin and celesticetin biosynthesis.

## Results and discussion

### New intermediates of lincomycin biosynthesis reveal conversion of 'waste product' **507** to antibiotic lincomycin

We have detected four intermediates of lincomycin biosynthesis in the culture broths of two mutant strains of the lincomycin producer, *Streptomyces lincolnensis*, with specific genes deleted (see Experimental in ESI†). Apart from the known intermediate **507** (Fig. 2C), three new hypothetical intermediates, **465**, **479** (Fig. 2D) and **521**, were found (compounds are named according to their nominal mass). The compounds mutually differ in the presence of the *N*-methyl group in their structures (present only in **479** and **521**) and in the presence of the intact *N'*-acetyl-*S*-cysteinyl 'detoxification label' (present only in **507** and **521**, whereas **465** and **479** have the 'detoxification label' deacetylated); see the graphic cartoon in Fig. 3 for the structural differences. We identified the compounds based on high resolution mass spectrometry (MS) and in-source collision-induced dissociation MS fragmentation and purified them from culture broths (Fig. S1†). The structures were further elucidated by NMR analysis (NMR data in Table S2,† <sup>1</sup>H and <sup>13</sup>C NMR spectra in Fig. S2†) and by incorporation experiments with *L*-cysteine-<sup>13</sup>C<sub>3</sub>, <sup>15</sup>N, which confirmed that *L*-cysteine is part of the



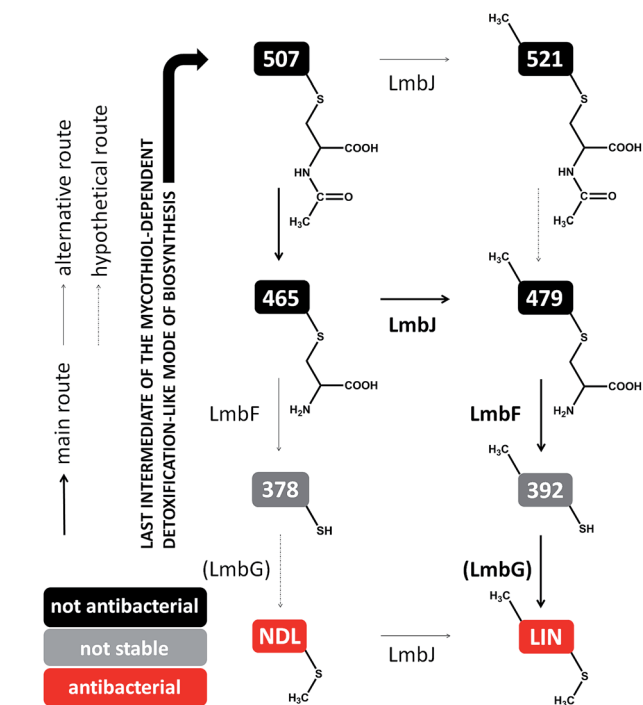


Fig. 3 Post-condensation steps of lincomycin biosynthesis leading from 507 (exhibiting no antimicrobial activity) to the final antibiotic.

molecular structure of the four intermediates (details in Fig. S3†).

The compounds were then tested *in vitro* as substrates of the recombinant proteins LmbJ and LmbF to interrogate the pathway from 507 to lincomycin (the purity and integrity can be assessed from SDS-PAGE gels in Fig. S4†). The cartoon in Fig. 3 summarizes the results presented and is discussed in more detail below. Specifically, we demonstrate that the main biosynthetic route proceeds as follows: first, compound 507, the proven product of the LmbE reaction,<sup>1</sup> is deacetylated by a yet unassigned enzyme to yield 465, then *N*-methylated by LmbJ to yield 479, which is subsequently converted into 392 by LmbF and finally *S*-methylated, presumably by LmbG, to give lincomycin. Additionally, we show that the relaxed substrate specificities of biosynthetic proteins enable either documented alternative or hypothetical bypasses (see Fig. 2D for the main stream; cartoon in Fig. 3 for all possible streams).

### Ornamentation by LmbJ *N*-methyltransferase is not the last biosynthetic step

In previous work by Najmanova *et al.*,<sup>6</sup> it has been shown that LmbJ *N*-methylates the chemically synthesized *N*-demethylincomycin (NDL), suggesting that *N*-methylation of the 4-propyl-L-proline moiety is the last step in lincomycin biosynthesis. However, the existence of the biosynthesized *N*-methylated intermediates 521 and 479 raises questions about whether the *N*-methylation occurs at an earlier stage. Indeed, we tested recombinant LmbJ with 507, 465 and NDL and revealed that all of these substrates can be converted into the *N*-methylated products 521, 479 and lincomycin, respectively (see Fig. 3 for the

scheme and Fig. S5† for the LC-MS data). However, the rapid and preferential conversion of 465 but not 507 or NDL in a competitive *in vitro* assay strongly suggests that 465 is the main natural substrate of LmbJ (Fig. 4). This finding also provides an explanation of the previously observed similar kinetic parameters of the homologous *N*-methyltransferases LmbJ and CcbJ for the conversion of NDL into lincomycin:<sup>6</sup> NDL is not the natural substrate for any of these two proteins. Moreover, these results confirm the broad substrate specificity of LmbJ, as predicted.<sup>7</sup>

### 'Detoxification label' is removed by LmbF only if the 'label' is first deacetylated

The hypothetical lincomycin intermediates 465 and 479 have their 'detoxification label' deacetylated, suggesting that deacetylation is involved in lincomycin maturation. Because we detected 479 as the major intermediate in the mutant strain of *S. lincolnensis* with an inactive *lmbF* gene, we considered that LmbF played a role in the subsequent conversion. Indeed, we detected significant consumption of 479 in the *in vitro* assay with recombinant LmbF (Fig. 5). The sequence analysis of LmbF shows that it belongs to the AAT\_I (aspartate aminotransferase, fold type I) superfamily of enzymes, which employs pyridoxal-5'-phosphate (PLP) as a cofactor. Since a typical reaction for PLP-mediated turnovers is transamination (and with respect to celastrol biosynthesis, as explained in the later text), we expected that LmbF would replace the primary amino group of 479 with the oxo functional group. Surprisingly, we did not detect a product corresponding to a simple transamination reaction. Instead, we found compound 392 (see Fig. 2D and 3) to be the product, which means that LmbF is responsible for the removal of the whole *S*-cysteine residue, *i.e.*, the complete deacetylated 'detoxification label' except for the sulfur atom (see LC-MS data in Fig. 5 and S6†). LmbF failed to carry out this

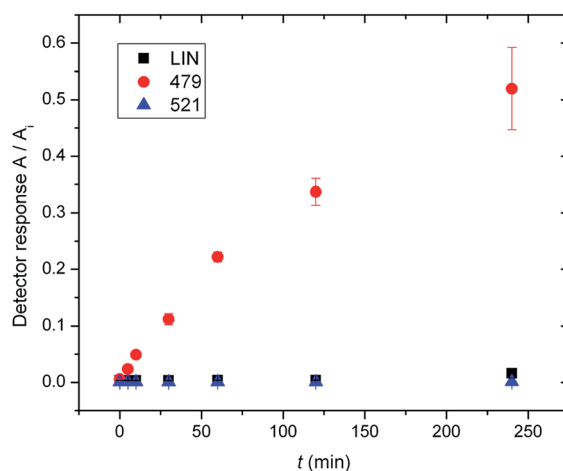


Fig. 4 Formation of 521, 479 and LIN products from 507, 465 and NDL substrates, respectively, in a competitive *in vitro* assay with LmbJ. Equimolar amounts of substrates were tested simultaneously in one reaction;  $n = 3$ .  $A$  – peak area of the product,  $A_i$  – peak area of the internal standard.

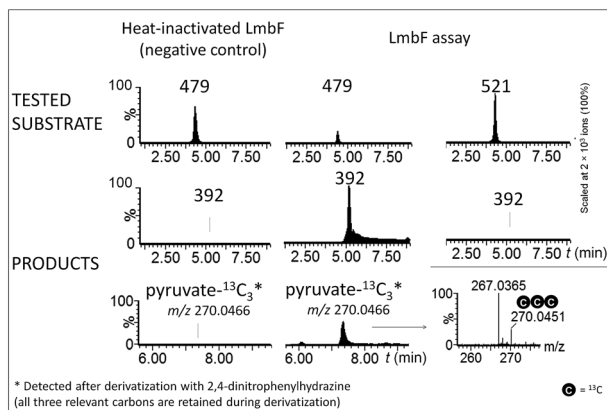


Fig. 5 Ion-extracted LC-MS chromatograms of LmbF *in vitro* assays with 479 and 521 as substrates. Pyruvate- $^{13}\text{C}_3$  was detected when 479- $^{13}\text{C}_3$ ,  $^{15}\text{N}$  was used as a substrate (see Incorporation experiments in ESI†).

reaction in the absence of PLP (Fig. S7†), confirming that the cofactor is essential for the turnover. Further, we observed that LmbF is also capable of removing the *S*-cysteine residue from 465 to give 378, but less readily than from 479 (Fig. S6 and S7†), indicating that 479 is the main natural substrate of LmbF (this is also supported by the predominant presence of 479 over 465 in the culture broth of the strain with the inactive *lmbF* gene depicted in Fig. S8†). On the other hand, we did not detect any turnover of 507 or 521 (LC-MS data in Fig. 5 and S7A†) where the primary amino group is not available due to its acetylation.

These findings provide insight into the reaction mechanism of the 'detoxification label' processing after it has been 'unlocked' by the deacetylation (Fig. 6): the unblocked primary amino group of 479 binds to PLP (a cofactor of LmbF) and LmbF catalyses the cleavage of the C–S bond by the  $\beta$ -elimination mechanism, which was described for PLP-dependent enzymes previously.<sup>12</sup> This results in the release of 392 and pyruvate, which was also detected as a product of the reaction (Fig. 5).

### Reactive sulfhydryl group is protected by methylation

LmbF products 378 and 392 contain the exposed sulfhydryl group, which is prone to oxidation. Indeed, we observed

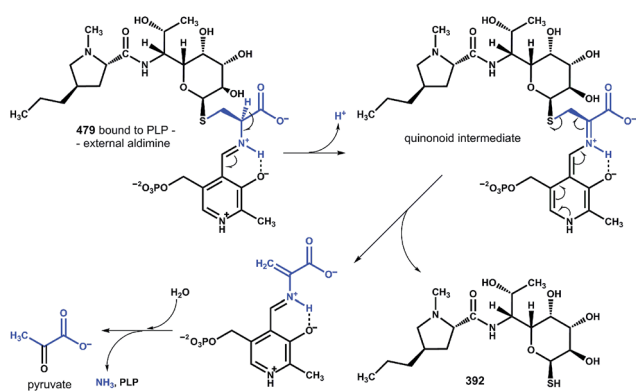


Fig. 6 Proposed mechanism of  $\beta$ -elimination catalysed by PLP-dependent LmbF. The fate of the 'detoxification label' is shown in blue.

a significant decrease of 378 and 392 along with the formation of dimers of these compounds (Fig. S9†) when the reaction time was prolonged from 2 h to 24 h. Therefore, we assume that the methylation of the sulfhydryl group occurs immediately after the removal of the *S*-cysteine residue to protect this reactive group. This reaction has been proposed to be catalysed by LmbG<sup>3</sup> because it represents the only putative methyltransferase encoded within the lincomycin gene cluster whose function has not been assigned yet. The remaining two methyltransferases LmbW and LmbJ were proved to act as a *C*-methyltransferase<sup>13</sup> and a *N*-methyltransferase,<sup>6</sup> respectively, modifying the lincomycin amino acid unit.

### 'Detoxification label' abolishes antibacterial activity

Furthermore, we tested whether the intermediates 507, 521, 465 and 479 exhibit antibacterial activity against the lincomycin-sensitive Gram-positive bacterium *Kocuria rhizophila* and we compared the results with those for NDL and lincomycin. None of the intermediates containing the complete or deacetylated 'detoxification label' exhibited antibacterial properties against *K. rhizophila* (Fig. S10†). A comparison of the results for NDL and lincomycin shows that the *N*-methyl group enhances the antimicrobial activity; however, it is not crucial. On the other hand, the presence of the 'detoxification label' completely abolishes the antibiotic activity of the compound. This observation is in accordance with previous studies indicating that mercapturic acid derivatives are less bioactive compared to the parent antibiotic.<sup>14</sup> Furthermore, not even deacetylation of the 'detoxification label' (intermediates 465 and 479) is a sufficient condition for antimicrobial activity.

### Lincomycin/celesticetin biosynthesis: homologous LmbF/CcbF catalyse a different type of reaction, forming a branch point in the related pathways

Lincomycin and celesticetin (Fig. 1) are the only known natural lincosamide antibiotics encoded by two independent but related biosynthetic gene clusters.<sup>3,15</sup> In contrast to lincomycin, with its methylated sulfur atom, celesticetin bears salicylate attached to sulfur *via* a two-carbon (2C) linker. Particularly, the origin of the 2C linker was unclear until the lincomycin intermediate 507 was revealed. We assume that condensation of celesticetin proceeds through a mechanism analogous to that of lincomycin, *via* a hypothetical intermediate 451 (Fig. 7), which would be analogous to 479; both compounds should bear the same deacetylated 'detoxification label'. However, the fate of the 'detoxification label' is probably different in the biosynthesis of celesticetin and lincomycin (see the proposed scheme in Fig. 7). Based on the celesticetin structure, it can be assumed that the 2C linker remains from the *S*-cysteine residue. Indeed, we have confirmed that these two carbons originate from *L*-cysteine since we achieved 78% incorporation of two  $^{13}\text{C}$  atoms into the celesticetin 2C linker when the culture medium was supplemented with *L*-cysteine- $^{13}\text{C}_3$ ,  $^{15}\text{N}$  (Fig. 7 and S11†). This means that CcbF does not catalyse the cleavage of the C–S bond by the  $\beta$ -elimination mechanism. Instead, CcbF presumably performs decarboxylation-dependent transamination, a reaction known





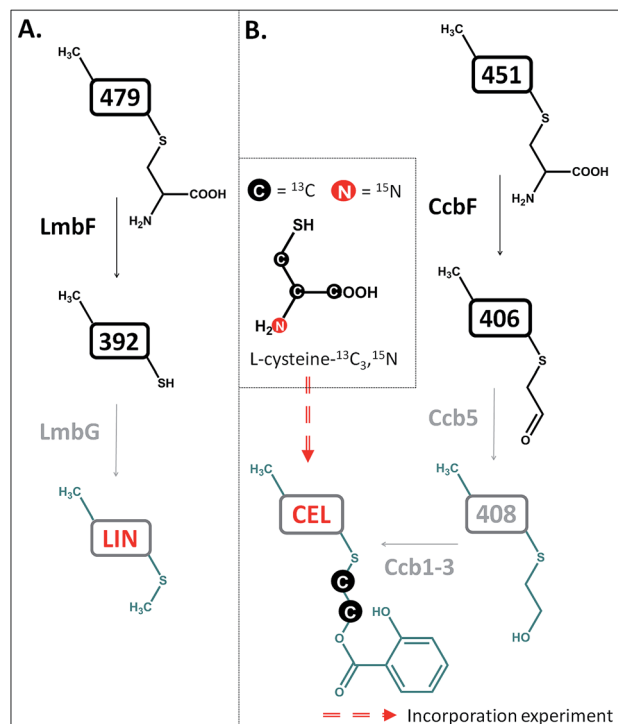


Fig. 7 Different processing of the 'detoxification label' by homologous LmbF and CcbF (steps in black) in lincomycin (A) and celesticetin (B) biosyntheses. Incorporation of two  $^{13}\text{C}$  atoms from L-cysteine reveals the origin of the 2C linker of celesticetin – see Fig. S11† for MS data. LIN – lincomycin; CEL – celesticetin; the grey colour indicates deduced biosynthetic steps.

to be catalysed by PLP-dependent enzymes as reviewed by John<sup>16</sup> and documented on dialkylglycine decarboxylase, for example.<sup>17</sup> The amino group would be replaced by an oxo group, which can be further reduced to a hydroxyl group (putative oxido-reductase encoded by the *ccb5* gene in the celesticetin gene cluster would be a suitable candidate for this reductase activity). Having completed this course of reactions, the intermediate is ready for conjugation with the acyl unit (already completed salicylate or its precursor) to form the final celesticetin scaffold (putative acyltransferase Ccb1, putative salicyl-AMP ligase Ccb2 and putative salicyl synthase Ccb3 can be expected to catalyse the formation and attachment of salicylate).

## Conclusion and perspectives

In this work, we present the final steps of lincomycin biosynthesis leading from the last proven intermediate, 507, to lincomycin. Interestingly, we have demonstrated that LmbJ *N*-methylates a different substrate at an earlier stage of the biosynthesis than previously suggested. Furthermore, we have revealed that deacetylation of the 'waste product' 507 represents the key step that switches the detoxification-like MSH-dependent process towards the biosynthetic mode. Deacetylation provides a free primary amino group for the PLP-dependent LmbF responsible for processing the deacetylated 'detoxification label'. PLP requires a primary amino group, to which it

binds to enable any possible PLP-dependent turnovers including  $\beta$ -elimination in lincomycin biosynthesis. Furthermore, based on the structures of lincomycin and celesticetin and  $^{13}\text{C}$  incorporation experiments, we predict that homologous CcbF from celesticetin biosynthesis performs a different type of reaction, decarboxylation-dependent transamination, which results in preservation of the 2C residue allowing the attachment of the salicylate unit. Such a surprising finding that two homologous PLP-dependent proteins from highly related biosynthetic pathways should exhibit different reaction specificity undoubtedly represents an exciting topic of further studies.

## Acknowledgements

This work was supported by the Ministry of Education, Youth and Sports of the Czech Republic project CZ.1.07/2.3.00/30.0003 and BIOCEV – Biotechnology and Biomedicine Centre of the Academy of Sciences and Charles University No. CZ.1.05/1.1.00/02.0109, from the European Regional Development Fund in the Czech Republic. The relevant part of the research was conducted within the "Prague Infrastructure for Structure Biology and Metabolomics", which has been built up by the financial support of the Operational Program Prague – Competitiveness (Project No. CZ.2.16/3.1.00/24023). The authors would also like to thank Dr Monika Cvancarova for advice on incorporation experiments and referees for their valuable comments, mainly for the suggestion of additional experiments improving the overall impact of the manuscript.

## References

- Q. F. Zhao, M. Wang, D. X. Xu, Q. L. Zhang and W. Liu, *Nature*, 2015, **518**, 115.
- S. Kadlcik, T. Kucera, D. Chalupska, R. Gazak, M. Koberska, D. Ulanova, J. Kopecky, E. Kutejova, L. Najmanova and J. Janata, *PLoS One*, 2013, **8**, e84902.
- J. Janata, S. Kadlcik, M. Koberska, D. Ulanova, Z. Kamenik, P. Novak, J. Kopecky, J. Novotna, B. Radojevic, K. Plhachova, R. Gazak and L. Najmanova, *PLoS One*, 2015, **10**, e0118850.
- G. L. Newton, Y. Av-Gay and R. C. Fahey, *Biochemistry*, 2000, **39**, 10739.
- G. L. Newton, N. Buchmeier and R. C. Fahey, *Microbiol. Mol. Biol. Rev.*, 2008, **72**, 471.
- L. Najmanova, E. Kutejova, J. Kadlec, M. Polan, J. Olsovska, O. Benada, J. Novotna, Z. Kamenik, P. Halada, J. Bauer and J. Janata, *ChemBioChem*, 2013, **14**, 2259.
- J. Bauer, G. Ondrovicova, L. Najmanova, V. Pevala, Z. Kamenik, J. Kostan, J. Janata and E. Kutejova, *Acta Crystallogr., Sect. D: Biol. Crystallogr.*, 2014, **70**, 943.
- N. M. Brahme, J. E. Gonzalez, J. P. Rolls, E. J. Hessler, S. Mizesak and L. H. Hurley, *J. Am. Chem. Soc.*, 1984, **106**, 7873.
- J. Novotna, J. Olsovska, P. Novak, P. Mojzes, R. Chaloupkova, Z. Kamenik, J. Spizek, E. Kutejova, M. Mareckova, P. Tichy, J. Damborsky and J. Janata, *PLoS One*, 2013, **8**, e79974.



- 10 E. Sasaki, C. I. Lin, K. Y. Lin and H. W. Liu, *J. Am. Chem. Soc.*, 2012, **134**, 17432.
- 11 C. I. Lin, E. Sasaki, A. S. Zhong and H. W. Liu, *J. Am. Chem. Soc.*, 2014, **136**, 906.
- 12 M. Ming, J. R. Lohman, T. Liu and B. Shen, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 10359.
- 13 A.-P. Pang, L. Du, C.-Y. Lin, J. Qiao and G.-R. Zhao, *J. Appl. Microbiol.*, 2015, **119**, 1064.
- 14 G. L. Newton and R. C. Fahey, *Arch. Microbiol.*, 2002, **178**, 388.
- 15 M. Koberska, J. Kopecky, J. Olsovska, M. Jelinkova, D. Ulanova, P. Man, M. Flieger and J. Janata, *Folia Microbiol.*, 2008, **53**, 395.
- 16 R. A. John, *Biochim. Biophys. Acta*, 1995, **1248**, 81.
- 17 G. B. Bailey and W. B. Dempsey, *Biochemistry*, 1967, **6**, 1526.

