



CrossMark  
 click for updates

Cite this: *RSC Adv.*, 2016, 6, 70414

## Seed mediated growth of silver nanoplates on glass: exploiting the bimodal antibacterial effect by near IR photo-thermal action and Ag<sup>+</sup> release†

A. D'Agostino,<sup>a</sup> A. Taglietti,<sup>\*a</sup> P. Grisoli,<sup>b</sup> G. Dacarro,<sup>c</sup> L. Cucca,<sup>a</sup> M. Patrini<sup>c</sup> and P. Pallavicini<sup>a</sup>

We developed a reproducible synthetic method to grow anisotropic silver nanoplates on glass, giving them the desired plasmonic features and tuning their size and LSPR absorption with growth time. Samples were fully characterized showing good stability, a strong photo-thermal effect and the ability to release controlled quantities of silver in water. These samples showed a strong antibacterial activity, which is based on two different mechanisms: silver ion release and hyperthermia caused by the photo-thermal effect under near-infrared laser irradiation. This behaviour allows us to foresee promising *in vivo* applications, ensuring a long-term antibacterial protection which can be reinforced, when needed, by a fast photo-thermal action which can be switched on by a NIR laser treatment.

Received 4th May 2016  
 Accepted 19th July 2016

DOI: 10.1039/c6ra11608f

[www.rsc.org/advances](http://www.rsc.org/advances)

### Introduction

The use of silver nanoparticles (AgNPs) as antibacterial agents has been one of the most studied nanotechnology issues of the last two decades. It is well known that bacterial infections, those involving the surface of medical devices (*e.g.* catheters, artificial prosthetics and subcutaneous implants) and more generally infections spread by materials used in the nosocomial environment, are still a very serious challenge for bio-medical scientists: bacterial and biofilm-associated infections involving these surfaces are probably the leading cause of medical implant failure, as well of other severe nosocomial problems.<sup>1,2</sup> As conventional antibiotics are not able to eradicate biofilms from the surfaces where they have formed, often surgical removal of the implant is the only affordable solution. A possible answer to these problems comes from coating of the described medical surfaces with AgNPs, which are quite effective as antibacterial, with a mechanism of action which is still debated by the scientific community. Anyway, it is generally accepted that the antibacterial action involves the release of Ag<sup>+</sup> ions and the following interaction of Ag<sup>+</sup> with bacteria. Recently, we and other authors formulated an hypothesis which suggests the coexistence of a short distance nanomechanical

action involving the direct interaction of AgNPs with the bacterial cell membrane.<sup>3</sup> In the last years we confirmed that AgNPs are effective towards Gram– bacterial strains but much less towards Gram+,<sup>4</sup> and that this action can be easily brought on a bulk glass surface by means of interactions of AgNPs surfaces with a –SH layer, after forming first a layer of mercaptopropyltriethoxy silane (MPTS).<sup>3,5</sup> Similar results were obtained attaching a monolayer of AgNPs on a layer of polyethyleneimine (PEI) polymer, a strategy which allows to establish electrostatic interactions between its amino groups and AgNPs surfaces.<sup>6</sup> Recently we also demonstrated that a monolayer of AgNPs obtained through preliminary silanization of the glass with 3-aminopropyltriethoxysilane (APTES) can be conveniently used against the biofilm-producer *Staphylococcus epidermidis* RP62A.<sup>7</sup> In all these cases, excellent stability in aqueous media, prolonged release of silver ions and high local concentration of Ag<sup>+</sup> are obtained, without any detaching of AgNPs and resulting in a strong antibacterial action with limited risks towards human health. Another way to use noble metal nanoparticles to fight bacteria presence and growth is to exploit their photo-thermal features:<sup>8</sup> for example, gold nanorods (GNRs)<sup>9</sup> and gold nanostars (GNSs)<sup>10</sup> have been used to destroy colonies of different bacterial strains. Gold is not intrinsically antibacterial, anyway anisotropic gold nano-objects featuring two or more localized surface plasmon resonances (LSPR) that undergo thermal relaxation when irradiated, can produce a localized hyperthermia which is used to kill bacteria and eradicate biofilms. Moreover, when at least one LSPR falls in the near-IR range (NIR, 750–950 nm) where tissues and blood are semi-transparent, one can imagine to use laser irradiation on subcutaneous devices coated with these kind of nano-materials. In a recent example,<sup>11</sup> we prepared active surfaces

<sup>a</sup>Department of Chemistry, University of Pavia, Viale Taramelli 12, 27100 Pavia, Italy.  
 E-mail: [angelo.taglietti@unipv.it](mailto:angelo.taglietti@unipv.it)

<sup>b</sup>Department of Drug Sciences, University of Pavia, Viale Taramelli 14, 27100 Pavia, Italy

<sup>c</sup>Department of Physics, University of Pavia, Via Bassi 6, 27100 Pavia, Italy

† Electronic supplementary information (ESI) available: Supplementary spectra, thermograms and SEM images of samples, photographs of samples, EDX data, XRD data, transmission curves of samples. See DOI: 10.1039/c6ra11608f



bearing controlled microgram quantities of gold as grafted monolayers of GNS. Laser irradiation of the GNS monolayer on their NIR LSPR results in efficient photo-thermal conversion, and we observed that the produced hyperthermia efficiently induced bacteria cell death in *S. aureus* biofilms. Photo-thermal effects have also been exploited to increase silver ions release from bimetallic nanostructures, and then considering this release for microbicidal tasks. In a recent example it was found that Pd@Ag nanosheets show a synergistic effect between silver ions and photo-thermal heating obtained from the plasmonic objects produced reducing silver on Pd nanosheet surfaces: upon irradiation in the NIR region, the hyperthermia caused an increase of silver ions release, resulting in an enhanced antibacterial action of the colloidal nanomaterial.<sup>12</sup> Other examples are reported of combined action between silver bactericidal action and gold photo-thermal features, obtained upon irradiation in the visible region in the case of gold core/silver shell spherical nanoparticles stabilized with aspartame,<sup>13</sup> or in the NIR in the case of gold core/silver shell nanorods.<sup>14</sup> Curiously, few examples are reported about the photo-thermal action of colloidal suspensions of nano-objects made of pure silver. The ability to give an efficient photo-to-heat conversion, in fact, is not only limited to gold and palladium nano-objects: silver itself can be used as well. Recent and pioneering reports from Astilean and coworkers<sup>15,16</sup> fully exploit this feature, showing the successful use of biocompatible chitosan-coated silver nanoplates for photo-thermal treatments of cancer cells when irradiated at 800 nm, a wavelength value where a plasmonic band can be easily placed by tuning morphology and dimensions of nanoplates themselves. It is well known that an LSPR band in the NIR can be obtained using anisotropic silver objects of different morphologies, exploiting the in plane (longitudinal) dipolar resonance.<sup>17</sup> To our knowledge, anyway, no examples of antibacterial action obtained with hyperthermia coming from pure silver plasmonic structures is reported, nor in colloidal suspension neither grafted to a surface. Seed-growth methods represent a very popular way to prepare anisotropic noble metal nano-objects, and are often based on the use of surfactant molecules to orient the anisotropic growth.<sup>18,19</sup> Unfortunately, it can be hard to remove them from the nanoparticles surfaces, thus complicating further chemistry which could be needed for further functionalizations.<sup>20</sup> Also, surfactants may have a harmful effect when used in situations connected to health, as they are in some cases considered toxic for human cells.<sup>21</sup> Seed-growth methods can be extended to grow anisotropic nano-objects directly on surfaces: a few examples, mainly in presence of surfactants, of seed-growth synthesis of silver anisotropic nano-objects on different kinds of substrates have been reported.<sup>20,22–26</sup> In a recent example, a seed mediated growth of silver nanoplates on a glass surface was obtained, with a certain degree of control, through several successive immersion steps, using citrate as orienting agent allowing only their longitudinal growth, and obtaining the generation of a very broad absorption covering the whole vis-NIR spectra.<sup>27</sup>

Here, we demonstrate how to prepare bulk glass materials bearing silver nanoplates, tailored to have a high absorption in the NIR region and having strong photo-thermal features, in

order to study their potential as model antibacterial surfaces. The long-term goal is the possibility of fabricating medical devices with coatings which could be active on long timescales as a result of the “classical” antibacterial effect due to silver ions release,<sup>3–7</sup> but with a further, additional antibacterial action which could be “switched on” on need by a NIR laser irradiation, producing localized hyperthermia,<sup>12</sup> and to evaluate the possible synergy between these two kind of actions. In order to minimize the use of possibly toxic chemicals and avoid the use of classical surfactants, we introduce here a surfactant free seed-growth method to be applied directly on surfaces, involving the use of citrate as a shape directing agent. The idea was to tune the geometric features of the nano-objects in a single growing step maintaining an easy synthetic setup and just controlling the growth time. In this way an easy control the LSPR features of the samples is obtained, and an intense absorption in the NIR region can be gained and exploited for efficient laser induced photo-thermal effect.

## Experimental

### Materials

Silver nitrate (>99.8%), sodium borohydride (>99.0%), sodium citrate (>99.0%), ascorbic acid ( $\geq 99\%$ ) were purchased from Sigma Aldrich. Trimethoxysilylpropyl(polyethylenimine) (50% in isopropanol) was purchased from Gelest Inc. Reagents were used as received. Solvents were purchased from Sigma-Aldrich and used as supplied. Microscopy cover glass slides (21 × 26 mm) were purchased from DelChimica. Water was deionized and then bidistilled.

Tryptone Soya Broth (TSB) and Tryptone Soya Agar (TSA) for bacteria culture were purchased from Oxoid, England. *S. aureus* ATCC 6538 and *E. coli* ATCC 10536 bacterial strains were used.

### Preparation of PEI-silane self-assembled monolayers on glass surface (SURF-PEI glasses)

The samples were prepared according a reported method.<sup>6</sup> Briefly: glass substrates were cleaned for 30 min in freshly prepared Piranha solution (3 : 1 v/v H<sub>2</sub>SO<sub>4</sub> : H<sub>2</sub>O<sub>2</sub> (30%)). **Caution!** Piranha solution is a strong oxidizing agent and should be handled with care., washed three times with ultrapure water in a sonic bath and oven-dried. The slides were then immersed for 6 min in a 4% (v/v) solution of PEI-silane in ethanol at room temperature. In a typical preparation, 8 glass slides were prepared at the same time, *i.e.* reacting in the same silane solution inside a 8-place holder (a microscope glass slides staining Jar). After this, the slides were washed two times with ethanol and one time with ultrapure water in a sonic bath and blow-dried with nitrogen.

### Seeds preparation

Seeds, spherical silver nanoparticles, were prepared according to a reported method.<sup>5</sup> Briefly, to 100 mL of ice cooled water the following solutions were added in sequence under vigorous stirring 1 mL of 1% (w/v) AgNO<sub>3</sub> solution, after a minute 1 mL of 1% (w/v) sodium citrate and, after a further minute, 0.500 mL of



an ice-cooled solution 0.075 wt% in NaBH<sub>4</sub> and 1 wt% in sodium citrate. After the last addition, stirring was immediately stopped, in order to avoid coagulation. The colloidal suspensions were stored in the preparation flask, maintained in the dark and used within 2 days from preparation.

### Preparation of SURF-PEI-SEEDS samples

The samples were prepared according a reported method.<sup>6</sup> PEI glasses were immersed in the colloidal suspension of seeds at room temperature for 15 min. In a typical preparation, 8 glass slides were prepared at the same time, *i.e.* they were made to react in the same seeds suspension (30 mL), inside a glass holder (Hellendhal staining Jars were used) where the slides were kept in vertical position. After this the obtained glass samples, showing a yellow colour, were placed in water and sonicated for 5 min. This procedure was repeated twice and then the glasses were blow dried with a nitrogen stream.

### Growth solution preparation

Growth solution was prepared in a flask: to 30 mL of sodium citrate  $8.5 \times 10^{-3}$  M were added, under vigorous stirring, 18 mL of AgNO<sub>3</sub>  $5 \times 10^{-4}$  M and 0.450 mL of ascorbic acid  $1 \times 10^{-2}$  M. After the last addition, stirring was immediately stopped.

### Preparation of SURF-PEI-PLATES samples

In a typical preparation, up to 8 SURF-PEI-SEEDS samples were immersed in the proper amount of growth solution for the desired time in a Hellendhal staining Jar. Changing in colour of sample was observed from yellow, characteristic colour of SURF-PEI-SEEDS glass, to orange, red and dark blue on increasing dipping times. After this, the slides were washed with bidistilled water two times and dried under nitrogen flux.

### Antibacterial activity tests

The antibacterial activity of functionalized cover glasses was investigated against *Staphylococcus aureus* ATCC 6538 (Gram+) and *Escherichia coli* ATCC 10356 (Gram-). The microorganisms were grown overnight in Tryptone Soya Broth (Oxoid; Basingstoke, Hampshire, England) at 37 °C. Washed cells were resuspended in Dulbecco's PBS and optical density (OD) was adjusted to 0.2 at 650 nm wavelength, corresponding approximately to  $1 \times 10^8$  Colony Forming Units (CFU) per mL. 10 μL of bacterial suspension was deposited on a standard glass slide (76 × 26 mm), then the microbial suspension was covered with a functionalized cover glass slide (21 × 26 mm), forming a thin film between the slides that facilitates direct contact of the microorganisms with the active surface. The two assembled glasses were introduced in a Falcon test-tube (50 mL) containing 1 mL of PBS to maintain a damp environment. In this test microbes are incubated in non-nutritive suspensions that do not give the microorganisms the potential to grow during the test. For each bacterial strain three equivalent modified glasses were prepared; the slides were maintained in contact with the liquid films containing bacteria at room temperature for 20 minutes, 5 and 24 h, respectively; for each time of contact an

unmodified glass slide was treated in the same way as control sample. After the times of contact, 9 mL of PBS were introduced in each Falcon test-tube under a gentle shaking to detach the assembled glass slides. Bacterial suspensions were then cultured on Tryptone Soya Agar (Oxoid; Basingstoke, Hampshire, England) to count viable cells. The decimal-log reduction rate, *i.e.* the Microbicidal Effect (ME), was calculated using the formula:

$$ME = \log N_C - \log N_E$$

( $N_C$  being the number of CFU mL<sup>-1</sup> developed on the unmodified control glasses, and  $N_E$  being the number of CFU mL<sup>-1</sup> counted after exposure to modified glasses). The results expressed as ME represent the average of three equivalent determinations. This method can be considered as a version of the Japanese Industrial Standard JIS Z 2811 developed to measure the antibacterial activity of plastic surfaces.

### Thermal microbicidal tests

Antibacterial activity due to photo-thermal effect of SURF-PEI-PLATES was investigated against *Staphylococcus aureus* ATCC 6538 (Gram+) and *Escherichia coli* ATCC 10356 (Gram-). One functionalized slide was cut in 4 sections of 10 × 10 mm in order to be completely irradiated by laser. A volume of 0.02 mL of bacterial suspension was deposited on 2 sections. For each pair of functionalized glasses, one was irradiated for 20 min whereas the other was not irradiated. After this time, the glass section covered with bacterial suspensions was suspended in 1 mL of sterile water, gently shaken and then water was suitable diluted in 3 different tubes: 1 : 100, 1 : 10 000, 1 : 100 000. From each tube, 1 mL of suspension was taken and then cultured on Tryptone Soya Agar (Oxoid; Basingstoke, Hampshire, England) to count viable cells. The decimal-log reduction rate, *i.e.* the "thermal microbicidal effect", ME<sub>T</sub> was calculated with the following formula

$$ME_T = \log N_C - \log N_T$$

where  $N_C$  is the number of CFU mL<sup>-1</sup> developed in contact with a not irradiated modified control glass sample, and  $N_T$  the number of CFU mL<sup>-1</sup> counted after exposure to modified glass samples and irradiation.

### Instrumentation and instrumental methods

Absorbance spectra of colloidal suspensions were taken with a Varian Cary 100 spectrophotometer in the 200–900 nm range and spectra of functionalized glasses were obtained placing the slides on the Varian Cary 100 spectrophotometer equipped with a dedicated Varian solid sample holder.

SEM images were taken from Tescan Mira XMU variable pressure Field Emission Scanning Electron Microscope – FEG SEM (Tescan USA Inc., USA) located at the Arvedi Laboratory, CISRiC, Pavia. Slides were mounted onto aluminum stubs using double sided carbon adhesive tape and were then made electrically conductive by coating in vacuum with a thin layer of Pt/Pd (3–5 nm). Observations were made in backscattered





the objects to give nanoplates.<sup>27</sup> These flat nano-objects typically show an in-plane dipole plasmon resonance band which position can be placed in the NIR region and tuned at a desired value just by regulating the aspect ratio of the objects.<sup>28,29</sup> Our expectations were that it was possible to obtain increasing anisotropic growth on surface just increasing immersion time in presence of high citrate concentration, so that control of immersion times could allow tuning of the morphology and consequently the LSPR features of plasmonic objects brought on glass samples.

Fig. 1 shows the UV-Vis-NIR spectra measured for samples extracted from the growth chamber at different interval times. The development of a broad, intense band which maximum, in about two hours, progressively shifts to NIR values, can be attributed to the growth of objects having different anisotropic shapes and increasing lateral dimensions.

When the experiment was repeated, in the same growth solution, on simple SURF-PEI samples, *i.e.* in absence of grafted seeds, no objects formation and growth was observed, yielding transparent and uncoated glass samples. It is important to stress one of the main results coming from this experiment: with this method the maximum of LSPR spectra of the obtained samples can be tuned to the desired value, in the range between 400 and 800 nm, just varying the immersion time of the seeds coated samples in the growth solution.

Increase of immersion times longer than two hours does not produce any noticeable change in spectra, as can be clearly seen in Fig. 1 (green dashed line spectrum) indicating that no further growth or shape changes are happening after this growing time.

SEM images were taken for the samples extracted after different immersion/growing times, and are reported in Fig. 2. Growth of objects can be immediately perceived moving from Fig. 2a (sample after 15 min immersion) to Fig. 2b (30 min immersion) to Fig. 2c (2 hours immersion).

In Fig. 2d, reporting a SEM image at an higher magnification, which is again relative to the two hours immersion time, the anisotropic growth of objects is evident: triangular and truncated triangular plates, with longitudinal dimensions

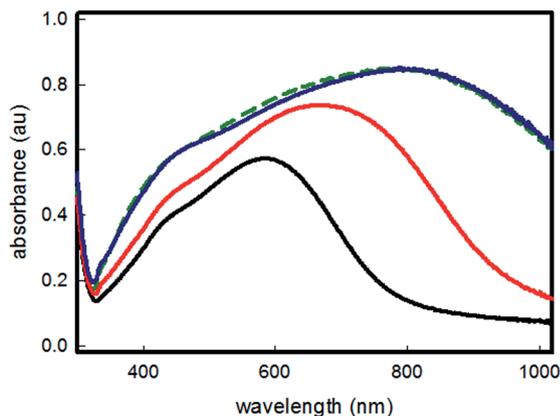


Fig. 1 UV-Vis-NIR spectra of samples after different times of seed mediated growth: 15 minutes (black line), 30 minutes (red line), two hours (blue line), 4 hours (green dashed line).

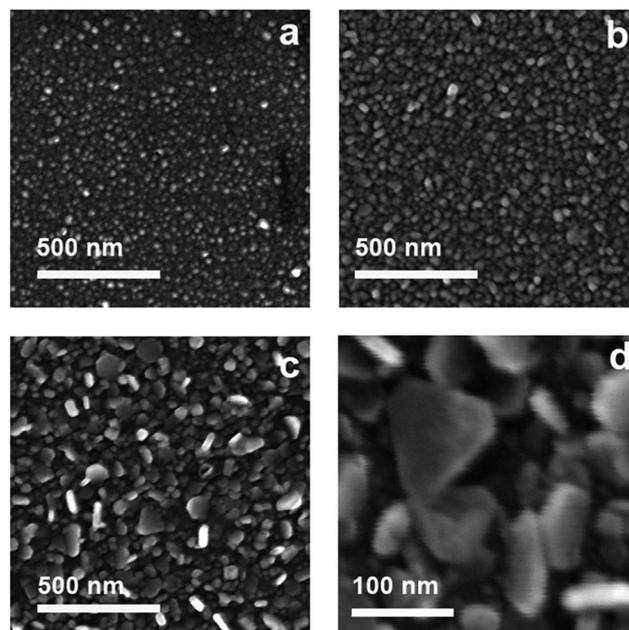


Fig. 2 SEM images of samples after different times of seed mediated growth: (a) 15 minutes, (b) 30 minutes, (c) 2 hours, (d) 2 hours, higher magnification.

ranging between 40 and 120 nm can be observed, all having a thickness not exceeding a value of 20 nm. The extremely high degree of poly-dispersion explains the very broad LSPR spectra observed for increasing growth time. Nevertheless, the NIR absorption spectra is explainable with the presence of a distribution of objects having different longitudinal dimensions, and a subsequent broad range of dipolar in-plane resonance wavelength.<sup>25–29</sup> The glass samples have an homogeneous dark blue-grey colour, which is consistent with the poly-dispersed array of grafted silver nanoplates and the LSPR features described: a photograph of a representative slide after two hours growth is reported in ESI† section.

As with two hours immersion the broad LSPR band was placed close to the desired wavelength value, we decided to use this growth time in the standard preparation for SURF-PEI-PLATES samples. The preparation of SURF-PEI-PLATES is reproducible: spectra of several different samples are given in Fig. S3 and S4 in the ESI† section, and evidence that no sensible differences in intensity and position of the LSPR peak can be observed among the samples obtained simultaneously from the same preparation (Fig. S3†) or coming from different preparation batches (Fig. S4†). It also ensures a very homogeneous coating of the surface, as can be seen from the SEM images taken on large area (see Fig. S5 in ESI† section). SURF-PEI-PLATES slides are stable in air, *i.e.* their spectra and colour did not change significantly for at least a 4-week period.

EDX analysis (see ESI†) were performed. Beyond the glass substrate components, the evidence of Ag L spectral line is consistent with the preparation procedure. We notice that EDX analysis in different points of the sample surface shows chemical homogeneity of Ag films (within  $\pm 5\%$ ), the intensity of any spectral line in EDX analysis being proportional to element



abundance. The presence of Ag plates is confirmed also from X-ray diffraction patterns (see ESI†). Only the peak of 111 reflection of a FCC structure is clearly evident, suggesting that the basal lattice plane of nano-objects is (111), as expected from the shape-directing action of citrate, and also that exists a preferred orientation of plates on the glass surface. The peak broadening confirms the nanostructured nature of silver.

We completed the characterization of the SURF-PEI-PLATES (samples c) by means of quantitative oxidation of the silver plates, and analysis of the obtained  $\text{Ag}^+$  solution by means of ICP. Data from eight experiments, coming from eight different samples preparations, we found an average of  $6.2 (0.6) \times 10^{-6} \text{ g cm}^{-2}$  of silver, a quantity which is considerably high, when compared to samples obtained grafting a monolayer of spherical seeds on a silanized glass, as we have described in the recent past.<sup>5-7</sup>

### Stability in water and silver release

The stability in water, as well their ability to release silver ions in water as a function of time was determined by experiments on several SURF-PEI-PLATES samples (see experimental part). The  $21 \times 26 \text{ mm}$  glass slides were immersed in 3 mL of ultrapure water for a given time interval, and then removed from the liquid. After registering their UV-Vis-NiR spectra, the solutions obtained after SURF-PEI-PLATES slides immersion in water were analysed with ICP. The data of silver concentration found in these solutions were expressed as  $\mu\text{g}$  of released silver per  $\text{cm}^2$  of exposed surface, as a function of time (see Fig. 3), showing a well-defined trend. We must stress the fact that, in analogy with our precedent investigations, these experiments were performed in ultrapure water.<sup>5-7</sup> Thus, the results cannot be of course simply transferred to more complex or biological environments, as silver ions can give poorly soluble salts or complexation with biomolecules when they are released in “real” media.<sup>30</sup>

Nevertheless these experiments allow to state that (i) no silver nano-objects were released in water, as Uv-Vis-NiR spectra did not show the presence of any absorption which could be related to presence of plasmonic objects detached from the

surface; (ii) silver is released in water as a function of immersion time, reaching a stationary concentration after approximately 2–3 days; (iii) as already observed in similar systems, the most of dissolved silver is released in the first 24–48 hours of immersion. As we have already demonstrated, silver release should involve the formation or the thickening of a  $\text{Ag}_2\text{O}$  layer on the water-exposed surface of silver nano-objects, with the formation of an oxidized silver layer in the initial 1–2 days period, reaching a steady-state in which the slowly released  $\text{Ag}^+$  ions are replaced by Ag oxidized from the bulk.<sup>5</sup>

As we start from a quite high surface concentration of silver, also the released amount is higher than the value which is usually released from monolayers made of small spherical silver nanoparticles grafted on glass.<sup>3,5-7</sup> As can be easily calculated, anyway, the amount of silver released in these conditions after 10 days is less than 5% (about  $2.5 \times 10^{-7} \text{ g cm}^{-2}$ ) of the total concentration of silver present on the surface. This allows to imagine surfaces retaining for a long time the possibility to release silver ions in an environment subjected to flow of aqueous media, a feature that is considered essential for fight bacterial proliferation leading to severe infection and/or biofilm formation: for example, recently it was demonstrated that biofilm growing on a biomaterial surface is a dynamic process able to reach a maximal thickness at 12–18 h and maturation by 24 h.<sup>31</sup> SEM images and UV-Vis-NiR spectra were taken for selected

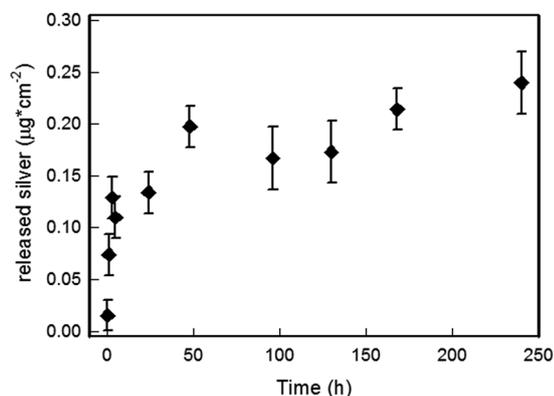


Fig. 3 Silver ion released from SURF-PEI-PLATES ( $\mu\text{g cm}^{-2}$  of exposed surface) versus time in 3 mL of water.

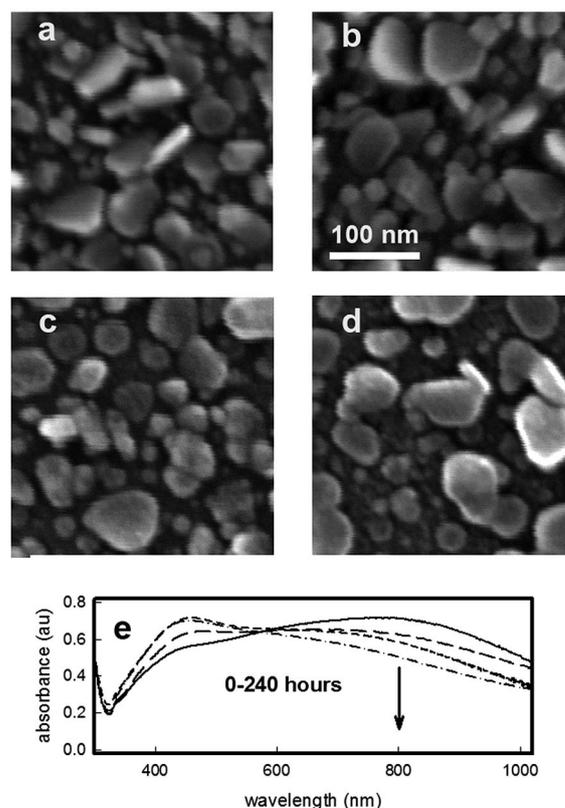


Fig. 4 (a–d) SEM images of SURF-PEI-PLATES samples after increasing immersion times in pure water: (a) 0 hours (as prepared), (b) 5 h immersion, (c) 24 h immersion, (d) 240 h immersion; (e) UV-Vis-NiR spectra of the same samples.



samples after the immersion in water and silver ion release. Fig. 4 shows the SEM images taken for samples after immersions of 5 (Fig. 4b), 24 (Fig. 4c) and 240 hours (Fig. 4d), compared with the untreated, as-prepared sample (Fig. 4a).

One can observe a progressive but limited consumption of the prismatic plates, causing a visible morphology change, from triangular like plates to more discoidal shapes. This of course modifies the UV-Vis-NIR spectra of the samples, causing a progressive depletion of the extinction at 800 nm, as reported in Fig. 4e: the consumption of the plates results in a progressive flattening of the absorption in the NIR nm zone, as reduction of longitudinal dimensions as well the snapping of edges are expected to reduce the intensity and blue shift the dipolar in-plane resonance.<sup>17</sup> On the other side, as can be clearly seen from large area SEM images (see Fig. S8 in ESI†), the overall homogeneity of the silver coating layer is preserved. Visually, the dark blue colour of the slides fades with prolonged immersion, as obvious consequence of the weakening of the broad band at 800 nm. In the same time a more yellowish aspect can be noticed, depending on the increase of the band in the 450–500 nm range, typical of discoidal and spheroidal silver nano-objects.<sup>17</sup> A photograph of a glass slide after 240 hours of immersion is reported in the ESI (see Fig. S9†): the dark blue colour has faded and a yellow *nuance* can be seen. Morphological changes caused by long immersion times in water can thus be perceived by the naked eye. This feature could also be exploited to assess the material shelf life and usability: the complete vanishing of the blue colour indicates the disappearing of the broad absorption at 800 nm, and excludes the possibility of any exploiting of NIR laser excitation.

### Photo-thermal studies

The photo-thermal behaviour of the samples obtained with different growth time has been measured by irradiation with a 808 nm continuous laser source (200 mW, spot diameter 1 cm,

resulting irradiance  $0.26 \text{ W cm}^{-2}$ ) on  $10 \times 10 \text{ cm}$  glass slides, which were obtained by cutting the larger slides typically used in the samples preparation. Thermograms (temperature *vs.* time graphs) were recorded using a thermocamera to register the temperatures reached by the glass samples while irradiated, observing in all cases an increase-plateau profile, reaching a stationary temperature within 40 seconds, as reported in Fig. 5a.

Temperature values recorded at plateau were of 37.8 (0.3), 53.8 (0.3) and 58.5 (0.3) degrees for the samples grown 15 minutes, 30 minutes and two hours, respectively. As can be seen from Fig. 5b, the maximum temperature obtained by the three different samples increases with the absorbance value of each sample at 808 nm, measured from spectra reported in Fig. 1, and the highest temperature is obtained when the laser source wavelength matches the  $\lambda_{\text{max}}$  of LSPR absorption. UV-Vis-NIR spectra were recorded before and after irradiation for each sample, showing that the described treatment and relative produced hyperthermia did not change the LSPR features of the samples (an example is given in Fig. S10 in ESI†).

A similar set of experiments was repeated on a sample of SURF-PEI-PLATES (two hours growth) after 24 hours of immersion in water, to check if the silver release, producing the objects consumption and changes in LSPR features described in Fig. 4, could change drastically the photo-thermal effect. Only a slight decrease is observed in the value of temperature reached after irradiation of these samples (about  $55 \text{ }^\circ\text{C}$ , see Fig. S11 in ESI† for the thermogram), as it could be expected from the LSPR absorption at 808 nm, which decreases only marginally after 24 hours of immersion in water as reported in Fig. 4e.

To our knowledge, there are no precise data about the temperatures needed to eliminate bacteria with photo-thermal effects. Moreover, temperature values measured by thermograms in the showed experiments describe temperature changes on bulk samples, a value which is a function of several experimental parameters,<sup>10</sup> giving no information on the temperatures changes in the close surroundings of the nano-objects, which should reach considerably higher values. Anyway, it is known that heating of *E. coli* cells at temperatures of  $52 \text{ }^\circ\text{C}$  for more than 5 minutes results in the destruction of bacteria permeability barrier.<sup>32</sup> Recently it was reported that irradiating for 5 minutes with a laser a cluster of gold nanorods it is possible to reach a temperature of about  $60 \text{ }^\circ\text{C}$  causing the death of most bacterial cells in a *E. coli* biofilm.<sup>33</sup> On the other side, we demonstrated that a few degrees of hyperthermia, caused by low irradiance laser excitation of a monolayer of GNSs on a glass bulk surface for longer times (30 min), is sufficient to almost completely eliminate the bacteria cells of a *S. aureus* biofilm layer.<sup>11</sup>

### Antibacterial tests

The antimicrobial activity of SURF-PEI-PLATES surfaces against *E. coli* and *S. aureus* was investigated. An established procedure allows the evaluation of the microbicidal effect (ME) in a thin liquid film in contact with the functionalized surface using the

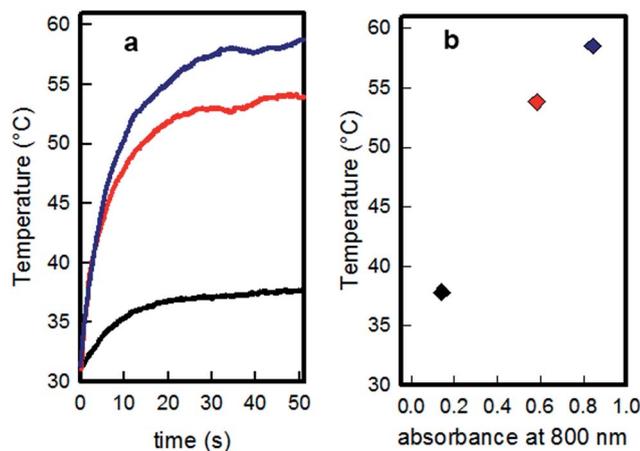


Fig. 5 (a) Temperature *versus* time thermograms obtained for samples under laser irradiation at 808 nm,  $0.26 \text{ W cm}^{-2}$ : samples grown 15 minutes (black line), 30 minutes (red line) and two hours SURF-PEI-PLATES (blue line); (b) maximum temperature reached by each sample *vs.* its absorbance at 808 nm.



Table 1 ME values<sup>a</sup> for SURF-PEI-PLATES samples

	20 min	5 h	24 h
<i>E. coli</i>	<0.3	0.9 (0.3)	>5
<i>S. aureus</i>	<0.3	0.8 (0.3)	>5

<sup>a</sup> Values are obtained as the average of three measurements. ME =  $\log N_C - \log N_E$ . A value higher than 5 indicates that no survived bacteria were found after the test.

following formula: ME =  $\log N_C - \log N_E$ , where  $N_C$  is the number of CFU mL<sup>-1</sup> developed on the unmodified control glasses, and  $N_E$  the number of CFU mL<sup>-1</sup> counted after exposure to modified glasses (CFU = colony forming unit).<sup>5</sup>

Experimental results after contact times of 20 minutes, 5 and 24 h are reported in Table 1, and show that SURF-PEI-PLATES glass slides display a significant bactericidal effect after a long contact time. 24 hours contact times are able to exert a complete microbicidal effect, eliminating at least 99.999% of bacteria.† The effect is much less intense with 5 hours of contact, while no microbicidal effect is present with very short contact times: it must be noted that silver release, which is considered as the main responsible of antibacterial action, is limited ( $0.015 \times 10^{-6}$  g cm<sup>-2</sup>, see Fig. 3) after a 20 min time of contact with water. Mechanisms by which AgNPs exert their antibacterial effect are still hugely debated, anyway Xiu *et al.*<sup>34</sup> showed that silver nano-objects are devoid of antibacterial action when used under anaerobic conditions which exclude oxidation to silver(i), thus preventing its release. SURF-PEI-PLATES samples action against *S. aureus* for long contact times seems much more intense than the one found for SURF-PEI-SEEDS samples bearing small spherical AgNPs,<sup>6</sup> and this should be due to the higher concentration of silver present on the former. Anyway, at this preliminary stage of investigation, one cannot exclude that also the different morphology of objects on surfaces is playing a complex role. Remarkably, in the tests proposed by CEN (European Committee for Standardization) in – EN 13697,§ the microbicidal activity of a disinfectant is considered acceptable when the decimal-log reduction rate (*i.e.* ME), is at least equal to 4 after 5 min of contact. In our case ME of the modified glass is superior to 4 after 24 h of contact for both strains, with a long term action which seems well suited to build antibacterial surfaces for medical devices.

Surfaces functionalization with silver nano-objects able to release controlled quantities of Ag(i) and imparting antibacterial features, is a technique that was widely investigated in the last decade. So, these first set of results was largely expected, on

† As the CFU counted in the control measures reach at least a value of  $10^5$ , although varying every time as consequence of differences in the experimental conditions, when no CFU can be found in the samples coming from SURF-PEI-PLATES we assign to ME a value of 5, to indicate that at least 99.999% of bacteria were eliminated during the test.

§ European Committee for Standardization, EN 13697, Chemical Disinfectants and Antiseptics – Quantitative Non-porous Surface Test for the Evaluation of Bactericidal and/or Fungicidal Activity of Chemical Disinfectants used in Food, Industrial, Domestic and Institutional Areas – Test Method and Requirements (Phase 2, Step 2), Brussels: CEN, 2002.

Table 2 ME<sub>T</sub> values<sup>a</sup> for SURF-PEI-PLATES samples

	20 min	20 min (after 24 h in water)
<i>E. coli</i>	>5	>5
<i>S. aureus</i>	3.7 (0.3)	3.8 (0.3)

<sup>a</sup> Values are obtained as the average of three measurements. ME<sub>T</sub> =  $\log N_C - \log N_T$ . A value higher than 5 indicates that no survived bacteria were found after the test.

the basis of several reported studies.<sup>3–7,35–37</sup> Anyway, in all the studies related to antibacterial effects based on silver nano-objects, hyperthermia caused by photo-thermal effects was never considered as an option to enhance their antimicrobial activity when grafted on surfaces. On the other side, hyperthermia based antimicrobial applications relies usually on magnetic and/or gold nanoparticles. The result is that no examples are reported about antimicrobial action caused by photo-thermal effects given by silver nanoplates, nor in colloidal suspension neither grafted to a surface. Our main interest, thus, was to obtain a novel and (until now) undescribed, antimicrobial action given by NIR laser excitation of anisotropic silver nano-objects. The final goal was to have an hyperthermia induced action which could be “switched on” as a result of NIR laser irradiation, in addition to the expected long term action due to silver release usually exhibited by silver AgNPs layers.

To obtain this, SURF-PEI-PLATES samples, opportunely cut in order to be completely covered by the light spot, were irradiated for 20 min with a 808 nm laser source, while in contact with the described bacteria suspensions, using an irradiance of 0.26 W cm<sup>-2</sup>. In this case the “thermal microbicidal effect”, ME<sub>T</sub> was calculated with the following formula:

$$ME_T = \log N_C - \log N_T$$

where  $N_C$  is the number of CFU mL<sup>-1</sup> developed after the contact with SURF-PEI-PLATES samples in absence of irradiation, taken as control, and  $N_T$  the number of CFU mL<sup>-1</sup> counted after exposure to SURF-PEI-PLATES sample in presence of irradiation.

It is important to stress that in these conditions ME<sub>T</sub> has to be ascribed only to the photo-thermal effect, as any possible influence from the limited silver release in absence of irradiation is considered in the control count. Moreover, as very limited silver ion release is observed after 20 minutes (see Fig. 3), no silver related ME is observed during this time (see Table 1). It is also important to stress that the ME<sub>T</sub> values do not change when unmodified, blank glass samples (both irradiated and not irradiated) are used to obtain control CFU values instead of not irradiated, functionalized samples. This clearly states that laser irradiation alone, in absence of plasmonic objects exerting photo thermal features, does not cause any harm to bacteria.

As reported in Table 2, in the case of ME<sub>T</sub> we observe an higher activity against *E. coli* than against *S. aureus*. This may be



due to the fact that Gram positive bacteria present a relatively thick (20–80 nm) and continuous cell wall consisting mainly of peptidoglycane, while Gram negative bacteria feature a thinner peptidoglycane layer (5–10 nm) surrounded by an outer phospholipidic membrane, which is expected to be less solid and resistant. The ME<sub>T</sub> test result show that the laser treatment kills more than 99.999% of *E. coli* and about 99.95% of *S. aureus* cells in 20 minutes of irradiation at 0.26 W cm<sup>-2</sup>, a value which is below the maximum allowed for exposure of the skin<sup>38</sup> and using a wavelength which is suitable for *in vivo* use.

At this point, one question arising was whether antibacterial effect given by laser irradiation was due only to hyperthermia or mediated by an augmented release of silver ions due the hyperthermia itself, as is reported for some described systems.<sup>12–14</sup> Thus, we measured the quantity of silver released in a fixed volume (3 mL) of water after 20 minutes of irradiation in the same conditions used for the ME<sub>T</sub> tests (laser spot 1 cm<sup>2</sup>, irradiance 0.26 W cm<sup>-2</sup>). After irradiation, the solution was brought to ICP, revealing that the quantity of silver ion released from the surface after 20 minutes of laser irradiation was identical, in the precision limits of the techniques, to the one found after 20 minutes of immersion in water in absence of irradiation (0.015 × 10<sup>-6</sup> g cm<sup>-2</sup>). Thus, we can state that the laser irradiation produces an antibacterial effect apparently due to hyperthermia only, and is not connected to an augmented silver release.

But what about the possibility to use the laser irradiation on SURF-PEI-PLATES samples after their immersion in water, to simulate a situation in which, in addition to a long-term microbicidal activity imparted by silver ions release, a reinforce of antibacterial action was desired? To answer this question, we repeated the ME<sub>T</sub> tests on SURF-PEI-PLATES samples after their immersion in 3 mL of water for 24 hours. As we have already described, thermograms did not show dramatic decrease of photo-thermal activity after immersion in water for 24 h, as UV-Vis-NIR spectra still show an intense absorption at 808 nm. As can be seen in Table 2, the ME<sub>T</sub> in these conditions is completely conserved, suggesting once again that the antibacterial action obtained within the 20 minutes irradiation is caused by hyperthermia from the silver nano-objects: as long as they absorb sufficient radiation at the proper wavelength, hyperthermia is produced and antibacterial action takes place.

## Conclusions

We demonstrated, for the first time, the coating of bulk surfaces with silver nanoplates which can exert an antibacterial action based on the photo-thermal effect, which can be switched on by a proper laser excitation. This novel kind of antimicrobial action is added to the well-known microbicidal abilities typically showed by AgNPs containing surfaces. The goal was reached by tuning the LSPR features of the nanoplates using a new seed growth synthesis, operated directly on bulk surfaces in absence of any surfactant or other harmful chemicals. We started by grafting small silver nanoparticles on bulk glass, previously treated with PEI-silane as grafting agent. This allowed to prepare stable (showing no release of nano-objects in

the environment) and reproducible samples bearing an homogeneous layer of flat silver nano-objects of various dimensions and shapes, with an overall broad absorption spectra having a maximum centered around 800 nm. Under laser irradiation at 808 nm and at safe irradiance values, in 20 minutes the almost complete elimination of bacteria *S. aureus* and *E. coli* is obtained *in vitro* through laser-induced hyperthermia. Of course, a complete microbicidal action is observed also in longer timescales, as a result of the contact with the surface, an expected<sup>3–7</sup> long-term antibacterial action which is explained with the slow and controlled release of silver ions from the nano-objects. Long time immersions in water produce morphological changes of the grafted object, reasonably related to silver ions release, but a 24 hours immersion do not affect seriously the antibacterial action obtainable by laser induced hyperthermia. We believe this new approach will have promising applications to several types of surfaces (prosthetic and subcutaneous devices, surgical sutures, wound dressings, to name a few examples), as it offers a long term antimicrobial protection which can be reinforced, on need, with the laser induced action.

## Acknowledgements

We gratefully acknowledge Prof. M. P. Riccardi, Arvedi Laboratory, CISRiC, Pavia, for carrying out SEM experiments. We also thank Prof. Marcella Bini, Department of Chemistry, University of Pavia, for XRD measures and fruitful discussions.

## Notes and references

- 1 D. Campoccia, L. Montanaro and C. R. Arciola, *Biomaterials*, 2013, **34**, 8018.
- 2 P. Pallavicini, G. Dacarro, Y. A. Diaz-Fernandez and A. Taglietti, *Coord. Chem. Rev.*, 2014, **275**, 38.
- 3 A. Taglietti, Y. A. Fernandez, E. Amato, L. Cucca, G. Dacarro, P. Grisoli, V. Necchi, P. Pallavicini, L. Pasotti and M. Patrini, *Langmuir*, 2012, **28**, 8140.
- 4 E. Amato, Y. A. Diaz-Fernandez, A. Taglietti, P. Pallavicini, L. Pasotti, L. Cucca, C. Milanese, P. Grisoli, C. Dacarro, J. M. Fernandez-Hechavarria and V. Necchi, *Langmuir*, 2011, **27**, 9165.
- 5 P. Pallavicini, A. Taglietti, G. Dacarro, Y. A. Diaz-Fernandez, M. Galli, P. Grisoli, M. Patrini, G. Santucci De Magistris and R. Zanoni, *J. Colloid Interface Sci.*, 2010, **350**, 110.
- 6 G. Dacarro, L. Cucca, P. Grisoli, P. Pallavicini, M. Patrini and A. Taglietti, *Dalton Trans.*, 2012, **41**, 2456.
- 7 A. Taglietti, C. R. Arciola, A. D'Agostino, G. Dacarro, L. Montanaro, D. Campoccia, L. Cucca, M. Vercellino, A. Poggi, P. Pallavicini and L. Visai, *Biomaterials*, 2014, **35**, 1779.
- 8 D. Jimenez de Aberasturi, A. B. Serrano-Montesadv and L. M. Liz-Marzán, *Adv. Opt. Mater.*, 2015, **3**, 602.
- 9 W. Jo and M. J. Kim, *Nanotechnology*, 2013, **24**, 195104.
- 10 S. A. Khan, A. K. Singh, D. Senapati, Z. Fan and P. C. Ray, *J. Mater. Chem.*, 2011, **21**, 17705.



- 11 P. Pallavicini, A. Donà, A. Taglietti, P. Minzioni, M. Patrini, G. Dacarro, G. Chirico, L. Sironi, N. Bloise, L. Visai and L. Scarabelli, *Chem. Commun.*, 2014, **50**, 1969.
- 12 S. Mo, X. Chen, M. Chen, C. He, Y. Lu and N. Zheng, *J. Mater. Chem. B*, 2015, **3**, 6255.
- 13 C. Fasciani, M. J. Silvero, M. A. Anghel, G. A. Arguello, M. C. Becerra and J. C. Scaiano, *J. Am. Chem. Soc.*, 2014, **136**, 17394.
- 14 K. C. L. Black, T. S. Sileika, J. Yi, R. Zhang, J. G. Rivera and P. B. Messersmith, *Small*, 2014, **10**, 169.
- 15 S. Boca-Farcau, M. Potara, T. Simon, A. Juhem, P. Baldeck and S. Astilean, *Mol. Pharm.*, 2014, **11**, 391.
- 16 S. C. Boca, M. Potara, A. M. Gabudean, A. Juhem, P. L. Baldeck and S. Astilean, *Cancer Lett.*, 2011, **311**, 131.
- 17 I. Pastoriza-Santos and L. M. Liz-Marzan, *J. Mater. Chem.*, 2008, **18**, 1725.
- 18 A. Casu, E. Cabrini, A. Donà, A. Falqui, Y. Diaz-Fernandez, C. Milanese, A. Taglietti and P. Pallavicini, *Chem.-Eur. J.*, 2012, **18**, 9381.
- 19 M. A. Mahmoud, C. E. Tabor, M. A. El-Sayed, Y. Ding and Z. L. Wang, *J. Am. Chem. Soc.*, 2008, **130**, 4590.
- 20 Y. Sun, *Chem. Mater.*, 2007, **19**, 5845.
- 21 A. M. Alkilany, P. K. Nalaria, C. R. Hexel, T. J. Shaw, C. J. Murphy and M. D. Wyatt, *Small*, 2009, **5**, 701.
- 22 K. H. Lee, K. M. Huang, W. L. Tseng, T. C. Chiu, Y. W. Lin and H. T. Chang, *Langmuir*, 2007, **23**, 1435.
- 23 K. Aslan, Z. Leonenko, J. R. Lakowicz and C. D. Geddes, *J. Phys. Chem. B*, 2005, **109**, 3157.
- 24 A. N. Sidorov, G. W. Sławinski, A. H. Jayatissa, F. P. Zamborini and G. U. Sumanasekera, *Carbon*, 2012, **50**, 699.
- 25 K. Nouneh, M. Oyama, R. Diaz, M. A. Lefdil, I. V. Kityk and M. Bousmina, *J. Alloys Compd.*, 2011, **509**, 2631.
- 26 H. Jia, J. Zeng, J. An, W. Song, W. Xu and B. Zhao, *Thin Solid Films*, 2008, **516**, 5004.
- 27 Y. K. Kim and D. H. Min, *RSC Adv.*, 2014, **4**, 6950.
- 28 D. Aherne, D. M. Ledwith, M. Gara and J. M. Kelly, *Adv. Funct. Mater.*, 2008, **18**, 2005.
- 29 D. M. Ledwith, A. M. Whelan and J. M. Kelly, *J. Mater. Chem.*, 2007, **17**, 2459.
- 30 J. Liu, D. A. Sonshine, S. Shervan and R. H. Hurt, *ACS Nano*, 2010, **4**, 6903.
- 31 G. Q. Zhao, L. H. Ye, Y. C. Huang, D. K. Yang, L. Li and G. Xu, *Cell Biochem. Biophys.*, 2011, **61**, 371.
- 32 N. Katsui, T. Tsuchido, R. Hiramatsu, S. Fujikawa, M. Takano and I. Shibasaki, *J. Bacteriol.*, 1982, **151**, 1523–1531.
- 33 W. Jo and M. J. Kim, *Nanotechnology*, 2013, **24**, 195104–195111.
- 34 Z. M. Xiu, Q. B. Zhang, H. L. Puppala, V. L. Colvin and P. J. Alvarez, *Nano Lett.*, 2012, **12**, 4271.
- 35 J. Chen, S. Li, J. Luo, R. Wan and W. Ding, *J. Nanomater.*, 2016, 7135852.
- 36 D. Paul, S. Paul, N. Roohpour, M. Wilks and P. Vadgama, *J. Funct. Biomater.*, 2013, **4**, 358–375.
- 37 H. Zhang, W. Meng and S. Ayusman, Silver nanoparticle antimicrobials and related materials, in *Nano-antimicrobials*, Springer, Berlin, Heidelberg, 2012, pp. 3–45.
- 38 ANSI, *American National Standard for Safe Use of Lasers*, Laser Institute of America, Orlando, FL, 2000.

