

Cite this: *RSC Adv.*, 2016, 6, 39688

# Nioplexes encapsulated in supramolecular hybrid biohydrogels as versatile delivery platforms for nucleic acids†

Santiago Grijalvo,<sup>abc</sup> Gustavo Puras,<sup>cd</sup> Jon Zárate,<sup>cd</sup> Ramon Pons,<sup>b</sup> Jose Luis Pedraz,<sup>cd</sup> Ramon Eritja<sup>bc</sup> and David Díaz Díaz<sup>\*ab</sup>

Supramolecular hydrogels based on N-protected phenylalanine (Fmoc-Phe-OH) were used to encapsulate non-ionic surfactant vesicles (niosomes). The niosomes consisted of an amphiphilic lipid mixed with polysorbate-80 and electrostatically complexed with a fluorescently labelled oligodeoxynucleotide (FITC-ODN) as a model nucleic acid derivative. The diffusion properties of the supramolecular hydrogel were conveniently tuned by adding a small amount of  $\kappa$ -carrageenan ( $\leq 1\%$  w/v) as a crosslinking agent. Interestingly, neither cationic niosomes nor the biopolymer additive significantly affected the hydrogelation properties of the amino acid-based low molecular weight (LMW) gelator. *In vitro* drug release experiments from Fmoc-Phe-OH hydrogels containing cationic niosomes were successfully carried out in the absence and in the presence of  $\kappa$ -carrageenan. The niosomal ODN liberation in solution was fitted using Higuchi, Korsmeyer-Peppas and Weibull drug release models, showing the prevalence of diffusion mechanisms in each case. Moreover, the time release was easily prolonged by increasing the concentration of  $\kappa$ -carrageenan. Preliminary transfection studies indicate the suitability of these supramolecular hybrid hydrogels to embed niosomal formulations and, consequently, for being used as tunable delivery vehicles for nucleic acids.

Received 12th January 2016  
Accepted 8th April 2016

DOI: 10.1039/c6ra01005a

www.rsc.org/advances

## 1. Introduction

Since Wichterle and Lim reported for the first time the synthesis of crosslinked poly(hydroxyethylmethacrylate) (pHEMA) for biological use,<sup>1</sup> synthetic and natural polymer-based hydrogels have gained substantial importance over the years in a good number of biomedical, pharmaceutical and biotechnological applications such as tissue engineering, drug delivery systems or cell culture scaffolds.<sup>2–5</sup> The research in this area has contributed to the development of novel smart polymer gel networks having specific responses to external stimuli such as pH, light, temperature or chemical additives,<sup>6–8</sup> which could facilitate the controlled liberation of embedded therapeutic drugs.<sup>9</sup>

In contrast to polymer or chemical gels<sup>10</sup> that are based on covalent bonds, physical or supramolecular gels<sup>11</sup> are typically made of low molecular weight (LMW) compounds self-assembled through non-covalent interactions (*e.g.*, hydrogen-bonding,  $\pi$ - $\pi$  stacking, van der Waals, dipole-dipole, charge-transfer and coordination interactions). In general, such non-covalent nature of supramolecular gels provides reversible *gel-to-sol* phase transitions as response to external stimuli. The solid-like appearance and rheological properties of the gels result from the immobilization of the liquid (major component) into the interstices of a solid matrix (minor component) mainly through capillary forces.<sup>12</sup> The formation of the 3D-network with numerous junction zones results from the entanglement of 1D-suprapolymeric strands of gelator molecules usually of nm diameters and mm lengths.<sup>13</sup> The synthetic versatility of many LMW gelators together with their biocompatible properties have allowed their use in a number of biomedical applications including cell adhesion, cell growth, tissue engineering and controlled drug delivery.<sup>14–17</sup> Among the plethora of LMW compounds that are able to form supramolecular hydrogel networks, amino acid derivatives and peptide conjugates<sup>18–23</sup> constitute one of the most studied groups. Within this context, the use of phenylalanine (Phe) derivatives as hydrogelator was reported for the first time in 2006.<sup>24,25</sup> In 2011, Roy and Banerjee<sup>26</sup> reported the ability of *N*-terminally

<sup>a</sup>Institute of Organic Chemistry, University of Regensburg, Universitätsstrasse. 31, D-93040 Regensburg, Germany. E-mail: David.Diaz@chemie.uni-regensburg.de; Fax: +49 941 943 4121; Tel: +49 941 943 4373

<sup>b</sup>Institute of Advanced Chemistry of Catalonia (IQAC-CSIC), Spain

<sup>c</sup>Biomedical Research Networking Center in Bioengineering, Biomaterials and Nanomedicine (CIBER BBN), Spain

<sup>d</sup>NanoBioCel group, University of the Basque Country (EHU-UPV), Spain

† Electronic supplementary information (ESI) available: Digital pictures, NMR spectroscopy, rheological measurements, fluorescence calibration curve, control experiments, fitting models, native PAGE gels and cell morphology images. See DOI: 10.1039/c6ra01005a



Fmoc protected Phe, Fmoc-Phe-OH, to form stable and transparent hydrogels at concentrations as low as 0.1% w/v.

On the other hand, the development of more efficient and safe strategies for the administration of a drug is a key step for the success in modern therapeutic processes. Besides the classical therapeutic approaches to find a drug, gene therapy and gene silencing strategies have emerged as promising treatment options for a number of diseases. However, the search of an optimal administration is a key to getting an efficient therapeutic action. Thus, colloidal drug delivery systems including nanoparticles, liposomes and hydrogels, among others, have experienced significant growth in recent years based on the necessity of optimizing the transportation of poorly-water soluble drugs as well as improving the bioavailability and pharmacokinetics of numerous nucleic acid therapeutics.<sup>27</sup> Within this context, niosomes (non-ionic surfactant vesicles) have become promising candidates for drug delivery applications<sup>28,29</sup> and an alternative to the use of liposomes, which are considered the keystone of particle-based carrier systems. This growing interest of niosomes is mainly due to their good physical stabilities, cost-effectiveness, easy large-scale synthesis and their structural similarities to liposomes. Moreover, they are able to efficiently entrap hydrophilic and lipophilic drugs either in aqueous compartments or in bilayer domains, respectively. This efficiency has been recently reported by some of us with the use of nioplexes, elaborated with cationic niosomes, for both *in vitro* and *in vivo* transfection experiments.<sup>30,31</sup>

Although liposomes, niosomes and other conventional lipid particles have mainly prepared to obtain more efficient non-druggable therapeutic formulations, the stability of such particles may be lessened by the action of macrophages and their rapid renal clearance. Several approaches have been reported to overcome this drawback, including variation of particle size, surface charge, lipid composition or the addition of different ligands and/or receptors.<sup>32,33</sup> Alternatively, the encapsulation of such colloidal particles either in synthetic<sup>34</sup> or natural polymers-based hydrogels<sup>35</sup> has also generated promising and fascinating hybrid materials that has helped to improve the efficacy of such liposomal formulations.<sup>36,37</sup> In fact, this kind of entrapment has generated efficient colloidal particle protections and thereby has reduced both undesirable burst release effects and improved drug delivery when therapeutic molecules have been administered<sup>38,39</sup> More recently, novel hybrid materials like injectable hydrogels containing liposomes with self-healing or shear-thinning properties have also proven to be another promising approach employed for therapeutic purposes.<sup>40</sup> In particular, hydrogel synthesis from polymer-nanoparticle interactions<sup>41</sup> or responsive hydrogel materials for intratumoral delivery have been designed for suitable *in vitro* and *in vivo* drug delivery processes and for some cancer treatments<sup>42</sup> by increasing the efficacy and reducing toxicity, respectively. Recently, multidomain peptides (MDPs), which have the ability to self-assemble and generate nanofiber hydrogels, have also been used to entrap liposomes containing several growth factors.<sup>43,44</sup>

In this work we describe the synthesis of lipid-based nioplexes bearing an oligodeoxynucleotide as a model biomolecule. These nioplexes were subsequently encapsulated within supramolecular hydrogels based on self-assembled Fmoc-Phe-OH molecules.<sup>23</sup> Some hydrogels were also physically cross-linked by the addition of  $\kappa$ -carrageenan. *In vitro* release kinetics of the niosomal formulation from these hydrogels was investigated and adjusted to several mathematical models. Finally, the suitability of hybrid materials in the transfection of nucleic acids (e.g. antisense oligonucleotide and plasmid DNA)

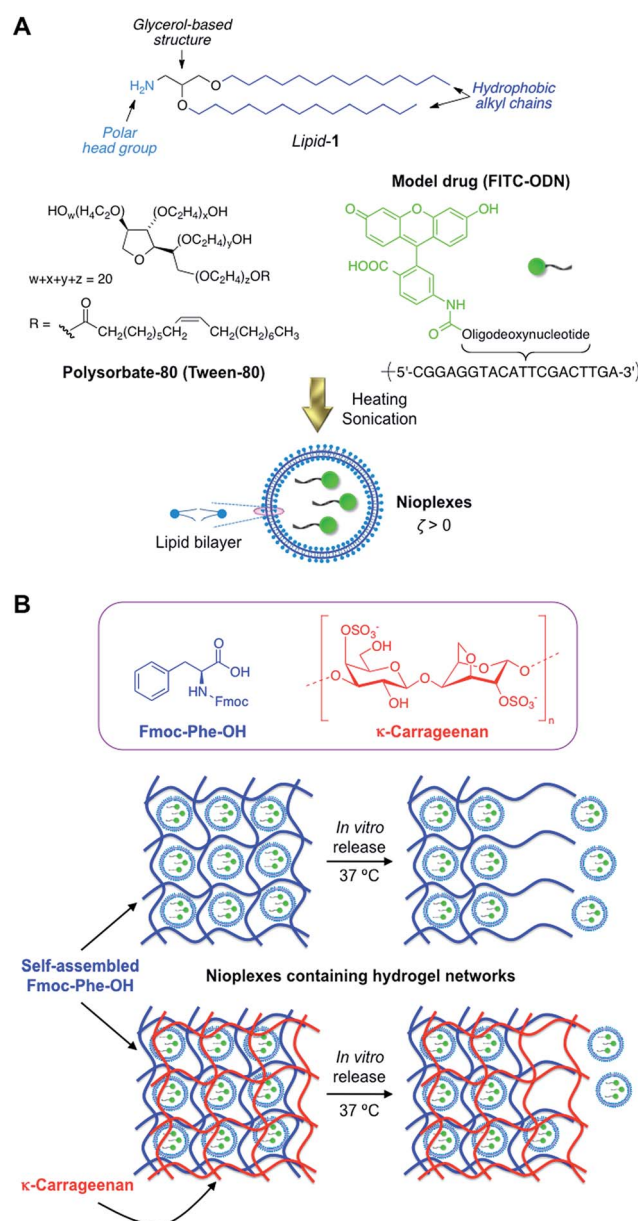


Fig. 1 Strategy used for the preparation of Fmoc-Phe-OH supramolecular hydrogels containing cationic nioplexes and FITC-ODN as a model nucleic acid. (A) Preparation of cationic nioplexes containing FITC-ODN. (B) Illustration of the encapsulation and *in vitro* release studies of niosomal FITC-ODN from Fmoc-Phe-OH-based hydrogels (hydrogel-1) and Fmoc-Phe-OH hydrogels physically crosslinked with  $\kappa$ -carrageenan (hydrogels-(2–4)).



in cell culture was also evaluated. The overall concept of this work is illustrated in Fig. 1. As far as we are aware, the use of LMW hydrogels to encapsulate nioplexes derived from cationic niosomes for the controlled delivery of therapeutic molecules has not yet been reported.

## 2. Results and discussion

### 2.1 Synthesis and characterization of hydrogels containing nioplexes

The synthesis of the amphiphilic *lipid-1* (ESI, Fig. S1†) and the subsequent preparation of the cationic niosomal formulation were successfully accomplished following the experimental procedures described in our previous studies.<sup>30,31</sup> Briefly, the cationic nioplexes used in this study were made upon mixing *lipid-1* with a non-ionic surfactant agent (polysorbate-80) and a fluorescently labelled oligodeoxynucleotide (FITC-ODN) as model antisense inhibitor (Fig. 1). The so obtained nioplexes were subsequently mixed with a suitable amount of the LMW

gelator (Fmoc-Phe-OH) in phosphate buffered saline (PBS, pH 7.4) solution and gently heated in a sealed tube.<sup>26</sup> After cooling the mixture down to room temperature, a transparent hydrogel was obtained (*hydrogel-1*; ESI, Fig. S2A†). Additionally, we prepared similar cationic nioplexes loaded-hydrogels but physically crosslinked with  $\kappa$ -carrageenan with the aim of tuning the diffusional properties of the matrix under physiological conditions. Thus, nioplexes-containing Fmoc-Phe-OH-based *hydrogels-2-4* crosslinked with increasing concentration of  $\kappa$ -carrageenan were prepared in PBS buffer (*i.e.*, *hydrogel-2*, *hydrogel-3* and *hydrogel-4* containing 0.5, 0.8 and 1% w/v of  $\kappa$ -carrageenan, respectively) (ESI, Fig. S2B†). All hydrogels (1–4) were able to encapsulate nioplexes with high efficiency (EE > 90%) and the gelation ability of Fmoc-Phe-OH was not significantly influenced by the presence of both  $\kappa$ -carrageenan and niosomes (ESI, Table S1†).

The mechanical properties of representative composite hydrogels (*i.e.*, *hydrogel-1* and *hydrogel-4*) were investigated by oscillatory rheological measurements and compared to control

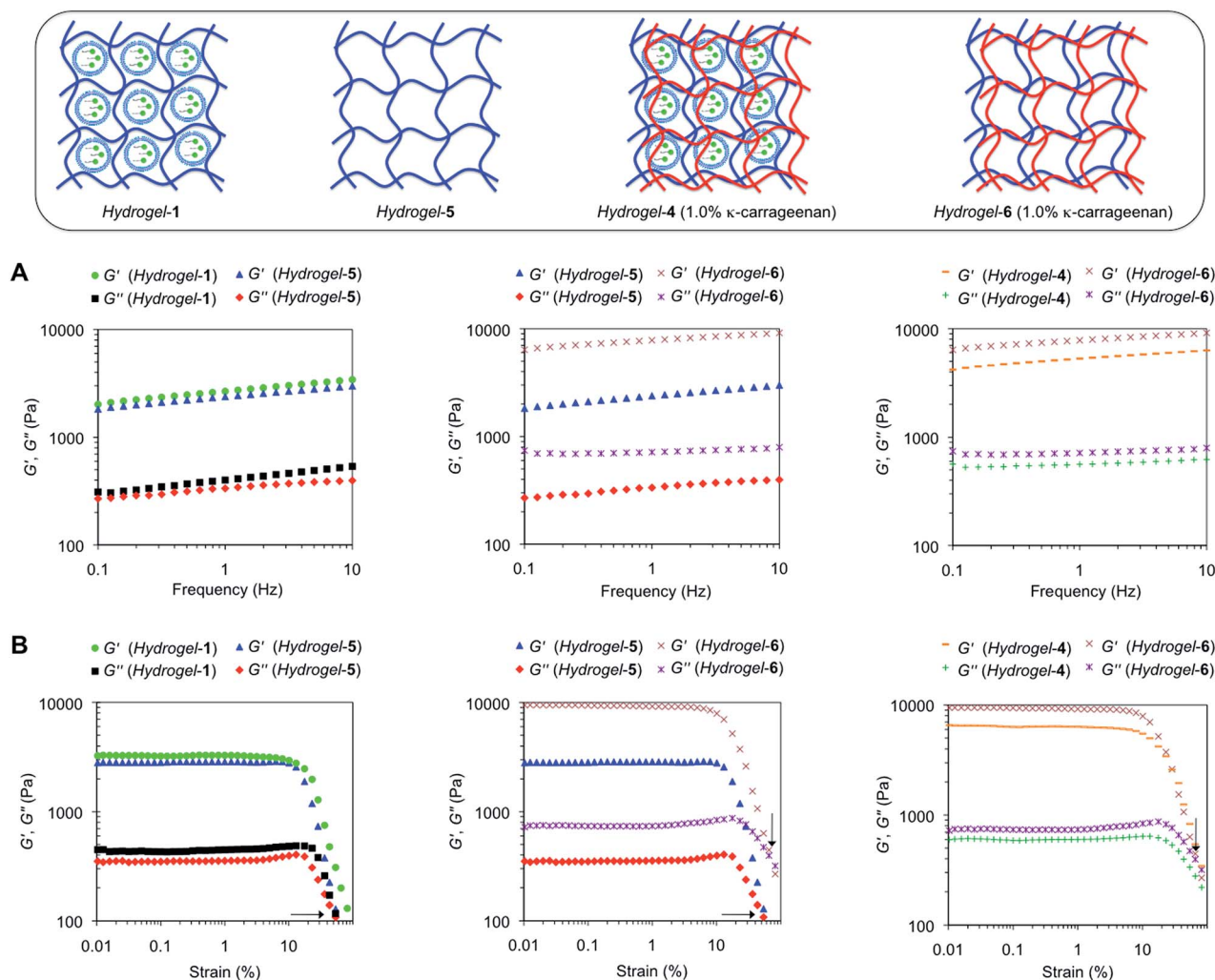


Fig. 2 Comparative rheological DFS (A) and DSS (B) measurements between *hydrogel-1* and native hydrogel (*hydrogel-5*) (left); native hydrogel (*hydrogel-5*; without cationic vesicles) and physically crosslinked hydrogel with  $\kappa$ -carrageenan (*hydrogel-6*; without cationic vesicles) (middle); *hydrogel-4* and *hydrogel-6* (right). The arrows in DSS plots indicate the cross-point between  $G'$  and  $G''$  ( $\gamma$  at break). The hydrogels were prepared in PBS (pH 7.4) and measured at room temperature (25 °C).





gels prepared in the absence of niosomes (*i.e.*, native *hydrogel-5* and *hydrogel-6*, which was only tuned with  $\kappa$ -carrageenan at 1% w/v) (Fig. 2). The storage modulus ( $G'$ ) and the loss modulus ( $G''$ ) were measured as a function of frequency (DFS, dynamic frequency sweep), strain (DSS, dynamic strain sweep) and time (DTS, dynamic time sweep). The viscoelastic nature of all samples was evidenced by  $G'$  values *ca.* one order of magnitude greater than  $G''$  and a low dependence of  $G'$  with the applied frequency. The addition of cationic niosomes to non-crosslinked hydrogels (*i.e.*, *hydrogel-1* vs. *hydrogel-5*) did not change the mechanical properties of the network, which displayed similar dissipation factor values ( $\tan \delta = G''/G' \approx 0.15$ ) (Fig. 2A; ESI, Table S1†), indicating a good response of both hydrogels to external forces. Interestingly, the incorporation of small amounts of  $\kappa$ -carrageenan (*hydrogel-6*) produced an increase in  $G'$  values compared to those obtained for the analogues non-crosslinked hydrogels (*i.e.*, *hydrogel-6* vs. *hydrogel-5*; *hydrogel-4* vs. *hydrogel-1*) (Fig. 2A).<sup>45</sup> Thus, the presence of  $\kappa$ -carrageenan seems to produce highly associated materials due to colloidal forces between the cationic niosomes and the anionic biopolymer. This was also reflected in the oligonucleotide release rates (*vide infra*). Again, the incorporation of cationic niosomes into hydrogels containing  $\kappa$ -carrageenan (*hydrogel-4*) did not induce any significant change in the rheological properties when compared to *hydrogel-6*.

On the other hand, DSS (dynamic strain sweep) measurements confirmed that all hydrogels remained intact while applied an oscillation stress within the linear viscoelastic regime (Fig. 2B; ESI, Fig. S3†). The presence of  $\kappa$ -carrageenan in the formulations produced only slight differences in the critical strain (ESI, Table S1†). Above the critical strain, crosslinked hydrogels induced a slighter resistance to the shear stress ( $\gamma$ ) until  $G''$  exceeded  $G'$  (*i.e.*,  $\gamma$  at break = 68% vs. 43% for *hydrogel-6* and *hydrogel-5*, respectively). The encapsulation of cationic niosomes into the gel matrices also increased slightly the critical strain  $\gamma_c$  values in comparison to native hydrogels (*i.e.*,  $\gamma_c$  = 16.6% and 20% for *hydrogel-1* and *hydrogel-4* vs. 13% and 13.4% for *hydrogel-5* and *hydrogel-6*, respectively) (ESI, Fig. S3 and Table S1†). A similar behaviour was observed in the resistance of the hybrid gels to shear stress (*e.g.*,  $\gamma$  at break = 87% vs. 68% for *hydrogel-4* and *hydrogel-6*, respectively). These differences were also observed for *hydrogel-1* when compared to the corresponding native material. These results suggest that the presence of  $\kappa$ -carrageenan and cationic niosomes within the supramolecular gel network could induce certain deformations in the systems under specific stress conditions, thus increasing the hydrogel rigidity and the corresponding shear stress resistance.<sup>46</sup> Finally, DTS (dynamic time sweep) measurements confirmed the temporal stability of the supramolecular hydrogels within the viscoelastic regime (0.1% strain, 1 Hz frequency) (ESI, Fig. S4†).

## 2.2 Small-angle X-ray scattering measurements

The behavior of *lipid-1* was investigated in aqueous media and their diffraction patterns (SAXS and WAXS experiments) were recorded at several temperatures (Fig. 3). SAXS curves were

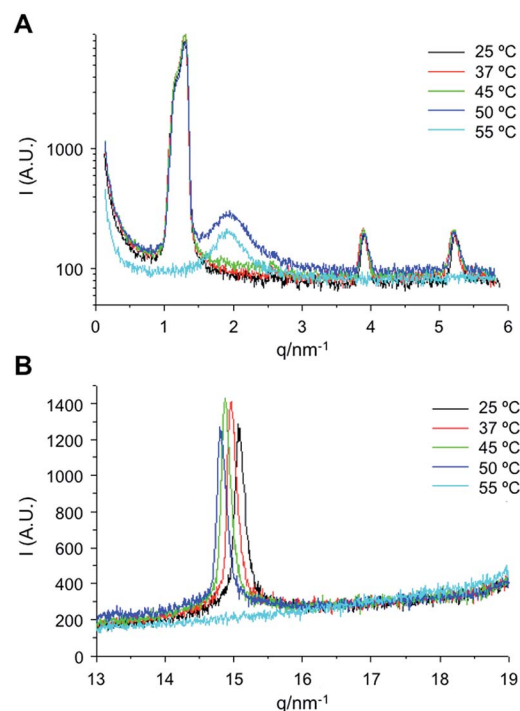


Fig. 3 (A) Scattered intensity of the fully hydrated lipid as a function of dispersion vector  $q$  at different temperatures. (B) WAXS spectra at several temperatures of the fully hydrated lipid as a function of scattering vector modulus  $q$ .

identical at both 25 °C and 37 °C whereas some increased background was appreciated around  $q = 2 \text{ nm}^{-1}$  at 45 °C (Fig. 3A). A partial fusion was observed at 55 °C with the presence of a band centered at  $q = 1.94 \text{ nm}^{-1}$  although the peaks did not show appreciable changes. At 55 °C, a complete fusion was observed and the band only remained in the SAXS domain. Importantly, the SAXS pattern at 50 °C did not change as a function of time (the three consecutive acquisitions were identical within experimental error), suggesting the coexistence of the lamellar and melted phases at this temperature. Therefore, we carried out the ODN encapsulation and release studies below this temperature in order to ensure the integrity of the vesicles during the experiments.

Using the same range of differing temperatures, the WAXS region (Fig. 3B) showed a single peak for the samples, which determined a lamellar order. This experiment confirmed a slight shift to smaller  $q$  values due to the increased mobility of the hydrophobic chains. This rise showed an interchain distance increase from 0.417 nm to 0.425 nm, which in turn implied a density decrease of about 3.5%.

Additionally, the fitting of the experimental scattering data recorded at 25 °C was consistent with a lamellar model (Fig. 4A). It is noteworthy the absence of the expected second peak for the lamellar arrangement. This is related to the electronic profile of the lamellae coupled with the interlamellar distance. This can be appreciated from the single lamellae curve shown in red. The first minimum coincides with the position where the second peak should appear. Fig. 4B shows the corresponding electronic density profile for the lamellae compared with the electronic



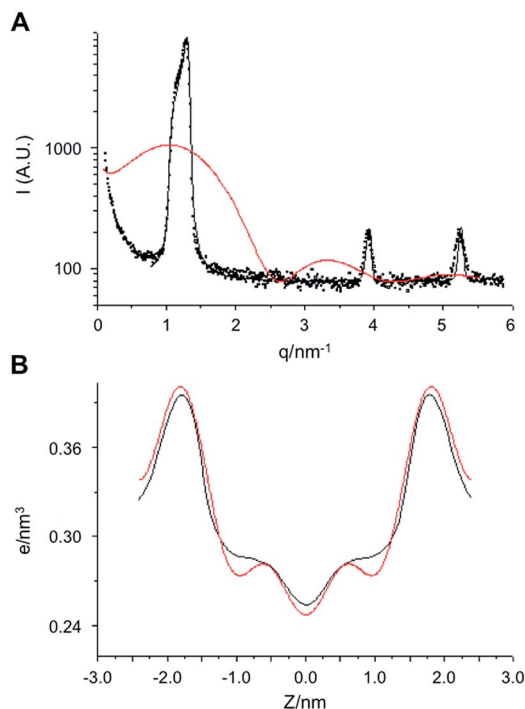


Fig. 4 (A) SAXS intensity as a function of dispersion vector  $q$  of the fully hydrated lipid at 25 °C (open black squares), fitted profile (black line) and bilayer form factor (red line). (B) Electronic density profile corresponding to the fitting in Fig. 3. Black line corresponds to the MCG profile and the red line to the Fourier reconstruction. The Fourier reconstruction was scaled to produce the same integral as the MCG model.

density profile obtained from the Fourier reconstruction using the three visible peaks. Although both electronic profiles are consistent, we believe that our model profile is more realistic in the sense that there is not a physical reason for the secondary minima around  $\pm 1$  nm from the bilayer center. From the electronic profile and the known volume and electron composition of the molecule we determined a hydrophobic bilayer half thickness of  $1.43 \pm 0.1$  nm, a total bilayer thickness of  $4.8 \pm 0.1$  nm, an area per molecule of  $0.541 \pm 0.003$  nm<sup>2</sup> and  $13 \pm 2$  water molecules per polar head. This amount of water roughly corresponds to a 30% hydration. Furthermore, the sharpness of the peaks suggests a strong correlation between the bilayers, which corresponds to  $28 \pm 2$  correlated bilayers, according to the model.

### 2.3 *In vitro* release studies

As mentioned above, in order to investigate and validate the efficacy of *hydrogels*-(1–4) as drug carriers, we studied the release kinetics of physiologically important biomolecules like oligonucleotides, which are involved in well-known gene regulation processes.<sup>47,48</sup> The interaction between fluorescently labelled oligodeoxynucleotide FITC-ODN<sup>49</sup> and cationic niosomes afforded the corresponding complexes (nioplexes) (Fig. 1A) with an optimal amine to phosphate (N/P) charge ratio of 14 as determined by the electrophoretic mobility shift assay (EMSA) (Fig. 5).

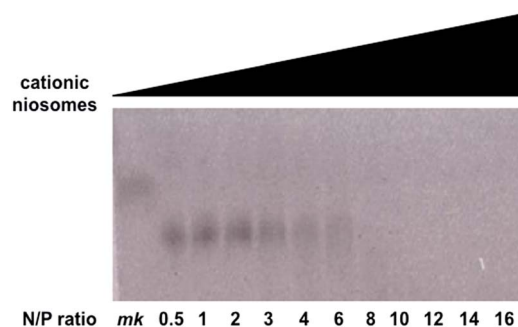


Fig. 5 Electrophoretic mobility shift assay (EMSA). FITC-ODN was complexed with growing cationic niosomes N/P charge ratios in 100 mM of PBS in a native polyacrylamide gel (20%) at 150 V and 25 °C. Bands were stained with SYBR-Green II and bromophenol blue was used as a marker (*mk*). The disappearance of the FITC-ODN band indicated the completion of the complex formation (*i.e.*, N/P ratio = 12).

*In vitro* release studies of the composite hydrogels made of Fmoc-Phe-OH and (FITC-ODN)-loaded cationic nioplexes (1–4) were performed at 37 °C to emulate physiological conditions. As a reservoir for the release, *hydrogels*-(1–4) were immersed in PBS solution and aliquots were removed at regular intervals and analyzed by fluorescence spectroscopy according to a proper standard curve of FITC-ODN in solution ( $r^2 > 0.99$ ) (ESI, Fig. S5†). The removed volume was always replaced by the same amount of fresh buffer. In the case of *hydrogel*-1, the release kinetics was controlled by erosion of the polymeric matrix and *ca.* 90% of the FITC-ODN was rapidly liberated in *ca.* 1.5 h (Fig. 6).

It is well-accepted that the rate of drug release can be modulated by adding additional ligands to the gel networks like synthetic molecules<sup>50</sup> and biopolymers<sup>51,52</sup> as crosslinking agents. This convenient strategy has helped to improve composite formulations and prolong the release of both small therapeutic drugs and macromolecules.<sup>53</sup> This encouraged us to incorporate in our hydrogel formulation growing concentrations of  $\kappa$ -carrageenan (*i.e.*, 0.5, 0.8 and 1%; *hydrogel*-2, *hydrogel*-3 and *hydrogel*-4, respectively)<sup>54</sup> that would act as a physical crosslinking agent. The total cumulative release profiles of niosomal FITC-ODN complexes from these hydrogels cross-linked with  $\kappa$ -carrageenan are shown in Fig. 7. In general, the incorporation of the biopolymer within the supramolecular gel network made of Fmoc-Phe-OH prolonged considerably the FITC-ODN releasing time compared to that observed for *hydrogel*-1. Specifically, *ca.* 80–90% of FITC-ODN was available in the receptor phase after 5 h of incubation of *hydrogels*-(2–4). This result suggests a fall in hydrogel erosion processes induced by the presence of the biopolymer.<sup>55</sup> Furthermore, it is worth mentioning that the presence of  $\kappa$ -carrageenan in the formulation was able to reduce the initial burst release during the first few minutes of the experiment (19% *vs.* 8% for *hydrogel*-1 and *hydrogels*-(2–4), respectively) (Fig. 6A and 7). This was probably due to the increase of the entanglement density of the non-covalently crosslinked hydrogels.



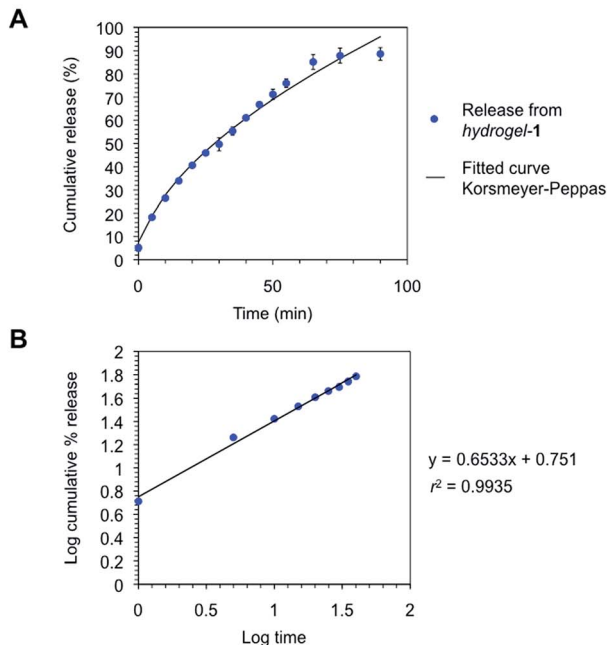
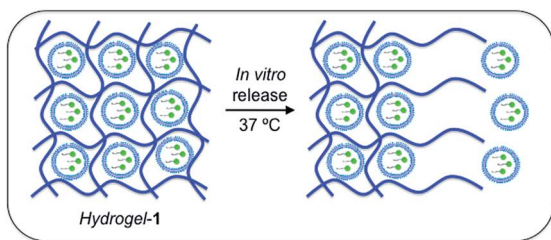


Fig. 6 (A) Cumulative release profile of the niosomal formulation containing FITC-ODN from *hydrogel-1* in PBS solution at 37 °C, along with the fitted curve ( $n = 2$ ) according to Korsmeyer–Peppas equation [eqn (4)]. (B) log–log plot showing the monomial relationship between FITC-ODN concentration and time. Model fitting parameters are given in the ESI (Table S3†).

Although the obtained niosomal FITC-ODN release profiles from physically crosslinked *hydrogels*-(2–4) were similar, the faster release was observed for *hydrogel-4* having the highest concentration of  $\kappa$ -carrageenan (1% w/v). This could be explained by the increase of the hydrophilic character of the *hydrogel-4* network in which additional biopolymer may contribute to a better dissolution of the swollen supramolecular matrix and thus promoting hydrogel erosion processes and thereby accelerating the drug release.

The analysis of the niosomal release was also studied by native polyacrylamide gel electrophoresis (PAGE) in order to clarify whether the presence of the cationic niosomes loaded in the supramolecular hydrogel could delay and control the oligonucleotide release from the supramolecular hydrogel.

The *in vitro* control release was carried out as described above and the collected aliquots in PBS were analyzed by PAGE (Fig. S7†). As a control, the same release experiment was also studied in the presence of 100 mM Triton-X100 (Fig. S7B†). As displayed in Fig. S7A,† the oligonucleotide is partially released from the hydrogel at different times which suggests the presence of cationic niosomes might control the FITC-ODN

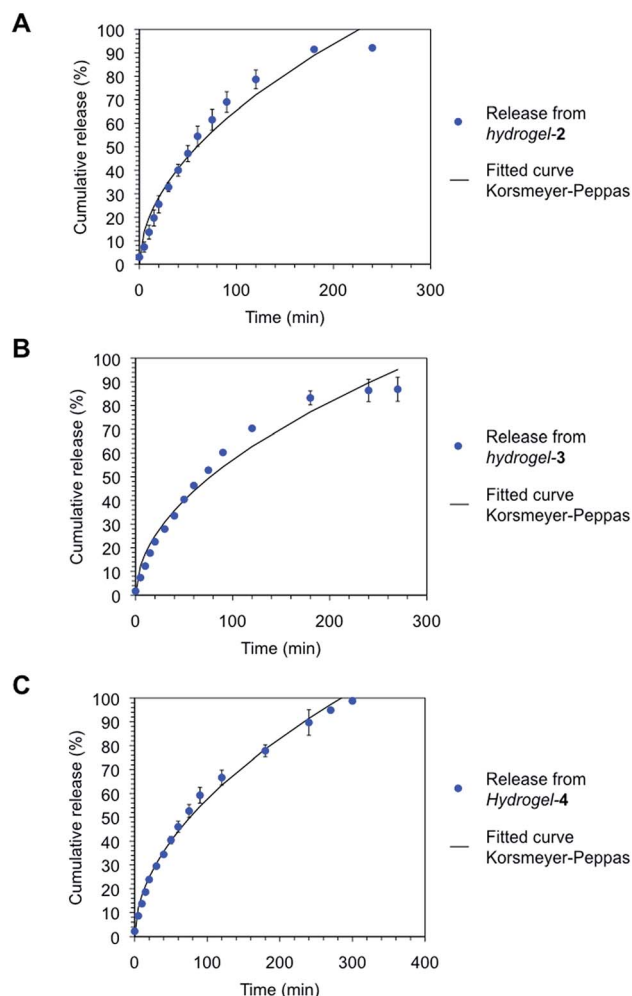
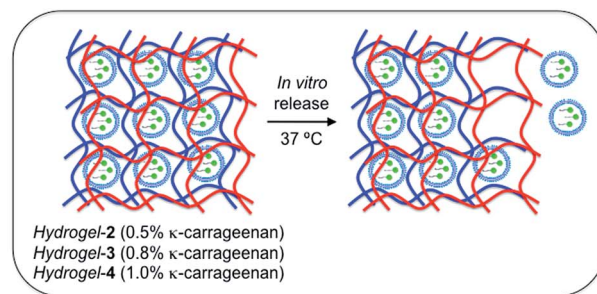


Fig. 7 Cumulative release profiles and corresponding fitted curves ( $n = 2$ ; according to Korsmeyer–Peppas equation [eqn (4)]) of the niosomal formulation containing FITC-ODN from (A) *hydrogel-2*, (B) *hydrogel-3* and (C) *hydrogel-4* in PBS solutions at 37 °C. Hydrogels were crosslinked with growing concentrations of  $\kappa$ -carrageenan (0.5, 0.8 and 1% w/v, respectively). Model fitting parameters and log–log plots are given in the ESI (Table S3 and Fig. S8†). The combination of the three release curves is displayed in the ESI (Fig. S6†).

liberation through the hydrogel. To corroborate these findings, Triton-X100 solution was added at the beginning of the experiment in order to solubilize the niosomal component and thus generate the unformulated FITC-ODN release in the receptor phase.<sup>56</sup> As expected, native PAGE gels confirmed the use of the surfactant generated a faster oligonucleotide release





at the first times of the release experiment (at  $t = 15$  min *et seq.*). This effect was also confirmed from the analyses of the anticipated band intensities. However, it was very difficult to visualize and confirm the presence of intact niosomal particles in the receptor phase by monitoring the release by native PAGE.

## 2.4 Drug release mathematical models

The release data obtained for *hydrogels*-(1–4) was fitted according to three theoretical models that describe the diffusion-related drug release from polymeric matrices. The selected models are defined by the Higuchi equation [eqn (3)], Korsmeyer–Peppas equation for burst release [eqn (4)] and Weibull equations for delayed release [eqn (5)].<sup>57–59</sup> The first two models are short time approximations and generally limited to the first 60% of the release.<sup>60</sup>

The total FITC–ODN cumulative release from all hydrogel matrices was found to follow a Korsmeyer–Peppas kinetic model (*i.e.*, their regression coefficients were greater ( $r^2 > 0.99$ ) than the rest of theoretical models (ESI, Tables S2–S4†)). According to Peppas' equation, the FITC–ODN release afforded  $n$  (diffusion coefficient) values ranging from 0.51 to 0.56. These exponent values are in close agreement with those predicted for non-Fickian diffusions.<sup>57</sup> This anomalous transport suggests the presence of specific interactions and/or erosion mechanisms between cationic niosomes and the fibrous gel matrices. Overall, the results of this study suggest a diffusion-controlled release as the main process that govern the liberation of FITC–ODN from the hydrogel matrices.<sup>61</sup>

## 2.5 MTT assay

To evaluate the cytocompatibility of Fmoc–Phe hydrogels crosslinked with  $\kappa$ -carrageenan in combination with cationic niosomes and nioplexes, a MTT reductase assay was carried out.<sup>62</sup> This assay is based on the tetrazolium salt reduction to formazan crystals by a dehydrogenase that is present in mitochondria. Then, untreated cells (*Blank*) and HeLa cells only treated with *hydrogel-3* (*Mock*) were selected as our negative controls. Furthermore, cationic niosomes-loaded *hydrogel-3* without forming complexes at two concentrations (1.7  $\mu\text{M}$  and 4.2  $\mu\text{M}$ , referred to lipid concentration) besides *hydrogel-3* containing cationic nioplexes (120 nM and 300 nM, referred to oligonucleotide concentration) were also evaluated.

When cultured for 24 h, *hydrogel-3* was totally degraded whereas both cationic niosomes and nioplexes were released from the hydrogel. As displayed in Fig. 8A, the anticipated hydrogel and cationic niosome hybrids did not show any evidence of toxicity obtaining good cellular viabilities (up to *ca.* 80%). These results also demonstrated normal cell morphology as illustrated in Fig. 8B–D which confirmed the suitability of the *hydrogel-3* to be used in cell culture.

## 2.6 Preliminary *in vitro* drug delivery studies

Firstly, we selected a phosphorothioate oligonucleotide complementary to the mRNA luciferase gene<sup>63</sup> to evaluate the gene silencing properties and therefore confirm the niosomal release from hydrogels but unfortunately the presence of Fmoc–

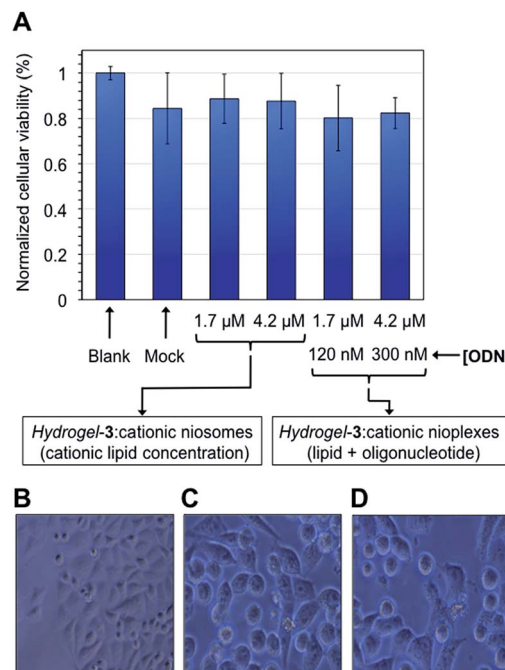


Fig. 8 (A) Cell viability studies according to the MTT assay in HeLa cells in the presence of *hydrogel-3* (*Mock*) and *hydrogel-3* containing both cationic niosomes (at 1.7  $\mu\text{M}$  and 4.2  $\mu\text{M}$ ) and cationic nioplexes (at 120 nM and 300 nM). Nioplexes were obtained from a N/P ratio of 14 according to EMSA assays. Blank corresponds to untreated HeLa cells. Data were means  $\pm$  SD of six independent experiments and the normalized cellular viability was expressed as a percentage to the untreated control cells. A phosphorothioate oligonucleotide complementary to the mRNA luciferase gene (5'-CGT TTC CTT TGT TCT GGA-3') was used as an antisense oligonucleotide model. (B) HeLa cell morphology images of untreated cells (blank). (C) HeLa cell morphology images in the presence of cationic niosomes. (D) HeLa cell morphology images in the presence of cationic nioplexes. HeLa cell images in the presence of mock (only hydrogel) are displayed in Fig. S9†

Phe inhibited the luciferase gene, as described for other Phe analogues.<sup>64</sup>

For this reason, we moved forward and selected pMax-GFP plasmid as a model to carry out transfection studies. For the preparation of the niosomal formulation, we used the same protocol as described by our research group previously.<sup>30</sup> After forming GFP nioplexes at two N/P ratio (4 and 6), particles were entrapped in *hydrogel-3* and placed in a transwell insert system. As a control experiment, GFP plasmid, in the absence of cationic niosomes, was encapsulated within the hydrogel. After 24 h of incubation, hydrogel was practically degraded and cellular uptake was analyzed by flow cytometry (Fig. 9). Firstly, we selected a normal gate (R1) for HeLa cells based on their light scatter properties, which distinguished single cells by relative size (Fig. 9A). This allowed us to analyze specific populations and selectively focus on the relationship between fluorescently labeled and unlabeled cell populations.

As displayed in Fig. 9E, our results suggest that GFP was taken more efficiently by HeLa cells when GFP-forming complexes with the *lipid-1* at N/P ratio of 6 were entrapped in *hydrogel-3* (17%) according to the gate positive population



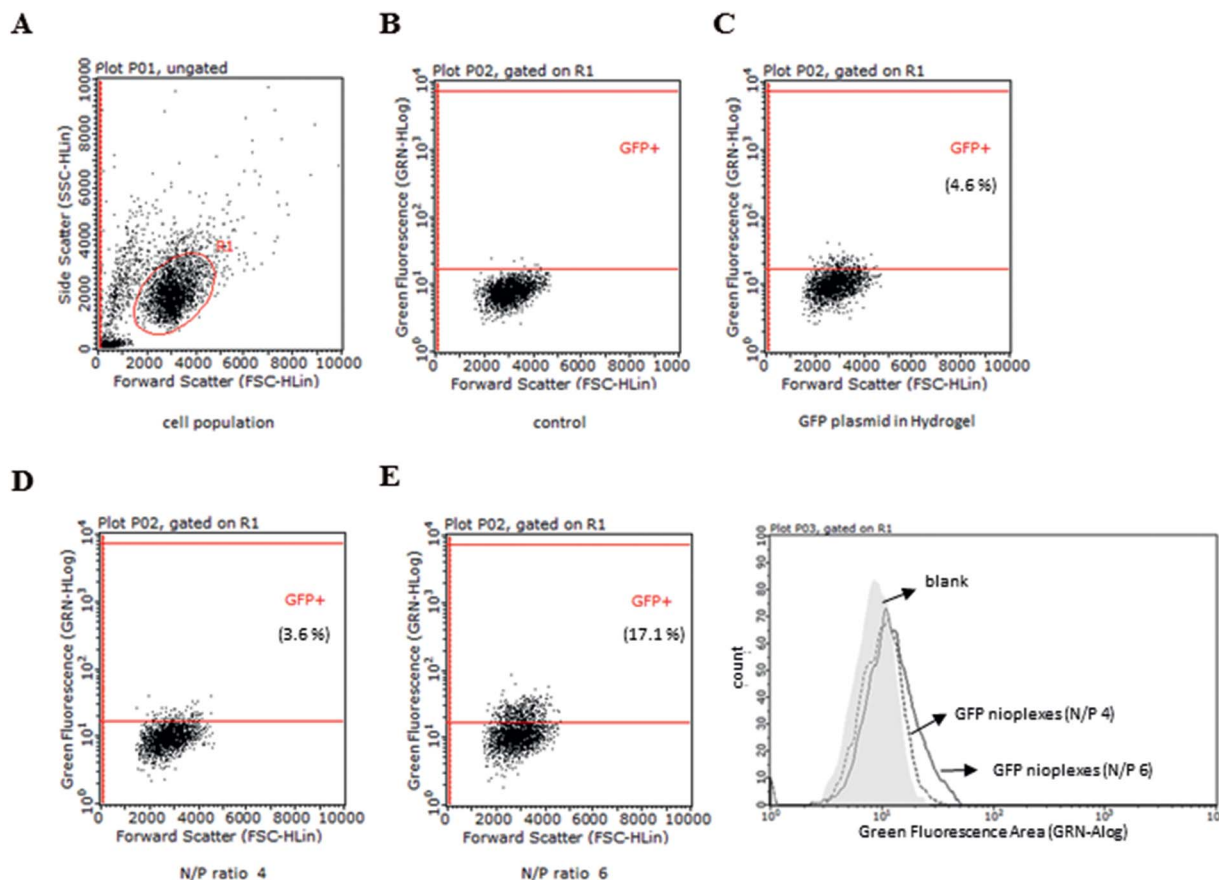


Fig. 9 Flow cytometry analysis of GFP expression in HeLa cells. (A) Normal cell population (negative control). (B) Control. (C) Flow cytometry analysis for GFP released from *hydrogel-3*. (D) Flow cytometry analysis for niosomal GFP complexes (N/P ratio of 4). (E) Flow cytometry analysis for niosomal GFP complexes (N/P ratio of 6). A flow cytometry histogram shows a slight shift between cell population containing GFP and untreated cells after 24 h incubation.

(Fig. 9E). When we compare this result with the transfection promoted by the unformulated GFP (Fig. 9C), the cell population was transfected roughly 4.6 per cent. Similar results were obtained in the case of GFP-forming complexes at N/P ratio of 4 (3.6%) (Fig. 9D). This difference can be also observed in the flow cytometry histogram, which shows a slight difference shift between fluorescent labelled and unlabelled cell populations.

This result suggests that part of the nioplexes integrity were unaltered when released from the hydrogel and continued to be active during the transfection process took place. These preliminary observations look promising and further studies involving hydrogelator concentration, optimized formulations and use of alternative crosslinking agents are currently investigated.

### 3. Experimental section

#### 3.1 Materials

All reagents employed in this work were used as received without further purification. Polysorbate-80 (Tween-80), Fmoc-L-phenylalanine (Fmoc-Phe-OH),  $\kappa$ -carrageenan and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT reagent) were purchased from Sigma-Aldrich. A fluorescently

labelled oligodeoxynucleotide sequence (5'-FITC-CGG AGG TAC ATT CGA CTT GA-3'; FITC-ODN) was synthesized *in house* in a DNA/RNA synthesizer ABI 3400 and purified by semi-preparative HPLC according to DMT *on*-based protocols. FITC (fluorescein isothiocyanate) was automatically coupled at the 5'-termini *via* phosphoramidite chemistry. Standard phosphoramidites and ancillary reagents used for the synthesis of FITC-ODN were purchased from Link Technologies. Phosphorothioate oligonucleotide of sequence 5'-CGT TTC CTT TGT TCT GGA-3' was purchased from Prologo (Sigma-Aldrich). PMax-GFP was purchased from Lonza. The amino lipid **1** was synthesized according to the literature (ESI, Fig. S1†).<sup>30,31</sup> UV-vis and fluorescence measurements were recorded on a Varian Cary BIO 50 UV/VIS/NIR Spectrometer and a Fluoromax-4 spectrofluorometer, respectively. Flow cytometer analyses were carried out in a Guava® easyCyte 8HT instrument (Millipore).

#### 3.2 Methods

**3.2.1 Electrophoretic mobility shift assay (EMSA).** FITC-ODN (0.5  $\mu$ M) was properly mixed with growing concentrations of nioplexes dispersed in PBS (working concentration = 500  $\mu$ M), which gave rise to nioplexes/FITC-ODN molar ratios





ranging from 0.5 to 16. The resultant complexes were analyzed by using electrophoresis on a 20% polyacrylamide gel at 150 V for 6 h at 25 °C in 1× TBE buffer. The polyacrylamide gel electrophoresis (PAGE) was immersed in 1× TBE buffer (200 µL) and stained with SYBR-Green (20 µL) (shaking 15 min at room temperature). Pictures were taken with a Fujifilm LAS-1000 Intelligent Dark Box II and IR-LAS-1000 Lite v1.2 instruments.

**3.2.2 Rheology.** Oscillatory rheological measurements were carried out in an AR 2000 Advanced rheometer (TA Instruments) equipped with a Julabo C cooling system. A plain-plate geometry (20 mm, stainless steel), a 1000 µm gap setting and a torque setting of 40 000 dynes cm<sup>-2</sup> at 25 °C were used for the measurements. After obtaining the supramolecular hydrogels in the absence and in the presence of nioplexes (total gel volume = 2 mL), three different experiments were carried: (1) Dynamic Frequency Sweep (DFS): variation of  $G'$  and  $G''$  with frequency (from 0.1 to 10 Hz at 0.1% strain); (2) Dynamic Strain Sweep (DSS): variation of  $G'$  and  $G''$  with strain (from 0.01 to 100%) and (3) Dynamic Time Sweep (DTS): variation of  $G'$  and  $G''$  with time maintaining both the strain and frequency values constant within the linear viscoelastic regime (strain = 0.1% strain; frequency = 1 Hz).

### 3.3 General procedures

**3.3.1 General procedure for the synthesis of cationic nioplexes-FITC-ODN complexes (lipoplexes).** Cationic nioplexes were made of *lipid-1* and non-ionic surfactant agent (polysorbate-80) in PBS (working concentration = 500 µM) at an optimal N/P ratio of 14 were mixed with a fixed amount of FITC-ODN. The mixture was vortexed and sonicated for 20 s. The resultant complexes were obtained by incubating the samples at 37 °C for 30 min.

**3.3.2 General procedure for supramolecular gel formation-loaded cationic nioplexes.** *Hydrogel-1* was obtained as follows: Fmoc-Phe-OH (10 mg; 0.026 mmol) was placed into a screw-capped glass vial (4 cm length × 1 cm diameter) and gently dissolved in PBS (3.5 mL, pH 7.4) at 70 °C. After the solution reached 40 °C, the hydrogelator (500 µL) together with cationic nioplexes:FITC-ODN nioplexes (0.08–0.1 µM) were carefully mixed for 5 min. The mixture was cooled down to room temperature affording the corresponding nioplexe-containing supramolecular hydrogel.

*Hydrogels-(2–4)* were obtained as follows: Fmoc-Phe-OH (10 mg; 0.026 mmol) and the corresponding amount of κ-carregeenan (0.5, 0.8 or 1% w/v, respectively) were placed into a screw-capped glass vial (4 cm length × 1 cm diameter) and gently dissolved in 100 mM PBS (3 mL, pH 7.4) at 70 °C. When the three solutions reached 45 °C, the mixture of hydrogelator and κ-carregeenan (500 µL) together with cationic nioplexes:FITC-ODN nioplexes (0.08–0.1 µM) were carefully mixed for 5 min. Finally, the three solutions were cooled down to room temperature affording the supramolecular hybrid hydrogels containing (FITC-ODN)-loaded cationic nioplexes (*hydrogel-2*, *hydrogel-3* and *hydrogel-4* for 0.5, 0.8 or 1% w/v, respectively).

The encapsulation efficiency (EE) of the supramolecular hybrid materials was calculated analyzing the initial FITC-ODN

amount (starting material) and the remaining mass of FITC-ODN in the receptor phase (at  $t = 0$ ) [eqn (1)].

$$EE = \frac{\text{mass(niosomal FITC-ODN aliquot)}}{\text{mass(niosome preparation)}} \times 100 \quad (1)$$

**3.3.3 Small-angle X-ray scattering (SAXS).** *Sample preparation for SAXS measurements:* *lipid-1* in its powder form was introduced into a glass capillary. A few microliters of PBS buffer was added on top and the ensemble was centrifuged to get the powder in contact with the buffer. Measurements were performed after 48 h incubation at room temperature. The reproducibility of the measurement was assessed by measuring again the sample after one week. *SAXS and WAXS analysis:* SAXS and WAXS patterns were obtained at the same time using a S3-MICRO (Hecus X-ray systems, Graz, Austria) coupled to a GENIX-Fox 3D X-ray source (Xenocs, Grenoble, France) working at 50 kV and 1 mA. This provides a detector focused X-ray beam with  $\lambda = 1.542 \text{ \AA}$  at Cu K $_{\alpha}$  line with more than 97% purity and less than 0.3% K $_{\beta}$ . The two detectors are PSD 50 (Hecus, Graz, Austria). The working  $q$ -range were  $0.1 \leq q \leq 6 \text{ nm}^{-1}$ , in the small angle range and  $13 \leq q \leq 19 \text{ nm}^{-1}$  in the wide angle range, where  $q = 4\pi/\lambda \sin(\theta/2)$  is the modulus of the scattering wave vector,  $\theta$  the scattering angle and  $\lambda$  the wavelength. SAXS patterns were analyzed in terms of a global model using the MCG model of Pabst and coworkers.<sup>65</sup> The analysis technique models the full  $q$ -range in the SAXS regime, including Bragg peaks and diffuse scattering. The bilayer electronic density profile was calculated as the sum of three Gaussian, one corresponding to the methyl contributions and the two others corresponding to the polar heads, as a difference with respect to the model of Pabst and co-workers.<sup>65,66</sup> We also incorporated a constant term through the hydrophobic part of the bilayer to give account of the methylene contribution.<sup>67</sup> The interbilayer ordering was taken into account considering the modified Caillé model as in the work of Pabst co-workers.

The volume of the lipid has been estimated to be  $0.911 \text{ nm}^3$  by using  $0.812 \text{ nm}^3$  for the hydrocarbon chains and  $0.099 \text{ nm}^3$  both obtained from partial group volumes: +0.009 per –O– (calculated from the density of 1,1-diethoxy butane); 0.0027 per NH $_2$  and 0 for trisubstituted C.<sup>68</sup> To properly scale the electronic density, we have considered the volume as well as the total number of electrons. From this calculation also the number of water molecules per polar head was obtained.

The Fourier reconstruction was performed using the same procedure as J. Pereira-Lachataignerais and co-workers<sup>69</sup> and using (- nil + -) for the phasing of the first 4 reflections. This phasing gives a sensible result and coincides with the results obtained by Pereira-Lachataignerais and co-workers.

**3.3.4 In vitro control release experiments.** Release experiments were carried out by adding 1 mL of PBS solution (pH 7.4) in 500 µL of supramolecular hydrogels (1–4). Hydrogels were placed into a screw-capped glass vial (4 cm length × 1 cm diameter) and were incubated at 37 °C. At specific time intervals, aliquots (1 mL) were carefully removed for subsequent fluorescence analysis. The removed volume was replaced with fresh PBS buffer (1 mL). The fluorescence of each aliquot (1 mL)



was measured by using excitation and emission wavelengths of 466 and 513 nm, respectively. The cumulative FITC-ODN release was obtained by calculating the total mass detected in the aliquots [eqn (2)]. The FITC-ODN concentration was determined *via* interpolation from a separately constructed standard curve of fluorescence *vs.* concentration (ESI, Fig. S5†).

*In vitro* control release experiment was also studied by native polyacrylamide electrophoresis (PAGE) gel (20%) (for further details, see ESI†). Cationic nioplexes (1  $\mu\text{M}$  FITC-ODN) were embedded and aliquots (50  $\mu\text{L}$ ) were taken at different times. Aliquots were loaded in the presence of 50% glycerol (2  $\mu\text{L}$ ) in  $1\times$  TBE and gels were run at 150 V and at 25  $^{\circ}\text{C}$  for six h. PAGE gel was stained with SYBR-Green (20  $\mu\text{L}$ ) and gel was stirred for 15 minutes. Images were analyzed using ImageJ software (US National Institute of Health)

Cumulative FITC-ODN =

$$\frac{\text{mass}(\text{total FITC-ODN, aliquots})}{\text{mass}(\text{initial FITC-ODN, loading})} \times 100 \quad (2)$$

### 3.4 Mathematical models

Release curves were fitted according to Higuchi [eqn (3)], Korsmeyer-Peppas [eqn (4)] and Weibull [eqn (5)] empirical equations. In Higuchi model, the fraction of drug released is proportional to the square root of time; where  $M_t$  and  $M_{\infty}$  are the cumulative and the maximal amounts of drug released at time  $t$ , respectively.  $k$  is a constant related to the formulation. In the case of Korsmeyer-Peppas equation,  $k$  is a rate constant and  $n$  is the release exponent that characterizes the release mechanism (*e.g.*,  $n = 0.5$  for Fickian diffusions and  $[1/2 < n < 1]$ ) for non-Fickian diffusions. For the Weibull model,  $b$  parameter describes the diffusional mechanism of the drug release. When  $b \leq 0.75$  the Fickian diffusion is the main mechanism while for  $b$  values  $0.75 < b < 1$  diffusion mechanism together with a contribution of another complex release processes can govern the liberation of the molecule.

$$\frac{M_t}{M_{\infty}} = k\sqrt{t} \quad (3)$$

$$\frac{M_t}{M_{\infty}} = kt^n \quad (4)$$

$$\frac{M_t}{M_{\infty}} = a\left(1 - \exp\left(-(kt)^b\right)\right) \quad (5)$$

### 3.5 MTT assay

HeLa cells (6000 cells per well) were cultured in a Transwell HTS 96-well plate at 37  $^{\circ}\text{C}$  with a humidified incubator, 5%  $\text{CO}_2$  overnight to achieve 60% confluence in DMEM supplemented with 10% FBS. Cells only treated with DMEM served as a negative control group (*Blank*). *Hydrogel-3* was previously heated up at 80  $^{\circ}\text{C}$  and filtered through 0.2  $\mu\text{m}$  membrane (15  $\mu\text{L}$  of *hydrogel-3*; *Mock*), *hydrogel-3* containing only cationic niosomes

at two cationic lipid concentrations (1.7  $\mu\text{M}$  and 4.2  $\mu\text{M}$ ) and cationic nioplexes-loaded *hydrogel-3* at two oligonucleotide concentration (120 nM and 300 nM) were placed on top of the transwell filters and were exposed to HeLa cells for 24 h (total volume = 200  $\mu\text{L}$ ). Cells only treated with DMEM served as a negative control group. After removing the supernatant and washed with PBS (200  $\mu\text{L}$ ), HeLa cells were incubated at 37  $^{\circ}\text{C}$  with fresh medium (200  $\mu\text{L}$ ) for 12 h. Then, 25  $\mu\text{L}$  of MTT solution (5  $\text{g L}^{-1}$  in PBS) were added and HeLa cells were incubated for another 2 h at 37  $^{\circ}\text{C}$ . The resultant formazan crystals were dissolved in DMSO (200  $\mu\text{L}$ ) and the absorbance was measured at 560 nm. All experiments were carried out in sextuplicate and normalized cellular viability was expressed as a percentage of the untreated control cells.

### 3.6 *In vitro* delivery studies and flow cytometry analysis

HeLa cells ( $10^5$  cells per well) were seeded in a 24 well-plate in a humidified incubator, 5%  $\text{CO}_2$  in DMEM supplemented with 10% FBS (1 mL) to achieve a 60% confluence. 24 h after seeding, fresh medium without antibiotic (600  $\mu\text{L}$ ) was replaced. Hydrogelator and  $\kappa$ -carrageenan at specific concentrations were dissolved in PBS as described previously and filtered through 0.2  $\mu\text{m}$  membrane. GFP plasmid (negative control) and cationic nioplexes (at N/P ratio of 4 and 6) containing GFP were entrapped within the preformed hydrogel (50  $\mu\text{L}$ ) the day before. A transwell insert system was introduced and the three hydrogels were placed in. After 24 hours of incubation, DMEM was discharged and cells were washed twice with PBS (500  $\mu\text{L}$ ). Cells were collected by trypsinization, centrifugated and resuspended in PBS (300  $\mu\text{L}$ ). Finally, transfection efficiencies of the three supramolecular hydrogels was analyzed by flow cytometry.

## 4. Conclusions

In conclusion, this work demonstrates the possibility of using supramolecular hydrogels as versatile carriers for oligonucleotide-loaded nioplexes. As a proof-of-concept, a fluorescently labelled oligodeoxynucleotide (FITC-ODN) was incorporated into cationic niosomes based on an amphiphilic lipid (*i.e.*, *lipid-1*) and subsequently embedded into biocompatible 3D supramolecular hydrogels based on Fmoc-Phe-OH (pH 7.4). In some cases, small amounts of  $\kappa$ -carrageenan were also added into the gel formulations in order to investigate the crosslinking effect on the drug diffusion through the matrix. Small-angle X-ray scattering (SAXS) measurements showed the formation of lamellar structures for the glycerol-based *lipid-1*, which remained practically constant in a temperature range between 25  $^{\circ}\text{C}$  and 45  $^{\circ}\text{C}$ . Furthermore, the strong correlation and low flexibility of the bilayers deduced from the SAXS curves supported the stability of the nioplexes under the conditions used in this work. It is worth mentioning that a complete fusion of the *lipid-1* was observed in the SAXS pattern when the temperature was increased at 55  $^{\circ}\text{C}$ . Interestingly, this property was previously not detected for the serinol-based lipid analogue.<sup>57</sup>

The results of this investigation showed that Fmoc-Phe-OH-based supramolecular hydrogels (without  $\kappa$ -carrageenan



additive) liberated almost the total amount of FITC-ODN in *ca.* 1.5 h. Nevertheless, physically crosslinked hydrogels with  $\kappa$ -carrageenan were able to prolong the release time of the niosomal oligonucleotide up to 5 h. This increase was presumably due to a reduction of both erosion processes and burst release effects by the presence of the hydrophilic biopolymer. In all cases, the niosomal oligonucleotide release was well fitted by using the Korsmeyer–Peppas kinetics model. The liberation of FITC-ODN was governed by non-Fickian diffusion, which suggested specific interactions between the cationic nioplexes and the gel networks. Moreover, these hybrid materials were found to be non-toxic in cell culture (*i.e.*, HeLa cells). To demonstrate whether nioplexes were still active when released from hydrogels, niosomal formulations containing GFP plasmid at their optimal N/P ratio were loaded within hydrogels and transfection efficiencies were evaluated by flow cytometry. These preliminary results showed better efficiencies for niosomal-loaded hydrogels than unformulated GFP-hydrogel materials, which indicated the presence of intact particles upon the release. We are currently investigating alternative approaches to improve this transfection efficiency by adding new crosslinking agents and the use of other peptide-based supramolecular gelators in order to obtain more robust and efficient vehicles for biomolecules.

## Acknowledgements

This work was supported by the Spanish Ministry of Education (Grants CTQ2013-41514-P, CTQ2014-52588 R, RTC-2014-2038-1), the University of Regensburg, the DFG (DI 1748/3-1), the Generalitat de Catalunya (2014/SGR/624) and the *Instituto de Salud Carlos III* (CB06\_01\_0019). S. G. thanks the Spanish Ministry of Education, Culture and Sports for a “Jose Castillejo” grant within the researcher mobility program (Programa Estatal de Promoción del Talento y su Empleabilidad en I+D+i, Subprograma Estatal de Movilidad, del Plan Estatal de Investigación Científica y Técnica y de Innovación 2013–2016). D. D. D. thanks the Deutsche Forschungsgemeinschaft (DFG) for the Heisenberg Professorship Award. We thank the Göpferich research group for assistance and granting us access to the rheometer. We would like to thank Dr Gemma Fabriàs and Mireia Casasampere for assistance and access to the cytometer and Dr Cristina Garrido for her help in the cytometer data processing. The authors also thank Jaume Caelles, at the SAXS-WAXS service at IQAC, for X-ray measurements.

## References

- O. Wichterle and D. Lim, *Nature*, 1960, **185**, 117–118.
- E. Bakaic, N. M. B. Smeets and T. Hoare, *RSC Adv.*, 2015, **5**, 35469–35486.
- H. Kamata, X. Li, U.-I. Chung and T. Sakai, *Adv. Healthcare Mater.*, 2015, **4**, 2360–2374.
- H. K. Lau and K. L. Kiick, *Biomacromolecules*, 2014, **16**, 28–42.
- S. Van Vlierberghe, P. Dubruel and E. Schacht, *Biomacromolecules*, 2011, **12**, 1387–1408.
- J. M. Knipe and N. A. Peppas, *Regenerative Biomaterials*, 2014, **1**, 57–65.
- M. C. Koetting, J. T. Peters, S. D. Steichen and N. A. Peppas, *Mater. Sci. Eng., R*, 2015, **93**, 1–49.
- C. Ma, Y. Shi, D. A. Pena, L. Peng and G. Yu, *Angew. Chem.*, 2015, **127**, 7484–7488.
- D. Roy, J. N. Cambre and B. S. Sumerlin, *Prog. Polym. Sci.*, 2010, **35**, 278–301.
- S.-k. Ahn, R. M. Kasi, S.-C. Kim, N. Sharma and Y. Zhou, *Soft Matter*, 2008, **4**, 1151–1157.
- X. Yang, G. Zhang and D. Zhang, *J. Mater. Chem.*, 2012, **22**, 38–50.
- Polymer Gels and Networks*, ed. Y. Osada and A. R. Khokhlov, Marcel Dekker, New York, 2002.
- A. Ajayaghosh, V. K. Praveen and C. Vijayakumar, *Chem. Soc. Rev.*, 2008, **37**, 109–122.
- J. Yu, W. Ha, J. Chen and Y. Shi, *RSC Adv.*, 2014, **4**, 58982–58989.
- F. Li, J. He, M. Zhang, K. C. Tam and P. Ni, *RSC Adv.*, 2015, **5**, 54658–54666.
- N. M. Sangeetha and U. Maitra, *Chem. Soc. Rev.*, 2005, **34**, 821–836.
- J. Li, Y. Gao, Y. Kuang, J. Shi, X. Du, J. Zhou, H. Wang, Z. Yang and B. Xu, *J. Am. Chem. Soc.*, 2013, **135**, 9907–9914.
- C. Yan and D. J. Pochan, *Chem. Soc. Rev.*, 2010, **39**, 3528–3540.
- T. Luo and K. L. Kiick, *Eur. Polym. J.*, 2013, **49**, 2998–3009.
- L. Chen, K. Morris, A. Layborn, D. Elias, M. R. Hicks, A. Rodger, L. Serpell and D. J. Adams, *Langmuir*, 2010, **26**, 5232–5242.
- R. Orbach, I. Mironi-Harpaz, L. Adler-Abramovich, E. Mossou, E. P. Mitchell, V. T. Forsyth, E. Gazit and D. Seliktar, *Langmuir*, 2012, **28**, 2015–2022.
- Y. Zhang, Z. Yang, F. Yuan, H. Gu, P. Gao and B. Xu, *J. Am. Chem. Soc.*, 2004, **126**, 15028–15039.
- D. J. Adams and P. D. Topham, *Soft Matter*, 2010, **6**, 3707–3721.
- V. Jayawarna, M. Ali, T. A. Jowit, A. F. Miller, A. Saiani, J. E. Gough and R. V. Ulijn, *Adv. Mater.*, 2006, **18**, 611–614.
- A. Mahler, M. Reches, M. Rechter, S. Cohen and E. Gazit, *Adv. Mater.*, 2006, **18**, 1365–1370.
- S. Roy and A. Banerjee, *Soft Matter*, 2011, **7**, 5300–5308.
- B. J. Boyd, *Expert Opin. Drug Delivery*, 2008, **5**, 69–85.
- I. F. Uchegbu and A. T. Florence, *Adv. Colloid Interface Sci.*, 1995, **58**, 1–55.
- H. Abdelkader, A. W. G. Alani and R. G. Alany, *Drug Delivery*, 2014, **21**, 87–100.
- G. P. Ochoa, J. Z. Sesma, M. A. Díez, A. Díaz-Tahoces, M. Avilés-Trigueros, S. Grijalvo, R. Eritja, E. Fernández and J. L. Pedraz, *Pharm. Res.*, 2014, **31**, 1665–1675.
- S. Grijalvo, A. Alagia, G. Puras, J. Zárate, J. L. Pedraz and R. Eritja, *Colloids Surf., B*, 2014, **119**, 30–37.
- T. Lian and R. J. Ho, *J. Pharm. Sci.*, 2001, **90**, 667–680.
- J. Parmentier, M. M. M. Becker, U. Heintz and G. Fricker, *Int. J. Pharm.*, 2011, **405**, 210–217.
- M. T. Popescu, S. Mourtas, G. Pampalakis, S. G. Antimisiaris and C. Tsitsilianis, *Biomacromolecules*, 2011, **12**, 3023–3030.





- 35 S. Grijalvo, J. Mayr, R. Eritja and D. Díaz Díaz, *Biomater. Sci.*, 2016, **4**, 555–574.
- 36 M. Glavas-Dodov, K. Goracinova, K. Mladenovska and E. Fredro-Kumbaradzi, *Int. J. Pharm.*, 2002, **1–2**, 381–384.
- 37 W. W. Gao, D. Vecchio, J. M. Li, J. Y. Zhu, Q. Z. Zhang, V. Fu, J. Y. Li, S. Thanphiwatana, D. N. Lu and L. F. Zhang, *ACS Nano*, 2014, **8**, 2900–2907.
- 38 Y. Liang and K. L. Kiick, *Biomacromolecules*, 2016, **17**, 601–614.
- 39 S. Mourtas, S. Fotopoulou, S. Duraj, V. Sfika, C. Tsakiroglou and S. G. Antimisiaris, *Colloids Surf., B*, 2007, **55**, 212–221.
- 40 N. El Kechai, A. Bochot, N. Huang, Y. Nguyen, E. Ferrary and F. Agnely, *Int. J. Pharm.*, 2015, **487**, 187–196.
- 41 E. A. Appel, M. W. Tibbitt, M. J. Webber, B. A. Mattix, O. Veisheh and R. Langer, *Nat. Commun.*, 2015, **6**, 6295–6304.
- 42 Y. Mao, X. Li, G. Chen and S. Wang, *J. Pharm. Sci.*, 2016, **105**, 194–204.
- 43 N. C. Wickremasinghe, V. A. Kumar and J. D. Hartgerink, *Biomacromolecules*, 2014, **15**, 3587–3595.
- 44 N. C. Wickremasinghe, V. A. Kumar, S. Shi and J. D. Hartgerink, *ACS Biomater. Sci. Eng.*, 2015, **1**, 845–854.
- 45 M. A. Khalily, M. Goktas and M. O. Guler, *Org. Biomol. Chem.*, 2015, **13**, 1983–1987.
- 46 S. Mourtas, M. Haikou, M. Theodoropoulou, C. Tsakiroglou and S. G. Antimisiaris, *J. Colloid Interface Sci.*, 2008, **317**, 611–619.
- 47 V. Baumann and J. Winkler, *Future Med. Chem.*, 2014, **6**, 1967–1984.
- 48 M. Wood, H. Yin and G. McClorey, *PLoS Genet.*, 2007, **3**, 845–854.
- 49 S. Pérez-Rentero, S. Grijalvo, G. Peñuelas, C. Fàbrega and R. Eritja, *Molecules*, 2014, **19**, 10495–10523.
- 50 M. Delgado, C. Spanka, L. D. Kerwin, P. Wentworth and K. D. A. Janda, *Biomacromolecules*, 2002, **3**, 262–271.
- 51 D. Das and S. Pal, *RSC Adv.*, 2015, **5**, 25014–25050.
- 52 J. Wang, Z. Wang, J. Gao, L. Wang, Z. Yang, D. Kong and Z. Yang, *J. Mater. Chem.*, 2009, **19**, 7892–7896.
- 53 W. E. Hennink and C. F. van Nostrum, *Adv. Drug Delivery Rev.*, 2012, **64**, 223–236.
- 54 Y. Liu, Y. Zhu, G. Wei and W. Lu, *Eur. J. Pharm. Sci.*, 2009, **37**, 306–312.
- 55 E. Ruel-Gariépy, G. Leclair, P. Hildgen, A. Gupta and J. C. Leroux, *J. Controlled Release*, 2002, **82**, 373–383.
- 56 M. Sila, S. Au and N. Weiner, *Biochim. Biophys. Acta, Biomembr.*, 1986, **2**, 165–170.
- 57 R. W. Korsmeyer, R. Gurny, E. Doelker, P. Buri and N. A. Peppas, *Int. J. Pharm.*, 1983, **15**, 25–35.
- 58 N. A. Peppas and J. J. Sahlin, *Int. J. Pharm.*, 1989, **57**, 169–172.
- 59 P. Costa and J. M. Sousa Lobo, *Eur. J. Pharm. Sci.*, 2001, **13**, 123–133.
- 60 A. Karewicz, K. Zasada, K. Szczubialka, S. Zapotoczny, R. Lach and M. Nowakowska, *Int. J. Pharm.*, 2010, **385**, 163–169.
- 61 A. K. Pannier and L. D. Shea, *Mol. Ther.*, 2004, **10**, 19–26.
- 62 T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55–63.
- 63 H.-Y. Zhang, J. Mao, D. Zhou, Y. Xu, H. Thonberg, Z. Liang and C. Wahlestedt, *Nucleic Acids Res.*, 2003, **31**, e72.
- 64 R. Lee and W. D. McElroy, *Biochemistry*, 1969, **8**, 130–136.
- 65 G. Pabst, M. Rappolt, H. Amenitsch and P. Laggner, *Phys. Rev. E: Stat. Phys., Plasmas, Fluids, Relat. Interdiscip. Top.*, 2000, **62**, 4000–4009.
- 66 E. Ojeda, G. Puras, M. Agirre, J. Zárate, S. Grijalvo, R. Pons and R. Eritja, *Org. Biomol. Chem.*, 2015, **13**, 1068–1081.
- 67 N. Kucerka, J. F. Nagle, J. N. Sachs, S. E. Feller, J. Pencer, A. Jackson and J. Katsaras, *Biophys. J.*, 2008, **95**, 2356–2367.
- 68 A. Pinazo, L. Pérez, M. Lozano, M. Angelet, M. R. Infante, M. P. Vinardell and R. Pons, *J. Phys. Chem. B*, 2008, **112**, 8578–8585.
- 69 J. Pereira-Lachataignerais, R. Pons, H. Amenitsch, M. Rappolt, B. Sartori and O. López, *Langmuir*, 2006, **22**, 5256–5260.

