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Macroporous three-dimensional graphene oxide foams for dye adsorption and antibacterial applications†

Swetha Jayanthi,^a Neerugatti KrishnaRao Eswar,^a Satyapaul A. Singh,^b Kaushik Chatterjee,^c Giridhar Madras^{*b} and A. K. Sood^d

Several reports illustrate the wide range applicability of graphene oxide (GO) in water remediation. However, a few layers of graphene oxide tend to aggregate under saline conditions thereby reducing its activity. The effects of aggregation can be minimized by having a random arrangement of GO layers in a three dimensional architecture. The current study emphasizes the potential benefits of highly porous, ultralight graphene oxide foams in environmental applications. These foams were prepared by a facile and cost effective lyophilization technique. The 3D architecture allowed the direct use of these foams in the removal of aqueous pollutants without any pretreatment such as ultrasonication. Due to its macroporous nature, the foams exhibited excellent adsorption abilities towards carcinogenic dyes such as rhodamine B (RB), malachite green (MG) and acriflavine (AF) with respective sorption capacities of 446, 321 and 228 mg g⁻¹ of foam. These foams were also further investigated for antibacterial activities against *E. coli* bacteria in aqueous and nutrient growth media. The random arrangement of GO layers in the porous foam architecture allowed it to exhibit excellent antibacterial activity even under physiological conditions by following the classical wrapping-perturbation mechanism. These results demonstrate the vast scope of GO foam in water remediation for both dye removal and antibacterial activity.

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1. Introduction

Environmental pollution caused by water-soluble toxic organic dyes released from the textile, paper and cosmetic industries has become a major concern in the environment.^{1,2} Various methods have been developed for treating these organic water pollutants. Among these techniques, adsorption is a simple, easy, inexpensive and efficient way of removing multiple components simultaneously.^{3,4} Several materials such as

activated carbon,⁵ conducting polymers,⁶ mesoporous silica⁷ and mesoporous hybrid xerogels⁸ have been employed as sorbent materials for the adsorption of various dyes. Nanoscale materials such as CNTs, carbon fibers and fullerenes^{9–12} are excellent adsorbents due to their high surface-to-volume ratio compared to bulk adsorbents. However, the preparation of most of these materials involves sophisticated techniques such as chemical vapor deposition and the requirement of high pressure and high temperature conditions¹³ that can limit their use in the large-scale applications.

In recent years, two-dimensional (2D) graphene-based materials such as graphene oxide (GO) and reduced graphene oxide (rGO) have gained significant interest due to their unique electronic, optical, mechanical and thermal properties.^{14–16} Moreover, the presence of surface functional groups and large surface area make these materials applicable in various fields such as sensors, water purification techniques, energy storage and catalysis.^{17–20} Ferrite nanoparticles decorated GO has been shown to be an excellent adsorbate for removing the heavy metals As(III), As(IV) and Pb(II).²¹ Hitherto, many GO and rGO-based materials have been employed as sorbent materials in dye removal applications.^{22–26} Although the single layered GO and rGO sheets are known to be efficient sorbent materials, the aggregation of the layers due to π - π interactions reduces the adsorption capacities. Furthermore, the requirement of centrifugation and magnetic separation techniques^{27–30} complicates

^aCentre for Nano Science and Engineering, Indian Institute of Science, Bangalore-560012, India

^bDepartment of Chemical Engineering, Indian Institute of Science, Bangalore-560012, India. E-mail: giridhar@chemeng.iisc.ernet.in; Fax: +91 80 23600683; Tel: +91 80 22932321

^cDepartment of Materials Engineering, Indian Institute of Science, Bangalore-560012, India

^dDepartment of Physics, Indian Institute of Science, Bangalore-560012, India

† Electronic supplementary information (ESI) available: (1) Structure of the dyes (Fig. S1), (2) (a) FT-IR spectrum and (b) XRD spectrum of GO foam (Fig. S2), (3) comparison plot of adsorption isotherms of (a) MG and (b) AF on GO foam and GO precipitate (Fig. S3), (4) photographs displaying the complete adsorption of RB dye on GO foam and GO precipitate. Concentration of the dye solution is 5 ppm and the amount of the material used is 6 mg (Fig. S4), (5) point of zero charge plot of GO foam under different pH conditions (Fig. S5), (6) UV-Vis spectra of GO foam before and after interactions with RB, MG and AF dye molecules (Fig. S6), (7) FT-IR spectrum of GO foam adsorbed with AF molecules under alkaline conditions (Fig. S7). See DOI: 10.1039/c5ra19925e

the material recovery processes. Therefore, three-dimensional graphene (3D) architectures are desired as they enhance the adsorption ability and also allow the effective reuse of the material by simple recovery processes without the loss of material due to its 3D structure. Many methods such as sol-gel, hydrothermal and template method have been developed in preparing graphene foams.^{31–34}

Among the several methods known, lyophilization is a simple, flexible, effective eco-friendly method that allows the preparation of various dimensions of foams depending on the dimensions of the containers used.³⁵ Due to its high surface-to-volume ratio and ease of material recovery, these ultra-light, highly porous graphene foams and their composites have been employed in water remediation. In this direction, Liu *et al.* reported the preparation of 3D graphene oxide sponges by a simple centrifugal vacuum evaporation method. These foams exhibited adsorption capacities 397 and 467 mg g^{−1} for methylene blue (MB) and methyl violet (MV) dyes, respectively.³⁶ Sui *et al.*³⁷ have reported the adsorption capacities of ~800 mg g^{−1} for 3D porous GO-PEI (polyethyleneimine) composite foams for acidic dyes such as Amaranth dyes. 3D reduced graphene oxide architectures are also known for the removal of methylene blue, rhodamine B and methyl orange dyes. These porous rGO sponges were prepared by the hydrothermal treatment of graphene oxide sheets with the assistance of thiourea.³⁸ These 3D architectures exhibited good adsorption abilities towards all the three dyes. However, to the best of our knowledge, the dye adsorption studies on GO foams prepared by lyophilization method have not been reported.

Along with the removal of toxic water-soluble organic dyes to prevent environmental pollution, degradation of water-borne microorganisms is also a crucial issue to be addressed. Microorganisms cause biofouling and eventually the degradation of metal surfaces interfering with many of the industrial applications.^{39,40} Several anti-biofouling coatings have been developed for controlling the microbial growth. But these coatings release toxic substances such as metals and antibiotics for the inhibition of microbial growth that causes secondary pollution.⁴¹ Therefore, the development of eco-friendly and effective anti-biofouling materials is a prerequisite. In this regard, graphene-based materials have been exploited for various applications such as biomedical,⁴² water disinfection⁴³ and in controlling crop diseases.⁴⁴ This is because of the direct contact mode of action of graphene-based materials against microbes and their excellent properties such as surface modification, good water dispersibility and good mechanical strength.^{45,46} Several studies have been conducted to understand the mechanism of antimicrobial activity of graphene-based materials.^{43,44,47–51} Graphene-based materials especially GO, loses its activity under physiological conditions due to the aggregation effects caused by π - π interactions.⁴⁷ Therefore, 3D architectures with interconnected layers in random arrangement can prevent the layer aggregation retaining the surface area of the material and its activity even under saline conditions. This motivated us to study the antibacterial activities of 3D macroporous GO foams prepared by lyophilization technique. To the best of our knowledge, antibacterial activity of 3D graphene architectures has not been reported till date.

In this present study, ultra-light, highly porous 3D GO foams were prepared by a simple and effective lyophilization technique. The porosity of the foams can be tuned by simply varying the concentrations of GO dispersions and the dimensions of the foams can be varied by choosing appropriate containers for lyophilization. The foams are used directly after lyophilization without pretreatment such as ultrasonication. The as-prepared GO foams were employed in the removal of three toxic cationic dyes (rhodamine B, malachite green and acriflavine) and for the degradation of *E. coli* bacteria. The antibacterial activities of GO foam and adsorption studies of acriflavine dye on graphene-based materials are reported for the first time. The adsorption capacities of these foams are found to be comparable to the GO precipitate, while the 3D foams exhibited higher antibacterial activities towards *E. coli* both in aqueous and nutrient growth media compared to the GO precipitate. This behavior can be attributed to the 3D porous architecture of the GO foam. The random arrangement of GO layers in the porous architecture of GO foams help in retain the activity even under saline conditions by reducing the extent of layer aggregation.

2. Experimental section

2.1. Synthesis of GO foam

Graphene oxide (GO) was synthesized by chemical oxidation of graphite using simplified Hummer's method.⁴⁶ Commercially procured graphite flakes (Superior Graphite Co.) (2 g) were added to the mixture of H₂SO₄ : H₃PO₄ (110 : 12 mL) and allowed to stir for 2–3 min. Subsequently, KMnO₄ (11.1 g) was slowly added and the resulting mixture was stirred for three days at room temperature. Finally, H₂O₂ (30%, 20 mL) was added to stop the reaction. The yellow precipitate obtained was washed repeatedly with 1 M aqueous HCl solution and distilled water until a pH of 4–5 was achieved. Dark brown gel of graphene oxide (GO) was obtained, which was collected by centrifuging the solution at 10 000 rpm. GO foams of weight ratio, 1 wt% were prepared by freeze-drying the dispersion of above-obtained GO precipitate (10 mg) in water (1 mL) in a lyophilizer³⁵ (Free zone 2.5 plus) for three days. The as-prepared GO foams were characterized using X-ray diffractometer (Rigaku instrument, Cu K α (λ = 1.5418 Å)), field emission scanning electron microscope (Zeiss Ultra FE-SEM), Fourier transform-infrared spectroscopy (Perkin Elmer FT-IR spectrometer), Raman spectroscopy (Witec spectrometer with 514 nm excitation), and porosity measurements (XRadia Versa XRM-500).

2.2. Adsorption studies

The adsorption characteristics of three cationic dyes were studied in the present work. The dyes chosen for present study are malachite green (MG), rhodamine B (RB) and acriflavine (AF) and structures of all three dyes are given in the Fig. S1 (see ESI†). For adsorption equilibrium studies, 3 mg of GO foam/GO precipitate (which were cut into fine number of pieces) was loaded in 10 mL of respective dye solution and kept in an incubator shaker at 37 °C for the period of 8 h. Contact time studies were performed in 30 mL dye solutions with the same

GO foam loading (0.3 g L^{-1}) under various pH conditions. The pH variation experiments were performed by using buffer capsules of $\text{pH } 4.0 \pm 0.1$, 7.0 ± 0.1 and 9.2 ± 0.1 respectively. The concentrations of all dyes were analyzed at their characteristic wavelength ($\lambda_{\text{max, MG}} = 617 \text{ nm}$, $\lambda_{\text{max, RB}} = 543 \text{ nm}$ and $\lambda_{\text{max, AF}} = 464 \text{ nm}$) by using UV-visible spectrometer (Shimadzu 1700). For kinetic studies, 9 mg of GO foam was loaded in 30 mL of respective dye solutions and kept in an incubator shaker at 37°C . The concentration of dye molecules were analyzed from the absorbance at the characteristic wavelength by recording absorption spectra at time intervals of 30 min over a period of 3 h.

2.3. Point of zero charge (PZC) studies

The point of zero charge of GO foam was determined by pH drift method. In this method, various pH solutions were prepared by adding 0.1 M HCl and 0.1 M NaOH solutions to 0.1 M NaCl solution. 9 mg of GO foam was dispersed in 10 mL of each pH solution. These solutions were well mixed and kept in dark for 24 h. The initial and final pH of the solutions were plotted, the point of intersection of the obtained experimental data and 45° line is noted as point of zero charge of the material.

2.4. Procedure for antibacterial evaluation

The accessories necessary for microbial growth and plating such as nutrient broth and nutrient agar solutions, Petri plates, micro tips, L-rods were autoclaved. *Escherichia coli* (MG 1655) was cultured in sterile liquid nutrient broth. The culture was centrifuged for 10 min at 5000 rpm to pellet the bacterial cells. The concentration of bacteria was maintained by re-suspending the pellet in sterile water. Sterile nutrient agar solution was equally poured into the Petri plates and solidified inside the laminar flow chamber. Experiments were conducted with 0.1 g L^{-1} of GO materials in bacterial suspension. For bacterial growth experiment, 0.1 g L^{-1} each of GO foam and GO precipitate were suspended into sterile nutrient broth separately without any pretreatment and inoculated with seed culture of bacteria obtained from its log phase of growth. To measure antibacterial activity, aliquots of suspensions were taken at specific intervals and its OD was measured using spectrophotometer at 600 nm. Similarly for plating, 100 μL of serial diluted samples were spread onto solidified nutrient agar plates and incubated at 37°C for overnight. The experiment was repeated and the bacterial colonies were counted and then evaluated for the bacterial degradation studies.

3. Results and discussion

Morphology of 2D GO flakes prepared by dispersing GO precipitate in water and 3D GO foam (1 wt%) prepared by lyophilization technique are characterized by FE-SEM. GO flakes have an average size of 10–20 μm , as shown in the Fig. 1a. Interconnected highly porous structure with pore size ranging from few microns to few tens of microns can be observed in the SEM image of GO foam (Fig. 1b).

Fig. 1 FE-SEM images of (a) GO flake and (b) GO foam of 1 wt%.

The incorporation of oxygen functional groups in GO foam during the oxidation of graphite is confirmed using FT-IR and XRD analysis. FT-IR spectrum of GO foam (Fig. S2a†) reveals the presence of the following functional groups: hydroxyl ($-\text{OH}$), carbonyl ($\text{C}=\text{O}$), aromatic $\text{C}=\text{C}$, carboxyl ($\text{C}-\text{OH}$), epoxy ($\text{O}-\text{C}-\text{O}$) and alkoxy ($\text{C}-\text{O}$) observed at 3427, 1730, 1626, 1384, 1223 and 1048 cm^{-1} wavenumbers, respectively.⁵² IR bands at 2852, 2924 and 2958 cm^{-1} are assigned to the symmetric and asymmetric stretching vibrations of CH_2 and asymmetric stretching vibrations of CH_3 groups, respectively. XRD spectrum of GO foam shows a sharp diffraction peak at 10.3° corresponding to the interlayer spacing of $8.6 \text{ \AA}$ (Fig. S2b†). The large d -spacing in GO foam compared to graphite ($3.4 \text{ \AA}$) also confirms the presence of oxygen functional groups in GO foam.⁵³

Porosity of the GO foam of 1 wt% is characterized using X-ray tomography. Three dimensional view of the bulk foam is shown in Fig. 2a. The volume of voids present in the bulk foam is found by scanning the foam in 900 frames in depth. From Fig. 2b, red color of the two-dimensional slices indicates the material, whereas the voids are represented by the dark background. The porosity of GO foam is found to be 86%. Further, the porous structure of the foam is characterized in detail using Avizo analysis model on micro CT images. The voids and the material are clearly marked in the Fig. 3a. Pore diameter distribution of the foam is analyzed by choosing a small volume of the foam as shown in the Fig. 3a. Equivalent diameter of the pores is found by processing the binary image after filtration process. The distribution of the equivalent diameter of the pores is given in the histogram (Fig. 3b) and the range is found to vary from a few microns to 1500 μm . Maximum number of pores have diameter below 100 μm . As the GO layers are separated far apart, the aggregation of GO layers is diminished under saline conditions. Therefore, more number of GO layers (*i.e.* high surface area) is exposed to bacteria and dye molecules, improving the activity of the GO foam. The 3D highly porous GO foams prepared by simple lyophilization technique have shown higher surface area in comparison to their 2D counterparts. Therefore, these macroporous GO foams can be employed as highly efficient materials in water purification applications.

3.1. Adsorption isotherms

Fig. 4a depicts the adsorption isotherms of all the three dyes on GO foam indicating the amount of dye adsorbed per g of GO foam *versus* the concentration of dye in the bulk solution over a period of 8 h. In order to understand the adsorption mechanisms and the affinity of dye molecules towards the adsorbent,

Fig. 2 (a) Three dimensional XT micrographs of GO foam (scale bar is 7000 μm), (b) two dimensional images of different slices of the above foam. Slice numbers are given above the images. Pseudo colors are given for the better visibility of the images.

experimental data are analyzed using Langmuir–Freundlich adsorption isotherm model. The Langmuir–Freundlich model describes both Langmuir-type and Freundlich-type adsorption behaviors⁵⁴ and it is expressed by the eqn (1),

$$q_e = \frac{QK_{\text{LF}}C_e^{1/n_F}}{(1 + K_{\text{LF}}C_e^{1/n_F})} \quad (1)$$

where Q represents the maximum adsorption capacity of adsorbent (mg g^{-1}), which is a measure of the total number of

Fig. 3 (a) 3D binary images of GO foam and the portion of the foam considered for the pore diameter analysis, (b) distribution of equivalent diameter of pores in GO foam.

Fig. 4 Adsorption isotherms and the corresponding fitting plots of RB, MG and AF dye molecules on (a) GO foam, and (b) GO precipitate at 37 °C and natural pH (~ 3). Concentration of GO foam is 0.3 g L^{-1} and adsorption time is 8 h. (c) Comparison plot of adsorption isotherms of RB dye on GO foam and GO precipitate.

binding sites available per g of the sorbent, K_{LF} is the affinity constant (L mg^{-1}) and n_F is the index of heterogeneity. The experimental results and corresponding fitting plots of all the three dyes are shown in the Fig. 4a. The parameters evaluated from the isotherms are tabulated in the Table 1.

Among the three dyes, equilibrium amount of dye adsorbed on the foam surface for a given concentration of dye molecules is observed to be more in the case of RB (446 mg g^{-1}) compared to MG (320 mg g^{-1}) and AF (228 mg g^{-1}) under acidic conditions ($\text{pH} \sim 3$) (Fig. 4a). This variation in the adsorption is due to the different interactions of various functional groups present on dye molecules and the GO surface. Similar experiments with

all three dyes are also carried out with GO precipitate dispersed in dye solution (represented as GO precipitate). The respective adsorption curves are shown in the Fig. 4b. Similar trend of dye adsorption is also observed with GO precipitate. Fig. 4c displays the comparison plot of GO foam and precipitate towards the dye adsorption of RB. Plots for other dyes MG and AF are shown in Fig. S3.† It is clearly seen from the plots of RB and AF that the equilibrium concentration of dye adsorbed on GO foam is lesser in comparison to GO precipitate at low dye concentrations. This can be attributed to the porosity effect in the case of foam that allows the dye molecules to diffuse slowly into the pores in contrast to the precipitate where GO flakes are well dispersed

Table 1 Isotherm parameters obtained using Langmuir–Freundlich isotherm model for the adsorption of RB, MG and AF dyes on GO foam and GO precipitate

Dye	Parameters	GO foam	GO precipitate
RB	Q (mg of dye per g of GO materials)	460 ± 13.0	474 ± 10
	n_F	1.20 ± 0.02	1.23 ± 0.08
	K_{LF}	0.141 ± 0.033	0.280 ± 0.039
MG	Q (mg of dye per g of GO materials)	351.4 ± 10.9	333.2 ± 29
	n_F	1.20 ± 0.03	1.20 ± 0.1
	K_{LF}	0.373 ± 0.098	0.15 ± 0.03
AF	Q (mg of dye per g of GO materials)	235.4 ± 21.6	231 ± 4
	n_F	1.20 ± 0.06	1.11 ± 0.06
	K_{LF}	0.099 ± 0.009	0.190 ± 0.045

in dye solution. The equilibrium amount adsorbed at higher concentrations is similar in both the cases. Even though the equilibrium dye adsorption is lower at low dye concentrations, the GO foam has an additional advantage over the GO precipitate in the recovery process, where there is no need of employing techniques such as ultrahigh centrifugation in recovering the material after dye adsorption.

We have also carried out the experiments to show the advantage of the GO foam over GO precipitate in recovering the material after adsorption process. GO foam of 1 wt% and GO precipitate of equal weight (6 mg) were added separately to the dye solutions (rhodamine B) of 5 ppm concentration. The material is allowed for the adsorption of dye until the solution became nearly colorless. GO foam and GO precipitate took approximately 24 and 48 h respectively for nearly complete adsorption. The corresponding photographs are shown in Fig. S4.† It is clearly evident from Fig. S4† that GO foams are intact whereas GO precipitate starts dispersing slowly in the solution making the material recovery process easy in the case of foams compared to precipitate.

Therefore, the foams can be recovered completely without any sophisticated techniques whereas in the case of GO precipitate, centrifugation is required for the complete recovery of the material. This makes the material recovery process easy in the case of foams compared to precipitate. Moreover, because of relatively high surface area, foams require less time for the adsorption of same amount of dye when compared to GO precipitate. The activity of GO can be increased by dispersing the precipitate in the dye solution using ultrasonication. However, the complete removal of material requires centrifugation for prolonged times. The relatively high surface area and the intactness of the structure increase the applicability of GO foams in dye removal processes. The usage of foams also prevents the secondary pollution caused by the residual GO particles that can still remain in the solution in the case of well dispersed GO precipitate even after prolonged centrifugation.³⁰

The adsorption mechanism of dye involves the following four steps: (1) dye diffusion in the bulk liquid phase, (2) diffusion through boundary layer, (3) intraparticle diffusion and (4) adsorption on active sites. Dye diffusion in the bulk liquid phase and within the particle determines the kinetics of adsorption of dye molecules. Fig. 5a–c display the adsorption plots of all three dyes at different time intervals under different pH conditions. The concentration of dye molecules in the solution phase declines sharply in the initial time periods that indicates faster adsorption kinetics. This is due to the availability of higher number of vacant sites for adsorption in the initial time period and these vacant sites saturate over time. The experimental data are analyzed using both pseudo-first order (PFO) and pseudo-second order (PSO) kinetic models⁵⁵ that are represented by the following eqn (2) and (3), respectively

$$C = C_0 e^{-k't} \quad (2)$$

$$C = \frac{C_0}{1 + k''C_0t} \quad (3)$$

C_0 represents the initial concentrations of the dye molecules, k' and k'' are the first and second order rate constants respectively. The corresponding fitting plots of all three dyes are given in the Fig. 5 and the rate constants are tabulated in Table 2. All three dyes follow pseudo-second order kinetics better than pseudo-first order kinetics. Under acidic pH, the rate of adsorption of RB molecules is faster than AF and MG molecules. But with increase in pH, rate of adsorption of AF and MG molecules increase whereas for RB molecules, the adsorption rate decreases.

The single resistance pore diffusion is expressed by Webber's pore diffusion model which is obtained from Fick's second law of diffusion.⁵⁵ The model is expressed by eqn (4)

$$q = k_i t^{0.5} + c \quad (4)$$

where, k_i is amount adsorbed (mg g^{-1}) at time t on the adsorbent, c is the intercept which represents the resistance to mass transfer in the external liquid film. Webber's pore diffusion model assumes that the external mass transfer is significant at the beginning of diffusion and the pore diffusion assumed to be constant. It is clear from Fig. 6 that the adsorption follows multi-linearity, but none of them have intercept equal to zero. This indicates that the rate of mass transfer during adsorption is solely controlled by film diffusion mechanism instead of pore diffusion.

3.2. Dye interactions with GO foam

The three cationic dyes RB, MG and AF chosen for the present study belong to different classes of functional groups: xanthene fluorine rhodamine, triaryl methane and acridine respectively, but of similar molecular size.⁵⁶ In order to understand the type of interactions present between dye molecules and the GO surface, pH dependent adsorption studies are carried out for all three dye molecules and the respective plots are shown in the Fig. 5. As the pH of the dye solution is increased, the adsorption capacity of RB molecules decreased, whereas the adsorption of AF and MG dye molecules increased.²⁷ This is because of the different functional groups present in dye molecules and on the GO surface that undergo protonation and de-protonation based on the pH of dye solution. Point of zero charge of GO is found to be 2.02 (Fig. S5†).⁵⁷ Under acidic conditions, as the pH of dye solution (~ 3) is greater than the pH_{PZC} , GO has negative charge on the surface. Therefore, all three dyes adsorb on the GO surface due to their cationic nature. These electrostatic interactions between the GO surface and dye molecules are also evident from the shifts observed in the Raman spectra of GO foam after dye adsorption. As known from the literature, in-plane bond stretching vibrations assigned as G-band in graphene is sensitive to the changes in electronic structure of graphene.⁵⁸ Therefore, charge transfer from any adsorbed molecules leads to the stiffening or softening of phonons which can be seen as a shift in the G-band of graphene. In all the three dyes, shifts in the G-band towards lower wavenumbers (Fig. 7) are observed indicating that the electron transfer has occurred from adsorbed dye molecules to the GO surface. A significant bathochromic shift is also observed in the characteristic

Fig. 5 Kinetic adsorption plots and the corresponding pseudo-second order fitting plots of RB, MG and AF dyes under (a) acidic pH, (b) neutral pH and (c) basic pH conditions. Concentration of GO foam is 0.3 g L^{-1} and adsorption time is 3 h.

absorption peaks of RB and MG dyes on adsorption to the GO surface (Fig. S6†). This shows that there is a possible π - π interaction between the phenyl rings of dye molecules and the GO surface. Apart from the electrostatic and π - π interactions, hydrogen-bonding interactions between the carboxyl groups of GO and RB molecules are also possible as the carboxyl groups remain protonated under acidic conditions.⁵⁹

Therefore, due to the additional hydrogen-bonding interactions along with π - π and electrostatic interactions, equilibrium adsorption follows the order of $\text{RB} > \text{MG} > \text{AF}$. Because of the presence of additional π - π interactions in MG molecules, adsorption is higher compared to AF molecules. The rate of adsorption in the initial time periods follows the order $\text{RB} > \text{AF} > \text{MG}$, under acidic conditions. Even though the equilibrium adsorption concentration is higher for MG molecules compared to AF molecules, the rate of adsorption is observed to be slightly faster in the case of AF molecules. This can be due to the presence of additional amine functional groups in AF

molecules, which remain protonated under acidic conditions. Therefore, extra positive charge from protonated amino groups on AF molecules leads to the higher rate of diffusion of AF

Fig. 6 Intraparticle diffusion plot for the adsorption of RB, AF and MG on GO foam under natural pH condition (initial concentration of dye = 50 ppm, GO foam loading 0.3 g L^{-1}).

Table 2 Rate constants of adsorption of RB, MG and AF dyes on GO foam under different pH conditions

Dye	PFO			PSO		
	pH = 3.7 ± 0.2	pH = 6.6 ± 0.1	pH = 9.5 ± 0.1	pH = 3.7 ± 0.2	pH = 6.6 ± 0.1	pH = 9.5 ± 0.1
MG	0.022 ± 0.001	$(2.28 \pm 0.03) \times 10^{-2}$	$(55.50 \pm 0.07) \times 10^{-3}$	$(57.40 \pm 1.6) \times 10^{-3}$	$(0.857 \pm 0.103) \times 10^{-3}$	$(1.94 \pm 0.02) \times 10^{-3}$
AF	0.032 ± 0.002	$(18.71 \pm 0.11) \times 10^{-2}$	0.183 ± 0.086	$(1.27 \pm 0.02) \times 10^{-3}$	$(14.45 \pm 0.98) \times 10^{-3}$	$(12.65 \pm 0.52) \times 10^{-3}$
RB	0.272 ± 0.013	$(9.29 \pm 0.24) \times 10^{-2}$	$(1.704 \pm 0.299) \times 10^{-4}$	$(0.807 \pm 0.010) \times 10^{-3}$	$(3.04 \pm 0.07) \times 10^{-3}$	$(3.462 \pm 0.031) \times 10^{-6}$

Fig. 7 Raman spectra of GO foam before and after adsorption of RB, MG and AF dyes.

molecules to GO surface from bulk solution compared to MG molecules. But the presence of limited number of carboxylate groups at the edges on GO surface limits the equilibrium concentration of adsorption of AF molecules.

In the basic conditions, the carboxyl groups on GO exists in de-protonated state that increases the negative charge on GO surface. This can be inferred from the presence of a new peak corresponding to carboxylate ions (1635 cm^{-1}) and the absence of -COOH peak at 1730 cm^{-1} in the FT-IR spectrum of GO (Fig. S7†). The higher negative charge on GO increases the adsorption of MG and AF dyes due to the increase in the electrostatic interactions and decrease in the interference of protons for adsorption at higher pH. However, in the case of RB, there is a negligible adsorption at higher pH because of the electrostatic repulsions between the negatively charged carboxylic groups present on both GO surface and RB molecules. Besides electrostatic interactions, the strong hydrogen bonding interactions between the amino groups on AF molecules and the carboxylate ions on GO favors the faster adsorption kinetics of AF on the GO surface compared to MG molecules even under basic pH conditions.

3.3. Antibacterial studies

Antibacterial activity of GO foam and GO precipitate towards *E. coli* bacteria is investigated by evaluating the growth curve and the degradation curve of bacteria in the presence and absence of GO materials. Loss of viability of bacterial cells is assessed by colony counting method after incubating the *E. coli* cells in aqueous medium containing either GO foam or GO precipitate for 5 h under shaking speed of 100 rpm. Fig. 8a shows the corresponding plot of bacterial degradation in the presence of GO materials. Bacterial loss gradually increased with time in the presence of both GO foam and GO precipitate. In the initial period, loss of cell viability is observed to be the same in both the cases but with progress in time, bacterial degradation is observed to be higher in the case of GO precipitate in comparison to GO foam. This can be attributed to the difference in the surface area of the GO exposed to the bacteria. As the solution is agitated, GO precipitate slowly disperses in water with time thus increasing the number of GO layers in the solution, whereas GO foam remained as such even after agitation.

Fig. 8b depicts the growth curve of *E. coli* cells in the presence and absence (control) of GO materials. Bacterial growth is monitored by growing the bacterial cells in nutrient broth medium and measuring the optical density value at 600 nm over different time intervals. Growth is observed to be relatively lower in the presence of GO foam compared to GO precipitate and the growth of the cells is almost similar in presence and absence of GO precipitate. This can be due to the aggregation of the GO layers under saline conditions, as the composition of nutrient broth medium consisted of salts. Cations in the salt medium neutralize the negatively charged carboxyl ions on the GO surface leading to the aggregation of GO layers because of π - π interactions. Therefore, GO precipitate is sparingly dispersed in saline solutions when compared to that in the distilled water.⁴⁷ But in the case of GO foam, comparatively low aggregation is observed because of the random arrangement of GO layers. Moreover, the size of bacteria is observed to be less than $3\text{ }\mu\text{m}$ in one dimension, as shown in the SEM images (Fig. 9a and b). Due to the 3D macroporous nature of GO foams of pore size ranging from few microns to few tens of microns, these foams allow more degree of freedom to the bacteria towards the interior surfaces of GO layers. However, this penetration of bacteria is not possible in the case of GO precipitate as the interlayer spacing is in the range of nanometers. Therefore, in the case of GO precipitate, only outer exposed surfaces are responsible for the bacterial growth inhibition. However, in the case of foam, besides the outer surfaces, inner surfaces of the GO layers embedded in the porous structure also contribute to the inhibition of the bacterial growth. Therefore, 3D macroporous GO foams show better antibacterial activity even under physiological conditions, where the 2D GO layers lose their activity due to the aggregation of layers.

Kinetic studies of bacterial degradation by GO foam and GO precipitate are also carried out by measuring the concentration of bacteria degraded at different time intervals for different initial bacterial concentrations. The respective plots are given in the Fig. 8c and d. The experimental data is found to fit with first order kinetic model. The rate constants evaluated for GO foam and GO precipitate are $(5.96 \pm 0.09)\text{ h}^{-1}$ and $(8.03 \pm 0.3)\text{ h}^{-1}$ respectively for the initial concentration of $2.7 \times 10^7\text{ CFU mL}^{-1}$. The values obtained for the initial concentration of $1.8 \times 10^8\text{ CFU mL}^{-1}$ are

Fig. 8 (a) Loss of cell viability of *E. coli* cells in the presence of GO foam and GO precipitate, (b) growth curve of *E. coli* bacteria in the absence (control) and presence of GO foam/GO precipitate, (c) and (d) first order kinetic plot for the bacterial degradation of GO materials at different initial concentrations of 1.8×10^8 and 2.7×10^7 CFU mL⁻¹ respectively. GO materials were incubated with *E. coli* cells (1.8×10^8 CFU mL⁻¹) at 37 °C for 5 h and 12 h for the evaluation of loss of cell viability and growth curves respectively. Concentrations of GO materials used in experiments are 0.1 g L⁻¹.

$(2.40 \pm 0.01) \text{ h}^{-1}$ and $(3.50 \pm 0.12) \text{ h}^{-1}$ respectively. The relatively lower rate of cell viability loss by GO foam is due to reduction in the accessible surface area for bacterial interaction.

3.4. Mechanism of antibacterial activity

Antibacterial mechanism and the interactions of bacterial cells with GO materials are further investigated by SEM analysis. FE-SEM images of *E. coli* cells before and after exposure to GO materials are shown in the Fig. 9a–f. It is clearly seen from the SEM images that the bacterial cells are intact in the absence of GO materials (Fig. 9a and b), whereas in the presence of either GO foam or GO precipitate, bacterial cells are wrapped by thin GO sheets and cell membranes are collapsed as shown in the Fig. 9c–f. Wrapping of bacterial cells by thin GO sheets isolates the cells from external growth medium interfering with the proton and nutrient uptake and also block the water channels of cells, as GO sheets are impermeable to many molecules.⁶⁰ This perturbs the membrane potential of bacterial cells that disturbs the cell metabolic activities which eventually disrupts the cell membrane leading to the death of cells.⁴⁴ It is reported that wrapping of the *E. coli* bacterial cells by graphene sheets biologically disconnect cells from growth medium preventing the cells to proliferate, resulting in the loss of cell viability.⁴³

It is also known that GO causes oxidative stress on the bacterial cells resulting in the disruption of cell membrane.^{47,48} Apart from the above mechanisms, bacterial cell death can also be due to the charge transfer from bacteria to sharp edges of the GO and graphene which act as good electron acceptors damaging the cell membrane during the direct contact interactions of bacteria with the graphene materials as reported earlier.^{50,51}

SEM images clearly show that the death of bacterial cells is due to the wrapping of cells by thin GO sheets in both the cases of GO foam as well as GO precipitate. The variation in the antibacterial activities of foam and precipitate in different media is attributed to the difference in the exposed surface area of GO materials to bacterial cells. GO foam exhibits antibacterial activity in both aqueous as well as in the nutrient broth medium due to its highly porous 3D structure which retains the GO layers intact even under the saline conditions. Moreover, during water purification after bacterial degradation, GO foam can easily be separated from the respective media without the need of any sophisticated techniques in contrast to GO precipitate. Therefore, these macroporous GO foams pave the way for the future antibacterial applications even under the physiological conditions, where GO precipitate shows lower activities due to aggregation issues.

Fig. 9 FE-SEM images of *E. coli* cells in the absence of GO materials (a and b), in the presence of GO precipitate (c and d), in the presence of GO foam (e and f). Arrows in the high magnification images show the disruption of cell membrane of *E. coli* cells in the presence of GO materials.

4. Conclusions

Macroporous, 3D GO foams were prepared by simple lyophilization technique from GO dispersions in water. These foams were further employed in dye adsorption and antibacterial applications. Because of its high surface area, these foams had shown efficient adsorption abilities towards the three cationic dyes such as RB, MG and AF. pH dependent adsorption studies resulted in the different sorption capacities of these foams based on the interactions with dye molecules. These foams also exhibited excellent antibacterial activities towards *E. coli* cells both in aqueous and saline environment unlike the GO precipitate which loses its activity in the presence of salt ions. The activity of GO foam even under saline conditions can be attributed to the 3D random arrangement of GO layers, which helps in decreasing the aggregation of GO layers. Porous foams have additional advantage over GO precipitate in material recovery processes. Due to 3D porous structure, GO foams can have better applicability in practical applications of water remediation, where most of the water bodies consist of salt ions. Further, the activities of these foams in environmental

applications can be improved by tuning the porosity of these foams which is feasible by lyophilization.

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