



Cite this: *Org. Biomol. Chem.*, 2016, **14**, 9639

Stereospecific prenylation of tryptophan by a cyanobacterial post-translational modification enzyme†

Masahiro Okada,* Tomotoshi Sugita, Kohei Akita, Yu Nakashima, Tian Tian, Chang Li, Takahiro Mori and Ikuro Abe*

Prenylation is a key post-translational reaction to increase the structural diversity and bioactivity of peptides and proteins. Until now, only one post-translational modification enzyme, ComQ, has been identified to mediate the prenylation of a tryptophan residue in ribosomally synthesized peptides. Here, we report the *in vitro* characterization of KgpF, a novel prenyltransferase which transfers dimethylallyl moieties to tryptophan residues during kawaguchipeptin A biosynthesis. The stereospecific prenylation by KgpF was determined by a combination of *in vitro* dimethylallylation of Fmoc-tryptophan by KgpF and chemical synthesis of dimethylallylated Fmoc-tryptophan diastereomers. KgpF modified the tryptophan derivative with a dimethylallyl group at the 3 position of its indole ring, resulting in the formation of a tricyclic structure with the same scaffold as prenylation by ComQ, but with the opposite stereochemistry.

Received 12th August 2016,
Accepted 15th September 2016

DOI: 10.1039/c6ob01759b

www.rsc.org/obc

Introduction

Numerous indole alkaloids are synthesized through tryptophan intermediates prenylated at each possible position of the indole ring.^{1–4} The prenylations of tryptophan contribute to the structural diversity and bioactivities of natural products. Indeed, several aromatic prenyltransferases have been identified as prenyl tryptophan synthases, and they are members of the ABBA prenyltransferase family that share a common structural motif known as the ABBA fold.⁵ In contrast, relatively few modification enzymes are known to mediate C–C bond formation for ribosomally synthesized and post-translationally modified peptides (RiPPs), even though new classes of post-translational modification enzymes have been reported in recent years.^{6,7} We first demonstrated post-translational prenylation of a tryptophan residue in a quorum sensing pheromone, ComX pheromone, produced by *Bacillus subtilis* and related bacilli.⁸ The tryptophan residue of ComX is modified by ComQ with either a geranyl or a farnesyl group at the gamma position, resulting in the formation of a tricyclic structure, including a newly formed pyrrolidine ring (Fig. 1).^{9–11} ComQ shows moderate similarity to isoprenyldiphosphate synthase, but no similarity to the ABBA family of aromatic prenyltransferases. Notably, in order to confirm the stereospecific

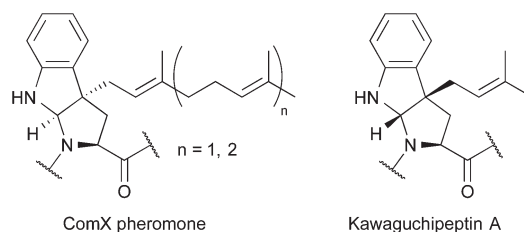


Fig. 1 Chemical structure of the modified tryptophan residues.

prenylation, the chemical synthesis of the ComX pheromone and its diastereomers was required because the stereochemistry of the modified tryptophan residue could not be determined by NMR analyses of the ComX pheromone, such as NOESY and ROESY measurements.^{9–12}

Another RiPP containing prenylated tryptophan residues, kawaguchipeptin A (**1**), was identified from the cyanobacterium *Microcystis aeruginosa* NIES-88 (Fig. 2).¹³ Kawaguchipeptin A (**1**), together with kawaguchipeptin B, are members of the cyanobactin family (cyclic RiPPs produced by cyanobacteria).¹⁴ Kawaguchipeptins A (**1**) and B are cyclic undecapeptides with very similar amino acid sequences. However, in contrast to kawaguchipeptin B, which possesses only L-amino acid residues (cyclic-[WLNGDNNWSTP]), kawaguchipeptin A (**1**) contains one D-leucine and two dimethylallylated tryptophan residues.^{13,14} Notably, it was suggested that the two tryptophan residues in **1** are modified with a dimethylallyl group at the C-3 position of their indole rings, resulting in the same scaffold as that of the ComX pheromone, but in the opposite

Graduate School of Pharmaceutical Sciences, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo, 113-0033, Japan. E-mail: abei@mol.f.u-tokyo.ac.jp

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c6ob01759b



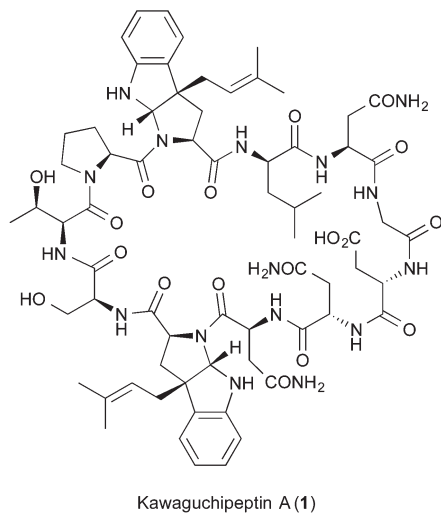


Fig. 2 Chemical structure of kawaguchipeptin A (1).

configuration (Fig. 1). However, the stereochemistry of C-3 in **1** has not been assigned unambiguously, because the stereochemistry of **1** was determined only by comparing the coupling constants of NMR spectra with those of similar synthetic prenylated tryptophan compounds.¹³ Here, we report the stereochemistry of the modified tryptophan residues, solved by using a combination of *in vitro* dimethylallylation of protected tryptophan derivatives and the chemical synthesis of the dimethylallylated Fmoc-tryptophan, Fmoc-Trp*(β -Pre) (**2b**), according to our strategy for the structure determination of ComX pheromones.^{9–12}

Results and discussion

In order to investigate the biosynthetic enzymes responsible for **1**, we first performed genome sequencing of the kawaguchipeptin A producing strain, *M. aeruginosa* NIES-88. As a result, we identified approximately 7 kb of a gene cluster flanked by two genes encoding two proteases at each end, which is an indicator of a cyanobactin biosynthetic gene cluster (Fig. S1†). A short peptide possessing the amino acid sequence of kawaguchipeptins [WLNGDNNWSTP] was encoded in this region. In addition, an ABBA prenyltransferase was encoded and located adjacent to the gene in this region. These results suggested that the gene cluster is responsible for kawaguchipeptin biosynthesis, and the ABBA prenyltransferase, KgpF, mediates the prenylation of the tryptophan residues as well as other prenyltransferases for tyrosine, serine, or threonine residues reported previously in the biosynthesis of cyanobactins.^{15,16} Very recently, Parajuli *et al.* have independently identified the kawaguchipeptin gene cluster, named *kgpA*-G, in *M. aeruginosa* NIES-88, and demonstrated its involvement in kawaguchipeptin production *in vivo*.¹⁷ Although they also reported the *in vitro* prenylation of several cyclic or linear peptides with dimethylallyldiphosphate (DMAPP) by

KgpF, the chemical structures of the modified tryptophan residues were never determined, much less the absolute configurations.¹⁷ To confirm the stereospecific prenylation of the tryptophan residues in **1** by KgpF, we investigated the *in vitro* prenylation of several protected tryptophan derivatives and precursor peptides. To our delight, we found that KgpF can prenylate Fmoc-tryptophan; however, prenylation was not observed at all when tripeptides [PWL] or [NWS], tryptophan, Boc-tryptophan, tryptophan methyl esters, Fmoc-Trp-OCH₃ or Trp-OCH₃, were used as substrates. In addition, Fmoc-tryptophan could not be modified by KgpF when GPP or FPP was used as an extender (Table 1).

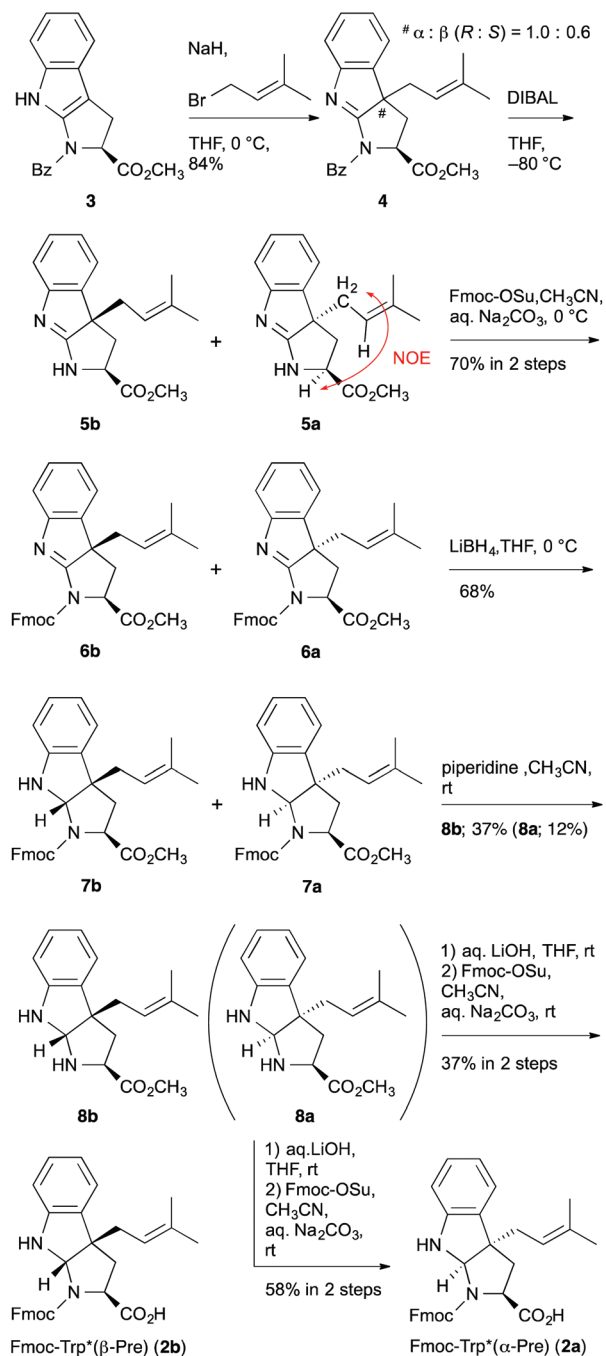
To accurately confirm the stereochemistry of the prenylation product, we synthesized Fmoc-protected C-3 dimethylallylated L-tryptophan diastereomers, Fmoc-Trp*(α -Pre) (**2a**) and Fmoc-Trp*(β -Pre) (**2b**), according to the synthesis of the ComX pheromone and its diastereomers (Scheme 1).^{9–12,18} Dimethylallylation of the tricyclic compound **3** was accomplished by treating it with NaH and 3,3-dimethylallyl bromide to give predominantly the C-dimethylallylated compounds **4a** and **4b**, as an inseparable mixture of diastereomers with a ratio of 1.0 : 0.6 (**4a** : **4b**, corresponding to *R* : *S*) in a high yield. Reductive cleavage of the benzoyl group of **4a** and **4b** was chemoselectively achieved by diisobutylaluminium hydride (DIBAL), although the reason for this highly selective reduction in the presence of the methyl ester was unknown. The diastereomers were separated by silica gel column chromatography to give analytically pure compounds **5a** and **5b**, and the stereochemistry was definitively determined by NOE analysis of **5a** (Fig. S4†). However, due to the instability of **5a**, the mixture was used in the next step reaction without purification. After Fmoc protection of **5a** and **5b** to give **6a** and **6b**, respectively, chemoselective reduction of the imines in **6a** and **6b** was achieved by lithium borohydride to give **7a** and **7b**, respectively, and then deprotection of Fmoc afforded the diastereomixture of **8a** and **8b**. The diastereomers were carefully separated by silica gel column chromatography to give the optically pure esters **8a** and **8b**. Although it was rather difficult to accurately determine the stereochemistry of **8a** (or **8b**) by NOE, except for the *cis* annulation, the small-scale synthesis of **8a** (or **8b**) using pure **5a** (or **5b**) as a reactant has made it possible for us to identify the absolute configurations of **8a** and **8b**.

Table 1 Substrates and extenders used for the *in vitro* reaction with KgpF

Substrate	Extender	Product
Pro-Trp-Leu	DMAPP	Not detected
Asn-Trp-Ser	DMAPP	Not detected
Trp	DMAPP	Not detected
Trp-OCH ₃	DMAPP	Not detected
Boc-Trp	DMAPP	Not detected ^a
Fmoc-Trp	DMAPP	2b
Fmoc-Trp-OCH ₃	DMAPP	Not detected
Fmoc-Trp	GPP	Not detected
Fmoc-Trp	FPP	Not detected

^a Others have independently confirmed the same result.¹⁶





Scheme 1 Synthesis of Fmoc-protected prenylated tryptophans.

Finally, hydrolysis of ester **8a** (or **8b**), and successive Fmoc protection afforded the desired compound **2a** (or **2b**). The chemical structures were scarcely determined by the NMR analyses of **2a** and **2b**, because the $^1\text{H-NMR}$ spectra of **2a** and **2b** were observed as 1 : 1 equilibrium mixtures between N-conformers, respectively (Fig. S14 and S15[†]). Therefore, due to the unambiguous confirmation of the synthesis of **2a** (or **2b**) without racemization, the small-scale synthesis of **8a** (or **8b**) using pure **2a** (or **2b**) via methyl esterification and Fmoc deprotection was carried out (data not shown).

The stereochemistry of the enzyme reaction was determined by comparing the retention times and mass fragmentations obtained from LC-MS with those of the optically pure synthetic standards **2a** and **2b**. The enzymatic product was consistent with **2b**, but not with **2a**, in LC-MS analysis (Fig. 3 and S16[†]). These results clearly demonstrated that KgpF modified Fmoc-tryptophan with a dimethylallyl group at the 3 position of its indole ring, resulting in the formation of a tricyclic structure with the same scaffold as ComX pheromones, but with the opposite stereochemistry.

In contrast to typical post-translational modifications, a specific amino acid sequence adjacent to the core peptide for directing KgpF is unlikely to be required. In addition, a specific amino acid motif in the core peptide is unlikely to be required for prenylation by KgpF, because there is no similarity between the sequences surrounding the two tryptophan residues prenylated by KgpF in kawaguchipeptin B. However, a recent paper reports that KgpF only modified the internal tryptophan residue of the two tryptophan residues in a linear undecapeptide [WLNGDNNWSTP], which possesses the same sequence as kawaguchipeptins from the N-terminal tryptophan, to the C-terminal proline.¹⁷ Considering the results together with our findings, N-terminal tryptophan residues and tryptophan derivatives with a free amino group are not recognized as a substrate by KgpF. More importantly, only one amino acid residue, Fmoc-tryptophan, was prenylated with DMAPP by KgpF. The substrate specificity of KgpF is similar to an ABBA prenyltransferase, LynF, for the biosynthesis of the cyanobactin, prenylagaramide B.¹⁶ LynF mediated the prenylation of not only a tyrosine residue of the original cyclic peptide, but also N-terminus protected tyrosine in the *in vitro* reaction.¹⁶ In a structural model of KgpF using the structures of a cyanobactin synthase, PatF (PDB ID codes 4BG2), and a dimethylallyl tryptophan synthase (PDB ID codes 3I4X) from

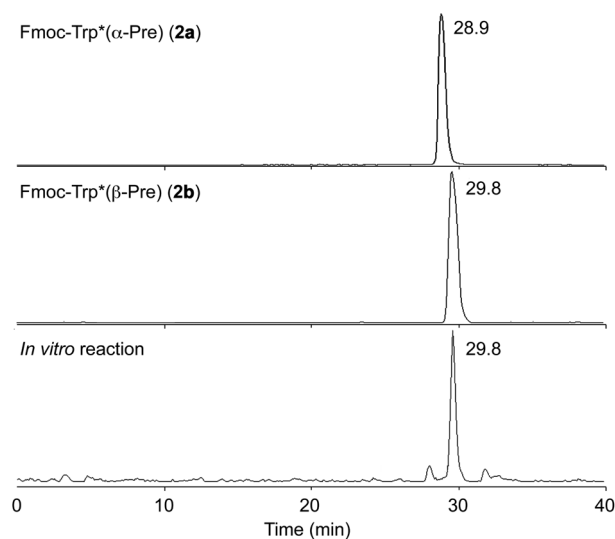


Fig. 3 LC-MS analysis of *in vitro* reaction of Fmoc-Trp with KgpF. Extracted ion chromatograms (positive) of synthetic **2a** (top), synthetic **2b** (middle), and the *in vitro* reaction product (bottom) at m/z 495.



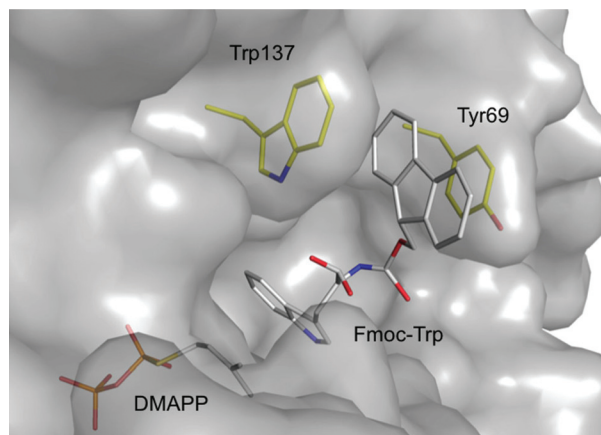


Fig. 4 A structural model of KgpF generated by using SWISS MODEL. The N-terminal Fmoc group of Fmoc-tryptophan was bound with aromatic amino acid residues in KgpF. The two aromatic amino acid residues, Tyr69 and Trp137, are highlighted in yellow.

Aspergillus fumigatus,^{19,20} the N-terminal Fmoc group of Fmoc-tryptophan was bound with aromatic amino acid residues, Tyr69 and Trp137 in KgpF (Fig. 4). It was suggested that these residues are important for its ability to bind Fmoc-tryptophan (Scheme 2). To obtain more detailed information about this enzyme, crystal structural studies of KgpF are needed in addition to *in vitro* prenylation studies.

Since the indole side chain of a tryptophan residue is an efficient nucleophile for C–C bond formation, most patterns of tryptophan posttranslational modifications involve C–C bond formation, even though there are fewer classes of posttranslational modifications for tryptophan than for other amino acid residues.^{9–11,13,21,22} Among the modification enzymes, KgpF accepted even Fmoc-tryptophan as a substrate and mediated regioselective and stereoselective dimethylallylation of Fmoc-tryptophan. Therefore, KgpF may exhibit relaxed substrate specificity toward diverse tryptophan residues in peptides, which in turn would expand the chemical diversity of peptides with prenylated tryptophan among the residues. KgpF may have great potential as a promising enzyme for the synthesis of prenylated tryptophan derivatives and peptide libraries containing prenylated tryptophan residues.

Conclusion

In this study, we analyzed the *in vitro* prenylation of tryptophan derivatives by KgpF. The stereospecific prenylation by KgpF was determined using a combination of *in vitro* dimethylallylation of Fmoc-tryptophan and chemical synthesis of dimethylallylated Fmoc-tryptophan diastereomers. KgpF modified the tryptophan derivative with a dimethylallyl group at the 3 position of its indole ring, resulting in the formation of a tricyclic structure with the same scaffold as ComX pheromones, but with the opposite stereochemistry. Since KgpF exhibited relaxed substrate specificity, KgpF may attract attention as a target for enzyme engineering for the synthesis of prenylated tryptophan derivatives.

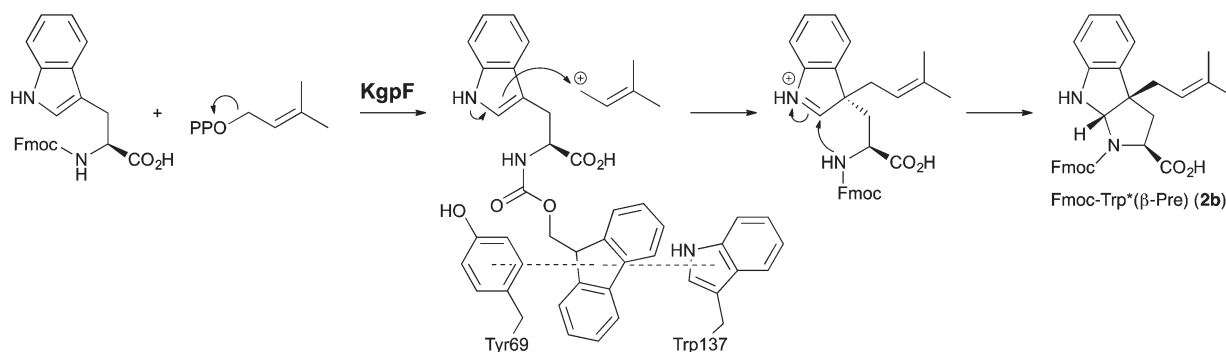
Experimental

Genomic analysis of *Microcystis aeruginosa* NIES-88

M. aeruginosa NIES-88 was obtained from the National Institute for Environmental Studies (Japan) collection. The *M. aeruginosa* NIES-88 genomic DNA was isolated according to the previously published method.²³ Sequencing of the genomic DNA was performed by an Ion PGM sequencer, with a total number of 618551 sequence reads (*ca.* 300 bp). These sequences were assembled *de novo* into contigs by using the Genious assembler (Biomatters), with the default medium sensitivity. Putative protein-coding sequences were determined by combining the prediction results from the FramePlot and Glimmer 3.02 programs into one large contig (7.4 kbp).^{24,25} The domain organizations were assessed by using BLASTP. The biosynthetic gene cluster of kawaguchipeptins is shown in Fig. S1.†

Heterologous expression and purification of KgpF

The KgpF gene was amplified from the genomic DNA of *M. aeruginosa* NIES-88 with a set of primers. After purification, the gene was ligated into the pET-22b(+) vector digested with *SacI* and *XhoI* by using an In-Fusion HD Cloning Kit. The constructed plasmid was transformed into the *Escherichia coli* BL21(DE3) strain for heterologous overexpression. When the



Scheme 2 Plausible mechanism for the synthesis of **2b** by KgpF.



transformant was grown at 37 °C in Luria-Bertani medium with ampicillin at an OD600 of 0.6–0.8, the culture was cooled at 20 °C, and then IPTG (final conc. 1.0 mM) was added and incubated for 12 h at 20 °C. The cells were collected and re-suspended in 100 mM Tris-HCl buffer (pH 7.5) containing 400 mM NaCl, 5 mM imidazole, and 10% (v/v) glycerol, and the cells were disrupted by ultrasonication. After centrifugation at 9000g for 20 min, the supernatant was loaded onto a Ni²⁺-NTA-agarose column. After washing the resin with the buffer containing 15 mM imidazole, KgpF with a C-terminal His tag was eluted with the buffer containing 300 mM imidazole. The eluate was filtered through a 10 kDa MWCO filter to give KgpF solution (500 µL at 44 g L⁻¹). The protein solution for the *in vitro* reaction was prepared before use. Forward primer; GGAGATTAAGTATGCGGATATAATTGATATACC, reverse primer; GGTGGTGGTCTCGAAGTTGGGAGAATCCCACC.

In vitro assay

To a 50 mM HEPES buffer (pH 7.5) containing 5 mM MgCl₂, 5 mM ZnCl₂, dimethylallyl diphosphate solution (final conc. 300 µM), substrate solution (final conc. 300 µM), and KgpF solution (final conc. 200 µM) were added, and the total volume was adjusted to 100 µL with the buffer solution. After the reaction mixture was incubated for 12 h at 37 °C, it was quenched and neutralized with 100 µL of acetonitrile. After centrifugation at 12 000g for 5 min, the supernatant was filtered through a membrane filter (0.45 µm) to prepare the sample for LC-MS analysis.

LC-MS analysis

LC-MS analysis was performed with a JMS-T100LP (JEOL) ESI-MS system and a 1100 series HPLC system (Agilent) with a X-select CSH C18 (Waters) column (2.1 × 150 mm). Each sample (10 µL) was injected into the LC column and eluted with a linear gradient of 58–95% CH₃CN/H₂O containing 0.5% acetic acid for 40 min at a flow rate of 0.1 mL min⁻¹. The enlarged extracted ion chromatograms and mass spectrum are shown in Fig. S17.†

Homology modeling

The structural model of KgpF was generated by the SWISS-MODEL web server (swissmodel.expasy.org) using the crystal structure of PatF (PDB ID; 4BG2), which belongs to the ABBA family of prenyltransferases,¹⁹ and dimethylallyl tryptophan synthase from *Aspergillus fumigatus* (PDB ID; 3I4X).²⁰

Acknowledgements

This work was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology, Japan (JSPS KAKENHI Grant Number JP15H01836, JP16H06443, and JP24688011) and the Takeda Science Foundation (to M. O.).

References

- 1 X. Yu and S.-M. Li, *Methods Enzymol.*, 2012, **516**, 259.
- 2 T. Walsh, *ACS Chem. Biol.*, 2014, **9**, 2718.
- 3 M. E. Tanner, *Nat. Prod. Rep.*, 2015, **32**, 88.
- 4 J. A. McIntosh, M. S. Donia and E. W. Schmidt, *Nat. Prod. Rep.*, 2009, **26**, 537.
- 5 M. Tello, T. Kuzuyama, L. Heide, J. P. Noel and S. B. Richard, *Cell. Mol. Life Sci.*, 2008, **65**, 1459.
- 6 L. M. Alkhalaf and K. S. Ryan, *Chem. Biol.*, 2015, **22**, 317.
- 7 P. G. Arnison, M. J. Bibb, G. Bierbaum, A. A. Bowers, T. S. Bugni, G. Bulaj, J. A. Camarero, D. J. Campopiano, G. L. Challis, J. Clardy, P. D. Cotter, D. J. Craik, M. Dawson, E. Dittmann, S. Donadio, P. C. Dorrestein, K. D. Entian, M. A. Fischbach, J. S. Garavelli, U. Goransson, C. W. Gruber, D. H. Haft, T. K. Hemscheidt, C. Hertweck, C. Hill, A. R. Horswill, M. Jaspars, W. L. Kelly, J. P. Klinman, O. P. Kuipers, A. J. Link, W. Liu, M. A. Marahiel, D. A. Mitchell, G. N. Moll, B. S. Moore, R. Muller, S. K. Nair, I. F. Nes, G. E. Norris, B. M. Olivera, H. Onaka, M. L. Patchett, J. Piel, M. J. Reaney, S. Rebuffat, R. P. Ross, H. G. Sahl, E. W. Schmidt, M. E. Selsted, K. Severinov, B. Shen, K. Sivonen, L. Smith, T. Stein, R. D. Sussmuth, J. R. Tagg, G. L. Tang, A. W. Truman, J. C. Vederas, C. T. Walsh, J. D. Walton, S. C. Wenzel, J. M. Willey and W. A. van der Donk, *Nat. Prod. Rep.*, 2013, **30**, 108.
- 8 R. Magnuson, I. Solomon and A. D. Grossman, *Cell*, 1994, **77**, 207.
- 9 M. Okada, I. Sato, S. J. Cho, H. Iwata, T. Nishio, D. Dubnau and Y. Sakagami, *Nat. Chem. Biol.*, 2005, **1**, 23.
- 10 M. Okada, H. Yamaguchi, I. Sato, F. Tsuji, J. Qi, D. Dubnau and Y. Sakagami, *Biosci. Biotechnol., Biochem.*, 2007, **71**, 1807.
- 11 M. Okada, H. Yamaguchi, I. Sato, F. Tsuji, D. Dubnau and Y. Sakagami, *Biosci. Biotechnol., Biochem.*, 2008, **72**, 914.
- 12 M. Okada, I. Sato, S. J. Cho, D. Dubnau and Y. Sakagami, *Tetrahedron*, 2006, **62**, 8907.
- 13 K. Ishida, H. Matsuda, M. Murakami and K. Yamaguchi, *Tetrahedron*, 1996, **52**, 9025.
- 14 K. Ishida, H. Matsuda, M. Murakami and K. Yamaguchi, *J. Nat. Prod.*, 1997, **60**, 724.
- 15 M. S. Donia, J. Ravel and E. W. Schmidt, *Nat. Chem. Biol.*, 2008, **4**, 341.
- 16 J. A. McIntosh, M. S. Donia, S. K. Nair and E. W. Schmidt, *J. Am. Chem. Soc.*, 2011, **133**, 13698.
- 17 A. Parajuli, D. H. Kwak, L. Dalponte, N. Leikoski, T. Galica, U. Umeobika, L. Trembleau, A. Bent, K. Sivonen, M. Wahlsten, M. Wang, E. Rizzi, G. De Bellis, J. Naismith, J. Jaspars, X. Liu, W. Houssen and D. P. Fewer, *Angew. Chem., Int. Ed.*, 2016, **55**, 3596.
- 18 F. Tsuji, K. Kobayashi, M. Okada, H. Yamaguchi, M. Ojika and Y. Sakagami, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 4041.
- 19 A. F. Bent, K. Jesko, W. E. Houssen, M. C. M. Smith, M. Jaspars and J. H. Naismith, *Acta Crystallogr., Sect. F: Struct. Biol. Commun.*, 2013, **69**, 618.



- 20 U. Metzger, C. Schall, G. Zoher, I. Unsöld, E. Stec, S.-M. Li, L. Heide and T. Stehle, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 14309.
- 21 W. S. McIntire, D. E. Wemmer, D. E. A. Chistoserdov and M. E. Lindstrom, *Science*, 1991, **252**, 817.
- 22 J. Hofsteenge, D. R. Müller, T. de Beer, A. Löffler, W. J. Richter and J. F. G. Vliegthart, *Biochemistry*, 1994, **33**, 13524.
- 23 N. Morin, T. Vallaey, L. Hendrickx, L. Natalie and A. Wilmotte, *J. Microbiol. Methods*, 2010, **80**, 148.
- 24 J. Ishikawa and K. Hotta, *FEMS Microbiol. Lett.*, 1999, **174**, 251.
- 25 A. L. Delcher, D. Harmon, S. Kasif, O. White and S. L. Salzberg, *Nucleic Acids Res.*, 1999, **27**, 4636.

