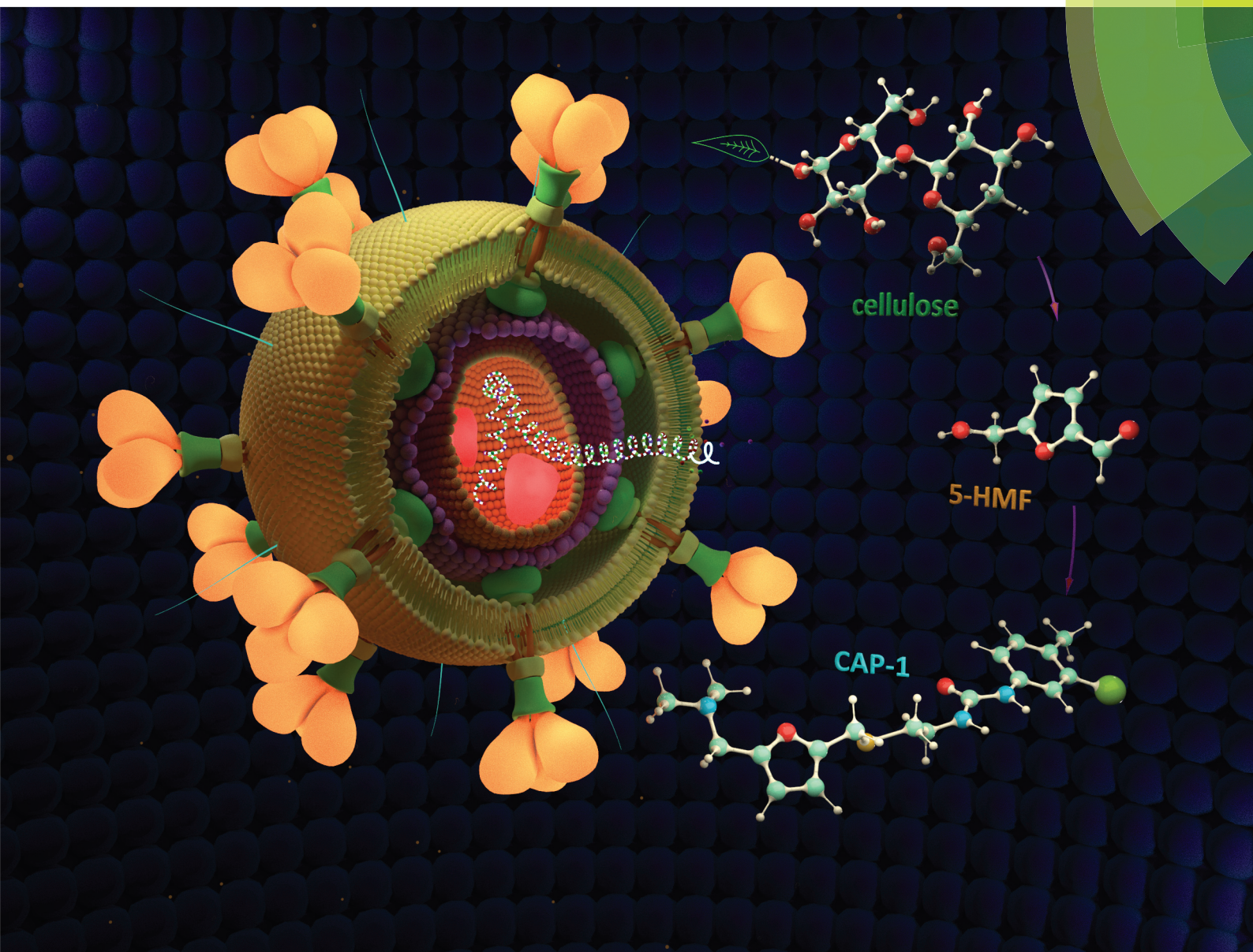


Organic & Biomolecular Chemistry

www.rsc.org/obc



ISSN 1477-0520



PAPER

Leonid V. Romashov and Valentine P. Ananikov
Synthesis of HIV-1 capsid protein assembly inhibitor (CAP-1) and its analogues based on a biomass approach

175 YEARS



Cite this: *Org. Biomol. Chem.*, 2016, **14**, 10593

Synthesis of HIV-1 capsid protein assembly inhibitor (CAP-1) and its analogues based on a biomass approach†

Leonid V. Romashov^a and Valentine P. Ananikov^{*a,b}

Received 10th August 2016,
Accepted 16th September 2016

DOI: 10.1039/c6ob01731b

www.rsc.org/obc

A biomass-derived platform chemical was utilized to access a demanded pharmaceutical substance with anti-HIV activity (HIV, human immunodeficiency virus) and a variety of structural analogues. Step economy in the synthesis of the drug core (single stage from cellulose) is studied including flexible variability of four structural units. The first synthesis and X-ray structure of the inhibitor of HIV-1 capsid protein assembly (CAP-1) is described.

Introduction

State-of-the-art healthcare urges for the on-demand production of pharmaceuticals¹ and access to personalized medicine.² The long term practical implementation of these goals is challenging to achieve due to unsustainable procedures of drugs synthesis, which are typically characterized by large overall production of toxic waste.³ In fact, the synthesis of drugs is accompanied by the largest amount of waste by-products among all chemical industry processes (25–100 kg of waste per 1 kg of drug product).³ A breakthrough concept in this area is to connect improved synthesis and step-economy⁴ with sustainable natural sources of chemicals.⁵ The use of biomass-derived chemicals has opened up new possibilities in the sustainable synthesis of bulk and fine chemicals.⁶ In this article, we describe a complete synthetic procedure to promising anti-HIV pharmaceutical substances starting from a low cost natural biomass (cellulose) and using a special optimized procedure for efficient fine organic synthesis applications.⁷

The search for effective anti-HIV drugs is one of the most important challenges in modern science and medicine.⁸ AIDS causes more than a million deaths annually.⁹ There is no cure or vaccine against AIDS, but antiretroviral therapy can slow the course of the disease and increase the survival time after infection.^{10,11} In this field, the rapid synthesis of candidate compounds with the possibility of synthesizing close structural analogues is of crucial importance.¹²

Currently available drugs against HIV are focused on the inhibition of the five stages of the HIV life cycle (Fig. 1) and include CCR5 coreceptor antagonists (*maraviroc*), which prevent HIV from binding to CD4 cells; fusion inhibitors (*enfuvirtide*), which block the HIV envelope from merging with the host CD4 cell membrane; nucleoside (*lamivudine*, *zidovudine*, *emtricitabine*, *abacavir*, *azidothymidine*, *didanosine*, etc.) and non-nucleoside (*rilpivirine*, *etravirine*, *delavirdine*, *efavirenz*, etc.) reverse transcriptase inhibitors, which block the conversion of HIV RNA into DNA; integrase strand transfer inhibitors (*raltegravir*, *dolutegravir*), which block the insertion of HIV DNA into CD4 cell DNA; and viral protease inhibitors (*tipranavir*, *indinavir*, *lopinavir*, *ritonavir*, *darunavir*, etc.), which prevent the cleavage of newly synthesized viral polyproteins at

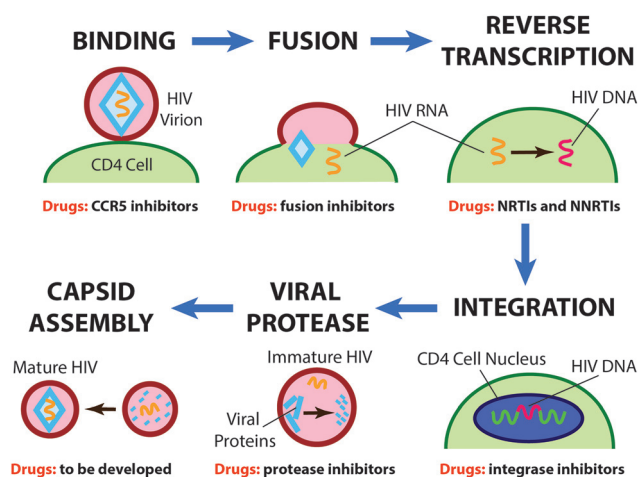


Fig. 1 Therapeutic target stages of HIV life cycle and the corresponding drugs.

^aZelinsky Institute of Organic Chemistry, Russian Academy of Sciences, Leninsky prospect 47, 119991 Moscow, Russia. E-mail: val@ioc.ac.ru

^bSaint Petersburg State University, Stary Peterhof, Saint Petersburg, 198504, Russia

† Electronic supplementary information (ESI) available: Experimental procedures and characterization data. CCDC 1497315. For crystallographic data in CIF or other electronic format see DOI: 10.1039/c6ob01731b

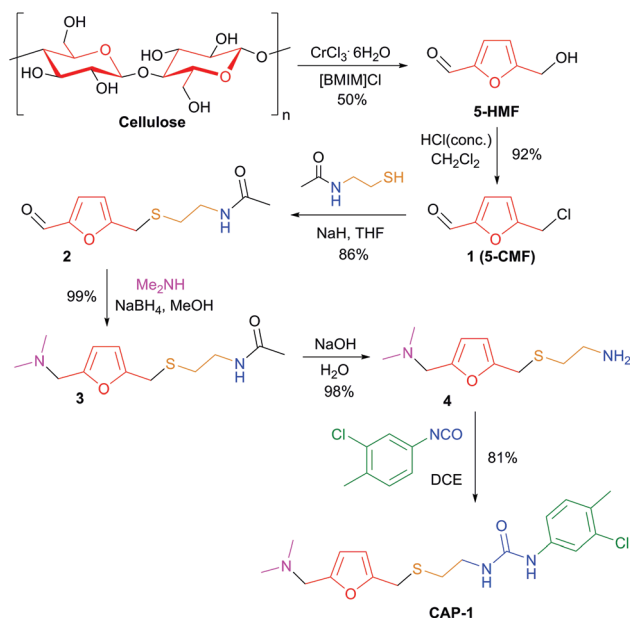


appropriate places for creating mature protein components of the infectious HIV virion.¹³

Recently, a novel type of anti-HIV activity – inhibition of HIV-1 capsid protein assembly, which is the last stage of HIV maturation – was described.¹⁴ *N*-(3-Chloro-4-methylphenyl)-*N'*-2-[5-[(dimethylamino)-methyl]-2-furylmethyl)sulfanyl]ethylurea (CAP-1) was one of the first compounds with such activity.^{14g,h} The structure of CAP-1 was disclosed in patents,¹⁵ however, to the best of our knowledge, the methods of synthesis have not been described and a 3D molecular structure was not determined.

Results and discussion

Here we report an efficient synthesis of the CAP-1 molecule which allows versatile structural variation (Fig. 2). The structure of CAP-1 can be subdivided into five fragments, which should be introduced into the target molecule as individual subunits. Retrosynthetic analysis of CAP-1 shows that this approach can be easily realized on the basis of the 2,5-disubstituted furan core (shown in red, Fig. 2), which is available from natural biomass involving the 5-HMF (5-(hydroxymethyl)furfural) platform-chemical^{7,16} pathway (Fig. 2). The first site, which can be varied easily, is an amine group (drug unit 1, Fig. 2). Variation at this position is possible *via* reductive amination of the 5-HMF aldehyde group with various primary and secondary amines. The second site is a spacer of variable length and flexibility (drug unit 2, Fig. 2). The third site (drug unit 3, Fig. 2) is a linkage group, which connects the spacer to an aromatic, aliphatic or heteroaromatic substituent (drug unit 4, Fig. 2) and can also be involved in hydrogen bonding with the substrate. The linkage can be composed of such important groups as urea, amide, ester, carbonate, carbamate,



Scheme 1 Synthesis of CAP-1 from cellulose as the starting material (similar color encoding of fragments was used, see Fig. 2).

sulphonamide, *etc.* Each of these four sites can be varied independently, and the final molecule can be assembled using efficient synthetic procedures. Thus, a library of CAP-1 structural analogues can be synthesized on the basis of this biomass-derived approach.

A practical synthesis procedure is shown in Scheme 1. The overall synthesis includes the following six steps: (1) CrCl_3 -catalysed conversion of cellulose into 5-HMF in an ionic liquid;^{7,17} (2) preparation of 5-(chloromethyl)furfural^{16b,c,18} (5-CMF) upon treatment of 5-HMF with HCl; (3) *S*-alkylation of *N*-acetylcysteine with 5-CMF; (4) reductive amination of the formyl group by using dimethylamine; (5) alkaline hydrolysis of the amide; (6) synthesis of the urea group by using arylisocyanate. Each step was conducted smoothly, and compound purification *via* chromatography was required only once, at the final step. The target product was isolated in 57% overall yield in a crystal form with >99.9% purity (see the ESI† for analysis).

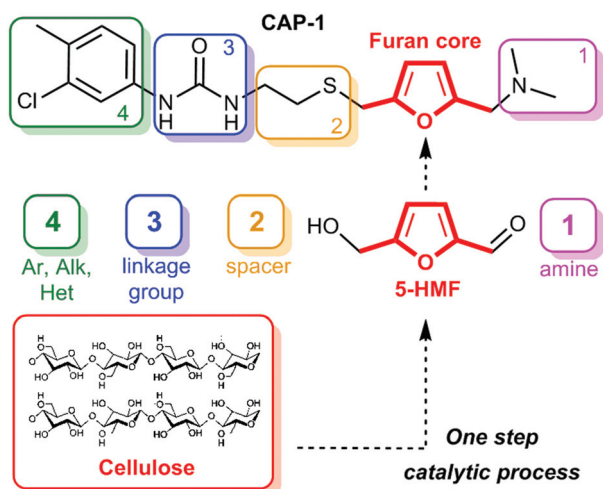


Fig. 2 Retrosynthetic analysis of CAP-1 and its partitioning into sites of structural variation: red – furan core, magenta – amine, orange – spacer, blue – linkage group, green – aromatic, heteroaromatic or aliphatic substituent.

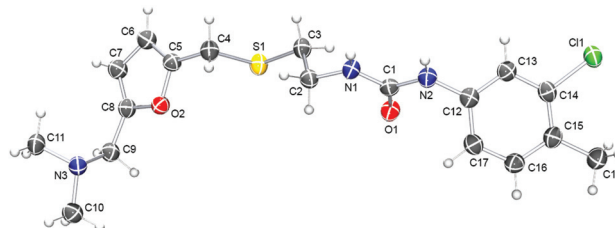
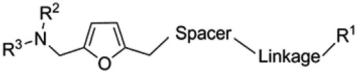
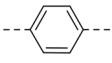


Fig. 3 Molecular view of CAP-1 determined by X-ray analysis. Selected geometry parameters: $\text{N}(3)–\text{C}(9) = 1.475 \text{ \AA}$, $\text{C}(9)–\text{C}(8) = 1.497 \text{ \AA}$, $\angle \text{N}(3)\text{C}(9)\text{C}(8) = 113.26^\circ$, $\text{C}(5)–\text{C}(4) = 1.487 \text{ \AA}$, $\text{C}(4)–\text{S}(1) = 1.843 \text{ \AA}$, $\angle \text{C}(5)\text{C}(4)\text{S}(1) = 107.61^\circ$, $\angle \text{C}(4)\text{S}(1)\text{C}(3) = 102.25^\circ$, $\angle \text{S}(1)\text{C}(3)\text{C}(2)\text{N}(1) = 168.33^\circ$, $\angle \text{C}(1)\text{N}(2)\text{C}(12)\text{C}(17) = 53.48^\circ$, and $\angle \text{N}(1)\text{C}(1)\text{N}(2) = 115.17^\circ$ (see the ESI† for complete description).



Table 1 Synthetic scope in the preparation of CAP-1 analogues

						
Entry	R ¹	R ²	R ³	Linkage	Spacer	Yield, %
1	4-Methyl-3-chlorophenyl	Me	Me	Urea		46
2	Ph	Me	Me	Urea	–SCH ₂ CH ₂ –	70
3	Me	Me	Me	Amide	–SCH ₂ CH ₂ –	79
4	1-Adamantan-methyl	Me	Me	Urea	–SCH ₂ CH ₂ –	72
5	1-Chloro-3-adamantyl	Me	Me	Urea	–SCH ₂ CH ₂ –	66
7	<i>p</i> -Tolyl	Me	Me	Sulfonamide	–SCH ₂ CH ₂ –	75
8	2-Nitrophenyl	Me	Me	Sulfonamide	–SCH ₂ CH ₂ –	65
9	H	Cyclo-propyl	H	Alcohol	–S–CH ₂ CH ₂ –	55
10	<i>t</i> -Bu	Me	Me	Carbamate	–SCH ₂ CH ₂ –	73

The critical point of the synthetic procedure was to ensure the use of purified 5-HMF (free from dimer and oligomer impurities) to avoid formation of by-products, simplify separation and to increase the yield.⁷ The solid-state structure of the synthesized CAP-1 molecule was determined by single crystal X-ray diffraction for the first time and revealed a twisted linear geometry (Fig. 3). The determined molecular structure (see the ESI† for complete details) gives key information for performing docking experiments, understanding the mechanism of action and the analysis of biological activity.

To demonstrate the applicability of the developed approach several structural analogues of CAP-1 were synthesized (Table 1). Variation in the substituents and in the nature of the linkage were successfully carried out giving the products in 46–79% yield (from 5-HMF). The developed biomass-derived approach was fully applicable for rapid access to various CAP-1 functionalized derivatives.

Conclusions

In conclusion, we have evaluated a convenient synthetic procedure for the preparation of the prominent HIV-1 capsid protein assembly inhibitor CAP-1 using cellulose as a renewable starting material. The structure of CAP-1 was determined by X-ray analysis. Using the developed procedure, several structural analogues of CAP-1 were successfully prepared and made available for high demand biochemical applications. Direct connection of the drug core to renewable natural sources of chemicals provides an efficient and green approach for the production of these pharmaceutical substances. Implementation of such methodologies into fine organic synthesis procedures is crucial to achieve long term sustainable drug development.

Conversion of natural carbohydrates into the 5-HMF platform chemical is a sustainable process,^{7a,16} and nowadays the production of 5-HMF approaches an industrial multi-ton scale.¹⁹ Therefore, a variety of efficient and practical implementations of biomass-derived drugs synthesis can be anticipated.

Experimental

5-(Hydroxymethyl)furfural was prepared from cellulose according to our recently published procedure.^{7a}

5-(Chloromethyl)furfural (1)²⁰

A 25 mL Erlenmeyer flask was loaded with 5-HMF (252 mg, 2 mmol), CH₂Cl₂ (10 mL), and conc. HCl (5 mL). The mixture was stirred at room temperature overnight. The aqueous layer was extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phase was dried over sodium sulfate. Charcoal (50 mg) was added and the mixture was stirred for 20 minutes and filtered through a Celite pad. The solvent was removed under reduced pressure to give 5-(chloromethyl)furfural (266 mg, 92%) as a yellow liquid. The product should be stored below +5 °C to prevent decomposition. ¹H NMR (CDCl₃, 400 MHz) δ = 9.61 (s, 1H), 7.18 (d, 1H, *J* = 3.5 Hz), 6.57 (d, 1H, *J* = 3.5 Hz), 4.59 (s, 2H). ¹³C{¹H} NMR (CDCl₃, 101 MHz) δ = 177.1, 156.0, 152.9, 121.6, 111.9, 36.5; Anal. Calcd for C₆H₅ClO₂: C, 49.85; H, 3.49; Cl, 24.52; Found: C, 49.84; H, 3.50, Cl, 24.49. HRMS (ESI): calcd [M + Na]⁺ 166.9870, Found: 166.9872.

5-[(2-Acetamidoethyl)thio]methyl]furfural (2)^{6b}

A Schlenk flask was loaded with *N*-acetylcysteamine (203 mg, 1.70 mmol) and a magnetic stirrer bar. The flask was filled with argon *via* three vacuum–argon cycles. 10 mL of absolute THF (distilled over sodium-benzophenone ketyl) was added. Sodium hydride (95%, 52 mg, 2.17 mmol) was added and the reaction mixture was stirred for 30 minutes. After that the solution of 5-(chloromethyl)furfural (256 mg, 1.77 mmol) in 5 mL of absolute THF was added dropwise. The reaction mixture was stirred overnight at room temperature. The solvent was removed under reduced pressure and brine (55 mL) was added. At this moment the color changed from light yellow to red-orange. The mixture was extracted with CH₂Cl₂ (4 × 40 mL), and the combined organic phase was washed with brine (100 mL) and dried over anhydrous Na₂SO₄. Charcoal (70 mg) was added and the mixture was stirred for another 20 minutes and filtered through a 15 mm Celite® pad. The



solvent was removed under reduced pressure to give 5-[[[2-acetamidoethyl]thio]methyl]furfural (332 mg, 86%) as a yellow oil. ^1H NMR (CDCl_3 , 400 MHz) δ = 9.57 (s, 1H), 7.19 (d, 1H, J = 3.5 Hz), 6.46 (d, 1H, J = 3.5 Hz), 6.17 (brs, 1H), 3.79 (s, 2H), 3.45 (q, 2H, J = 6.2 Hz), 2.73 (t, 2H, J = 6.5 Hz), 2.03 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 101 MHz) δ = 177.2, 170.3, 158.8, 152.4, 122.7, 110.5, 38.3, 32.1, 28.0, 23.2. Anal. Calcd for $\text{C}_{10}\text{H}_{13}\text{NO}_3\text{S}$: C, 52.85; H, 5.77; N, 6.16; Found: C, 52.81; H, 5.80, N, 6.14. HRMS (ESI): calcd $[\text{M} + \text{Na}]^+$ 250.0508, Found: 250.0515.

5-[[[2-Acetamidoethyl]thio]methyl]-*N,N*-dimethyl-2-furanmethanamine (3)^{6b}

The liquid dimethylamine (1 mL, prepared from dimethylamine hydrochloride and solid NaOH) was added to a solution of 5-[[[2-acetamidoethyl]thio]methyl]furfural (410 mg, 1.804 mmol) in 60 mL of absolute methanol and the resulting mixture was stirred at room temperature for 40 minutes. The solution became an intense orange. The reaction mixture was cooled to 0 °C and sodium borohydride (103 mg, 2.706 mmol) was added portionwise. The mixture was stirred at 0 °C for 20 minutes and then warmed up to room temperature. The solvent was removed under reduced pressure and the residue was dissolved in CH_2Cl_2 , filtered from inorganic impurities, and evaporated to dryness to give 5-[[[2-acetamidoethyl]thio]methyl]-*N,N*-dimethyl-2-furanmethanamine (459 mg, 99%) as a yellow oil. ^1H NMR (CDCl_3 , 400 MHz) δ = 6.27 (brs, 1H), 6.11 (s, 2H), 3.69 (s, 2H), 3.41 (s, 2H), 3.30 (q, 2H, J = 6.2 Hz), 2.65 (t, 2H, J = 6.5 Hz), 2.24 (s, 6H), 1.95 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 101 MHz) δ = 170.2, 151.9, 151.4, 109.8, 108.2, 56.0, 45.1, 38.5, 31.8, 28.3, 23.3. Anal. Calcd for $\text{C}_{12}\text{H}_{20}\text{N}_2\text{O}_2\text{S}$: C, 56.61; H, 6.33; N, 11.00; Found: C, 56.21; H, 7.91, N, 10.90. HRMS (ESI): calcd $[\text{M} + \text{H}]^+$ 257.1318, Found: 257.1321.

5-[[[2-Aminoethyl]thio]methyl]-*N,N*-dimethyl-2-furanmethanamine (4)

5-[[[2-Acetamidoethyl]-thio]methyl]-*N,N*-dimethyl-2-furanmethanamine (44 mg, 0.172 mmol) was added to 2 mL of 2N NaOH. The mixture was heated at reflux for 2.5 hours, then cooled down to room temperature and extracted with CH_2Cl_2 (3 \times 5 mL). The combined organic phase was washed with brine (15 mL) and dried over anhydrous Na_2SO_4 . The solvent was removed under reduced pressure, and the residue was dried *in vacuo* to give 5-[[[2-aminoethyl]thio]methyl]-*N,N*-dimethyl-2-furanmethanamine (36 mg, 98%) as a pale yellow oil. ^1H NMR (CDCl_3 , 400 MHz) δ = 6.09 (s, 2H), 3.67 (s, 2H), 3.40 (s, 2H), 2.81 (t, 2H, J = 6.4 Hz), 2.59 (t, 2H, J = 6.4 Hz), 2.23 (s, 6H), 1.90 (brs, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 101 MHz) δ = 151.9, 151.3, 109.4, 108.0, 55.9, 45.0, 40.8, 35.7, 28.1. Anal. Calcd for $\text{C}_{12}\text{H}_{18}\text{N}_2\text{OS}$: C, 56.04; H, 8.47; N, 13.07; Found: C, 56.01; H, 8.48, N, 13.02. HRMS (ESI): calcd $[\text{M} + \text{H}]^+$ 239.1213, Found: 239.1217.

1-(3-Chloro-4-methylphenyl)-3-((5-((dimethylamino)methyl)furan-2-yl)methyl)thio)ethylurea (CAP-1)

To a solution of 5-[[[2-aminoethyl]thio]methyl]-*N,N*-dimethyl-2-furanmethanamine (110 mg, 0.513 mmol) in 5 mL of 1,2-

dichloroethane, 3-chloro-4-methylphenylisocyanate (90.3 mg, 0.539 mmol) was added and the mixture was heated at 50 °C overnight. The solvent was removed under reduced pressure and the residue was purified by column chromatography (eluent CHCl_3 :MeOH = 8:1 (v/v)). The product was dried *in vacuo* to obtain CAP-1 (159 mg, 81%) as a white solid. ^1H NMR (CDCl_3 , 500 MHz) δ = 7.51 (s, 1H), 7.38 (d, 1H, J = 1.9 Hz), 7.13 (dd, 1H, J = 8.2, 1.9 Hz), 7.07 (d, 1H, J = 8.2 Hz), 6.15 (d, 1H, J = 3.0 Hz), 6.11 (d, 1H, J = 3.0 Hz), 5.82 (t, 1H, J = 5.8 Hz), 3.70 (s, 2H), 3.49 (s, 2H), 3.84 (q, 2H, J = 6.0 Hz), 2.69 (t, 2H, J = 6.0 Hz), 2.31 (s, 6H), 2.28 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 126 MHz) δ = 155.9, 152.2, 150.6, 138.2, 134.5, 131.1, 130.3, 120.6, 118.4, 110.7, 108.3, 55.9, 45.0, 39.4, 33.0, 28.7, 19.4. Anal. Calcd for $\text{C}_{18}\text{H}_{24}\text{ClN}_3\text{O}_2\text{S}$: C, 56.61; H, 6.33; N, 11.00; Found: C, 56.33; H, 6.36, N, 10.72. HRMS (ESI): calcd $[\text{M} + \text{H}]^+$ 382.1351, Found: 382.1350. Crystals suitable for X-ray analysis were grown using a vapor diffusion method in a chloroform/hexane system.

Synthesis of CAP-1 derivatives

The detailed description of the synthesis and characterization of structural analogues of CAP-1 is provided in the ESI.† Adamantane-containing isocyanates for the synthesis of adamantyl derivatives were obtained according to the literature.²¹

Acknowledgements

This work was supported by the Russian Science Foundation (RSF Grant 14-13-01030). The authors thank Dr Victor Khurstalev for carrying out X-ray analysis and Dr Viktor Burmistrov and Prof. Gennady Butov for precursors of adamantyl derivatives.

Notes and references

- (a) A. Adamo, R. L. Beingessner, M. Behnam, J. Chen, T. F. Jamison, K. F. Jensen, J.-C. M. Monbaliu, A. S. Myerson, E. M. Revalor, D. R. Snead, T. Stelzer, N. Weeranoppanant, S. Y. Wong and P. Zhang, *Science*, 2016, **352**, 61–67; (b) R. E. Martin, *Science*, 2016, **352**, 44–45.
- (a) M. A. Hamburg and F. S. Collins, *N. Engl. J. Med.*, 2010, **363**, 301–304; (b) A. D. Weston and L. Hood, *J. Proteome Res.*, 2004, **3**, 179–196; (c) P. C. Ng, S. S. Murray, S. Levy and J. C. Venter, *Nature*, 2009, **461**, 724–726; (d) L. Chin, J. N. Andersen and P. A. Futreal, *Nat. Med.*, 2011, **17**, 297–303; (e) R. L. Schilsky, *Nat. Rev. Drug Discovery*, 2010, **9**, 363–366; (f) G. S. Ginsburg and J. J. McCarthy, *Trends Biotechnol.*, 2001, **19**, 491–496.
- (a) R. A. Sheldon, *Green Chem.*, 2007, **9**, 1273–1283; (b) R. A. Sheldon, *Chem. Commun.*, 2008, 3352–3365.
- (a) P. A. Wender, V. A. Verma, T. J. Paxton and T. H. Pillow, *Acc. Chem. Res.*, 2008, **41**, 40–49; (b) P. A. Wender and B. L. Miller, *Nature*, 2009, **460**, 197–201; (c) P. A. Wender, *Nat. Prod. Rep.*, 2014, **31**, 433–440.



- 5 (a) C. H. Christensen, J. Rass-Hansen, C. C. Marsden, E. Taarning and K. Egeblad, *ChemSusChem*, 2008, **1**, 283–289; (b) P. Anastas and N. Eghbali, *Chem. Soc. Rev.*, 2010, **39**, 301–312; (c) P. J. Dunn, *Chem. Soc. Rev.*, 2012, **41**, 1452–1461; (d) J. A. Geboers, S. Van de Vyver, R. Ooms, B. Op de Beeck, P. A. Jacobs and B. F. Sels, *Catal. Sci. Technol.*, 2011, **1**, 714–726; (e) S. Van de Vyver, J. Geboers, P. A. Jacobs and B. F. Sels, *ChemCatChem*, 2011, **3**, 82–94.
- 6 (a) M. E. Zakrzewska, E. Bogel-Lukasik and R. Bogel-Lukasik, *Chem. Rev.*, 2011, **111**, 397–417; (b) M. Mascal and S. Dutta, *Green Chem.*, 2011, **13**, 3101–3102; (c) M. Mascal and E. B. Nikitin, *Green Chem.*, 2010, **12**, 370–373; (d) F. Chang, W.-H. Hsu and M. Mascal, *Sustainable Chem. Pharm.*, 2015, **1**, 14–18; (e) F. Chang, S. Dutta, J. J. Becnel, A. S. Estep and M. Mascal, *J. Agric. Food Chem.*, 2014, **62**, 476–480.
- 7 (a) K. I. Galkin, E. A. Krivodaeva, L. V. Romashov, S. S. Zalesskiy, V. V. Kachala, J. V. Burykina and V. P. Ananikov, *Angew. Chem., Int. Ed.*, 2016, **55**, 8338–8342; (b) A. S. Kashin, K. I. Galkin, E. A. Khokhlova and V. P. Ananikov, *Angew. Chem., Int. Ed.*, 2016, **55**, 2161–2166.
- 8 (a) L. O. Kallings, *J. Intern. Med.*, 2008, **263**, 218–243; (b) G. H. Friedland and R. S. Klein, *N. Engl. J. Med.*, 1987, **317**, 1125–1135; (c) C. Flexner, *Nat. Rev. Drug Discovery*, 2007, **6**, 959–966; (d) E. D. Clercq, *Nat. Rev. Drug Discovery*, 2007, **6**, 1001–1018; (e) P. Piot, M. Bartos, P. D. Ghys, N. Walker and B. Schwartlander, *Nature*, 2001, **410**, 968–973; (f) D. D. Richman, *Nature*, 2001, **410**, 995–1001; (g) V. Hughes, *Nature*, 2010, **466**, S11–S13; (h) P. M. Sharp and B. H. Hahn, *Cold Spring Harbor. Perspect. Med.*, 2011, **1**, a006841.
- 9 (a) MDG 6: 15 YEARS, 15 LESSONS OF HOPE FROM THE AIDS RESPONSE, UNAID <http://www.unaids.org>; (b) U. Weiss, *Nature*, 2001, **410**, 961–961.
- 10 (a) M. P. Girard, S. Osmanov, O. M. Assossou and M.-P. Kieny, *Vaccine*, 2011, **29**, 6191–6218; (b) L. Menendez-Arias, *Antiviral Res.*, 2010, **85**, 210–231; (c) J. N. Blankson, D. Persaud and R. F. Siliciano, *Annu. Rev. Med.*, 2002, **53**, 557–593; (d) C. C. J. Carpenter, D. A. Cooper, M. A. Fischl, J. M. Gatell, B. G. Gazzard, S. M. Hammer, M. S. Hirsch, D. M. Jacobsen, D. A. Katzenstein, J. S. G. Montaner, D. D. Richman, M. S. Saag, M. Schechter, R. T. Schooley, S. Vella, P. G. Yeni and P. A. Volberding, *J. Am. Med. Assoc.*, 2000, **283**, 381–390; (e) M. A. Fischl, D. D. Richman, M. H. Grieco, M. S. Gottlieb, P. A. Volberding, O. L. Laskin, J. M. Leedom, J. E. Groopman, D. Mildvan, R. T. Schooley, G. G. Jackson, D. T. Durack and D. King, *N. Engl. J. Med.*, 1987, **317**, 185–191.
- 11 (a) M. S. Cohen, Y. Q. Chen, M. McCauley, T. Gamble, M. C. Hosseinipour, N. Kumarasamy, J. G. Hakim, J. Kumwenda, B. Grinsztejn, J. H. S. Pilotto, S. V. Godbole, S. Mehendale, S. Chariyalertsak, B. R. Santos, K. H. Mayer, I. F. Hoffman, S. H. Eshleman, E. Piwowar-Manning, L. Wang, J. Makhema, L. A. Mills, G. de Bruyn, I. Sanne, J. Eron, J. Gallant, D. Havlir, S. Swindells, H. Ribaud, V. Elharrar, D. Burns, T. E. Taha, K. Nielsen-Saines, D. Celentano, M. Essex, T. R. Fleming and H. S. Team, *N. Engl. J. Med.*, 2011, **365**, 493–505; (b) J. K. Rockstroh, J. L. Lennox, E. DeJesus, M. S. Saag, A. Lazzarin, H. Wan, M. L. Walker, X. Xu, J. Zhao, H. Teppler, M. J. DiNubile, A. J. Rodgers, B.-Y. Nguyen, R. Leavitt, P. Sklar and S. Investigators, *Clin. Infect. Dis.*, 2011, **53**, 807–816; (c) E. M. Connor, R. S. Sperling, R. Gelber, P. Kiselev, G. Scott, M. J. Osullivan, R. Vandyke, M. Bey, W. Shearer, R. L. Jacobson, E. Jimenez, E. Oneill, B. Bazin, J. F. Delfraissy, M. Culnane, R. Coombs, M. Elkins, J. Moye, P. Stratton and J. Balsley, *N. Engl. J. Med.*, 1994, **331**, 1173–1180.
- 12 (a) A. Persidis, *Nat. Biotechnol.*, 2000, **18**, 466–467; (b) M. J. Dapp, L. Bonnac, S. E. Patterson and L. M. Mansky, *J. Virol.*, 2014, **88**, 354–363.
- 13 (a) Overview of HIV Treatments, <https://www.aids.gov/hiv-aids-basics/just-diagnosed-with-hiv-aids/treatment-options/overview-of-hiv-treatments/>; (b) Antiretroviral drugs used in the treatment of HIV infection, FDA <http://www.fda.gov/ForPatients/Illness/HIVAIDS/Treatment/ucm118915.htm>.
- 14 (a) T. Dorfman, A. Bukovsky, A. Ohagen, S. Hoglund and H. G. Gottlinger, *J. Virol.*, 1994, **68**, 8180–8187; (b) L. S. Ehrlich, B. E. Agresta and C. A. Carter, *J. Virol.*, 1992, **66**, 4874–4883; (c) B. M. Forshey, U. von Schwedler, W. I. Sundquist and C. Aiken, *J. Virol.*, 2002, **76**, 5667–5677; (d) T. Fricke, A. Brandariz-Nunez, X. Wang, A. B. Smith, III and F. Diaz-Griffero, *J. Virol.*, 2013, **87**, 10587–10597; (e) N. Goudreau, C. T. Lemke, A.-M. Faucher, C. Grand-Maitre, S. Goulet, J.-E. Lacoste, J. Rancourt, E. Malenfant, J.-F. Mercier, S. Titolo and S. W. Mason, *ACS Chem. Biol.*, 2013, **8**, 1074–1082; (f) Y. Jin, Z. Tan, M. He, B. Tian, S. Tang, I. Hewlett and M. Yang, *Bioorg. Med. Chem.*, 2010, **18**, 2135–2140; (g) B. N. Kelly, S. Kyere, I. Kinde, C. Tang, B. R. Howard, H. Robinson, W. I. Sundquist, M. F. Summers and C. P. Hill, *J. Mol. Biol.*, 2007, **373**, 355–366; (h) C. Tang, E. Loeliger, I. Kinde, S. Kyere, K. Mayo, E. Barklis, Y. N. Sun, M. J. Huang and M. F. Summers, *J. Mol. Biol.*, 2003, **327**, 1013–1020; (i) U. K. von Schwedler, K. M. Stray, J. E. Garrus and W. I. Sundquist, *J. Virol.*, 2003, **77**, 5439–5450.
- 15 (a) A. Agarwal, X. Chen, M. Deshpande and M. Summers, WO 2006053323, 2006; (b) A. Agarwal, X. Chen, M. Deshpande and M. Summers, US 20060100232, 2006; (c) A. Agarwal, X. Chen, M. Deshpande and M. Summers, EP 1809380, 2007.
- 16 (a) F. S. Asghari and H. Yoshida, *Ind. Eng. Chem. Res.*, 2006, **45**, 2163–2173; (b) J. N. Chheda, Y. Roman-Leshkov and J. A. Dumesic, *Green Chem.*, 2007, **9**, 342–350; (c) R. Karinen, K. Vilonen and M. Niemela, *ChemSusChem*, 2011, **4**, 1002–1016; (d) E. A. Khokhlova, V. V. Kachala and V. P. Ananikov, *ChemSusChem*, 2012, **5**, 783–789; (e) S. Lima, M. M. Antunes, M. Pillinger and A. A. Valente, *ChemCatChem*, 2011, **3**, 1686–1706; (f) J. Lewkowski, *ARKIVOC*, 2001, **2**, 17–54; (g) R. A. Sheldon, *J. Mol. Catal. A: Chem.*, 2016, **422**, 3–12; (h) M. Besson, P. Gallezot and C. Pinel, *Chem. Rev.*, 2014, **114**, 1827–1870; (i) J. D. Lewis,



- S. Van de Vyver, A. J. Crisci, W. R. Gunther, V. K. Michaelis, R. G. Griffin and Y. Román-Leshkov, *ChemSusChem*, 2014, 7, 2255–2265; (j) P. C. A. Bruijninx and Y. Roman-Leshkov, *Catal. Sci. Technol.*, 2014, 4, 2180–2181.
- 17 K. I. Galkin, E. A. Krivodaeva and V. P. Ananikov, *Russ. Chem. Bull.*, 2015, 2954–2957.
- 18 S. Dutta, L. Wu and M. Mascal, *Green Chem.*, 2015, 17, 3737–3739.
- 19 (a) T. Kläusli, *Green Process. Synth.*, 2014, 3, 235; (b) S. Krawielitzki and T. M. Kläusli, *Ind. Biotechnol.*, 2015, 11, 6–8; (c) E. de Jong, M. A. Dam, L. Sipos and G.-J. M. Gruter, in *Biobased Monomers, Polymers, and Materials*, ed. P. B. Smith and R. A. Gross, American Chemical Society, Washington, DC, 2012, pp. 1–13; (d) A. Scott, *Chem. Eng. News*, 2016, 94, 15.
- 20 A. Bredihhin, U. Mäeorg and L. Vares, *Carbohydr. Res.*, 2013, 375, 63–67.
- 21 (a) V. Burmistrov, C. Morisseau, K. S. S. Lee, D. S. Shihadih, T. R. Harris, G. M. Butov and B. D. Hammock, *Bioorg. Med. Chem. Lett.*, 2014, 24, 2193–2197; (b) V. Burmistrov, C. Morisseau, D. Danilov, T. R. Harris, I. Dalinger, I. Vatsadze, T. Shkineva, G. M. Butov and B. D. Hammock, *Bioorg. Med. Chem. Lett.*, 2015, 25, 5514–5519.

