Ravynic acid, an antibiotic polynene tetramic acid from *Penicillium* sp. elucidated through synthesis†

J. D. Myrtle, A. M. Beekman‡ and R. A. Barrow*

A new antibiotic natural product, ravynic acid, has been isolated from a *Penicillium* sp. of fungus, collected from Ravensbourne National Park. The 3-acylpolyene tetramic acid structure was definitively elucidated via synthesis. Highlights of the synthetic method include the heat induced formation of the 3-acylphosphorane tetramic acid and a selective Wittig cross-coupling to efficiently prepare the natural compounds carbon skeleton. The natural compound was shown to inhibit the growth of *Staphylococcus aureus* down to concentrations of 2.5 µg mL⁻¹.

Introduction

The search for new antibiotic compounds that possess novel modes of action and inhibit new targets is an on-going challenge in the struggle against infectious disease. One class of natural products that has attracted much attention are those that bear the tetramic acid moiety.¹ Some years ago we reported the isolation and elucidation of a tetramic acid containing natural product, ravenic acid, 1 (tetramic acid moiety highlighted in red, Fig. 1), from a *Penicillium* sp. collected from Ravensbourne National Park in south-east Queensland, Australia.² Ravenic acid possessed moderate antibiotic activity against a clinical strain of methicillin resistant *Staphylococcus aureus* (MRSA). Re-examination of the fungal extract has revealed a very minor co-metabolite isolated from the *Penicillium* sp., that was shown to possess significantly greater antibiotic activity than ravenic acid, but was present in microgram quantities from large scale fermentation, effectively precluding study by NMR. However, a mass spectrum was recorded, and the CID ESI-MS fragmentation pattern displayed many similarities to 1, but with a m/z ratio of the deprotonated molecule (m/z 256, −ve ESI) along with those of some daughter ions two units lower. Given ravenic acid’s structure, and following careful analysis of the mass spectrum we reasoned the most probable modification accounting for a loss of two mass units represents an oxidation of a double bond to afford a triple bond in the polynene side chain. As such, tetramic acids 2 and 3 were identified as synthetic targets to allow the definitive elucidation of the co-metabolite of ravenic acid (Fig. 1).

Results and discussion

Pathways presented previously by Ley³ and Moloney⁴ offered promise for the preparation of 2 and 3, but had been shown to be unsuccessful with unsubstituted amides and unsaturated side chains. During our studies of ravenic acid and its potential co-metabolites, Schobert and co-workers published an elegant synthesis of ravenic acid, representing a new synthetic procedure to prepare 3-acyl tetramic acids in a highly efficient

---

†Electronic supplementary information (ESI) available: ¹H and ¹³C NMR spectra of new compounds. See DOI: 10.1039/c6ob00938g
‡Current address: School of Pharmacy, University of East Anglia, Norwich Research Park, Norwich, Norfolk, UK NR4 7TJ.
manner. Thus, we modelled a retrosynthetic pathway on the procedure outlined by Schobert (Fig. 2). Late stage Wittig olefination would allow a divergent strategy to the preparation of 2 and 3 from the common intermediate 4 and aldehydes 5 and 6. Compound 4 could be prepared by the thermal coupling of tetaenic acid 7 and phosphorane 8. The use of the Boc protecting group is notable, as it had been employed successfully by Schobert, in contrast to the di-methoxybenzyl protecting group which had been the downfall of our previous attempts to synthesize ravenic acid. Despite numerous attempts employing a variety of deprotection protocols starting material consistently decomposed.

In a forward manner, tetaenic acid 7 was prepared in one step from N-Boc-glycine (9) by coupling with Meldrum’s acid mediated by EDC and DMAP, followed by heat induced rearrangement, in 83% yield (Scheme 1). Consistent with previously reported data, 7 exists as a mixture of keto and enol tautomers in CDCl₃ in the approximate ratio of 1:1, while in deuterated methanol, the enol form is exclusively observed.

Phosphorane 8 was formed by treatment of phosphorane 10 with sodium hexamethyldisilazide. In spite of the structural similarity of 8 with ketenes, it is relatively stable, not displaying a tendency to dimerise. Phosphorane 8 and tetaenic acid 7 were coupled under thermal conditions to form key intermediate 4. Heat facilitates nucleophilic addition of the enol tautomer of 7 on 8, with subsequent proton transfer and tautomerism leading to the formation of 4. In CDCl₃, tetaenic acid 4 exists in two forms, which have been assigned as the phosphorane and the tautomeric/resonance betaine form.

Preparation of the requisite aldehydes 5 and 6 was achieved in a rapid and efficient manner (Scheme 2). Propargyl alcohol was brominated by treatment with potassium hydroxide and bromine to form 11, and subsequent copper-mediated coupling with propyne led directly to alcohol 12. Stereoselective reduction of the triple bond proximal to the alcohol with LiAlH₄ afforded an alcohol, where the stereochemistry of the newly created alkene was deduced from the coupling constant (J = 15.6 Hz) between the olefinic protons at δ 5.68 and 6.15, indicative of the trans relationship of substituents. Oxidation employing Dess–Martin periodinane yielded aldehyde 13 which was employed directly in the following step. Sonogashira coupling of E-1-bromoprop-1-ene with propargyl alcohol successfully afforded alcohol 14, with subsequent oxidation using DMP providing aldehyde 15, with a yield of 77% over the 2 steps. Isomerization was not observed over either of the two steps, with the stereochemistry of alcohol 14 determined through analysis of the olefinic coupling constants (J = 15.8 Hz). Examination of conditions for the Horner–Wadsworth–Emmons olefination between triethyl-2-phosphonopropionate and aldehydes 13 and 15 found that sodium hydride as the base increased the yield and selectively giving the desired E isomer. The proton NMR spectra and GCMS data indicated that one stereoisomer was formed preferentially (>95%), with literature comparisons, and the HWE reaction’s known stereochemical outcomes, supporting the assignment of the E configuration. Attempts were made to proceed directly to aldehydes 5 and 6 in one step using DIBAL-H as the reducing agent. Treatment of the resulting esters 16–17 with one equivalent of DIBAL-H resulted in a 1:1 mixture of the alcohol and the starting ester, indicating that the intermediate aldehyde is reduced more rapidly than the ester. Thus three equivalents of DIBAL-H were used to ensure complete conversion of the esters to conjugated alcohols, and the alcohols subsequently oxidised to aldehydes 5 and 6 using DMP. With the synthesis
of the target aldehydes complete, attention turned to completion of the final steps in the synthesis of the ravenic acid analogues.

The common tetramic acid phosphorane intermediate 4 was coupled with aldehydes 5 and 6 via the Wittig olefination to give Boc-protected ravenic acid analogues 18 and 19 in yields of 52% and 62%, respectively (Scheme 3). With either silica or alumina column chromatography resulting in decomposition, size exclusion chromatography (Sephadex LH-20) was employed to purify the polyene tetramic acids. In each case, the product with the E stereochemistry about the newly created olefinic double bond was formed as deduced by analysis of the olefinic coupling constants ($^{3}J = 15.6$ Hz) in the $^{1}H$ NMR spectrum. The concluding step involved treatment with TFA in dichloromethane resulting in the cleavage of the Boc protecting group, providing the ravenic acid analogues 2 and 3.

Finally, comparison of the mass spectral data as well as co-injection on a HPLC column allowed the assignment of the co-metabolite as structure 3, named here as ravynic acid. The synthesis of this newly discovered natural product was performed in a convergent manner with the longest linear sequence of steps counting nine. Ravenic acid had been shown to inhibit the growth of a methicillin-resistant Staphylococcus aureus strain down to 25 µg mL$^{-1}$. Ravynic acid has been shown to inhibit the growth of the same strain down to approximately 2.5 µg mL$^{-1}$. Further biological testing will allow for the assessment of the spectrum of activity and determination of the mode of action of this antibiotic.

Conclusions

In conclusion, we have isolated and unambiguously identified a new antibacterial tetramic acid natural product from a Penicillium sp. of fungus. Ravynic acid is a 3-acyltetramic acid, which has been synthesized from Boc-glycine and propargyl alcohol, to allow definitive elucidation of the structure. Notable features of the synthesis include the thermally induced formation of the 3-acylphosphorane tetramic acid and a selective Wittig cross-coupling to efficiently prepare the natural compounds carbon skeleton.

Experimental

General information

$^{1}H$, $^{13}C$ NMR spectra were recorded at 298 K, at 300 MHz, 400 MHz or 500 MHz and 75 MHz, 100 MHz or 125 MHz respectively, on an Inova 300, Varian MR-300, Varian MR-400 or Inova 500 instruments. Chemical shifts are reported in ppm ($\delta$). NMR experiments were run in deuterated chloroform (CDCl$_3$), or methanol (CD$_3$OD) as indicated; $^{1}H$ NMR spectra are referenced to the resonance from residual CHCl$_3$ at 7.26 ppm, or CHD$_2$OD at 3.31 ppm, $^{13}C$ NMR spectra are referenced to the central peak in the signal from CDCl$_3$ at 77.0 ppm, or CD$_3$OD at 49.0 ppm. The appearance and multiplicities of $^{1}H$ resonances are expressed by the abbreviations: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) and
combinations thereof for more highly coupled systems. $^{13}$C NMR were run as proton decoupled spectra. $^1$H signals and $^{13}$C signals, where appropriate, are described by chemical shift $\delta$ (multiplicity, $J$/Hz, integration, assignment). Where the spectral characteristics ($^1$H and $^{13}$C NMR) agree with published data, a literature reference is cited. Where no literature is available, including known but insufficiently characterised compounds, data ($^1$H NMR, $^{13}$C NMR, IR, MS, HRMS) are presented. Assignments were supported by 2D NMR analysis including HSQC, HMBC and NOESY data and by literature precedence. EI-MS and HREI-MS were recorded on a VG autospect mass spectrometer, operating at 70 eV. ESI-MS and HREI-MS were recorded on a Bruker Apex 3. Positive ionisation was detected unless otherwise indicated. Mass/charge ratios ($m$/z) are reported and relative abundance of the ions is as percentage of base peak intensity. IR spectra were recorded on a Bruker Alpha-P as neat solid or film. Diagnostic peaks are presented in wavenumbers (cm$^{-1}$). Peak strengths are expressed by the abbreviations: s (strong), m (medium), w (weak) and b (broad). Melting points were determined on a Reichert Thermogalen hot stage microscope and are uncorrected. Flash chromatography was performed under pressure using silica gel (230–400 mesh Scharlau 60, 40–60 micron). Analytical thin layer chromatography (TLC) was performed on Merck silica gel 60 F254 aluminium backed plates. Plates were viewed under UV light (254 nm) and/or by developing with ceric phosphomolybdate (C-4), 63.0 (C-1), 4.2 (C-6). IR:

$\nu$ (cm$^{-1}$) 3301 (s, br), 1445 (s), 1347 (s), 1021 (s), 625 (s). HREI-MS: calc 94.0419 (C6H6O) found 94.0419.

(E)-Hex-2-en-4-yn-1-ol. A solution of hexa-2,4-diyn-1-ol (300 mg, 3.19 mmol, 1.0 eq.) in THF (6 mL) was added dropwise to a suspension of LiAlH4 (132 mg, 3.30 mmol, 1.03 eq.) in THF (6 mL) at 0 °C. After 4 h the mixture was cooled to 0 °C and EtOAc (4 mL) was added followed by sat. NH4Cl (4 mL) carefully added. The mixture was filtered through celite and the residual solids washed with EtOAc. The filtrate was transferred to a separating funnel with additional sat. NH4Cl (50 mL). The mixture was extracted with EtOAc (3 × 45 mL) and the combined organic extracts dried over MgSO4 and concentrated in vacuo. Purification via flash column chromatography on silica using 60% Et2O:petroleum ether as eluent afforded the title compound as a clear oil (198 mg, 65%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 6.15 (dt, $J = 15.6, 5.6$ Hz, 1H, H-2), 5.68 (dd, $J = 15.6, 2.4$ Hz, 1H, H-3), 4.16 (dd, $J = 6.0, 5.6$ Hz, 2H, H-1), 1.94 (t, $J = 6.0$ Hz, 1H, OH). $^{13}$C (75 MHz, CDCl$_3$) $\delta$ 140.2 (C-2), 111.2 (C-3), 86.8 (C-5), 77.4 (C-4), 63.0 (C-1), 4.2 (C-6). IR: $\nu$ (cm$^{-1}$) 3317 (br s), 2917 (w), 1432 (m), 1091 (s), 1008 (s), 951 (s). EI-MS $m$/z (%) 95 (28, M$^+$), 84 (17), 81 (100), 67 (29), 53 (30). HREI-MS: calc 95.0497 (C$_6$H$_6$O) found 95.0497.

(E)-Hex-2-en-4-yn-1-ol. Freshly prepared Dess–Martin periodinane (920 mg, 2.16 mmol, 1.05 eq.) was added to a stirred solution of (E)-hexa-2,4-diyn-1-ol (198 mg, 2.06 mmol, 1.0 eq.) in CH$_2$Cl$_2$ (6 mL) at room temperature. After 90 min the mixture was diluted with Et$_2$O (5 mL) and sat. NaHCO$_3$ (10 mL), and sodium thiosulfate (1.0 g) added. After 30 min the mixture was filtered through celite and the aqueous layer re-extracted with Et$_2$O (15 mL). The combined organic layers were washed with sat. NaHCO$_3$ (10 mL), dried over MgSO$_4$ and concentrated in vacuo. The crude product obtained was a volatile yellow oil (197 mg, crude yield 99%) that was used directly in the next step without further purification. $^1$H NMR (300 MHz, CDCl$_3$) $\delta$ 9.53 (d, $J = 7.5$ Hz, 1H, H-1), 6.57 (dq, $J = 15.7, 2.4$ Hz, 1H, H-3), 6.38 (dd, $J = 15.7, 7.5$ Hz, 1H, H-2), 2.09 (d, $J = 2.4$ Hz, 3H, H-6). $^1$H NMR (300 MHz, CDCl$_3$) $\delta$ 6.429 (s, 2H, H-1), 1.94 (br s, 1H, OH).

Hexa-2,4-diyn-1-ol (12). Proprylene (~5 mL) was condensed into a RB flask using a dry ice condenser. The flask was slowly warmed by raising above the dry ice/acetone bath to allow the slow transfer of the propylene via a cannula to RB flask charged with n-butylamine/H$_2$O (15 mL, 30% butyramine) and CuCl (35 mg, cat). After complete addition, 3-bromoprop-2-yn-1-ol (11) (2.41 g, 17.8 mmol, 1.0 eq.) was added dropwise and as the mixture turned green a few drops of hydroxylamine solution added from a dropping funnel. The mixture was stirred for 25 min and then extracted with Et$_2$O (3 × 20 mL). The combined organic extracts were washed with brine (20 mL), dried over Na$_2$SO$_4$ and concentrated in vacuo. Purification by column chromatography using 50% Et$_2$O:petroleum ether yielded the title compound as an unstable white solid (644 mg, 38%). $^1$H NMR (300 MHz, CDCl$_3$) $\delta$ 4.31 (s, 2H, H-1), 1.94 (s, 3H, H-6), 1.10 (s, 2H, H-1), 6.57 (dq, $J = 15.7, 2.4$ Hz, 1H, H-3), 6.38 (dd, $J = 15.7, 7.5$ Hz, 1H, H-2), 2.09 (d, $J = 2.4$ Hz, 3H, H-6).
cation via flash column chromatography on silica using 40% EtO\(_2\)P: petroleum ether as eluent gave the desired product as a pale yellow oil (1.78 g, 79%). \(^1\)H NMR (300 MHz, CDCl\(_3\)) \(\delta\) 6.16 (dq, \(J = 15.8, 6.9\) Hz, 1H, H-5), 5.50 (dt, \(J = 15.8, 2.1\) Hz, 1H, H-4), 4.36 (dd, \(J = 2.1, 2H, H-1\)), 1.78 (dd, \(J = 6.9, 3H, H-6\)). \(^13\)C (75 MHz, CDCl\(_3\)) \(\delta\) 140.1 (C-5), 110.0 (C-4), 85.4, 84.0, 51.0 (C-1), 18.4 (C-6). IR: \(\nu\) (cm\(^{-1}\)) 3345 (br s), 2380 (m), 1443 (m), 1163 (m), 1013 (s), 953 (s). EI-MS m/z (%) 96 (85, \(M^+\)), 81 (100), 77 (35), 67 (31), 53 (45), 39 (52). HREI-MS: calc 960575 (C\(_2\)H\(_6\)O\(_2\)) found 960578.

**Page 2**

**Triethyl-2-phosphonopropanoate.** Ethyl-2-bromopropanoate (3.59 mL, 27.6 mmol) and triethylphosphite (4.73 mL, 27.6 mmol) were stirred at reflux for 36 h. Purification by Kugelrohr distillation gave phosphonate as a clear oil (6.19 g, 94%). \(^1\)H NMR (300 MHz, CDCl\(_3\)) \(\delta\) 4.10–4.30 (m, 6H, 3 × OCH\(_2\)CH\(_3\)), 3.02 (dd, \(J_{PH} = 23.3\) Hz, \(J_{HH} = 7.2\) Hz, 1H, H-2), 1.44 (dd, \(J_{PH} = 18.2\) Hz, \(J_{HH} = 7.2\) Hz, 3H, H-3), 1.25–1.28 (m, 9H, 3 × OCH\(_2\)CH\(_3\)).

**(2E,4E)-Ethyl-2-methylocta-2,4-dien-6-ynoate (16).** Freshly prepared triethyl-2-phosphonopropanoate (0.64 mL, 2.80 mmol, 1.4 eq.) was added to a stirred suspension of NaH (113 mg, 2.80 mmol, 1.4 eq., 60% in mineral oil) in THF (5 mL) at 0 °C. After 1 h a solution of aldehyde 13 (188 mg, 2.00 mmol, 1.0 eq.) in THF (2 mL) was added and the mixture allowed to warm to room temp. and stirred for 16 h. The reaction was quenched with sat. NH\(_4\)Cl (15 mL), transferred to a separating funnel and extracted with EtO\(_2\) (3 × 15 mL). The combined ethereal layers were washed with water (2 × 15 mL) followed by brine (2 × 15 mL), dried over MgSO\(_4\) and concentrated in vacuo. The crude product was purified via flash column chromatography on silica using 7% EtO\(_2\): petroleum ether as eluent to yield ester 16 (35 mg, 73%) from ester 15 (40 mg, crude yield 88%), from ester 17 (73 mg, 0.41 mmol).

**Page 3**

\(^1\)H NMR (300 MHz, CDCl\(_3\)) \(\delta\) 7.13 (\(d\), \(J = 12.0\) Hz, 1H, H-3), 6.76 (dd, \(J = 15.3, 12.0\) Hz, 1H, H-4), 5.86 (dq, \(J = 15.3, 2.1\) Hz, 1H, H-5), 4.18 (q, \(J = 7.2\) Hz, 2H, OCH\(_2\)CH\(_3\)), 1.99 (dd, \(J = 2.1\) Hz, 3H, H-8), 1.92 (s, 3H, C-2 Me), 1.27 (t, \(J = 7.2\) Hz, 3H, OCH\(_2\)CH\(_3\)). \(^13\)C (75 MHz, CDCl\(_3\)) \(\delta\) 167.9 (C-1), 136.9, 135.4, 128.5 (C-2), 118.4 (C-5), 92.3 (C-7), 79.1 (C-6), 60.6 (OCH\(_2\)CH\(_3\)), 14.2 (OCH\(_2\)CH\(_3\)), 12.7 (C-2 Me), 4.6 (C-8). IR: \(\nu\) (cm\(^{-1}\)) 2980 (w), 1703 (s), 1368 (m), 1283 (s), 1227 (s), 1101 (m), 959 (m). EI-MS m/z (%) 178 (98, \(M^+\)), 163 (96), 149 (93), 135 (100), 121 (95). HREI-MS: calc 1780994 (C\(_{11}\)H\(_{14}\)O\(_2\)) found 1780992.

**Page 4**

**Page 5**
(2E,6E)-2-methylocta-2,6-dien-4-ynol (49 mg, 0.36 mmol). 1H NMR (300 MHz, CDCl3) δ 9.46 (s, 1H, H-1H), 6.46 (s, 1H, H-3), 6.35 (d, J = 15.6, 6.9 Hz, 1H, H-7), 5.76 (d, J = 15.6, 1.8 Hz, 1H, H-6), 1.94 (s, 3H, C-2 Me), 1.87 (d, J = 6.9 Hz, 3H, H-8).

tert-Butyl-2,4-dioxopyrrolidine-1-carboxylate (7). EDC-HCl (805 mg, 4.20 mmol, 1.5 eq.) was added to a mixture of N-Boc-glycine (500 mg, 2.80 mmol, 1.0 eq.), Meldrum’s acid (489 mg, 4.20 mmol, 1.5 eq.) and DMAP (513 mg, 4.20 mmol, 1.5 eq.) in CH2Cl2 (30 mL) at 0 °C. The mixture was allowed to warm to room temperature over 15 min, then stirred for an additional 4 h. After diluting with EtOAc (100 mL) the organic layer was washed with brine (2 × 50 mL), 5% citric acid (2 × 50 mL), and brine (50 mL), dried over Na2SO4 and concentrated under reduced pressure. The crude oil obtained was dissolved in EtOAc (40 mL) and heated at reflux for 1 h. The solvent was removed under reduced pressure (toluene added as a co-solvent to aid in the removal of EtOAc) to afford the title compound as a white solid (164 mg, 83%).

Keto : enol = 53 : 47 in CDCl3, only enol form in CD3OD. Note: H-3 and C-3 peaks not observed in spectra when run in CD3OD. 1H NMR (300 MHz, CDCl3) δ keto: 4.24 (s, 2H, H-5), 3.25 (s, 2H, H-3), 1.54 (s, 9H, C(CH3)3), 1H NMR (300 MHz, CD3OD) δ keto: 4.23 (s, 2H, H-5), 1.53 (s, 9H, C(CH3)3). 13C (100 MHz, CDCl3) δ 143.6 (s, C-4), 132.3 (d, JPC = 10.3 Hz), 129.3 (d, JPC = 2.4 Hz), 123.9 (s, C-6), 115.2 (s, C-2), 110.5 (d, JPC = 12.2 Hz), 104.7 (C-3), 83.0 (C(CH3)3), 53.6 (C-5), 28.1 (C(CH3)3). 13C (100 MHz, CD3OD) δ 145.6 (d, JPC = 12.9 Hz), 131.5 (s, C(CH3)3), 129.6 (s, C-6), 115.2 (s, C-2), 110.5 (d, JPC = 12.2 Hz), 104.7 (C-3), 83.0 (C(CH3)3), 53.6 (C-5), 28.1 (C(CH3)3). IR: ν (cm⁻¹) 2978 (m), 1755 (s), 1597 (s). 1H NMR (500 MHz, CDCl3): 2 : 1 mixture of phosphoranes (A) and (B). In CD3OD: 4 : 1 mixture of tautomers major (a) and minor (b). Form A: 7.45–7.75 (m, 15H, PPh3), 5.10 (d, J = 12.0 Hz, 2H, CH2P), 3.78 (s, 2H, H-5), 1.48 (s, 9H, C(CH3)3). 13C (100 MHz, CDCl3) δ 192.2 (C-4), 182.3 (C-6), 171.2 (C-2), 149.9 (NCO2Bu), 134.33, 134.29, 133.8 (d, J = 10.3 Hz), 133.3 (d, J = 2.6 Hz), 132.9 (d, J = 10.5 Hz), 132.3 (d, J = 2.4 Hz), 131.6 (d, J = 10.1 Hz), 129.6 (d, J = 12.8 Hz), 129.3 (d, J = 12.7 Hz), 128.5 (d, J = 12.2 Hz), 123.6 (d, J = 92.0 Hz), 119.6 (d, J = 88.0 Hz), 104.7 (C-3), 83.0 (C(CH3)3), 53.6 (C-5), 28.4 (C(CH3)3). IR: ν (cm⁻¹) 1768 (s), 1720 (m), 1620 (s), 1555 (s), 1436 (s), 1327 (s), 1153 (s), 1102 (s), 844 (s), 689 (s). ESI-MS m/z (%): 501 (14, M⁺), 401 (65), 301 (90), 277 (100), 183 (61), 57 (74). HREI-MS: calc 501.1705 (C29H34NO3P) found 501.1710.

(Z)-tert-Butyl-3-(1-hydroxy-2-(triphenylphosphoranylidene)-ethylidene)-2,4-dioxopyrrolidine-1-carboxylate (4). A solution of (Triphenylphosphoranylidene)ketene (8) (1.82 g, 9.13 mmol, 1.0 eq.) in THF (30 mL) was added dropwise over 10 min to a refluxing solution of tetratomic acid 7 (2.76 g, 9.13 mmol, 1.0 eq.) in THF (90 mL) under argon. After refluxing overnight the mixture was allowed to cool to room temperature and the solvent removed in vacuo. The crude product was purified by flash column chromatography using CH2Cl2 → 10% MeOH: CH2Cl2 as eluent to yield the desired product as a white solid (3.80 g, 84%). In CDCl3: 2:1 mixture of phosphorane (A) and betaine (B). In CD3OD: 4:1 mixture of tautomers major (a) and minor (b). Note: peaks associated with C-7 and H-7 not observed in CD3OD. 1H NMR (500 MHz, CDCl3): δ Form A: 7.45–7.75 (m, 15H, PPh3), 5.44 (d, J = 19.5 Hz, 1H, P(CH3)), 3.94 (s, 2H, H-5), 1.49 (s, 9H, C(CH3)3), 1H NMR (300 MHz, CDCl3) δ Form A: 7.45–7.75 (m, 15H, PPh3), 5.10 (d, J = 12.0 Hz, 2H, CH2P), 3.78 (s, 2H, H-5), 1.48 (s, 9H, C(CH3)3). 13C (100 MHz, CDCl3) δ 192.2 (C-4), 182.3 (C-6), 171.2 (C-2), 149.9 (NCO2Bu), 134.33, 134.29, 133.8 (d, J = 10.3 Hz), 133.3 (d, J = 2.6 Hz), 132.9 (d, J = 10.5 Hz), 132.3 (d, J = 2.4 Hz), 131.6 (d, J = 10.1 Hz), 129.6 (d, J = 12.8 Hz), 129.3 (d, J = 12.7 Hz), 128.5 (d, J = 12.2 Hz), 123.6 (d, J = 92.0 Hz), 119.6 (d, J = 88.0 Hz), 104.7 (C-3), 83.0 (C(CH3)3), 53.6 (C-5), 28.4 (C(CH3)3). IR: ν (cm⁻¹) 1768 (s), 1720 (m), 1620 (s), 1555 (s), 1436 (s), 1327 (s), 1153 (s), 1102 (s), 844 (s), 689 (s). ESI-MS m/z (%): 501 (14, M⁺), 401 (65), 301 (90), 277 (100), 183 (61), 57 (74). HREI-MS: calc 501.1705 (C29H34NO3P) found 501.1710.
crude product was purified by column chromatography on Sephadex using 50% CH2Cl2:MeOH to yield the target compound as an orange oil (74 mg, 52%). Tautomeric ratio Z:E = ∼:2:1 in CDCl3. For 1H NMR spectrum the major tautomer is assigned, for the 13C NMR spectrum, both tautomers are assigned. 1H NMR (400 MHz, CDCl3) δ 7.79 (d, J = 15.6 Hz, 1H, H-8), 7.31 (d, J = 15.6 Hz, 1H, H-7), 6.29 (dq, J = 15.6, 6.8 Hz, 1H, H-14), 6.10 (s, 1H, H-10), 5.74 (dq, J = 15.6, 2.0 Hz, 1H, H-13), 4.06 (s, 2H, H-5), 3.11 (s, 3H, C-9 Me), 1.86 (dd, J = 6.8, 2.0 Hz, 3H, 3H-15), 1.56 (s, 9H, C(CH3)3), 13C (100 MHz, CDCl3) δ 197.7 (C-4 minor), 189.3 (C-4 major), 174.8 (C-6 minor), 176.2 (C-6 major), 173.6 (C-2 major), 164.8 (C-2 minor), 149.8 (OCOCBu minor), 149.4 (OCOCBu major), 148.7 (C-8), 144.8 (C-14), 142.1 (C-14), 124.2 (C-10), 118.6 (C-7 major), 118.0 (C-7 minor), 110.9 (C-13), 105.3 (C-3 minor), 102.9 (C-12), 101.5 (C-3 major), 86.7 (C-11), 84.0 (C(CH3)3 major), 83.4 (C(CH3)3 minor), 54.7 (C-5 major), 52.3 (C-5 minor), 28.0 (C(CH3)3), 19.0 (C-15), 14.8 (C-9 Me). IR: ν (cm⁻¹) 1743 (m), 1613 (s), 1551 (m), 1343 (s), 1310 (s), 1156 (s). ESI-MS m/z (%) 357 (3, M⁺), 301 (7), 257 (19), 2257 (43), 126 (54), 91 (100). HREI-MS: calc 357.1576 (C20H23NO5) found 357.1566.

(2Z,4E,6E)-1-Hydroxy-4-methyldeca-2,4,8-trien-6-yn-1-ylidene)pyrrolidine-2,4-dione (2). Prepared in a similar manner to tetratomic acid 3, from tetratomic acid 4 (251 mg, 0.50 mmol, 1.0 eq.) and aldehyde 5 (50 mL of malt extract broth (MEB). Each flask was inoculated with two 1 cm² blocks of fungal culture of the malt extract agar (PDA) plate (an initially white culture developed which turned green upon sporulation). Yellow globules developed after 4 days. A sterile inoculating loop was used to transfer fungal material from the radius of the loop to a new PDA plate which grew to give a pure culture of the fungus sp. MINAP-9902, which was identified as a Penicillium species.

Culturing of MINAP-9902

The fungus was grown on 110 mm malt extract agar plates at 25 °C for 4 days. Eight 250 mL conical flasks were prepared, four with 50 mL potato dextrose broth (PDB) and four with 50 mL of malt extract broth (MEB). Each flask was inoculated with two 1 cm² blocks of fungal culture of the malt extract plate. These flasks were maintained at 25 °C, and agitated on an orbital shaker at 120 rpm for 4, 6, 8 and 10 days (two conical flasks, one each of PDB and MEB, were removed periodically every two days from the four day mark). For each of the conical flasks, the mycelia were mechanically separated from the broth by filtration and the mycelia were homogenised in a solution of CH2Cl2:EtOH (1:4). The crude extracts were concentrated, then suspended in 50 mL of deionised water, and extracted successively with hexane (2×50 mL), CH2Cl2 (2×50 mL) and EtOAc (2×50 mL). Previous studies indicated that the bioactive metabolites of interest were to be found in...
the CH₂Cl₂ layer. Thus, the CH₂Cl₂ layers were dried over MgSO₄, filtered, and concentrated in vacuo to provide samples that were subjected to HPLC.

**HPLC**

Analytical HPLC analyses were conducted on the eight fungal extracts prepared. HPLC conditions:

Analytical and semi-preparative HPLC was performed using an Agilent 1100 system, utilising a Phenomenex LUNA 3 μ C-18 (2) 250 × 4.60 mm column (analytical) or a Phenomenex LUNA 5 μ C-18(2) 250 × 10.00 mm column (semi-preparative) and an Agilent 1100 series diode array detector for elution detection. Elution was carried out using 0.1% v/v TFA in Milli-Q water and pre-filtered (0.45 μm nylon membrane filter) MeCN, with detection at 254 nm and 300 nm or 400 nm. Elution was carried out with 60% MeCN/H₂O (0.1% TFA), increasing to 80% MeCN/H₂O (0.1% TFA) over 30 min, with flow rates of 1.5 mL min⁻¹ (analytical) or 2.5 mL min⁻¹ (semi-preparative). In the first stage of HPLC work, analytical HPLC was carried out on synthetic analogues 2 and 3 to determine the retention times of these compounds. Subsequent analytical runs were conducted on the eight fungal extracts prepared from MINAP-9902, to probe for retention times similar to those for the prepared synthetic analogues. Our analysis of these runs revealed that the fungal extracts that were cultured for 6 days (both PDB and MEB) contained the metabolite of interest in a ratio that was yet to be overwhelmed by a suite of compounds produced by this fungus upon prolonged culture. This was based on the presence of peaks with similar retention times to our synthetic analogues and the comparison of the UV spectra of these peaks to our synthetic analogues. In particular, we decided to focus on the PDB broth that was cultured for 6 days (sample name: PDB6), as the signals of interest appeared stronger in the PDB HPLC trace, in comparison to that of the MEB HPLC trace. In PDB6, a strong peak observed at 14.4 min (analytical HPLC) was assigned to be ravenic acid. Peaks with retention time in the vicinity of analogue 3 (16.0 min) were also present (15.9 min and 17.6 min), but peaks with retention times close to that of 2 (13.2 min) were notably absent. Co-injection of 3 with PDB6 resulted in peak enhancement, strongly indicative of matching structures. At this stage semi-preparative HPLC was performed on PDB6 to isolate the metabolite of interest. Small quantities of the metabolite were isolated and mass spectral data recorded: EI-MS m/z (%) 257 (93, M⁺), 242 (10), 129 (48), 126 (46), 84 (75), 69 (76), 57 (100). HREI-MS: calc 257.1052 (C₁₅H₁₅NO₃) found 257.1052. The EI-MS of the metabolite isolated from PDB6 closely resembled the EI-MS of 3, with matching molecular ions (m/z 257), as well as a number of daughter ions (m/z 126, 84, 69) observed in both spectra. Re-injection of the metabolite of interest to the HPLC returned a peak with a retention time matching that of synthetic analogue 3 (see ESI†), confirming that no decomposition or structural modification had taken place. The matching retention times and mass spectral data of the unknown metabolite with analogue 3, provide strong evidence that the structure of the metabolite corresponds to that of 3.

**Determination of the sensitivity of compounds against *Staphylococcus aureus***

Kirby Bauer bioassays were performed routinely to identify fractions containing an active compound(s). This was achieved by inoculating an 83 mm nutrient agar plate with 200 μL suspension of *Staphylococcus aureus* cells in sterilised deionised water, and spreading this suspension over the agar surface with a flame sterilised glass spreader. The plate was dried for 10 min in a laminar flow hood, while 5 mm paper disks were soaked in a solution of the fractions or compounds of interest, dissolved in a volatile solvent. The disks were allowed to dry and then lightly pressed onto the agar surface. The plates were then incubated at 37 °C for 16 hours and examined for growth inhibition around the disks. The sensitivity of a clinical strain of *Staphylococcus aureus* was assessed by a turbidity assay OD600 (optical density at λ 600 nm) as described in a previous publication.¹⁷

**Acknowledgements**

JDM acknowledges the ANU for an Australian Postgraduate Award. This research was undertaken with the support of the ANU and an NHMRC project grant (1028092).

**Notes and references**


