



Synthetic approaches towards alkaloids bearing α -tertiary amines

Anastasia Hager,[†] Nina Vrielink,[†] Dominik Hager, Julien Lefranc and Dirk Trauner*

Covering: up to August 2015

Received 15th August 2015

DOI: 10.1039/c5np00096c

www.rsc.org/npr

Alkaloids account for some of the most beautiful and biologically active natural products. Although they are usually classified along biosynthetic criteria, they can also be categorized according to certain structural motifs. Amongst these, the α -tertiary amine (ATA), *i.e.* a tetrasubstituted carbon atom surrounded by three carbons and one nitrogen, is particularly interesting. A limited number of methods have been described to access this functional group and fewer still are commonly used in synthesis. Herein, we review some approaches to asymmetrically access ATAs and provide an overview of alkaloid total syntheses where those have been employed.

1 Introduction

- 2 Methods used for the installation of α -tertiary amines
- 2.1 A C,C-bond is formed in the step that generates the ATA
- 2.2 A C,N-bond is formed in the step that generates the ATA
- 3 Homotropane alkaloids
- 4 Histrionicotoxins
- 5 *Lycopodium* alkaloids
- 6 Hasubanan alkaloids
- 7 *Stemona* alkaloids
- 8 Indole alkaloids
- 9 Cephalotaxines
- 10 *Erythrina* alkaloids
- 11 Indolizidine and quinolizidine alkaloids
- 12 Lactacycline and salinosporamide
- 13 Manzacidins
- 14 Tetrodotoxin
- 15 Miscellaneous alkaloids
- 16 Conclusions
- 17 Acknowledgements
- 18 Notes and references

1 Introduction

Alkaloids have played an important role in the development of synthetic organic chemistry, pharmacology and medicine. Once considered to be metabolic waste products, they are now known to benefit their producers in various ways, *e.g.* as antimicrobials,

antifeedants or as mediators of ecologically beneficial interactions.¹ Though a limited number of amino acids are involved in their biosynthesis, alkaloids exhibit enormous structural variability, which is often increased through the incorporation of terpenoid and polyketide components and late-stage oxidative transformations.² Reflecting their structural diversity and relatively weak basicity, alkaloids interact with a large variety of biological targets and have found many uses in human medicine.^{3,4} In addition, they have provided inspiration for countless synthetic drugs that borrow structural motifs from their natural counterparts.

The α -tertiary amine (ATA) stands out among the structural features frequently found in alkaloids.^{5–8} For the purposes of

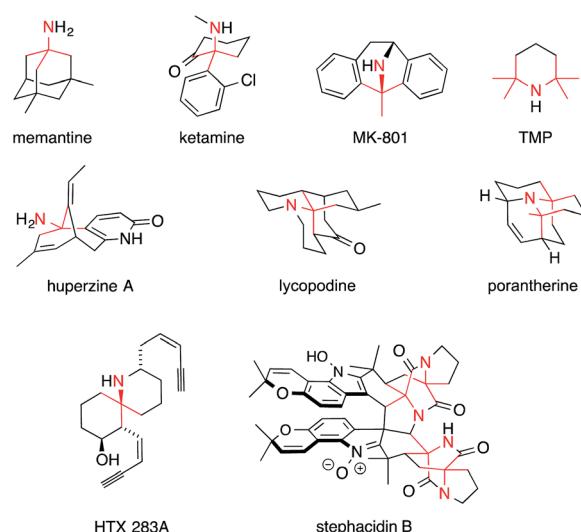


Fig. 1 Alkaloids and synthetic molecules that contain the ATA motif.

Fakultät für Chemie und Pharmazie, Ludwig-Maximilians-Universität München, and Munich Center for Integrated Protein Science, Butenandtstr. 5 – 13, 81377 München, Germany. E-mail: Dirk.Trauner@lmu.de

[†] The authors contributed equally to the review.



this review and in keeping with the literature, we define an ATA as a nitrogen atom bound to a sp^3 -hybridized carbon that bears three additional carbon–carbon bonds. The nitrogen itself can be sp^3 -hybridized as part of a primary, secondary and tertiary amine. Broadening our definition, it can also be sp^2 - or sp -hybridized, *e.g.* in an amide or isonitrile. The tetrasubstituted carbon from which the C,N-bond branches out is often stereogenic, which makes ATAs particularly interesting from a synthetic point of view. Our definition puts emphasis on this particular C,N-bond and avoids the confusion that is often associated with the term ‘quaternary stereocenter’, which, strictly speaking, refers only to a carbon atom surrounded by four other carbons.

Fig. 1 shows some alkaloids and drugs with alkaloid-like properties that illustrate our definition and demonstrate that the nitrogen in ATAs (highlighted in red) can be substituted to various degrees. Memantine and huperzine A contain primary ATAs, whereas ketamine, MK-801 and histrionicotoxin 283A (HTX

283A) feature secondary ATAs, and lycopodine is representative of molecules containing a tertiary ATA. 2,2,6,6-Tetramethylpiperidine (TMP) and the alkaloid porantherine are examples for molecules featuring a twofold ATA. The dimeric alkaloid staphacidin B contains no fewer than four ATAs. Notably, the α -carbons are stereogenic in the majority of these compounds.

In this review, we wish to provide a brief survey of synthetic methods used to install the ATA motif and discuss their application in the total synthesis of alkaloids. The syntheses included here have been selected based on their historical significance, the intriguing structure of their target molecule, and the elegance and efficiency of the method used. The order of their presentation is somewhat arbitrary, mixing biosynthetic and taxonomic categories (such as *Lycopodium* alkaloids), with purely structural ones (such as quinolizidine alkaloids). Generally, we have aimed to proceed from simpler target molecules to more complex ones. While our review is by no means comprehensive, we hope to feature the most instructive examples for the



Anastasia Hager studied chemistry in Würzburg, conducting her Diploma thesis on natural product isolation and synthesis with Prof. G. Bringmann. After joining Prof. D. Trauner's research group for an internship at UC Berkeley, she moved to Munich for her PhD work that was centered on the development of biomimetic syntheses of complex natural products implementing cascade reactions.

From 2013 until 2015 she was a postdoctoral fellow with Prof. D. Fiedler at Princeton University developing novel chemical tools to allow for investigations of inositol pyrophosphate signaling pathway. She now works as a laboratory head in process research at Bayer in Wuppertal.



Dominik Hager studied chemistry in Würzburg, and in 2007, he joined the group of Prof. K. P. C. Vollhardt at UC Berkeley as a visiting researcher. Dominik performed his Diploma studies with Prof. G. Bringmann, working on the development of an axial-chirality transfer concept. For his graduate work he joined the laboratory of Prof. D. Trauner at the LMU. His research projects were centered on natural product synthesis. Between 2013 and 2015 he was a postdoctoral researcher with Prof. D. W. C. MacMillan at Princeton University developing new photochemical protocols. Currently, Dominik works as laboratory head for Bayer CropScience AG.



Nina Vrielink was born in 1987 in Nordhorn, Germany. She studied chemistry and biochemistry at LMU. During her Master's studies in chemistry in Munich, she joined the group of Prof. E. Tate at the Imperial College London for a research internship. In 2012, she joined the laboratory of Prof. D. Trauner for her Master's thesis and returned to conduct her Ph.D. studies starting in 2013. In

2014, she visited the laboratory of Prof. B. M. Stoltz for three months. She is currently pursuing natural product synthesis of α -tertiary-amine-containing alkaloids, supported by a scholarship from the Deutsche Telekom Foundation.



Julien Lefranc was born in 1984 in Marseille, France. He studied chemistry at the Université de la Méditerranée, Marseille, and then moved to the Ecole Nationale Supérieure de Chimie de Montpellier in 2006. In 2008, he joined the group of Prof. J. Clayden at the University of Manchester where he obtained his PhD working on the syntheses of α -tertiary amines. In 2012, Julien joined the group of Prof. D. Trauner as a postdoctoral researcher where he worked on Natural Product synthesis. Since 2014, Julien has been working as a laboratory leader in Medicinal Chemistry for Bayer Healthcare in Berlin.



establishment of ATAs and thus provide inspiration and valuable lessons for future work. We also hope that this review will benefit the design of synthetic pathways toward drugs and synthetic building blocks that contain α -tertiary amines.

2 Methods used for the installation of α -tertiary amines

Many approaches toward the installation of ATAs have been developed but only a relatively small subset of these has proven popular in alkaloid total synthesis. Here, we provide a brief survey of these methods, discussing them in general terms. We classify them according to the bond that is formed in the key step and the electronic nature of the nitrogen and carbon, respectively. However, it should be noted that not all of the syntheses discussed in this review fall into this simplified organizational scheme.

2.1 A C,C-bond is formed in the step that generates the ATA

- The α -carbon is electrophilic. Some of the most commonly encountered methods involve the addition of carbon nucleophiles to activated imines and iminium ions (Scheme 1). They include Mannich reactions, Strecker reactions, aza-Prins reactions and the 1,2-addition of organometallic reagents to C,N-double bonds. *N*-Acyliminium ions are particularly powerful electrophiles in reactions of this type. A variant of the Heck olefination that involves enamines also falls into this category.

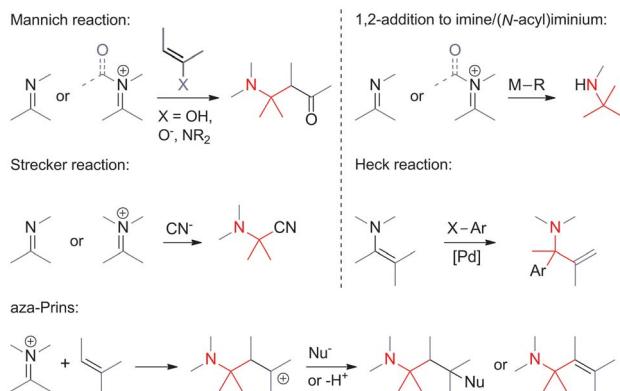
- The α -carbon is nucleophilic. In an Umpolung of the above situation, the α -carbon can also serve as a nucleophile (Scheme 2). For instance, the alkylation of branched nitroalkanes or of deprotonated amino acid derivatives can be used to establish ATAs. Insertions of carbons into nucleophilic C,H-bonds next to a C,N-bond are a member of this general category as well.

- Pericyclic reactions. Pericyclic reactions that form a C,C-bond in the key step have occasionally been employed to form



Dirk Trauner was born and grew up in Linz, Austria. After studying biology and biochemistry at the University of Vienna, he joined Professor Johann Mulzer's group at the Free University of Berlin to pursue natural product synthesis. Subsequently, he became a postdoctoral fellow with Professor Samuel J. Danishefsky at the Memorial Sloan-Kettering Cancer Center in New York City. In 2000, Dirk joined the University of

California, Berkeley, where he became an Associate Professor of chemistry (with tenure). In the summer of 2008, he moved to the University of Munich, where he currently resides as a Professor of Chemistry and Chemical Biology. His research interests range from organic synthesis and natural product chemistry to chemical neurobiology, optogenetics and photopharmacology.



Scheme 1 C,C-bond formation involving electrophilic α -carbons.

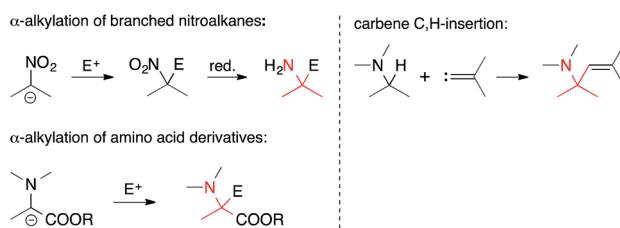
ATAs (Scheme 3). They include Diels–Alder cycloadditions involving 1-aminodienes or 2-azadienes, as well as certain amino dienophiles, [2+2] cycloadditions, and 1,3-dipolar cycloadditions involving nitrones and azomethine imines.

- Radical reactions. Radical reactions establishing ATAs are relatively rare, but not unprecedented (Scheme 4). 5-*endo*-Trig and 6-*endo*-trig cyclizations as well as radical transfer allylations belong to this category.

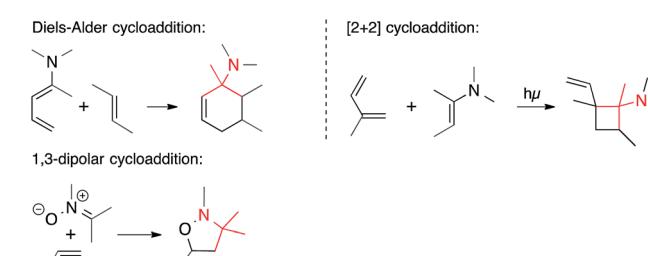
2.2 A C,N-bond is formed in the step that generates the ATA

- The nitrogen is electrophilic. Rearrangements that involve electron-deficient nitrogen atoms are often encountered in the formation of ATAs (Scheme 5).

They include the Curtius, Schmidt, Hofmann, Beckmann and Stieglitz rearrangements.⁵ Often, these reactions can be classified as [1,2] sigmatropic rearrangements. Related

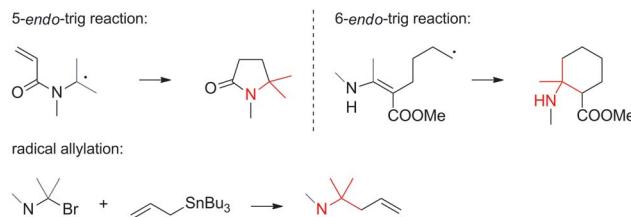


Scheme 2 C,C-bond formation involving nucleophilic α -carbons.

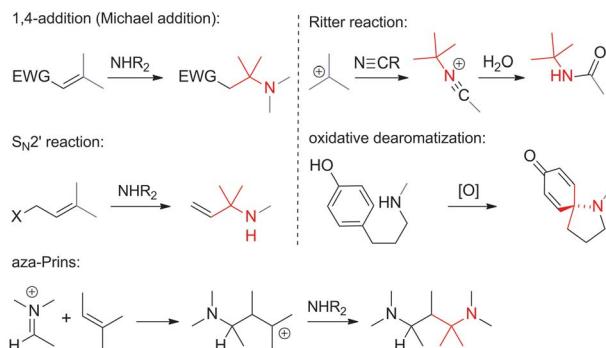


Scheme 3 C,C-bond formation involving pericyclic reactions.





Scheme 4 C,C-bond formation via radical reactions.

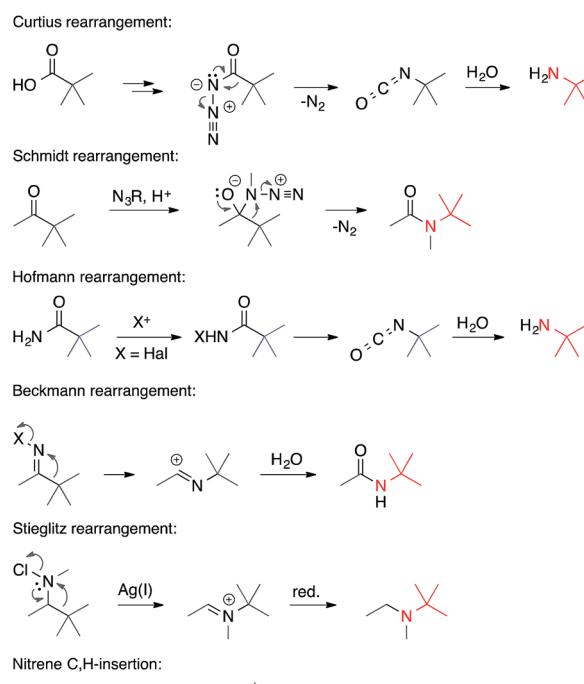


Scheme 5 C,N-bond formation involving nucleophilic nitrogens. EWG = electron-withdrawing group.

nucleophilic substitutions involving *N*-haloamines have been used as well. An electron-deficient nitrogen atom also plays a role in the insertion of nitrenes into C,H-bonds.

• *The nitrogen is nucleophilic.* The formation of ATAs through nucleophilic additions or substitutions involving nitrogen is fairly common (Scheme 6). The classical Michael addition falls into this category, as do S_N2' reactions and haloaminations. For obvious reasons, S_N2 reactions are rare and mostly confined to intramolecular cases. Carbocations that react with a nucleophilic nitrogen occur in the aza-Prins reaction and the Ritter reaction. Oxidative dearomatizations have also been used in a few cases to establish ATAs.

• *Pericyclic reactions.* Pericyclic reactions in which a C,N-bond is formed provide powerful means to establish ATAs (Scheme 7). Overman, Kazmaier–Claisen and [3,3] sigmatropic rearrangements of allylic isocyanides belong to this category. Divinyl cyclopropane rearrangements have also been used to establish ATAs.⁵



Scheme 5 C,N-bond formation involving electrophilic nitrogens.

Scheme 7 C,N-bond formation involving pericyclic reactions.

Many more methods have emerged in recent years that can be used to create ATAs, such as reactions proceeding *via* C,H-activation⁹ and hydroaminations.¹⁰ Since they have not yet been employed in the total synthesis of alkaloids, they are not featured in this review. Other methods, such as the Mannich reaction, Curtius rearrangement and Michael reaction, have proven to be so popular in the total synthesis of alkaloids that we cannot include all instances where they have been employed in this review.

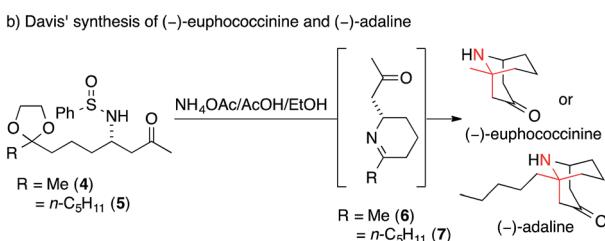
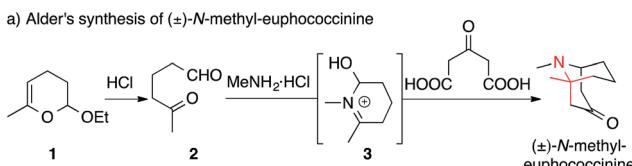
3 Homotropane alkaloids

One of the first applications of Mannich reactions in the construction of ATAs occurred during the synthesis of certain homotropane alkaloids. Three representatives, euphococcinine, *N*-methyl euphococcine and adaline, feature an ATA in the bridgehead position of a bicyclic framework (Fig. 2). These simple natural products are excreted by lady beetles (coccinellids) when threatened.^{11,12}





Fig. 2 Representative ATA-containing homotropane alkaloids.



Scheme 8 Homotropane alkaloid syntheses by Alder (1959) and Davis (2010). Ac = acetyl.

In 1959, Alder synthesized *N*-methyl-euphococcinine using a protocol analogous to the famous tropinone syntheses of Robinson^{13,14} and Schöpf¹⁵ (Scheme 8a).¹⁶ Dehydropyran **1** was converted into ketoaldehyde **2**, which was then transformed into *N*-methyl-euphococcinine in a one-pot process (*via* iminium-intermediate **3**).^{17–19} A similar strategy was later adopted to synthesize the structurally related alkaloid adaline.²⁰

Throughout the years, this biomimetic Mannich strategy was adopted in other syntheses of euphococcinine and adaline.^{21–23} Alternative approaches involved a 1,3-dipolar cycloaddition,²⁴ addition to an *N*-acyliminium ion,²⁵ Michael addition^{26,27} and allylic rearrangement of a cyanate to an isocyanate.^{28,29}

In 2010, Davis published a biomimetic synthesis of $(-)$ -euphococcinine and $(-)$ -adaline in enantiopure form (Scheme 8b).³⁰ The key steps of these syntheses involved the stereoselective formation of piperideine **6** and **7** from the enantiomerically pure *N*-sulfinyl aminoketones **4** and **5**, respectively.³¹ An ensuing intramolecular Mannich reaction afforded the azabicyclononane natural products.

4 Histrionicotoxins

In 1971, Daly isolated six different alkaloids, termed histrionicotoxins (HTXs), from skin extracts of the Colombian poison arrow frog *Dendrobates histrionicus* (Fig. 3).^{32,33} They all contain a unique spirocyclic piperidine core and differ mostly in the length and the degree of saturation of the two side chains. Several histrionicotoxins were identified as inhibitors of nicotinic acetylcholine receptors,^{34–38} which, together with their attractive structures, prompted significant attention from the synthetic community.³⁹ The low natural abundance of these

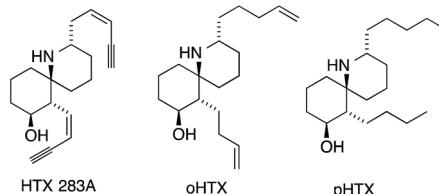


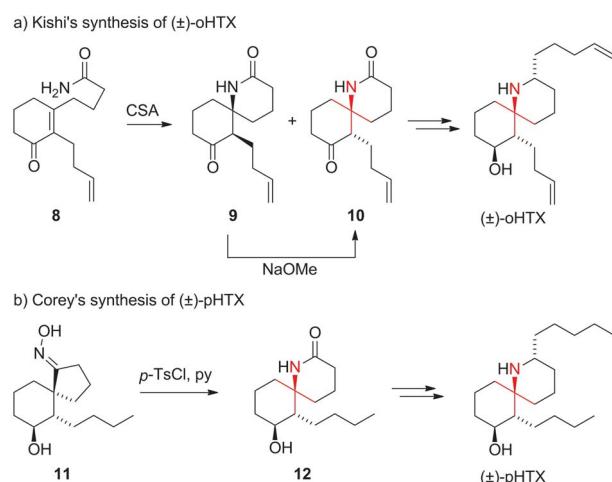
Fig. 3 The major members of the histrionicotoxin family.

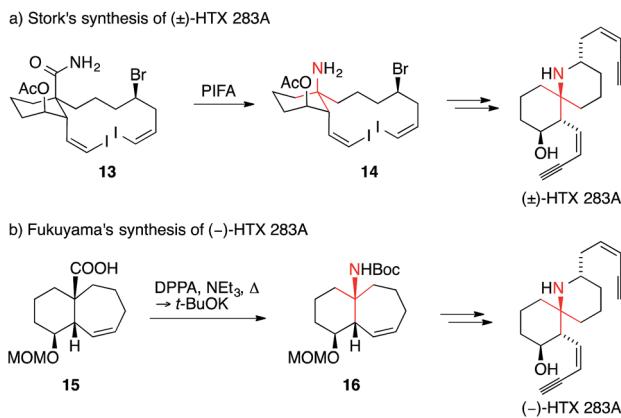
alkaloids and the fact that the frogs do not secrete HTXs in captivity made an efficient synthetic approach all the more desirable.

The first total synthesis of histrionicotoxin alkaloids was reported by Kishi in 1975 (Scheme 9a).^{40–42} His synthesis of octahydrohistrionicotoxin (oHTX) utilized an intramolecular acid-catalyzed aza-Michael addition to set the ATA. Amide **8** was converted to a 2 : 1 mixture of epimeric spiroketolactams **9** and **10**. It was possible to transform **9** into the desired diastereoisomer **10** upon treatment with sodium methoxide.

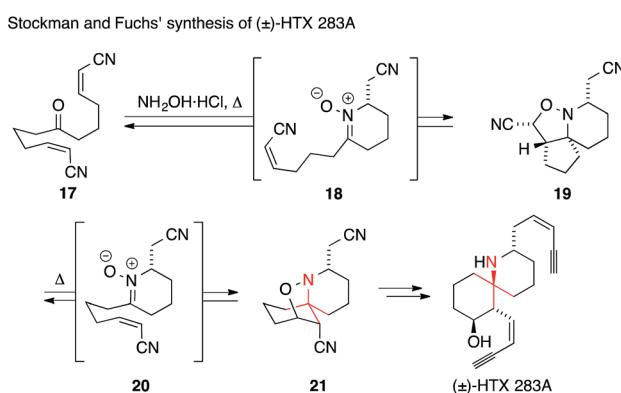
In the same year, Corey reported the first racemic synthesis of perhydrohistrionicotoxin (pHTX), a synthetic HTX derivative (Scheme 9b).⁴³ For the installation of the ATA, Corey used a Beckmann rearrangement that expanded spirocyclic oxime **11** to spirocyclic amide **12**. Several other groups subsequently employed related ring expansion strategies.^{44–48}

Stork reported a synthesis of HTX 283A using a Hofmann rearrangement to set the ATA (Scheme 10a).⁴⁹ During this transformation, amide **13** was oxidized with bis(trifluoroacetoxy)iodobenzene to promote the alkyl migration, giving amine **14** after decarboxylation. More recently, Fukuyama reported an asymmetric synthesis of HTX 283A (Scheme 10b).^{50,51} The key carboxylic acid **15** underwent a stereospecific Curtius rearrangement to yield bicyclo[5.4.0]undecane **16**, which could be converted into HTX 283A.

Scheme 9 Early synthetic approaches to HTXs by Kishi (1975) and Corey (1975). CSA = camphorsulfonic acid, HTX = histrionicotoxin, *p*-TsCl = *para*-toluenesulfonyl chloride.



Scheme 10 HTX 283A syntheses by Stork (1990) and Fukuyama (2011). HTX = histrionicotoxin, Ac = acetyl, Boc = *tert*-butyloxycarbonyl, *t*-Bu = *tert*-butyl, DPPA = diphenylphosphoryl azide, MOM = methoxy-methyl, PIFA = bis(trifluoroacetoxy)iodobenzene.



Scheme 11 HTX 283A synthesis by Stockman and Fuchs (2006). HTX = histrionicotoxin.

A particularly short and efficient synthesis of racemic HTX 283A was reported in 2006 by Stockman and Fuchs (Scheme 11).⁵² In their approach, the key intermediate 21 was formed from the symmetric ketodinitrile 17 using a cascade reaction. Ketone 17 was condensed with hydroxylamine yielding nitron 18 after intramolecular Michael addition. Subsequent intramolecular [3+2] cycloaddition afforded isoxazolidine 19. Following its isolation, 19 was converted to its more stable regioisomer 21 through a retro-[3+2]/[3+2] cycloaddition process (*via* intermediate 20). This so-called 'Holmes dinitrile' (21) had been previously converted into HTX 283A.^{53,54}

Additional strategies to set the ATA in the histrionicotoxins involved Michael reactions,^{41,55,56} Tsuji–Trost amination,^{57,58} iodoetherification,^{59–61} oxidative dearomatizations,⁶² Henry reactions,⁶³ diastereoselective aziridation,⁶⁴ 1,2-carbon migration of hydroxyimines,⁶⁵ [2,3] sigmatropic rearrangements,⁶⁶ [3,3] sigmatropic rearrangements (Kazmaier–Claisen rearrangement)⁶⁷ and [2+2] cycloadditions.^{68,69}

5 *Lycopodium* alkaloids

The genus *Lycopodium* comprises more than 200 species of moss-like plants that have been used in traditional Chinese medicine for centuries.⁷⁰ They have yielded a variety of ATA-containing alkaloids, some of which are featured in Fig. 4. Lycopodine, the first member of the family described in the literature,⁷¹ has proven to be a particularly attractive target for chemical synthesis. In 1968, only seven years after its structural elucidation, Stork⁷² and Ayer^{73,74} simultaneously published the first syntheses of this natural product. Ayer established the ATA using an intermolecular addition of a racemic Grignard reagent to tricyclic, C_V -symmetric iminium ion 22 to obtain 23 (Scheme 12a).

Stork's synthesis involved a Pictet–Spengler type reaction, furnishing tetracyclic amide 25 ($24 \rightarrow 25$, Scheme 12b). A similar cyclization approach was also utilized almost 30 years later in Padwa's asymmetric synthesis of lycopodine (not shown).⁷⁵ Wiesner published a synthesis of 12-*epi*-lycopodine and annotinine using an innovative photochemical strategy (Scheme 12c).^{76–79} This work involved the conversion of vinylogous imide 26 into *exo*-methylene cyclobutane 27 *via* a photochemical [2+2] cycloaddition of allene.

Heathcock established one of the most elegant and influential routes to lycopodine in 1982 (Scheme 13a).^{80–82} In a remarkable sequence, intermediate 28 underwent deprotection, condensation and intramolecular Mannich reaction to yield secondary amine 30, presumably *via* iminium ion 29. The installation of the α -tertiary amine and the formation of two out of four rings thus occurred in a single step, mimicking the proposed biosynthesis of this natural product. Subsequent optimization led to the shortest synthesis of lycopodine to date,

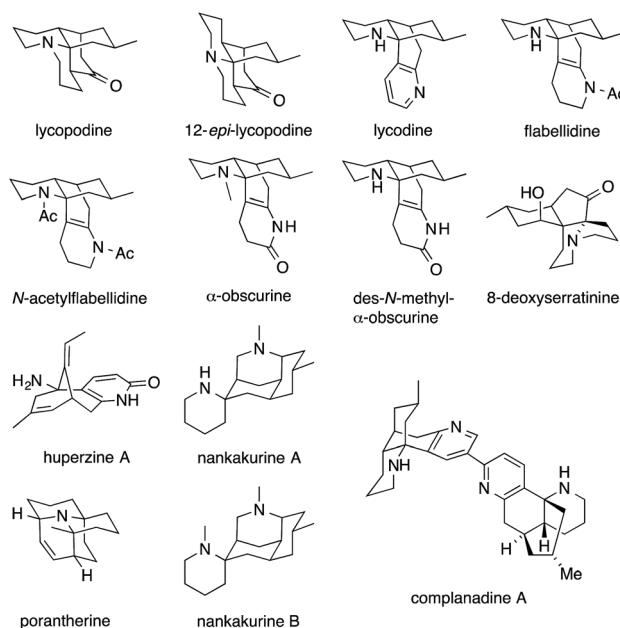
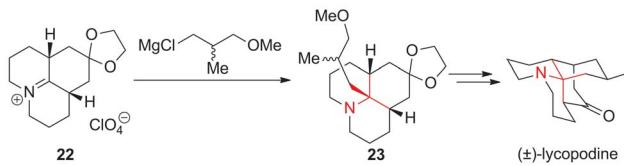
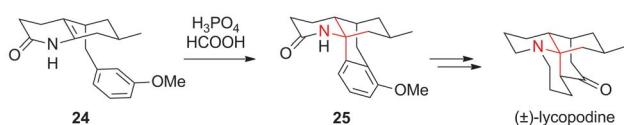


Fig. 4 Representative *Lycopodium* alkaloids.

a) Ayer's synthesis of (\pm)-lycopodineb) Stork's synthesis of (\pm)-lycopodinec) Wiesner's synthesis of (\pm)-12-*epi*-lycopodine

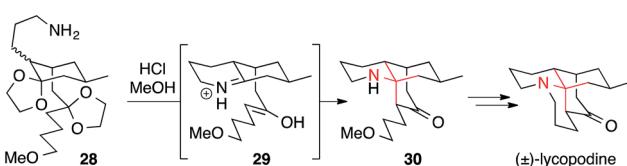
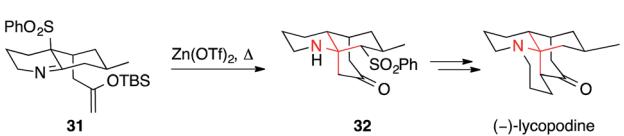
Scheme 12 Representative lycopodine syntheses by Ayer (1968), Stork (1968) and Wiesner (1967). Bn = benzyl.

consisting of only eight steps.⁸² Using a similar sequence, lycodine and lycodoline were prepared as well.⁸²

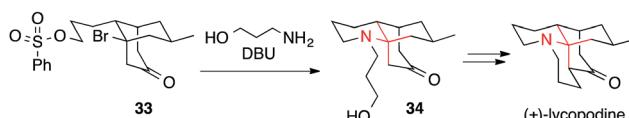
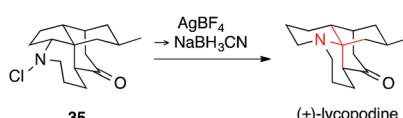
Variations of Heathcock's strategy have been used in other synthetic approaches toward *Lycopodium* alkaloids, *e.g.* in syntheses of clavoloneine by Evans (2005)⁸⁴ and Fujioka (2011).⁸³

One drawback of intramolecular Mannich reactions, however, is the need to simultaneously form an iminium ion and an enol. Thus, long reaction times of up to 18 days were needed.⁸² Recently, this problem was solved in an elegant way by Carter (Scheme 13b).^{85,86} Using an aza-Wittig approach, Carter was able to prepare and isolate the TBS-enol ether imine 31. Treatment of 31 with zinc triflate furnished the ATA and concomitantly resulted in the rearrangement of the sulfinyl residue yielding lycopodine precursor 32.

In 1985, Kraus published a route towards lycopodine that was based on the formation of a bridgehead olefin (Scheme 14a).⁸⁷ Tertiary alkyl bromide 33 was treated with DBU and 3-

a) Heathcock's synthesis of (\pm)-lycopodineb) Carter's synthesis of ($-$)-lycopodine

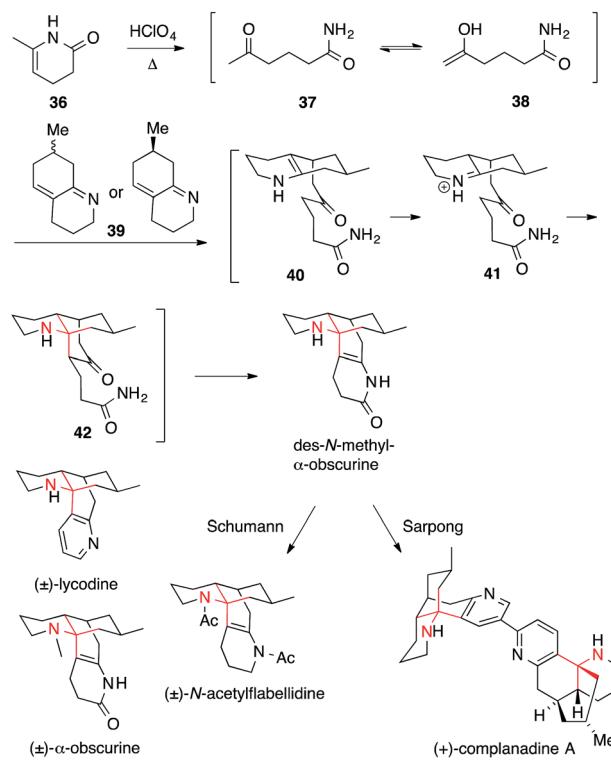
Scheme 13 Lycopodine syntheses by Heathcock (1982) and Carter (2008). OTf = trifluoromethanesulfonate, TBS = *tert*-butyldimethylsilyl.

a) Kraus' synthesis of (\pm)-lycopodineb) Grieco's synthesis of (\pm)-lycopodine

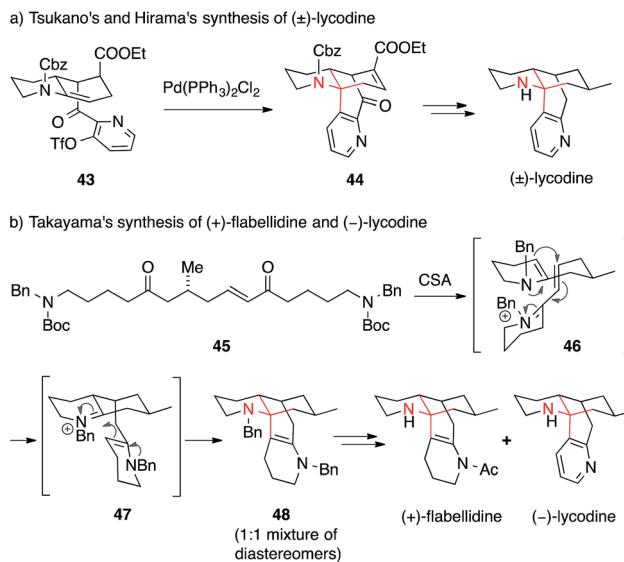
Scheme 14 Lycopodine syntheses by Kraus (1985) and Grieco (1998). DBU = 1,8-diazabicyclo[5.4.0]undec-7-ene.

amino-1-propanol to install amino ketone 34, which could be further transformed into the natural product in two additional steps using Heathcock's protocol. An equally unusual approach was reported by Grieco, who employed a Stieglitz rearrangement (Scheme 14b).⁸⁸ To effect the reaction, *N*-chloroamine 35 was treated with silver tetrafluoroborate followed by cyanoborohydride. Many other syntheses of lycopodine have been accomplished utilizing different strategies, such as Michael additions, for the assembly of the ATA.⁸⁹⁻⁹¹

Members of the lycodine class of natural products feature an ATA and a pyridine or pyridone moiety. The parent compound, lycodine,⁹² was first isolated from *L. annotinum* in 1958.⁹³ Apart

Schumann's syntheses of (\pm)-lycodine, (\pm)- α -obscurine, (\pm)-*N*-acetylflabellidine and Sarpong's synthesis of (+)-complanadine A

Scheme 15 Synthesis of various *Lycopodium* alkaloids by Schumann (1982) and Sarpong (2010). Ac = acetyl.



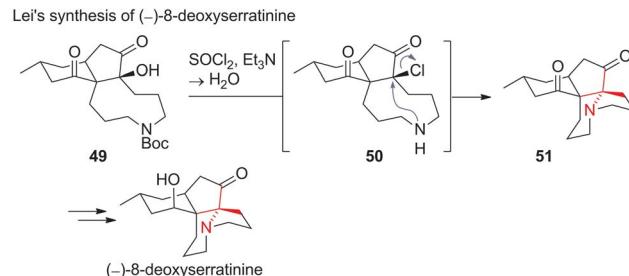
Scheme 16 Syntheses of various *Lycopodium* alkaloids by Tsukano (2010) and Takayama (2014). Cbz = benzyloxycarbonyl, CSA = camphorsulfonic acid, Boc = *tert*-butyloxycarbonyl, Bn = benzyl, OTf = trifluoromethanesulfonate.

from the Heathcock synthesis mentioned above,⁸² several additional syntheses of lycodine have been achieved to date.^{94–97}

Schumann used a classical Mannich strategy to access racemic lycodine, α -obscurine and *N*-acetylflabellidine (Scheme 15).^{98–100}

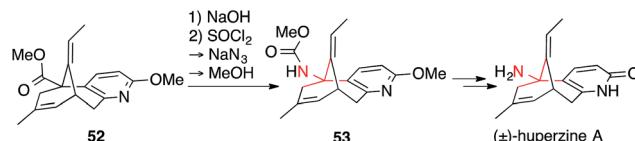
The mechanism of the key double Mannich reaction cascade was further explored almost 30 years later by Sarpong.⁹⁴ He used the same cascade as an opening sequence in an asymmetric synthesis of enantiomerically pure (+)-complanadine A, a lycodine dimer, which was shown to enhance expression of nerve growth factor in human cells.¹⁰¹ It was found that cyclic enamide 36 opens to ketone 37 or enol 38 under acidic conditions, which adds to the unsaturated bicyclic imine 39. Protonation of the resulting enamine 40 triggers a second, intramolecular Mannich reaction to afford tricycle 42 via the iminium ion 41. Finally, an intramolecular enamide formation furnished tetra-cyclic des-*N*-methyl- α -obscurine, containing the entire lycodine framework.

In an unusual approach, Tsukano and Hirama applied an intramolecular palladium-mediated Heck reaction between



Scheme 17 Lei's synthesis of $(-)$ -8-deoxyserratinine (2014). Boc = *tert*-butyloxycarbonyl.

Kozikowski's synthesis of (\pm) -huperzine A



Scheme 18 Huperzine A synthesis by Kozikowski (1989).

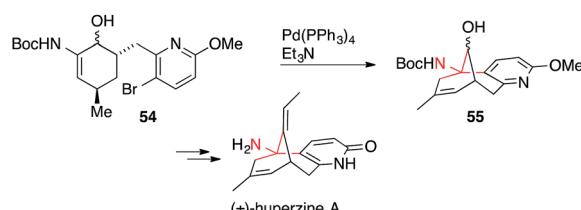
enecarbamate and pyridine triflate 43 to form the ATA, which yielded lycodine precursor 44 (Scheme 16a).⁹⁵

Recently, another very short synthesis of $(-)$ -lycodine as well as the closely related (+)-flabellidine was accomplished by Takayama (Scheme 16b).⁹⁷ Starting from a linear precursor 45, he was able to assemble the whole tetracyclic skeleton 48 of both alkaloids in a cascade reaction involving a double condensation (45 → 46), a conjugate addition (46 → 47) followed by a Mannich reaction (47 → 48). In addition, Shair published an approach towards several members of the 7-membered-ring-containing *Lycopodium* alkaloids using a trans-annular Mannich reaction (not shown).^{102,103}

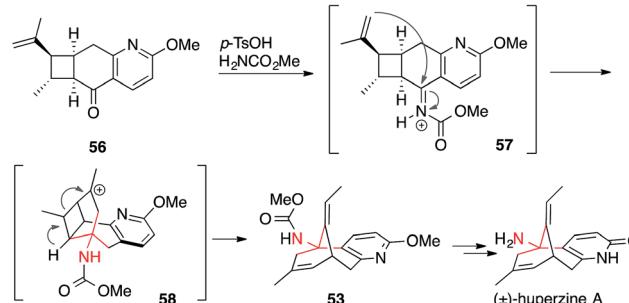
One of the rare cases of an S_N2 reaction in ATA formation can be found in Lei's recent synthesis of $(-)$ -8-deoxyserratinine (Scheme 17).¹⁰⁴ Tertiary alcohol 49 was converted into chloride 50, which was attacked intramolecularly by the free secondary amine (50 → 51). In 2014, Lei extended his strategy to a synthesis of the oxidised congener $(-)$ -serratinine.¹⁰⁵ Other approaches towards 8-deoxyserratinine and related alkaloids include a Schmidt rearrangement and an intramolecular epoxide opening.^{106–108}

In contrast to the multiple strategies used for the installation of ATAs in the *Lycopodium* alkaloids mentioned above, the

a) Sun's and Lin's synthesis of huperzine A



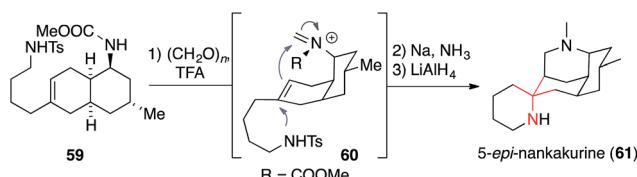
b) White's synthesis of (\pm) -huperzine A



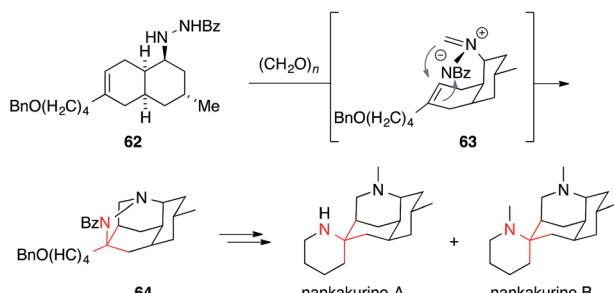
Scheme 19 Huperzine A syntheses by Sun/Lin (2012) and White (2013). Boc = *tert*-butyloxycarbonyl, *p*-TsCl = *para*-toluenesulfonic acid.



a) Overman's synthesis of misassigned nankakurine A (61)



b) Overman's synthesis of (+)-nankakurines A and B

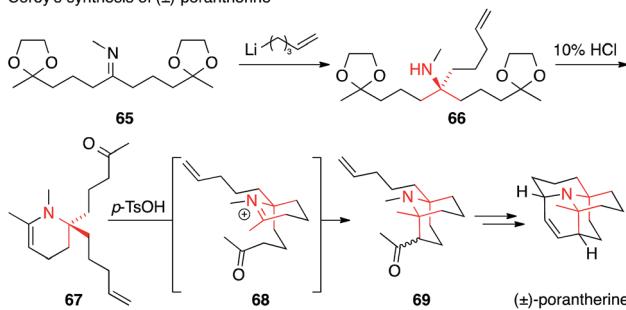


Scheme 20 Overman's syntheses of misassigned nankakurine A (2008) and revised nankakurines A and B (2010). Bn = benzyl, Bz = benzoyl, TFA = trifluoroacetic acid, Ts = para-toluenesulfonyl.

methods used to access the medicinally important acetylcholine esterase inhibitor huperzine A are less diverse. Since the ATA in huperzine A is primary, it can be efficiently installed using a Curtius rearrangement. Indeed, synthetic efforts towards huperzine A were almost exclusively focused on carboxylic acid precursors, such as 52.^{109–116}

The first synthesis of huperzine A was published by Koziowski in 1989 (Scheme 18).¹⁰⁹ First, he completed the core 52 wherein the primary amine is replaced by a methyl ester. After saponification, Curtius rearrangement (52 → 53) followed by double deprotection provided racemic huperzine A. In the following years, many huperzine A syntheses and several semi-syntheses were published.^{117,118} All of them featured a racemic or enantiomerically pure carboxylic acid derivative of precursor 53, keeping the Curtius rearrangement as the key step for the formation of the ATA.^{109–116} These efforts culminated in the recently published large-scale asymmetric synthesis of huperzine A.¹¹⁹

Corey's synthesis of (±)-porantherine



Scheme 21 Porantherine synthesis by Corey (1974). p-TsOH = para-toluenesulfonic acid.

A few groups, however, have been able to avoid Curtius rearrangements in the synthesis of huperzine A. Sun and Lin accessed the alkaloid using an intramolecular Heck reaction (54 → 55) (Scheme 19a),¹²⁰ whereas the White group performed an elegant tandem intramolecular aza-Prins cyclization/cyclobutane fragmentation (56 → 53) to set the ATA in 53 (Scheme 19b).¹²¹

Two *Lycopodium* alkaloids recently isolated from *Lycopodium hamiltonii*, *viz.* the nankakurines A and B, have attracted broad interest in the synthetic community (Fig. 4).^{122,123} So far, two syntheses of these natural products have been reported. In 2008, Overman published an enantioselective synthesis of the misassigned original structure of nankakurine A (61) (Scheme 20a) followed by the syntheses of the reassigned structures of nankakurine A and B in 2010 (Scheme 20b).^{124,125}

In the case of 5-epi-nankakurine (61), an aza-Prins reaction (59 → 60) was used, which allowed for the direct formation of both piperidine rings in 61 in one step starting from bicyclic 59.¹²⁴ This strategy, however could not be applied for the formation of actual nankakurine A. Its synthesis was accomplished utilizing an intramolecular 1,3-dipolar cycloaddition of an azomethine imine 63, formed *in situ* *via* condensation of 62 with formaldehyde. This reaction provided access to tetracyclic pyrazolidine 64, which, after SmI2 mediated N,N-bond cleavage, gave rise to nankakurines A and B.¹²⁵ Two years later, Waters reported a racemic synthesis of nankakurines A and B using a Grignard addition to an iminium species derived from luciduline, which is easily accessible by total synthesis (not shown).¹²⁶

Porantherine, the major alkaloid of the poisonous woody shrub *Poranthera corymbosa*, is structurally similar to the *Lycopodium* alkaloids, although not a member of the family (Fig. 4).^{127,128} Possessing two tertiary carbons attached to the same amine (twofold ATA), porantherine is a considerable synthetic challenge that has been met only twice thus far.^{129,130} Both syntheses are racemic and based on similar strategies for the assembly of the ATA motif, namely an addition to a ketimine followed by Mannich reaction. Corey published his synthesis of the natural product in 1974 (Scheme 21),¹²⁹ only three years after its isolation. The first ATA was installed through addition of an organolithium compound to imine 65 to form 66, which then cyclized to the corresponding enamine 67 upon treatment with acid. The formation of the second ATA center through an intramolecular Mannich addition (*via* iminium ion 68) furnished ketone 69, which was eventually converted to the natural product.

A second synthesis of porantherine, published by Stevens in 1987, involved the addition of two alkylolithium compounds to an iminoether (not shown).¹³⁰

6 Hasubanan alkaloids

The hasubanan alkaloids, isolated from various plant sources, are structurally related to the better-known morphine alkaloids but feature a pyrrolidine ring instead of a piperidine ring. They are comprised of over 40 family members, all of which share the same aza-propellane skeleton (Fig. 5).¹³¹



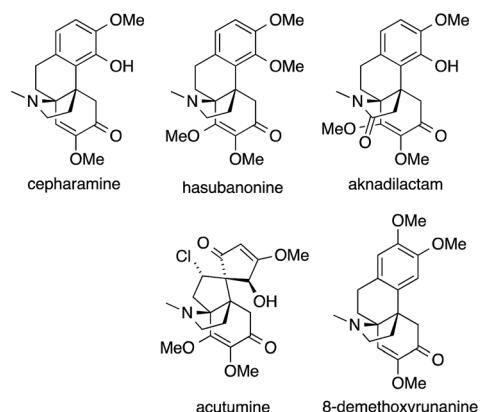
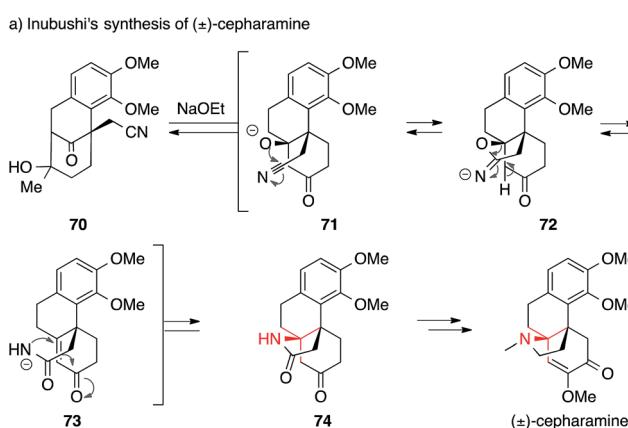
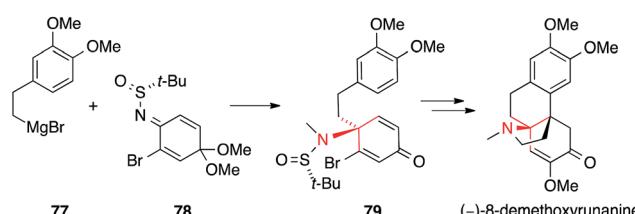
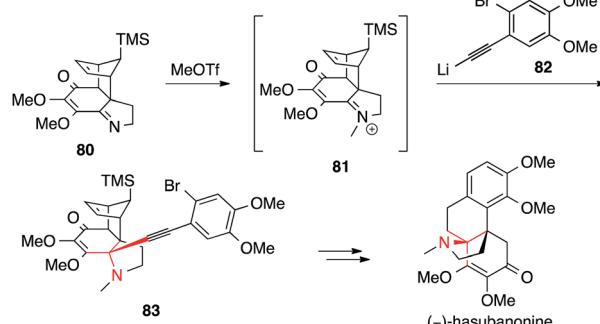


Fig. 5 Representative hasubanan alkaloids.

Although the physiological effects of the hasubanans are of less significance compared to the related morphine alkaloids, their beautiful structures stimulated numerous synthetic studies and several total syntheses have been reported to date.^{132–146}

The first successful syntheses of (\pm) -cepharamine,^{132,135} (\pm) -hasubanone,^{134,137} (\pm) -aknadilactam¹³⁴ and (\pm) -metaphanine^{136,138} were published by Inubushi in the 1970s (Scheme 22a). Starting from tricyclic β -tetralone 70, the ATA was set *via* a cascade reaction involving an aza-Michael addition (71 \rightarrow 74). Almost 30 years later, Schultz prepared $(+)$ -cepharamine using a Hofmann-type rearrangement to introduce the ATA (not shown).¹³⁹

Recent years have seen revived interest in hasubanans. An elegant method for the installation of the ATA was developed by

Scheme 22 Syntheses of hasubanan alkaloids by Inubushi (1969) and Castle (2009). Bn = benzyl, TBS = *tert*-butyldimethylsilyl.a) Reisman's synthesis of $(-)$ -8-demethoxyrunanineb) Herzon's synthesis of $(-)$ -hasubanoneScheme 23 Syntheses of hasubanan alkaloids by Reisman (2011) and Herzon (2011). *t*-Bu = *tert*-butyl, TMS = trimethylsilyl, OTf = trifluoromethanesulfonate.

Castle, who closed the pyrrolidine ring of isohasubanone through a S_N2' -reaction.^{140,147} Subsequently, this strategy was adapted to access $(-)$ -acutumine (Scheme 22b).¹⁴² To this end, amine 75 was exposed to a Lewis acid to generate an allylic cation that was intercepted by the appended secondary amine. The resulting ATA-containing pyrrolidine 76 was then converted into $(-)$ -acutumine.

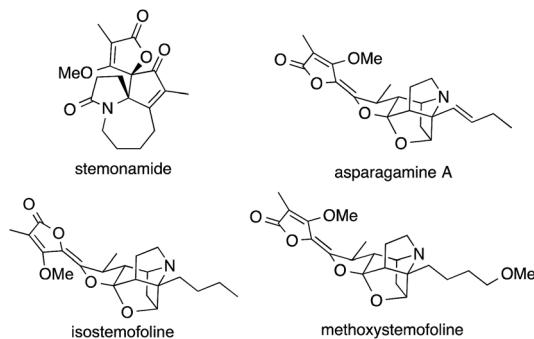
In contrast to this approach, which sets the ATA at a relatively late stage in the synthesis, Reisman installed it at the beginning (Scheme 23a).¹⁴⁴ Reaction of the chiral *N*-*tert*-butanesulfinimine 78 with phenethyl Grignard 77 provided sulfonamide 79 with a high degree of diastereoselectivity. Subsequently, 79 was converted into a series of hasubanan alkaloids such as $(-)$ -8-demethoxyrunanine.

The first enantioselective synthesis of hasubanone was published by Herzon (Scheme 23b).¹⁴³ Methylation of imino-quinone Diels–Alder adduct 80 (80 \rightarrow 81), followed by addition of alkynyl lithium 82 gave amine 83, which was eventually transformed into optically pure $(-)$ -hasubanone. This strategy proved to be versatile, as many more hasubanan alkaloids, including $(-)$ -runanine, $(-)$ -delavayne, $(+)$ -periglaucine B and $(-)$ -acutumine, could be accessed by variation of the alkynyl species.^{143,145,146}

7 *Stemona* alkaloids

Plants belonging to the family *Stemonaceae*, which are mostly found in Southeast Asia, have been used for centuries as insecticides and for the treatment of respiratory diseases.^{148–150} Phytochemical investigations led to the isolation of a variety of natural products known as *Stemona* alkaloids (Fig. 6).^{151,152} These polycyclic natural products possess highly complex



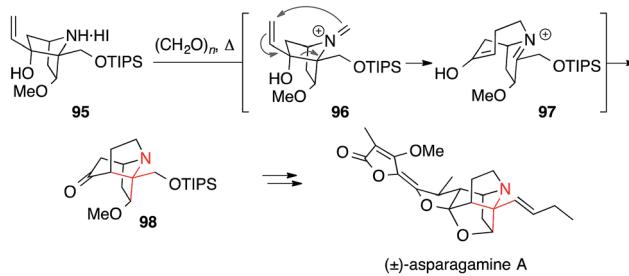
Fig. 6 Representative *Stemonota* alkaloids.

structures weaving together pyrrolidines and butenolides, often through *spiro* fusions that contain ATAs.

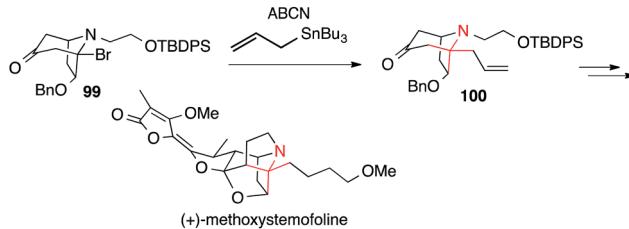
The structural beauty of these molecules generated considerable interest in the synthetic community and stimulated the development of new synthetic methods for the installation of ATAs.^{151,152} The strategies employed range from classical additions to imines,^{153,154} to radical cyclization cascades,^{155,156} radical allylations,¹⁵⁷ semipinacol-Schmidt cascades,^{158,159} Schmidt reactions,¹⁶⁰ aza-Cope–Mannich reactions,¹⁶¹ cyclopropane-Cope rearrangements¹⁶² and catalytic 1,3-dipolar cycloadditions.¹⁶³

The first synthesis of a *Stemonota* alkaloid, *viz.* isostemofoline, was published by Kende in 1999 and employed a highly unusual and elegant approach.¹⁶² The ATA was formed *via* rhodium-catalyzed reaction of pyrrole **84** with vinyl diazoester **85**. The

a) Overman's synthesis of (±)-asparagamine A



b) Huang's synthesis of (+)-methoxystemofoline

Scheme 25 Syntheses of *Stemonota* alkaloids by Overman (2003) and Huang (2015). ABCN = 1,1'-azobis(cyclohexanecarbonitrile), Bn = benzyl, TBDPS = *tert*-butyldiphenylsilyl, TIPS = triisopropylsilyl.

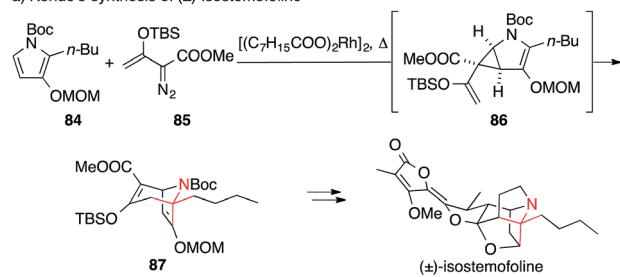
resultant divinyl cyclopropane **86** underwent Cope rearrangement *in situ* to afford bicyclic **87**, which was then used as a key intermediate in the further assembly of the natural product (Scheme 24a).

More recently, two synthetic approaches aimed at members of the stemonamine group were published. Ishibashi developed an entry to racemic stemonamide and isostemonamide as well as their reduced derivatives stemonamine and isostemonamine, based on a radical cascade as the key step for the formation of the ATA (Scheme 24b).^{155,156} Treatment of the achiral precursor **88** with tributyltin hydride and 1,1'-azobis(cyclohexanecarbonitrile) (ABCN) at elevated temperatures effected a 7-*endo*-trig cyclization that likely yielded radical **89** as the proposed intermediate, which in turn underwent an unusual 5-*endo*-trig cyclization providing access to a separable mixture of isomers **90** and **91**. Further transformations of these tricyclic compounds furnished stemonamide and some of its congeners.

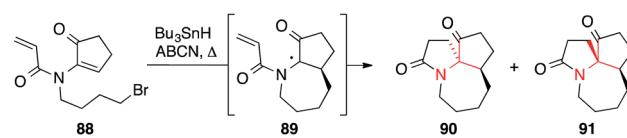
An alternative approach to stemonamide and related *Stemonota* alkaloids was published by Zhang (Scheme 24c).¹⁵⁹ Based on his systematic studies on the reactivity of α -hydroxy epoxides such as **92**,¹⁶⁴ he developed a powerful cascade that combines a semipinacol rearrangement with an Aubé–Schmidt reaction (**92** \rightarrow **94**). The resulting amide **94** was obtained as a 5 : 1 mixture of diastereomers, reflecting the diastereomeric mixture of propargylic azides employed as substrates. Using this strategy and variations thereof, Zhang was able to synthesize stemonamide and three additional *Stemonota* alkaloids, *viz.* maistemonine, stemonamine, and isomaistemonine.^{158–160}

The only synthesis of asparagamine A, an unsaturated derivative of stemonamide, was achieved by Overman in 2003.¹⁶¹ He installed the ATA using his signature aza-Cope–Mannich cascade (Scheme 25a). The synthesis of a precursor molecule

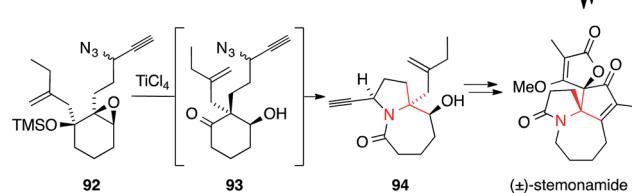
a) Kende's synthesis of (±)-isostemofoline



b) Ishibashi's synthesis of (±)-stemonamide



c) Zhang's synthesis of (±)-stemonamide

Scheme 24 Syntheses of *Stemonota* alkaloids by Kende (1999), Ishibashi (2008) and Zhang (2011). ABCN = 1,1'-azobis(cyclohexanecarbonitrile), Boc = *tert*-butyloxycarbonyl, MOM = methoxymethyl, TBS = *tert*-butyldimethylsilyl, TMS = trimethylsilyl.

suitable for this transformation commenced with a Diels–Alder reaction, forming an ATA that later took part in the rearrangement. Treatment of **95** with excess paraformaldehyde at elevated temperatures generated iminium ion **96**, which underwent the aza-Cope–Mannich sequence to afford the ATA of asparagamine A (**98**, *via* intermediate **97**). More recently, a radical allylation (**99** → **100**) was utilized by Huang to set the ATA in (+)-methoxystemofoline at an early stage of the synthesis (Scheme 25b).¹⁵⁷

8 Indole alkaloids

Indole alkaloids are a structurally and biosynthetically heterogeneous class of natural products characterized by an indole nucleus or derivative thereof. Several of them, albeit not the best known ones, contain ATAs (Fig. 7).

Kopsine, the first member of the so-called *Kopsia* alkaloids, was isolated as early as 1890,¹⁶⁵ but it took several decades before its complex structure, and those of its congeners, could be elucidated.^{166–173} All members of this family possess an ATA incorporated in a bicyclo[2.2.2]octane system. Thus, the kopsanes seem predestined for Diels–Alder reactions, and few syntheses fail to employ a [4+2] cycloaddition strategy.^{174–176} The routes used can be divided into two main categories: (a) intermolecular Diels–Alder reactions^{177–180} and (b) intramolecular Diels–Alder reactions.^{181–185}

The very first synthesis of (±)-aspidofractinine, completed in 1976, introduced an intermolecular Diels–Alder reaction to set the ATA using nitroethylene as a dienophile (not shown).¹⁷⁷ Over time, phenyl vinyl sulfone emerged as a more practical dienophile^{178,179} and in 2009 the first enantioselective synthesis of (+)-aspidofractinine was reported by Spino using this reagent (Scheme 26a).¹⁸⁰ In this case, imine **101** thermally isomerized to diene **102**, which then underwent cycloaddition from the sterically more accessible convex side to afford sulfone **103**.

The first successful intramolecular Diels–Alder approach to (±)-kopsanone and (±)-10,22-dioxokopsane was reported in 1983 by Magnus (Scheme 26b).^{181,182} They synthesized sulfide **104** as a suitable precursor, with the dienophile placed in the concave position. The cycloaddition reaction proceeded at 100 °C and provided intermediate **105**, which was transformed into (±)-kopsanone in a few steps. Using a similar strategy, other indole alkaloids, (±)-kopsijasmine and (±)-kopsine, were prepared as racemates,^{184,185} as well as (–)-kopsinilam and (–)-kopsinine in enantiomerically pure form.¹⁸³

In a recent example for an alternative approach by Boger, a powerful radical transannular cyclization was applied to install the ATA of kopsinine (Scheme 26c).¹⁷⁶ Upon treatment of xanthate **106** with SmI₂, ATA **108** was formed as a single diastereomer. Presumably, a primary radical intermediate **107** is formed, which undergoes a radical cyclization followed by reduction and diastereoselective protonation of the ester enolate.

Lapidilectine B and lundurine A are two structurally related *Kopsia* alkaloids that contain two ATAs. Although not originating from the same organism, they show a similar scaffold with a bridged 8-membered ring fused to an indoline on one

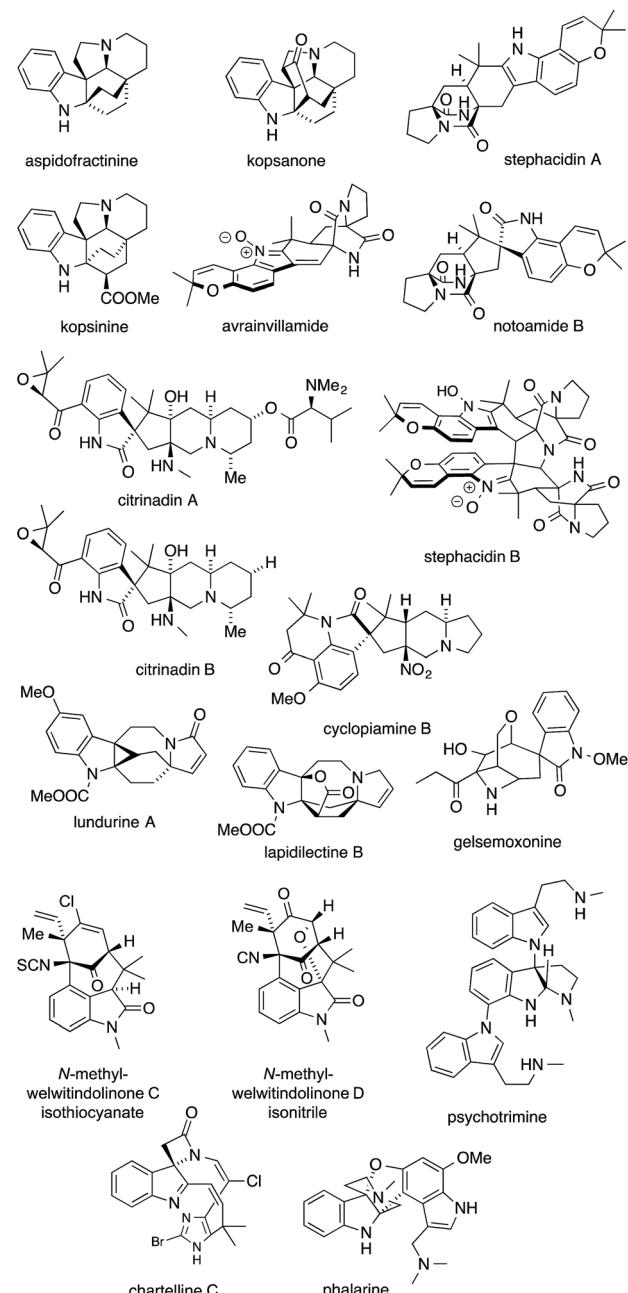
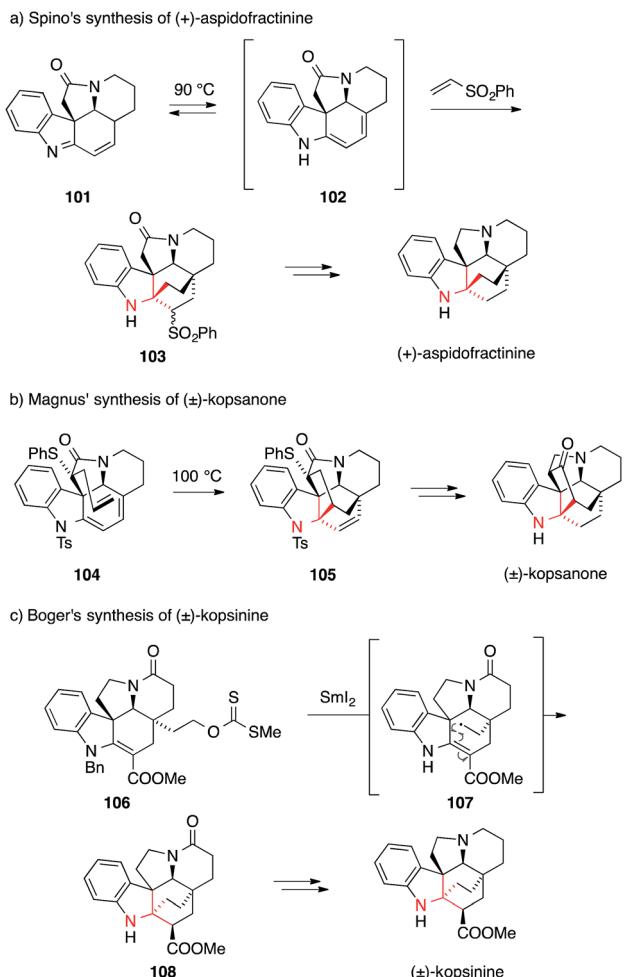


Fig. 7 Indole alkaloids featuring ATAs.

side and a 5-membered ring on the other. Lapidilectine A was isolated by Awang from the leaves of the tree *Kopsia lapidilecta* in 1992.^{186,187} Lundurines A–D were isolated from the Malaysian tree *Kopsia tenuis*,¹⁸⁸ and shown to be effective at bypassing multidrug resistance in vincristine-resistant KB cells.¹⁸⁹

Qin accomplished the first enantioselective synthesis of (–)-lundurine A in 2014 (Scheme 27a).¹⁹⁰ The first ATA was established *via* the addition of allylmagnesium bromide to an iminium ion generated by *in situ* alkylation of imine **109** to form tetracycle **110**. In order to establish the two fully substituted stereocenters on the indoline of **112**, Qin resorted to an unusual intramolecular Simmons–Smith cyclopropanation of diiodide



Scheme 26 *Kopsia* alkaloid syntheses by Spino (2009), Magnus (1983) and Boger (2013). Bn = benzyl, Ts = *p*-toluenesulfonyl.

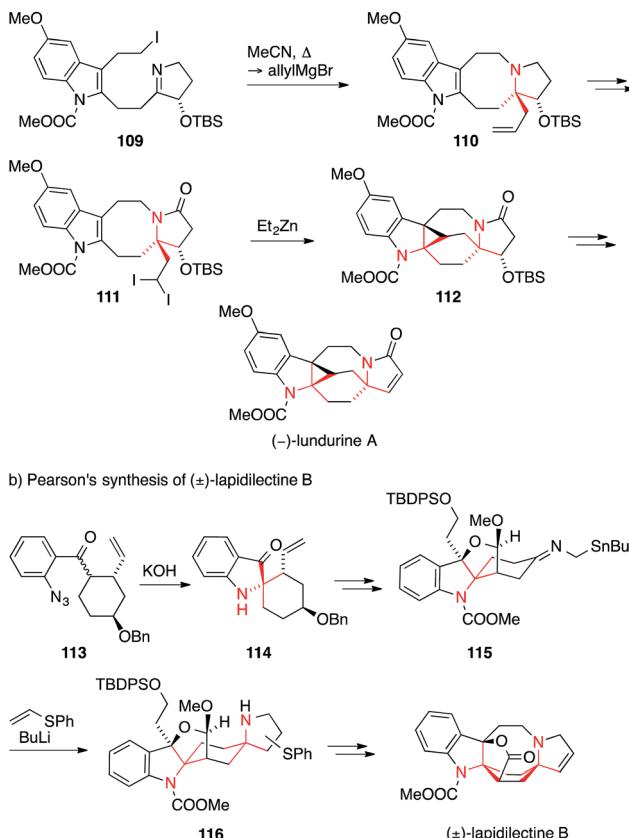
111. Two other racemic syntheses of lundurine A and B have been reported by Nishida.^{191–193} He employed a Curtius rearrangement and a 1,2-addition to an iminium ion for lundurine B¹⁹³ and a Tsuji–Trost amination and an indoxyl bisalkylation for the synthesis of lundurine A (not shown).^{191,192}

In 2001, Pearson employed a Smalley cyclization of aryl ketone azide 113 to furnish the spiroindoxyl 114 (Scheme 27b).^{194,195} In the final steps of his lapidilectine B synthesis, he then used his trademark azaallyl anion [3+2] cycloaddition to establish the pyrrolidine ring (115 → 116) as an inconsequential mixture of regioisomers.

The cycloaddition approach has not been limited to the kopsane alkaloids. Other indole alkaloids, such as stephacidin A and the notoamides, which bear two ATAs, were prepared by a presumably biomimetic [4+2] cycloaddition.

Williams synthesized stephacidin A and notoamide B starting from imidate 117, which underwent base-mediated isomerization to 118 followed by intramolecular Diels–Alder reaction to afford diazabicyclo[2.2.2]octane 119 (Scheme 28).¹⁹⁶ This remarkable reaction sets both ATAs in a single step. Later that

a) Qin's synthesis of (−)-lundurine A



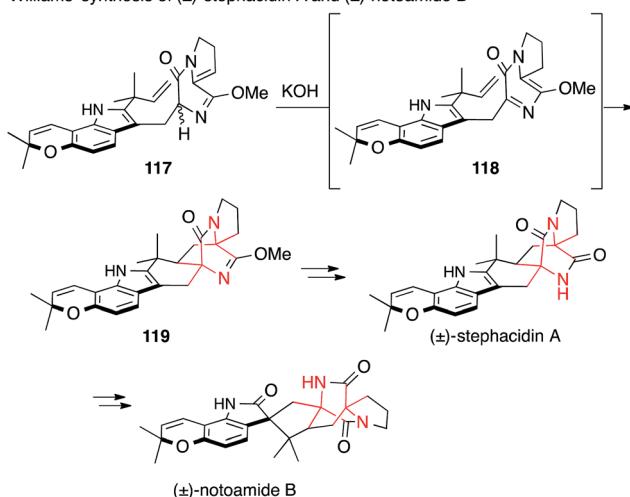
Scheme 27 Syntheses of *Kopsia* alkaloids by Qin (2014) and Pearson (2001). Bn = benzyl, TBDPS = *tert*-butyldiphenylsilyl, TBS = *tert*-butyldimethylsilyl.

year, stephacidin B was accessed *via* avrainvillamide using the same strategy.¹⁹⁷

In 2005, Baran used the α -alkylation of proline derivative 120 with complete chirality transfer, a method developed by Seebach,¹⁹⁸ to set the first ATA of stephacidin A in 121 (Scheme 29).¹⁹⁹ The second ATA present in 123 was installed by an intramolecular, stereocontrolled oxidative enolate coupling starting from diketopiperazine 122. Baran was then able to convert stephacidin A into avrainvillamide and stephacidin B following a biosynthetic proposal.²⁰⁰

A second synthesis of avrainvillamide and stephacidin B was accomplished concurrently by Myers (Scheme 30).²⁰¹ In this case, the first ATA was installed by a Strecker-type addition of TMS cyanide to enamine 124 to form the *N*-Boc amino nitrile 125. The second ATA was then set by a very unusual radical transfer cyclization. Abstraction of a hydrogen atom in 126, followed by loss of toluene, generates an aminoacyl radical which attacks the enamide double bond and ejects a phenylthiyl radical to form the diketopiperazine 127.

Several members of a related family of prenylated spirooxindole alkaloids, namely cyclopamine, citrinadin A and citrinadin B, also feature an asymmetric ATA.^{202–204} In 2013, Martin and Wood reported the first syntheses of citrinadin A and B.^{205,206}

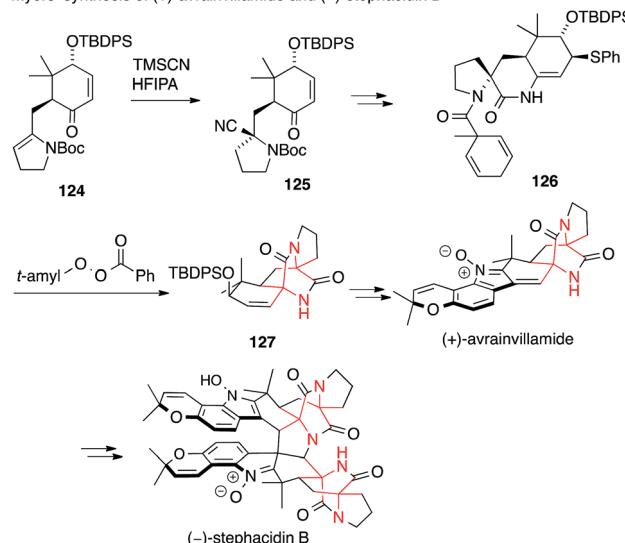
Williams' synthesis of (\pm)-stephacidin A and (\pm)-notoamide B

Scheme 28 Synthesis of prenylated indole alkaloids by Williams (2007).

In the case of citrinadin A, epoxide **128** was heated in the presence of methylamine to provide 1,2-amino alcohol **129** (Scheme 31a).²⁰⁵ Wood's approach employed an azide-mediated opening of epoxide **130** to establish the ATA in **131** (Scheme 31b).²⁰⁶ Both reactions are rare examples where an ATA has been set through a S_N2 reaction.

More recently, Sarpong published his entry to the prenylated indole alkaloids cyclopamine B and *ent*-citrinalin B (Scheme 32). The first ATA was set *via* a Hofmann rearrangement (**132** \rightarrow **133**).²⁰⁷ The second, not asymmetric, ATA center was established by treating *ent*-citrinalin B with sodium hydride to effect the rearrangement of the chromanone to the tetrahydroquinolone

Myers' synthesis of (+)-avravillamide and (-)-stephacidin B



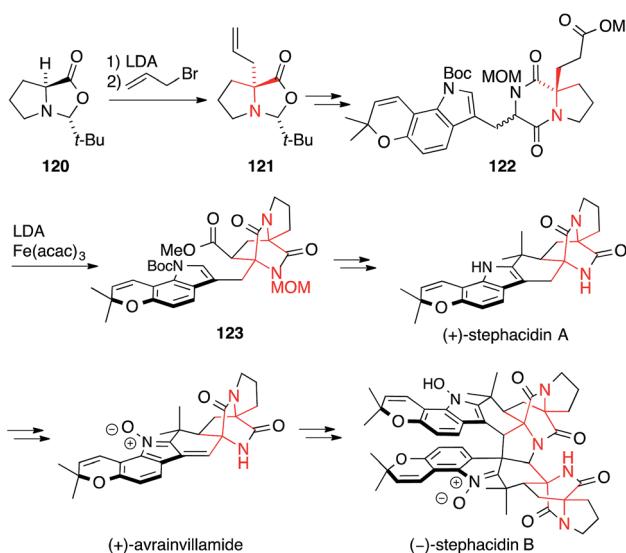
Scheme 30 Synthesis of prenylated indole alkaloids by Myers (2005). *t*-amyl = *tert*-amyl (2-methylbutyl), Boc = *tert*-butyloxycarbonyl, HFIPA = 1,1,1,3,3,3-hexafluoroisopropyl alcohol, TBDPS = *tert*-butyl-diphenylsilyl, TMSCN = trimethylsilyl cyanide.

moiety present in cyclopamine *via* retro-Michael/Michael addition. Using a similar approach, he was then able to synthesize the structurally related alkaloids stephacidin A and notoamide B.²⁰⁸

Two alkaloids closely related to notoamide B, marcfortine B and C, were synthesized by Trost using a Michael addition and a radical cyclization to set the two ATAs (not shown).^{209,210}

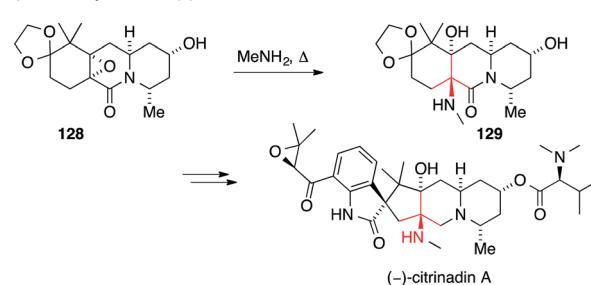
Gelsemoxonine is an indole alkaloid with an ATA that is part of a azetidine, a rare structural motif. It is also a member of the

Baran's synthesis of (+)-stephacidin A, (+)-avravillamide and (-)-stephacidin B

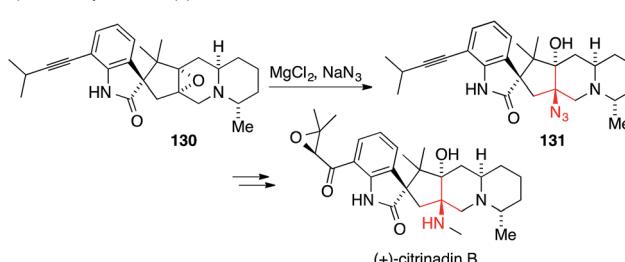


Scheme 29 Synthesis of prenylated indole alkaloids by Baran (2005). acac = acetylacetone, Boc = *tert*-butyloxycarbonyl, *t*-Bu = *tert*-butyl, LDA = lithiumdiisopropylamide.

a) Martin's synthesis of (-)-citrinadin A



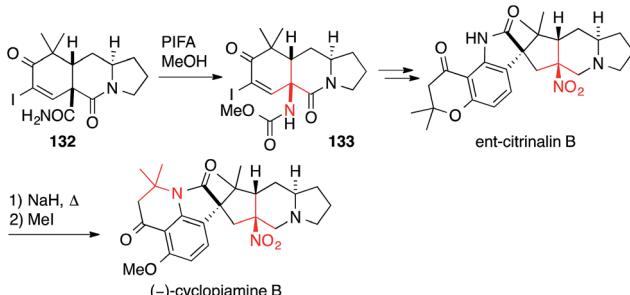
b) Wood's synthesis of (+)-citrinadin B



Scheme 31 Syntheses of prenylated indole alkaloids by Martin (2013) and Wood (2013).



Sarpong's synthesis of (−)-cyclopamine B

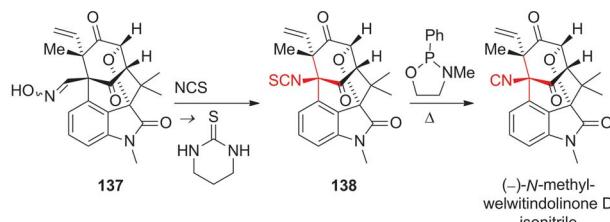
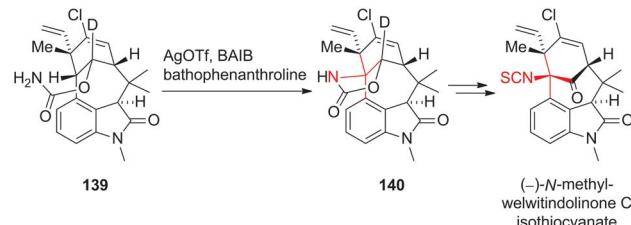


Scheme 32 Synthesis of *ent*-citrinalin B and (−)-cyclopamine B by Sarpong (2015). PIFA = (Bis(trifluoroacetoxy)iodo)benzene.

Gelsemium spirooxindole family, a large alkaloid family with highly compact, strained and complex structures, which have attracted considerable synthetic activity.^{211–213} In 2011, Fukuyama accomplished a total synthesis of gelsemoxonine that employed an intramolecular epoxide opening of 134 to install the ATA (Scheme 33a).²¹⁴ Recently, Carreira published an elegant entry to gelsemoxonine, setting the ATA 136 *via* a diastereoselective propynyllithium addition to isoxazoline 135 (Scheme 33b).²¹⁵

The welwitindolinones are another class of indole alkaloids with an ATA that is not part of the indole-derived moiety itself. The first welwitindolinone natural products (Fig. 7) were isolated by Moore in 1994 from the cyanobacteria *Hapalosiphon welwitschii* and *Westiella intracta*.²¹⁶ All welwitindolinones known so far feature a [4.3.1] bicyclic framework, which, in some cases, contains a modified ATA that bears an isothiocyanate or isonitrile functional group.^{217,218}

Being a considerable challenge for total synthesis, the welwitindolinones have become popular targets.²¹⁹ The first total synthesis of *N*-methylwelwitindolinone D isonitrile was accomplished by Rawal in 2011^{220–222} using Kim's oxime rearrangement to install the isothiocyanide (137 → 138, Scheme 34a).^{223,224} Desulfurization of 138 then gave the naturally occurring

a) Rawal's synthesis of (−)-*N*-methylwelwitindolinone D isonitrileb) Garg's synthesis of (−)-*N*-methylwelwitindolinone C isothiocyanate

Scheme 34 Syntheses of welwitindolinones by Rawal (2011) and Garg (2011). BAIB = (diacetoxymido)benzene, NCS = *N*-chlorosuccinimide, OTf = trifluoromethanesulfonate, Ph = phenyl.

isonitrile. Martin completed a synthesis that intercepts Rawal's synthesis in 2012.²²⁵

Garg's total synthesis of *N*-methylwelwitindolinone C isothiocyanate used an intramolecular Ag-mediated nitrene C,H-insertion of amide 139 as the critical step, which furnished carbamate 140 (Scheme 34b).^{226,227} To improve the regioselectivity and yield of this late stage transformation, the authors beautifully exploited the deuterium kinetic isotope effect.²²⁸

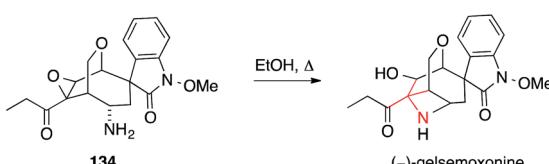
Both Rawal and Garg were able to subsequently synthesize several members of the welwitindolinone family by varying their initial strategies.^{222,228–230} In addition, Hatakeyama recently accomplished another synthesis of (−)-*N*-methylwelwitindolinone C isothiocyanate using an endgame similar to Rawal's.²³¹

Two examples of reactions which have been specifically developed to set an ATA, both explored by the Baran laboratory, are shown in Scheme 35.

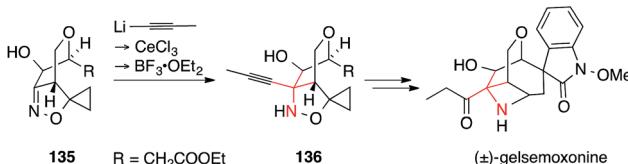
In the synthesis of chartelline C, the ATA was set *via* a cascade reaction initiated by the bromination of indole 141 at 185 °C resulting in 142 (Scheme 35a). Amide attack furnished intermediate 143, which then rearranged in a 1,5-shift to give the ring contracted spiro-β-lactam 144.^{232,233} For the synthesis of psychotrimine, a coupling of indole 147 with 2-iodoaniline (146) was developed to yield 148, which then underwent further cyclization to give 149 (Scheme 35b).²³⁴ This method was also used for the syntheses of psychotetramine,²³⁵ kapakahine B and kapakahine F.^{236,237}

Another interesting way to install an ATA in a structurally complex indole alkaloid was published by Danishefsky (Scheme 36).²³⁸ In his synthesis of the furanobisindole alkaloid phalarine, amino acid derivative 150 was treated with formaldehyde and acid to set the ATA in a diastereoselective fashion (150 → 152). It is not clear, however, whether this reaction proceeds *via* a 1,2-Wagner–Meerwein shift (possible intermediate 153) or

a) Fukuyama's synthesis of (−)-gelsemoxonine

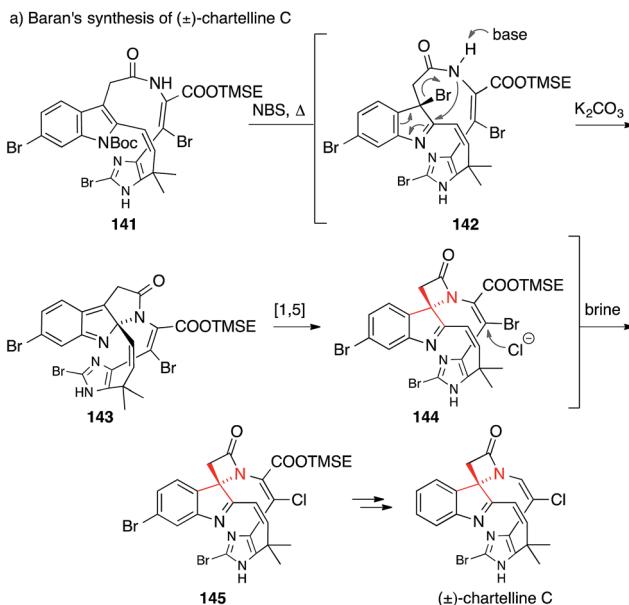
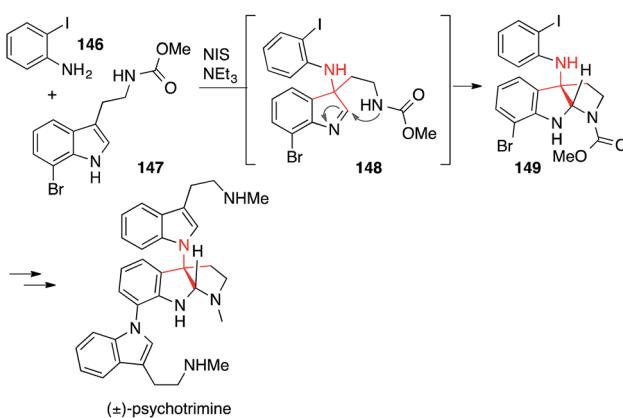


b) Carreira's synthesis of (±)-gelsemoxonine



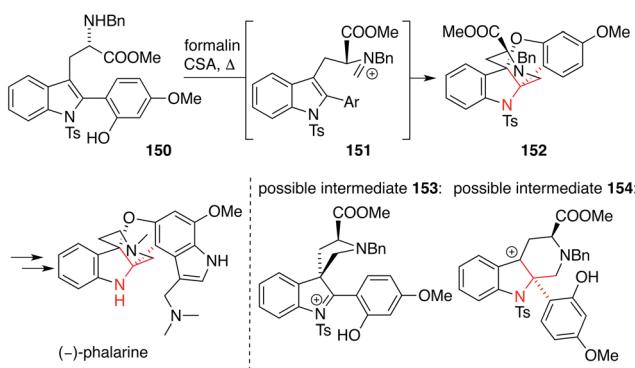
Scheme 33 Syntheses of gelsemoxonine by Fukuyama (2011) and Carreira (2013). Boc = *tert*-butyloxycarbonyl.



b) Baran's synthesis of (\pm)-psychotrimine

Scheme 35 Baran's syntheses of chartelline C (2006) and psychotrimine (2008). Bn = benzyl, Boc = tert-butyloxycarbonyl, NBS = N-bromosuccinimide, NIS = N-iodosuccinimide, TMSE = 2-(trimesilyl)ethyl.

Danishefsky's synthesis of (-)-phalarine



Scheme 36 Danishefsky's synthesis of phalarine (2010). Bn = benzyl, CSA = camphorsulfonic acid.

a Pictet–Spengler reaction (possible intermediate 154) starting from iminium intermediate 151.

9 Cephalotaxines

Due to their interesting chemical structure and antileukemic activities, the cephalotaxines, isolated from the Japanese plum yew (*Cephalotaxus harringtonii*), have emerged as popular targets for natural product synthesis (Fig. 8).²³⁹

The first synthesis of cephalotaxine itself was reported by Weinreb in 1972 (Scheme 37a).²⁴⁰ Conversion of enamine 155 into diketone 156 set the stage for a Lewis-acid catalyzed cyclization to yield tertiary amine 158 (*via* intermediate 157). Weinreb was able to synthesize cephalotaxine in six additional steps with an overall yield of 20%, setting a high bar for the following syntheses.

In 1988, Fuchs utilized an intramolecular [4+2] nitroso-Diels–Alder cycloaddition to assemble the benzazepine 161 from hydroxamic acid 159 (*via* intermediate 160, Scheme 37b).²⁴¹

Tietze published a formal asymmetric synthesis of ($-$)-cephalotaxine in 1999 that is based on palladium catalysis

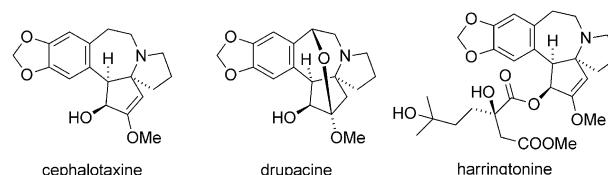
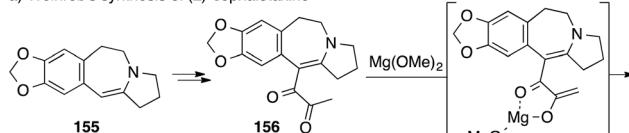
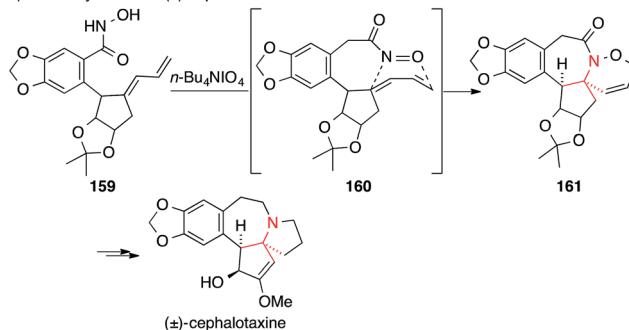
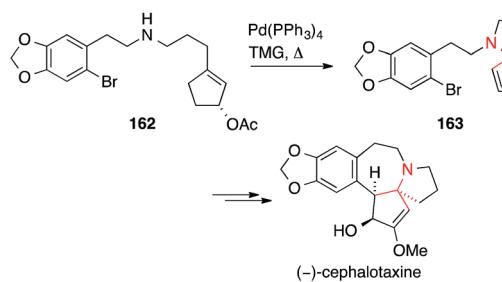


Fig. 8 Representative cephalotaxines.

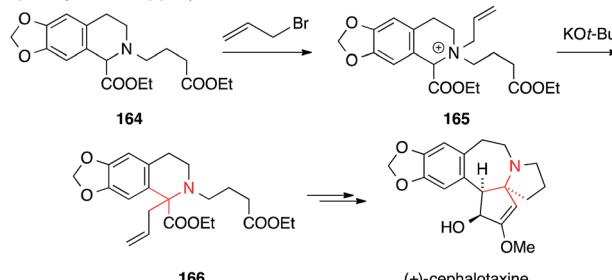
a) Weinreb's synthesis of (\pm)-cephalotaxineb) Fuchs' synthesis of (\pm)-cephalotaxine

Scheme 37 Syntheses of Cephalotaxine by Weinreb (1972) and Fuchs (1988).

a) Tietze's synthesis of (-)-cephalotaxine



b) Li's synthesis of (±)-cephalotaxine



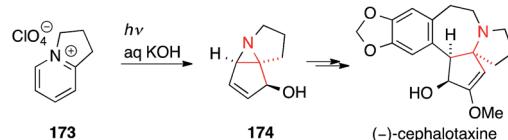
Scheme 38 Syntheses of cephalotaxine by Tietze (1999) and Li (2003).
Ac = acetyl, TMG = 1,1,3,3-tetramethylguanidine.

(Scheme 38a).²⁴² A Tsuji–Trost allylation on secondary amine 162 established the ATA and afforded 163 which was converted into cephalotaxine *via* a subsequent Heck reaction. A similar Tsuji–Trost allylation was used by Stoltz in 2007 in his synthesis of drupacine and cephalotaxine.²⁴³

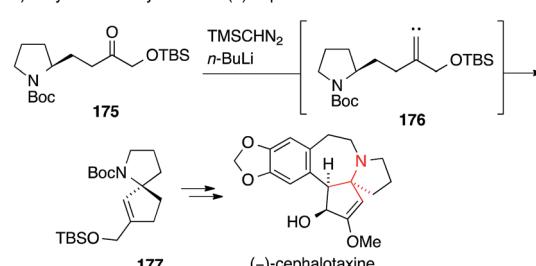
A [2,3] sigmatropic rearrangement was utilized by Li to convert quaternary ammonium ion 165 to the α -tertiary amino ester 166 (164 \rightarrow 166, Scheme 38b).²⁴⁴

A rather unusual approach for the asymmetric synthesis of (–)-cephalotaxine was pursued by Royer, who introduced the

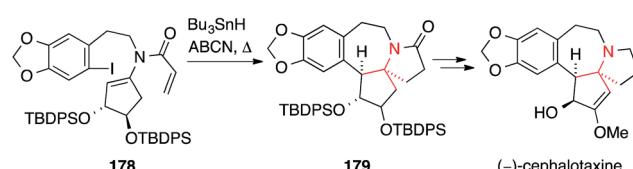
a) Mariano's formal synthesis of (–)-cephalotaxine



b) Hayes' formal synthesis of (–)-cephalotaxine

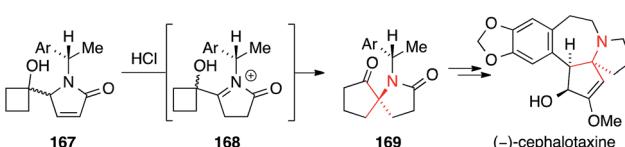


c) Ishibashi's synthesis of (–)-cephalotaxine

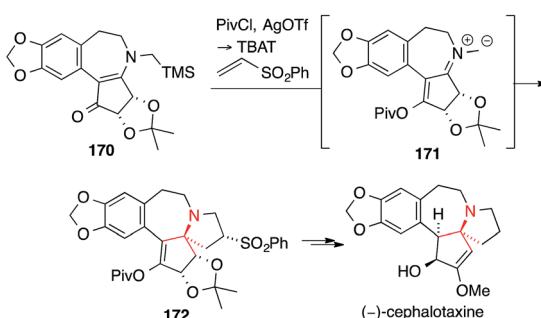


Scheme 40 Syntheses of cephalotaxine by Mariano (2006), Hayes (2008) and Ishibashi (2008). ABCN = 1,1'-azobis(cyclohexanecarbonitrile), Boc = *tert*-butyloxycarbonyl, TBS = *tert*-butyldimethylsilyl, TBDPS = *tert*-butyldiphenylsilyl, TMS = trimethylsilyl.

a) Royer's synthesis of (–)-cephalotaxine



b) Gin's synthesis of (–)-cephalotaxine



Scheme 39 Syntheses of cephalotaxines by Royer (2004) and Gin (2006). Ar = aryl, OTf = trifluoromethanesulfonate, Piv = pivaloyl, TBAT = tetrabutylammonium difluorotriphenylsilicate.

ATA on key intermediate 169 *via* semipinacol rearrangement of chiral α -hydroxyiminium 168. The latter was generated by acid-catalyzed isomerization and protonation of pyrrolinone 167 (Scheme 39a).²⁴⁵

Another synthesis was developed by Gin, who transformed vinylogous amide 170 into azomethine ylide 171 which then underwent 1,3-dipolar cycloaddition with phenyl vinyl sulfone to yield 172 (Scheme 39b).^{246,247} The unexpected yet advantageous stereochemical outcome of this cycloaddition was confirmed by X-ray analysis.

In 2006, Mariano used a photochemical cyclization of a bicyclic pyridinium ion 173 to afford aziridine 174, which was then transformed into the natural product (Scheme 40a).²⁴⁸ Hayes' synthetic route towards (–)-cephalotaxine made use of an alkylidene carbene 1,5-C,H-insertion starting from ketone 175 to furnish spiro[4.4]azanonane 177, which corresponds to the skeleton of cephalotaxine (175 \rightarrow 177, Scheme 40b).²⁴⁹

Ishibashi chose a radical cyclization approach to synthesize cephalotaxine (Scheme 40c).²⁵⁰ Treatment of aryl iodide 178 with tributyltin hydride and a radical initiator resulted in a 7-*endo* cyclization followed by a 5-*endo* cyclization that, after hydrogen transfer, yielded 179. Additional functional group manipulations allowed them to intercept an intermediate that had previously been used to synthesize (–)-cephalotaxine.²⁵¹

A host of other methods have been used for the installation of the ATA in cephalotaxines.²³⁹ These include alkylation,^{252,253} Claisen rearrangement,²⁵⁴ Michael addition,^{255,256} Schmidt



reaction,²⁵⁷ addition to an imine,²⁵⁸ transannular cyclization²⁵⁹ and oxidative rearrangement.²⁵¹

10 Erythrina alkaloids

Erythrina alkaloids were discovered at the end of the 19th century, when extracts of *Erythrina* trees were found to possess curare-like neuromuscular activities.²⁶⁰ Due to their biological activities and interesting structures (Fig. 9), several total syntheses of these natural products have been carried out and many creative ways to install ATAs have been developed in this context.²⁶¹

In 1990, the group of Ishibashi published the synthesis of (\pm) -3-demethoxyerythratidinone using an intramolecular Pummerer-like rearrangement of the enamine **180**, setting the stage for a Pictet–Spengler-type reaction (**181** \rightarrow **182**) to furnish the ATA (Scheme 41a).²⁶² Thirteen years later, the same group published an oxidative radical cyclization starting from enamine **183** to obtain the skeleton of 3-demethoxyerythratidinone **182** (Scheme 41b).²⁶³

Tsuda's approach featured an intermolecular photochemical [2+2] cyclization to install the ATA, starting from bicyclic **184** and diene **185** (Scheme 41c). In the following steps, a ring expansion of the four-membered ring in **186** furnished the six-membered ring by a formal 1,3-migration of a vinylcyclobutane, affording the scaffold of erysotrine.²⁶⁴

Funk accomplished the synthesis of isophellibiline *via* an approach that relies on pericyclic reactions (Scheme 42a).²⁶⁵ Heating of dioxine **187** resulted in retro-Diels–Alder reaction to afford dehydroalanine derivative **188**, which then underwent intramolecular [4+2] cycloaddition to yield lactam **189**. The latter was converted into isophellibiline in a few steps.

Recently, Sarpong developed a new methodology to furnish ATAs and applied it to the synthesis of cocculolidine (Scheme 42b).²⁶⁶ Propargylic alcohol **190** underwent cycloisomerization upon heating to form benz[g]indolizinone **191** which was then transformed to cocculolidine in two additional steps.

A short and elegant synthesis of 3-demethoxyerythratidinone was accomplished by Streuff in 2015 (Scheme 43a). He used a titanium(III)-catalyzed reductive Umpolung (**192** \rightarrow **194**) to assemble the 1,1-disubstituted tetrahydroisoquinoline core of the *Erythrina* alkaloids.²⁶⁷ In the same year, Ciufolini set the same ATA *via* an oxidative dearomatizative cyclization of an oxazoline **195**, yielding spiropiperidine **197** presumably *via* intermediate **196** (Scheme 43b).^{268,269}

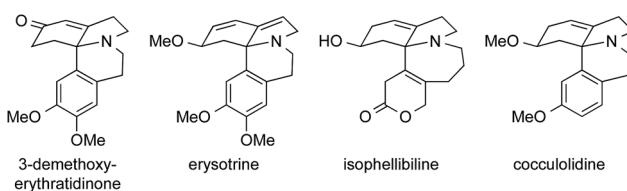
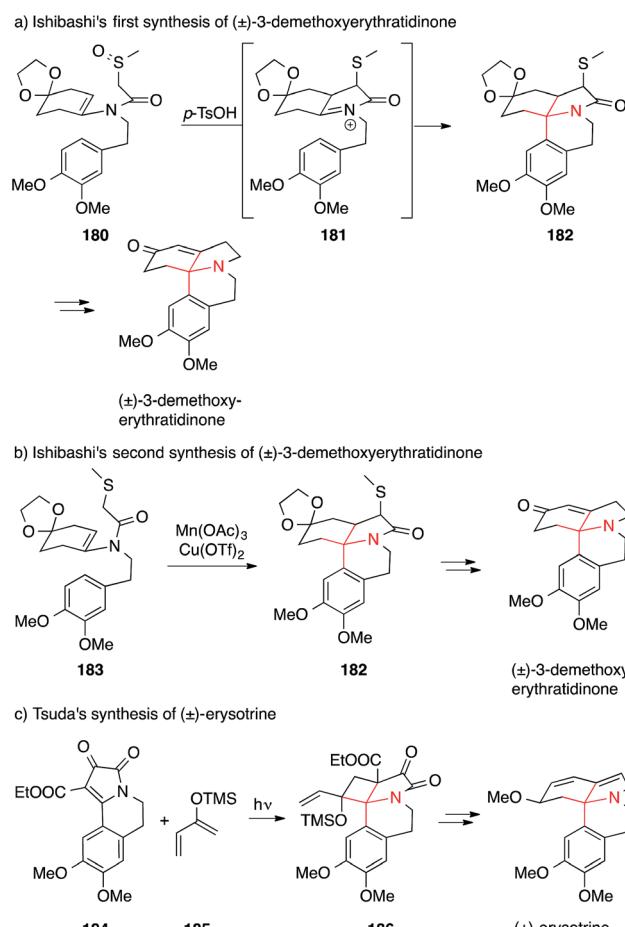
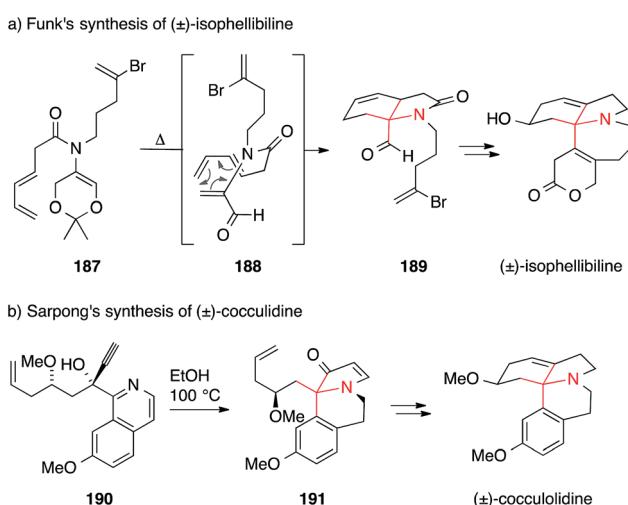


Fig. 9 Representative *Erythrina* alkaloids.



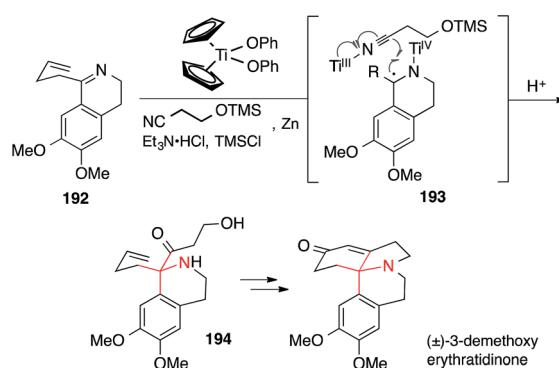
Scheme 41 Syntheses of *Erythrina* alkaloids by Ishibashi (1990 and 2003) and Tsuda (1992). Ac = acetyl, OTf = trifluoromethanesulfonate, TMS = trimethylsilyl, *p*-TsOH = *para*-toluenesulfonic acid.



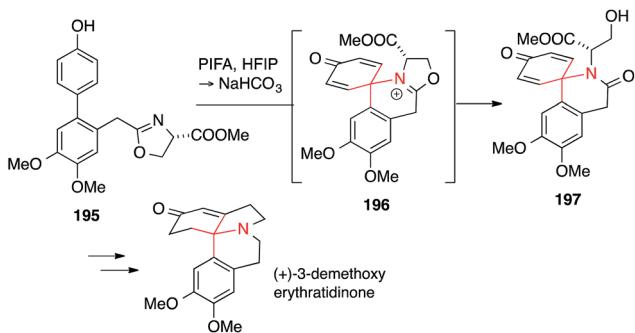
Scheme 42 Syntheses of *Erythrina* alkaloids by Funk (2012) and Sarpong (2013).



a) Streuff's synthesis of (\pm) -3-demethoxyerythridinone



b) Ciufolini's synthesis of $(+)$ -3-demethoxyerythridinone



Scheme 43 Syntheses of *Erythrina* alkaloids by Streuff (2015) and Ciufolini (2015). HFIP = 1,1,1,3,3,3-hexafluoroisopropyl alcohol, PIFA = (bis(trifluoroacetoxy)iodo)benzene, TMS = trimethylsilyl.

Other methods used to install the ATA in *Erythrina* alkaloids include 1,2-addition of organometallic reagents to sulfinimines or iminium ions,^{270–272} Schmidt reaction,²⁷³ Diels–Alder reactions,²⁷⁴ Michael addition,²⁷⁵ *N*-acyliminium Pictet–Spengler reaction^{276–278} and Heck reaction.²⁷⁹

11 Indolizidine and quinolizidine alkaloids

A range of alkaloids that belong to the indolizidine and quinolizidine structural class feature an ATA in their carbon skeleton.²⁸⁰ They include natural products as diverse as the cylindricines,^{280,281} FR901483,²⁸² himandrines, lepidiformines²⁸³ and halichlorine²⁸⁴ (Fig. 10).

The first synthesis of cylindricine alkaloids (*viz.* cylindricine A, D and E) was accomplished by Snider utilizing a double Michael addition of ammonia to divinylketone 198 which gave the ATA 199, a direct precursor of cylindricine A (Scheme 44a).²⁸⁵ Variations of this approach have been used several times in the synthesis of cylindricines.²⁸⁶

In 2003, Padwa published a synthesis featuring a Michael addition/dipolar cycloaddition cascade between butadiene 201 and oxime 200 to form 203 *via* intermediate 202 (Scheme 44b).²⁸⁷

The Hsung synthesis of enantiomerically pure cylindricine C relies on a nucleophilic attack of a diene on *N*-acyliminium ion 205 starting from ketone 204 (Scheme 45).^{288,289} This vinylogous

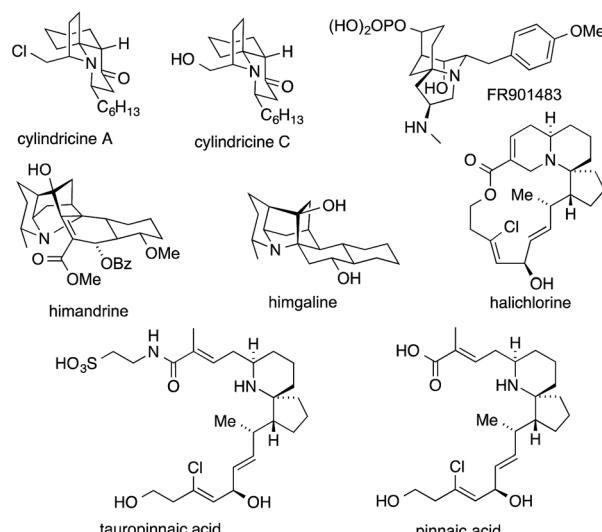


Fig. 10 Selection of quinolizidine alkaloids, including halichlorine and the closely related alkaloids taupinnic acid and pinnic acid.

aza-Prins approach was based on a synthesis published by Kibayashi in 2005.²⁹⁰

Additional strategies to synthesize the ATA in cylindricine alkaloids involve mainly Michael additions,^{286,291,292} Grignard additions to an imine,²⁹³ a cycloaddition of an alkyne to a pyrrole derivative,²⁹⁴ and carboazidation.²⁹⁵

FR901483, an alkaloid isolated from the fermentation broth of a *Cladobotryum* species with an intricate tricyclic structure,²⁸² proved to be an equally popular synthetic target. A biomimetic approach was employed by Sorensen in his enantioselective synthesis (Scheme 46a).²⁹⁶

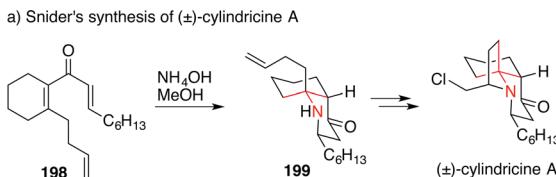
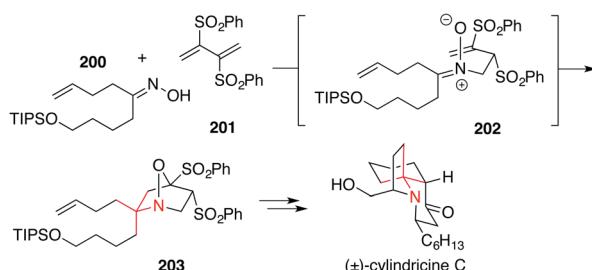
The oxidative azaspiroannulation of amine 207 promoted by (diacetoxymido)benzene resulted in the formation of spiroamine 208, an intermediate on the way to the natural product. The same year, Ciufolini set the ATA *via* a closely related oxidative spiroannulation (not shown, for an example of the methodology see Scheme 43b).²⁹⁷

An alternative to this strategy was found by Wang.²⁹⁸ In this case, an Aubé–Schmidt reaction of azide 209 provided access to lactam 212, featuring the ATA of FR901483 (209 → 212, Scheme 46b). Additional synthetic strategies to set the ATA in FR901483 include a triple Michael addition,²⁹⁹ a one-pot bisalkylation,^{300,301} an aza-Cope rearrangement/Mannich cyclization^{302,303} and an oxidative dearomatization.²⁹⁷

The members of the galbulimima alkaloid family, such as himgaline and himandrine, also possess an ATA-containing quinolizidine core (Fig. 10). Exploring a biosynthetic hypothesis, Chackalamannil used an intramolecular Michael addition to convert GB 13 to himgaline *via* ketone 213 (Scheme 47a).^{304,305} In an interesting variation of this approach, Movassaghi converted enone 214 *via* its α -chloroester 215 to hexacyclic amine 216, which could then be transformed into himandrine (Scheme 47b).³⁰⁴

In 1996, Uemura disclosed a small series of unusual marine alkaloids featuring ATAs. One of these compounds,

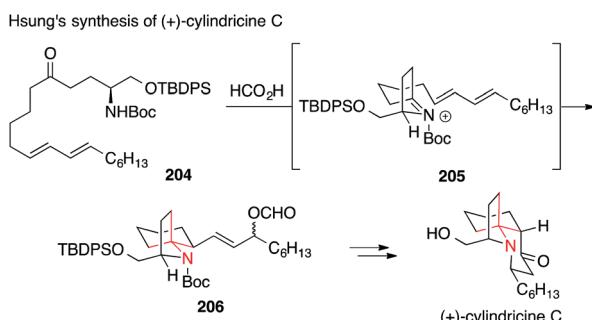


b) Padwa's synthesis of (\pm)-cylindricine CScheme 44 Syntheses of cylindricines by Snider (1997) and Padwa (2003). Boc = *tert*-butyloxycarbonyl, TBDS = *tert*-butyldiphenylsilyl.

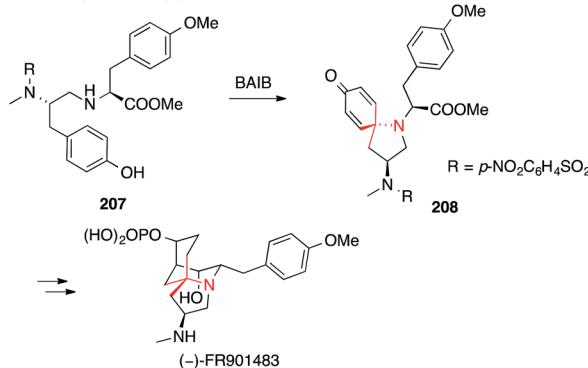
halichlorine, was isolated from the marine sponge *Halichondria okadai* and was found to selectively inhibit the induction of vascular cell adhesion molecule-1 (VCAM-1).³⁰⁶ Pinnaic acid and tauropinnaic acid were recovered from bivalve *Pinna muricata*.^{307,308} All three molecules present a challenging 6-aza-spiro [4.5]decane core containing the ATA. The latter two lack a quinolizidine moiety, but are included in this chapter due to their close structural relationship.

Danishefsky and Trauner were the first to report the synthesis of (+)-halichlorine in 1999³⁰⁹ followed by a synthesis of pinnaic acid in 2001 (Scheme 48).^{310,311} They used Meyers' lactam 217 as a chiral precursor, which was combined with allyl-trimethylsilane in a Sakurai reaction to install the ATA in 218. Intermediate 219 could be diversified to reach both halichlorine and pinnaic acid. These syntheses established the absolute configuration of halichlorine and confirmed the stereochemistry at C-14 and C-17 of pinnaic acid.

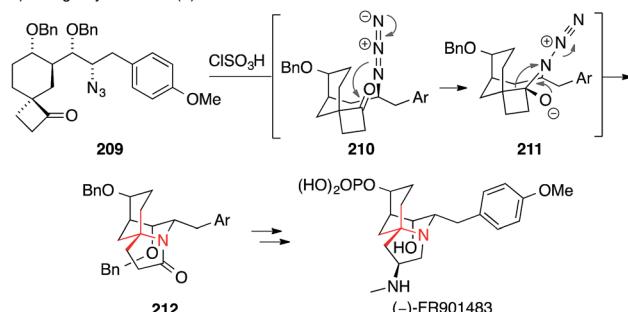
A related approach employing a different type of *N*-acyl iminium ion was used by Heathcock in 2004 for the synthesis of halichlorine, pinnaic acid and tauropinnaic acid (Scheme 49a).³¹² Treatment of carbamate acetal 220 with allyl trimethylsilane and titanium tetrachloride furnished ATA-bearing

Scheme 45 Synthesis of cylindricine C by Hsung (2004). Boc = *tert*-butyloxycarbonyl, TBDS = *tert*-butyldiphenylsilyl.

a) Sorensen's synthesis of (-)-FR901483



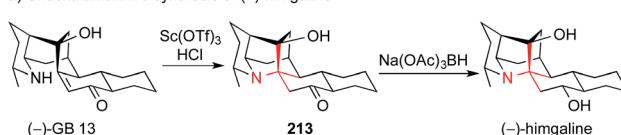
b) Wang's synthesis of (-)-FR901483

Scheme 46 Syntheses of FR901483 by Sorensen (2000) and Wang (2012). Ar = aryl, BAIB = (diacetoxymido)benzene, Bn = benzyl, TBDS = *tert*-butyldiphenylsilyl.

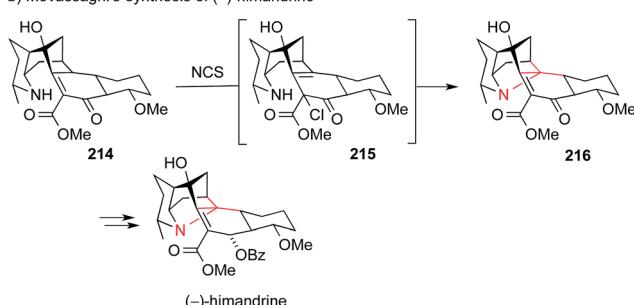
carbamate 221 with a high degree of stereoselectivity. This key intermediate could be transformed into all three natural products.

In 2007, Arimoto reported his version of an asymmetric synthesis of pinnaic acid using a Beckmann rearrangement to install the ATA (Scheme 49b).³¹³ The enantiomerically pure bicyclic ketone 222 was treated with a bulky hydroxylamine

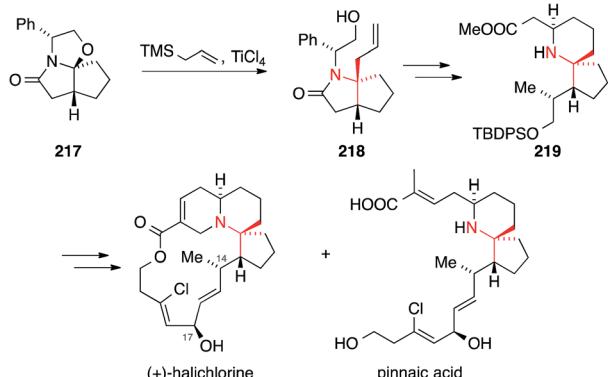
a) Chackalamannil's synthesis of (-)-himgaline



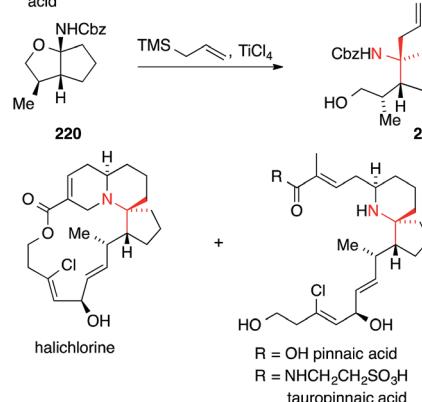
b) Movassaghi's synthesis of (-)-himandrine

Scheme 47 Synthesis of galbulimima alkaloids by Chackalamannil (2009) and Movassaghi (2009). Ac = acetyl, Bz = benzoyl, NCS = *N*-chlorosuccinimide, OTf = trifluoromethanesulfonate.

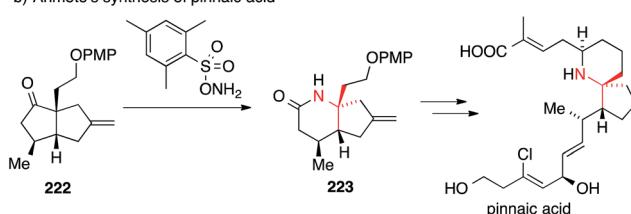
Danishefsky-Trauner synthesis of (+)-halichlorine and pinnaic acid

Scheme 48 Synthesis of halichlorine and pinnaic acid by Danishefsky and Trauner (1999). TBDPS = *tert*-butyldiphenylsilyl, TMS = trimethylsilyl.

a) Heathcock's synthesis of (±)-halichlorine, (±)-pinnaic acid and (±)-tauropinnaic acid



b) Arimoto's synthesis of pinnaic acid

Scheme 49 Syntheses of halichlorine, pinnaic acid and tauropinnaic acid by Heathcock (2004) and Arimoto (2007). Cbz = benzyloxycarbonyl, PMP = *p*-methoxyphenyl, TMS = trimethylsilyl.

reagent to afford the desired lactam 223, which was then converted into the natural product.

12 Lactacystine and salinosporamide A

In 1991, Omura isolated the unusual natural product lactacystin from *Streptomyces* sp. OM-6519 and identified it as a proteasome inhibitor (Fig. 11).^{314,315} A structurally related β -lactone, salinosporamide A, which shows similar biological activity, was subsequently isolated from a marine bacterium, *Salinispora tropica*.³¹⁶

Both compounds possess a densely functionalized γ -lactam core with three contiguous stereocenters, one of which is of the

ATA type. Their significant biological activity has stimulated a large number of total syntheses,³¹⁷ and a variety of methods for the installation of the ATA motif have been applied.^{318–345}

In pioneering work, Corey reported five total syntheses of lactacystin between 1992 and 1998.^{318–320,326,327} The Corey group showed that the ATA can be installed using an aldol addition of α -amino acid derivative 224 (*via* intermediate 225, Scheme 50a). Other groups also contributed to this field in the 1990s.^{321–325,328} In most cases, the strategy applied for the installation of the ATA motif involved an alkylation or aldol reaction of an α -amino acid derivative.^{318–322,324,326–328} By contrast, Shibasaki introduced the ATA with a catalytic enantioselective Strecker reaction (Scheme 50b).^{317,334} In this work, phosphinoylimine 226 was converted to aminonitrile 227 using a gadolinium catalyst and the chiral ligand A.

Another unusual approach was taken by Wardrop³⁴⁶ in his formal synthesis and Hayes³³⁸ in his total synthesis of lactacystin (Scheme 50c). Both groups explored an intramolecular carbene insertion into a C,H-bond to form the five-membered heterocyclic core. Hayes converted the enantiomerically pure

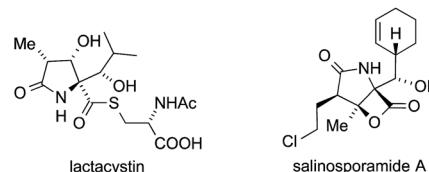
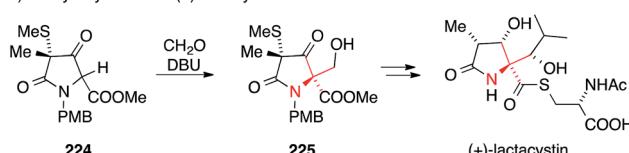
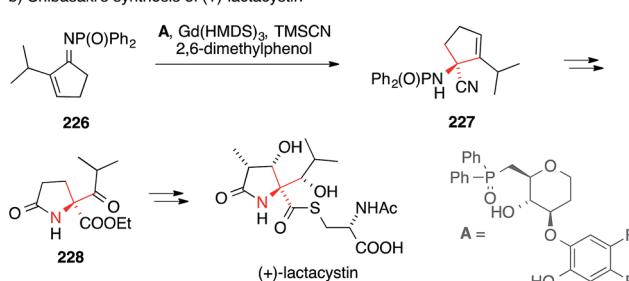


Fig. 11 Lactacystin and salinosporamide A.

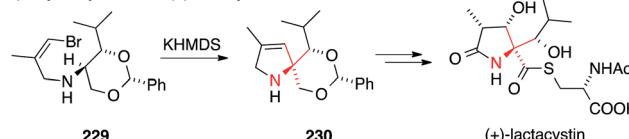
a) Corey's synthesis of (+)-lactacystin



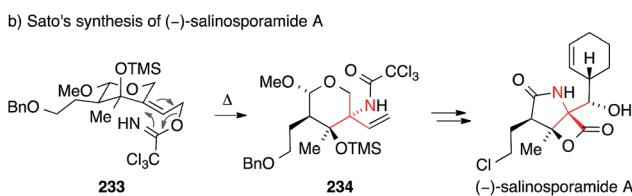
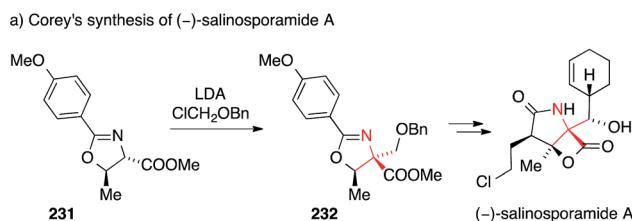
b) Shibasaki's synthesis of (+)-lactacystin



c) Hayes' synthesis of (+)-lactacystin



Scheme 50 Syntheses of lactacystin by Corey (1992), Shibasaki (2006) and Hayes (2008). Ac = acetyl, DBU = 1,8-diazabicycloundec-7-ene, HMDS = bis(trimethylsilyl)amine, PMB = paramethoxybenzyl, TMS = trimethylsilyl.



Scheme 51 Syntheses of salinosporamide A by Corey (2004) and Sato (2011). Bn = benzyl, LDA = lithium diisopropylamide, TMS = trimethylsilyl

v vinyl bromide 229 to the corresponding vinylidene carbene, which underwent cyclization to afford 230 in high yield.

The first synthesis of salinosporamide was reported by Corey in 2004.^{330,331} In this case, the ATA was installed by alkylation of threonine-derived oxazoline 231 with chloromethyl benzyl ether (*via* intermediate 232, Scheme 51a).

A more recent synthesis of salinosporamide A published by Sato and Chid uses a stereoselective Overman rearrangement to install the ATA (Scheme 51b).³⁴³ Heating of the highly functionalized trichloroacetimidate 233 provided trichloroacetamide 234 as a key intermediate.

Other syntheses of salinosporamide and lactacystine used different strategies for the formation of the ATA, such as intramolecular aldol reactions,^{336,337} intramolecular acylations,³³² acid catalyzed cyclizations of malonates,^{335,339} indium-catalyzed Conia-ene reactions,³⁴¹ C_H-alkynylation³⁴⁷ and enzymatic desymmetrizations of prochiral ATAs.³⁴⁰

13 Manzacidins

The manzacidins, a small family of bromopyrrole alkaloids, have attracted considerable attention from the synthetic community despite, or maybe because of their relatively simple structures. Manzacidins A-C (Fig. 12) were first isolated from the Okinawan sponge *Hymeniacidon* sp. by Kobayashi in 1991,³⁴⁸ followed by the isolation of manzacidin D from the 'living fossil' sponge *Astrosclera willeyana*³⁴⁹ and *N*-methylmanzacidin C from *Axinella brevistyla*.³⁵⁰

In 2000, Ohfune reported the synthesis of manzacidins A and C *via* a Strecker reaction and assigned the absolute configuration of these natural products (not shown).³⁵¹ In 2002, DuBois synthesized manzazidin C using an elegant oxidative C,H-insertion that involved sulfamate 235 (*via* intermediate 236, Scheme 52a).³⁵² One year later, he used a similar strategy to set the ATA in tetrodotoxin (see chapter 14, Scheme 54c).³⁵³

Leighton accomplished the synthesis of manzacidin C employing their asymmetric silane-promoted [3+2]

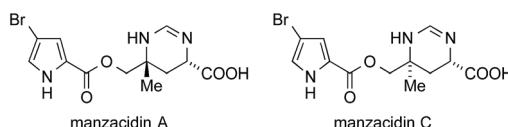
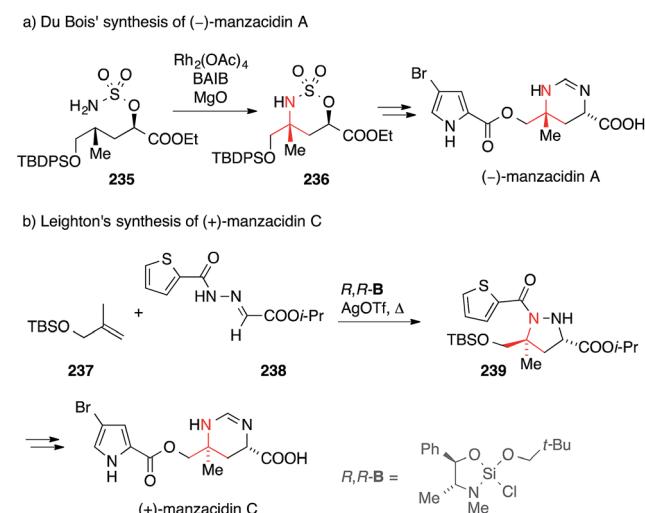


Fig. 12 Manzacidin A and C.

cycloaddition methodology (Scheme 52b).³⁵⁴ Exposure of alkene 237 and hydrazone 238 to chiral silane *R,R*-B gave pyrazolidine 239, thus setting both stereocenters of the target molecule, including the ATA, in a single step. Intermediate 239 was subsequently converted to manzacidin C via reductive N,N-bond cleavage.

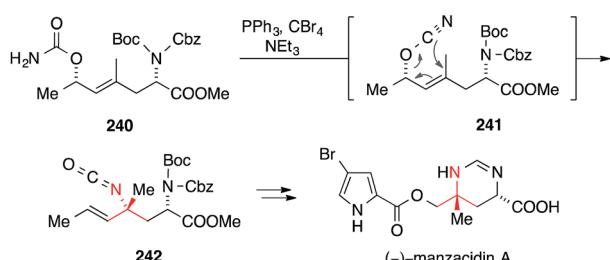
A more recent formal synthesis of manzacidins A and C, published by Ichikawa, features a rare allyl cyanate/isocyanate rearrangement as the key step (Scheme 53).³⁵⁵ To this end, he synthesized carbamate **240**, which was converted to allyl cyanate **241** by *in situ* dehydration. The subsequent rearrangement with chirality transfer gave isocyanate **242**, which was then transformed into manzacidin A. The synthesis of manzacidin C was accomplished analogously from a diastereoisomer of carbamate **240**.³⁵⁵

Several other synthetic approaches toward these molecules have been reported. These strategies for the installation of the ATA moiety involve diastereoselective nitrene insertion,³⁵² 1,3-dipolar cycloaddition,^{356,357} Hofmann rearrangement,³⁵⁸ dia stereoselective iodocyclization,^{359,360} Grignard addition to an imine³⁶¹ and a variety of other methods.^{362–366} Indeed, manzacidines remain targets of great interest for synthetic chemists. In 2015, Inoue published a synthesis of manzacidin A using a radical-based decarbonylative coupling (not shown).³⁶⁷ Recently the relative stereochemistry of manzacidin B, which possesses an additional stereocenter, was revised using total synthesis.^{362,363,365}



Scheme 52 Syntheses of manzacidins by DuBois (2002) and Leighton (2008). Ac = acetyl, BAIB = (diacetoxyiodo)benzene, OTf = trifluoromethanesulfonate, *i*-Pr = *iso*-propyl, TBDPS = *tert*-butyldiphenylsilyl.

Ichikawa's synthesis of (-)-manzacidin A

Scheme 53 Synthesis of manzacidin A by Ichikawa (2012). Boc = *tert*-butyloxycarbonyl, Cbz = benzyloxycarbonyl.

14 Tetrodotoxin

Tetrodotoxin (TTX) was first isolated from the *Fugu* puffer fish in 1909.^{368,369} Its structure was independently reported by Hirata-Goto,³⁷⁰ Tsuda³⁷¹ and Woodward³⁷² in the 1960s. Their assignment was confirmed by X-ray crystallography, which also established the absolute configuration of the molecule.^{373,374} TTX features a highly functionalized heteroadamantane framework that contains an *ortho*-acid and is fused to a cyclic guanidinium moiety *via* an ATA motif. The molecule is an extremely powerful and selective blocker of voltage-gated sodium channels and is widely used as a research tool in neuroscience.³⁷⁵⁻³⁷⁹ Due to its intriguing structure and bioactivity, attempts to synthesize TTX have been made from an early stage and activity in this field has recently increased significantly.³⁸⁰

The first total synthesis of TTX was accomplished by Kishi in 1972 (Scheme 54a).³⁸¹⁻³⁸⁴ In his approach, the ATA was formed using a Beckmann rearrangement of oxime 243, which was synthesized using a regioselective Diels–Alder reaction. The resulting key intermediate 244 was converted into TTX using a series of stereoselective redox transformations, ring cleavage and the installation of the cyclic guanidine with newly developed methodology. Although the Kishi synthesis was not enantioselective, it still stands as one of the strategically most elegant approaches to a natural product featuring an ATA motif.

After a 30 year lull, Isobe published the first enantioselective synthesis of TTX wherein the ATA motif was installed with a stereoselective Overman rearrangement (Scheme 54b).³⁸⁵⁻³⁸⁷ To this end, an allylic alcohol was converted to trichloroacetimidate 245, which underwent rearrangement to yield trichloroacetamide 246. Compound 246 bears all the carbon atoms of TTX and could be converted into the natural product in a series of steps.

Shortly thereafter, DuBois developed an enantioselective approach to TTX that involved his signature nitrene insertion chemistry (Scheme 54c).³⁸⁸ Exposure of the key intermediate, carbamate 247, to a hypervalent iodine reagent and magnesium oxide in the presence of a rhodium catalyst led to the formation of oxazolidinone 248, which bears the ATA motif. Insertion into other possible C,H-bonds was largely avoided through careful engineering of the substrate.

15 Miscellaneous alkaloids

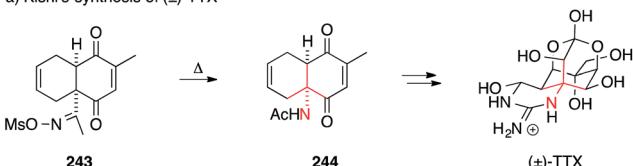
ATA's occur in many other alkaloids that cannot easily be categorized along the biosynthetic and structural lines shown above. An example is gracilamine, which was isolated in 2005 by Ünver and Kaya from the Amaryllidaceae species *Galanthus gracilis*.³⁸⁸ In 2012, the first synthesis of gracilamine was disclosed by Ma (Scheme 55a).³⁸⁹ It relies on a potentially biomimetic, stereoselective and intramolecular [3+2] cycloaddition, transforming 249 into the highly functionalized pyrrolidine 250.

In a recent synthesis, Gao set the ATA *via* an intramolecular Mannich annulation (Scheme 55b).³⁹⁰ First, α -ketoester 252 was condensed with amine 251. The resulting iminium ion 253 then underwent a diastereoselective Mannich reaction to furnish the hexacyclic scaffold 254 of gracilamine.

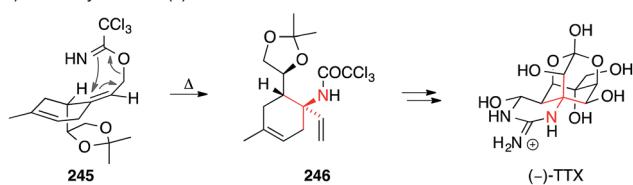
The amathaspiramides A-F are a family of marine alkaloids isolated from the bryozoan *Amathia wilsoni* in 1999 (Fig. 13).³⁹¹ They feature an unusual spirocyclic core consisting of a pyrrolidine fused to a pyrrolidinone moiety.³⁹¹

The first total synthesis of a member of this family, *viz.* amathaspiramide F, was disclosed by Trauner in 2002 (Scheme 56a).³⁹² In this work, the proline-derived *N,N*-acetal 255 was converted to the corresponding silyl ketene acetal, which underwent a diastereoselective Michael addition to the nitro olefin 256, establishing the ATA of 257. Subsequently, Ohfune published his approach to amathaspiramide F which utilizes an enolate Claisen rearrangement for the same purpose (not shown).³⁹³

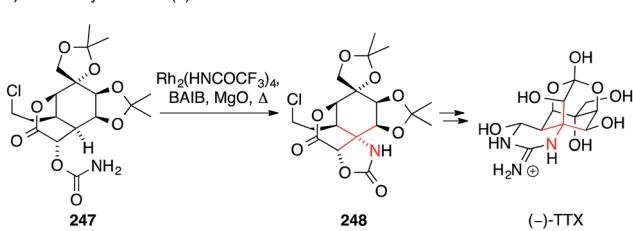
In 2012, Fukuyama reported the asymmetric synthesis of the entire amathaspiramide family (Scheme 56b).³⁹⁴ In their work,

a) Kishi's synthesis of (\pm)-TTX

b) Isobe's synthesis of (-)-TTX

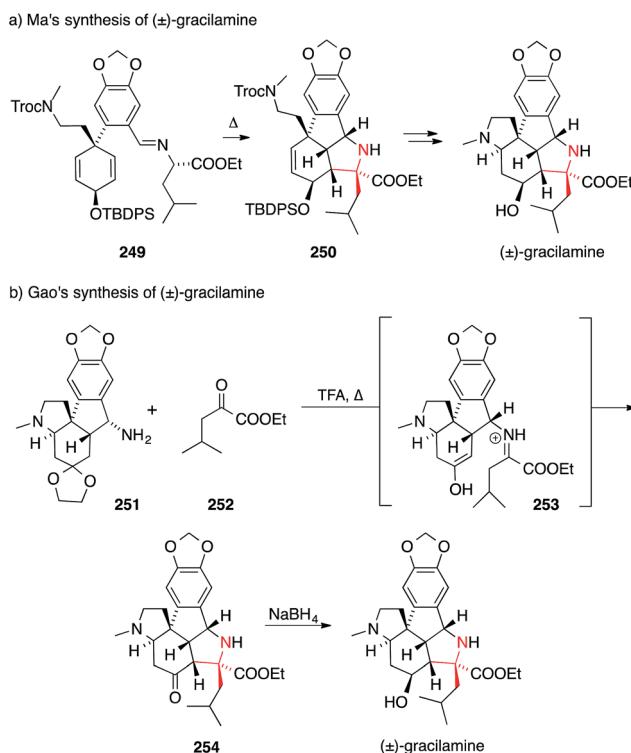


c) DuBois' synthesis of (-)-TTX



Scheme 54 Syntheses of tetrodotoxin (TTX) by Kishi (1972), Isobe (2003) and DuBois (2003). Ac = acetyl, BAIB = (diacetoxymido)benzene, Ms = methanesulfonyl, Ph = phenyl.





Scheme 55 Syntheses of gracilamine by Ma (2012) and Gao (2014). TBDPS = *tert*-butyldiphenylsilyl, Troc = 2,2,2-trichlorethoxycarbonyl, TFA = trifluoroacetic acid.

the benzyl ester **258** bearing a quaternary stereocenter was first deprotected and the resulting acid converted to the corresponding amine *via* Curtius rearrangement. After hydrolysis of the resulting isocyanate, the intermediate amino ester underwent cyclization to afford the pyrrolidinone **259**, which could be converted into all members of the family.

In Tambar's asymmetric synthesis of amanthaspiramide F, proline derivative **260** underwent a palladium-catalyzed allylic substitution of carbonate **261** to yield an intermediary quaternary ammonium ion **262**, which, after deprotonation, engaged in a stereoselective [2,3] Stevens rearrangement *via* **263** (Scheme 57a).³⁹⁵ The resulting ATA-containing pyrrolidine **264** was converted into amathaspiramide F.

More recently, Lee used a formal [3+2] cycloaddition between lithium(trimethylsilyl)diazomethane **266** and α,β -unsaturated ester **265** to set the ATA in amanthaspiramide C.

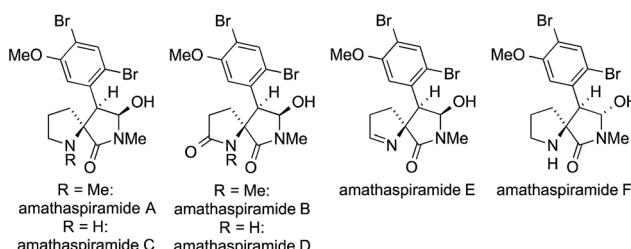
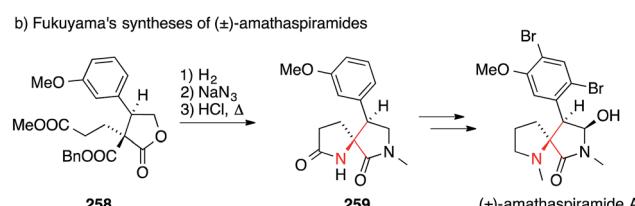
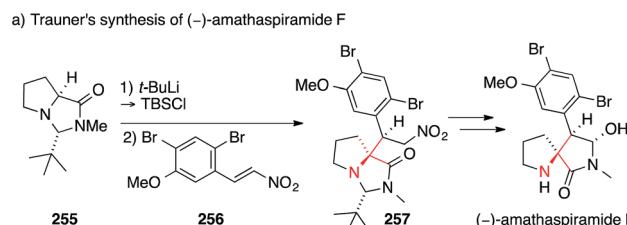
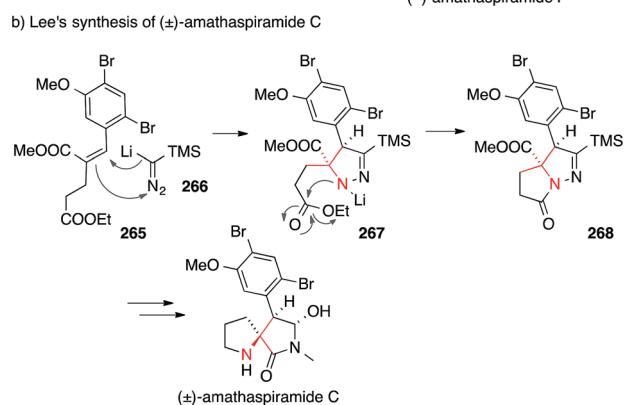
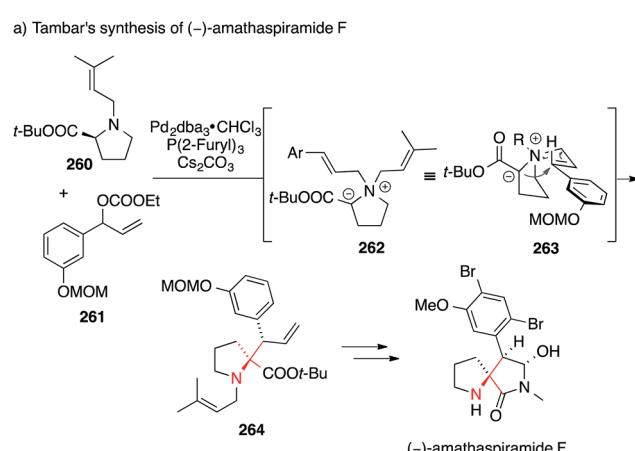


Fig. 13 The amathaspiramides A–F.



Scheme 56 Syntheses of amathaspiramides by Trauner (2002) and Fukuyama (2012). Bn = benzyl, *t*-Bu = *tert*-butyl, TBS = *tert*-butyldimethylsilyl

(via intermediate 267, Scheme 57b).³⁹⁶ The N,N-bond in pyrazoline 268 was cleaved by treatment with *p*-TsOH and additional transformations led to the total synthesis of amathaspiramide C and the formal synthesis of all the other amathaspiramides.



Scheme 57 Syntheses of amathaspiramides by Tambar (2013) and Lee (2015). *t*-Bu = *tert*-butyl, dba = dibenzylideneacetone, MOM = methoxymethyl, TMS = trimethylsilyl.

16 Conclusions

Herein, we have provided a survey of syntheses that feature the installation of an α -tertiary amine (ATA) as a common thread. This structural motif is widespread amongst alkaloids and has physicochemical consequences, such as increased lipophilicity and chromatographic mobility that distinguishes its bearers from other basic amines. Since ATAs also occur in drug candidates and building blocks for functional materials, our review is intended to provide a useful reference for medicinal chemists and colleagues active in the material sciences. It may also provide a baseline for the development of additional and hopefully more efficient methods for the synthesis of target molecules containing α -tertiary amines.

17 Acknowledgements

We thank the Deutsche Forschungsgemeinschaft (SFB 749) for financial support. We are also grateful to the Deutsche Telekom Foundation (Ph.D. Scholarship to N. V.). We thank Felix Hartmann, Benjamin Williams, Daniel W. Terwilliger, Dr Henry Toombs Ruane and David Konrad for helpful discussions and for proofreading the manuscript.

18 Notes and references

- 1 G. A. Cordell, *The Alkaloids*, Elsevier, London, 1998.
- 2 P. M. Dewick, *Medicinal Natural Products; A Biosynthetic Approach*, Wiley-VCH, Weinheim, third edn, 2009.
- 3 M. Wink, in *Studies in Natural Products Chemistry*, Elsevier, 2000, vol. 21, ch. Interference of Alkaloids with Neuroreceptors and Ion Channels, pp. 3–122.
- 4 G. A. Cordell, M. L. Quinn-Beattie and N. R. Farnsworth, *Phytother. Res.*, 2001, **15**, 183–205.
- 5 J. Clayden, M. Donnard, J. Lefranc and D. J. Tetlow, *Chem. Commun.*, 2011, **47**, 4624–4639.
- 6 G. Dake, *Tetrahedron*, 2006, **62**, 3467–3492.
- 7 S. A. A. El Bialy, H. Braun and L. F. Tietze, *Synthesis*, 2004, 2249–2262.
- 8 S. H. Kang, S. Y. Kang, H.-S. Lee and A. J. Buglass, *Chem. Rev.*, 2005, **105**, 4537–4558.
- 9 A. Sharma and J. F. Hartwig, *Nature*, 2015, **517**, 600–604.
- 10 L. Huang, M. Arndt, K. Gooßen, H. Heydt and L. J. Goosßen, *Chem. Rev.*, 2015, **115**, 2596–2697.
- 11 A. Glisan King and J. Meinwald, *Chem. Rev.*, 1996, **96**, 1105–1122.
- 12 P. Laurent, B. Lebrun, J.-C. Braekman, D. Daloz and J. M. Pasteels, *Tetrahedron*, 2001, **57**, 3403–3412.
- 13 R. Robinson, *J. Chem. Soc., Trans.*, 1917, **111**, 762–768.
- 14 B. K. Blount and R. Robinson, *J. Chem. Soc.*, 1933, 1511–1512.
- 15 C. Schöpf and G. Lehmann, *Liebigs Ann. Chem.*, 1935, **518**, 1–37.
- 16 K. Alder, H. Betzing, R. Kuth and H.-A. Dortmann, *Liebigs Ann. Chem.*, 1959, **620**, 73–87.
- 17 R. I. Longley and W. S. Emerson, *J. Am. Chem. Soc.*, 1950, **72**, 3079–3081.
- 18 K. Alder and H. A. Dortmann, *Chem. Ber.*, 1953, **86**, 1544–1554.
- 19 K. Alder, H. Wirtz and H. Koppelberg, *Liebigs Ann. Chem.*, 1956, **601**, 138–154.
- 20 B. Tursch, C. Chome, J. C. Braekman and D. Daloz, *Bull. Soc. Chim. Belg.*, 1973, **82**, 699–703.
- 21 D. H. Gnecco Medina, D. S. Grierson and H.-P. Husson, *Tetrahedron Lett.*, 1983, **24**, 2099–2102.
- 22 C. Yue, J. Royer and H.-P. Husson, *J. Org. Chem.*, 1992, **57**, 4211–4214.
- 23 M. F. Mechelke and A. I. Meyers, *Tetrahedron Lett.*, 2000, **41**, 4339–4342.
- 24 E. C. Davison, A. B. Holmes and I. T. Forbes, *Tetrahedron Lett.*, 1995, **36**, 9047–9050.
- 25 T. Itoh, N. Yamazaki and C. Kibayashi, *Org. Lett.*, 2002, **4**, 2469–2472.
- 26 R. K. Hill and L. A. Renbaum, *Tetrahedron*, 1982, **38**, 1959–1963.
- 27 T. C. Coombs, Y. Zhang, E. C. Garnier-Amblard and L. S. Liebeskind, *J. Am. Chem. Soc.*, 2009, **131**, 876–877.
- 28 S. Roy and C. Spino, *Org. Lett.*, 2006, **8**, 939–942.
- 29 M. Arbour, S. Roy, C. Godbout and C. Spino, *J. Org. Chem.*, 2009, **74**, 3806–3814.
- 30 F. A. Davis and R. Edupuganti, *Org. Lett.*, 2010, **12**, 848–851.
- 31 F. A. Davis, *J. Org. Chem.*, 2006, **71**, 8993–9003.
- 32 J. W. Daly, I. Karle, C. W. Myers, T. Tokuyama, J. A. Waters and B. Witkop, *Proc. Natl. Acad. Sci. U. S. A.*, 1971, **68**, 1870–1875.
- 33 J. W. Daly, *J. Nat. Prod.*, 1998, **61**, 162–172.
- 34 E. X. Albuquerque, K. Kuba and J. Daly, *J. Pharmacol. Exp. Ther.*, 1974, **189**, 513–524.
- 35 A. J. Lapa, E. X. Albuquerque, J. M. Sarvey, J. Daly and B. Witkop, *Exp. Neurol.*, 1975, **47**, 558–580.
- 36 A. T. Eldefrawi, M. E. Eldefrawi, E. X. Albuquerque, A. C. Oliveira, N. Mansour, M. Adler, J. W. Daly, G. B. Brown, W. Burgermeister and B. Witkop, *Proc. Natl. Acad. Sci. U. S. A.*, 1977, **74**, 2172–2176.
- 37 K. Takahashi, A. E. Jacobson, C. P. Mak, B. Witkop, A. Brossi, E. X. Albuquerque, J. E. Warnick, M. A. Maleque, A. Bavoso and J. V. Silverton, *J. Med. Chem.*, 1982, **25**, 919–925.
- 38 C. E. Spivak, M. A. Maleque, A. C. Oliveira, L. M. Masukawa, T. Tokuyama, J. W. Daly and E. X. Albuquerque, *Mol. Pharmacol.*, 1982, **21**, 351–361.
- 39 A. Sinclair and R. A. Stockman, *Nat. Prod. Rep.*, 2007, **24**, 298–326.
- 40 T. Fukuyama, L. V. Dunkerton, M. Aratani and Y. Kishi, *J. Org. Chem.*, 1975, **40**, 2011–2012.
- 41 M. Aratani, L. V. Dunkerton, T. Fukuyama, Y. Kishi, H. Kakoi, S. Sugiura and S. Inoue, *J. Org. Chem.*, 1975, **40**, 2009–2011.
- 42 S. C. Carey, M. Aratani and Y. Kishi, *Tetrahedron Lett.*, 1985, **26**, 5887–5890.
- 43 E. J. Corey, J. F. Arnett and G. N. Widiger, *J. Am. Chem. Soc.*, 1975, **97**, 430–431.
- 44 T. Ibuka, H. Minakata, Y. Mitsui, E. Tabushi, T. Taga and Y. Inubushi, *Chem. Lett.*, 1981, **10**, 1409–1412.



45 T. Ibuka, Y. Mitsui, K. Hayashi, H. Minakata and Y. Inubushi, *Tetrahedron Lett.*, 1981, **22**, 4425–4428.

46 T. Ibuka, H. Minakata, Y. Mitsui, K. Hayashi, T. Taga and Y. Inubushi, *Chem. Pharm. Bull.*, 1982, **30**, 2840–2859.

47 N. Maezaki, H. Fukuyama, S. Yagi, T. Tanaka and C. Iwata, *J. Chem. Soc., Chem. Commun.*, 1994, 1835–1836.

48 D. Kim and C. Won Park, *Chem. Commun.*, 1997, 2263–2264.

49 G. Stork and K. Zhao, *J. Am. Chem. Soc.*, 1990, **112**, 5875–5876.

50 Y. Adachi, N. Kamei, S. Yokoshima and T. Fukuyama, *Org. Lett.*, 2011, **13**, 4446–4449.

51 Y. Adachi, N. Kamei, S. Yokoshima and T. Fukuyama, *Org. Lett.*, 2014, **16**, 1273.

52 M. S. Karatholuvhu, A. Sinclair, A. F. Newton, M.-L. Alcaraz, R. A. Stockman and P. L. Fuchs, *J. Am. Chem. Soc.*, 2006, **128**, 12656–12657.

53 G. M. Williams, S. D. Roughley, J. E. Davies, A. B. Holmes and J. P. Adams, *J. Am. Chem. Soc.*, 1999, **121**, 4900–4901.

54 E. C. Davison, M. E. Fox, A. B. Holmes, S. D. Roughley, C. J. Smith, G. M. Williams, J. E. Davies, P. R. Raithby, J. P. Adams, I. T. Forbes, N. J. Press and M. J. Thompson, *J. Chem. Soc., Perkin Trans. 1*, 2002, 1494–1514.

55 M. Glanzmann, C. Karalai, B. Ostersehlt, U. Schön, C. Frese and E. Winterfeldt, *Tetrahedron*, 1982, **38**, 2805–2810.

56 J. J. Venit, M. DiPierro and P. Magnus, *J. Org. Chem.*, 1989, **54**, 4298–4301.

57 S. A. Godleski, D. J. Heacock, J. D. Meinhart and S. van Wallendael, *J. Org. Chem.*, 1983, **48**, 2101–2103.

58 W. Carruthers and S. A. Cumming, *J. Chem. Soc., Chem. Commun.*, 1983, 360–361.

59 D. Tanner and P. Somfai, *Tetrahedron Lett.*, 1985, **26**, 3883–3886.

60 D. Tanner and P. Somfai, *Tetrahedron*, 1986, **42**, 5657–5664.

61 D. Tanner, M. Sellen and J. E. Baeckvall, *J. Org. Chem.*, 1989, **54**, 3374–3378.

62 D. J. Wardrop, W. Zhang and C. L. Landrie, *Tetrahedron Lett.*, 2004, **45**, 4229–4231.

63 R. W. Fitch and F. A. Luzzio, *Ultrason. Sonochem.*, 1997, **4**, 99–107.

64 O. Y. Provoost, S. J. Hedley, A. J. Hazelwood and J. P. A. Harrity, *Tetrahedron Lett.*, 2006, **47**, 331–333.

65 P. Compain, J. Goré and J.-M. Vatèle, *Tetrahedron*, 1996, **52**, 6647–6664.

66 D. Tanner, L. Hagberg and A. Poulsen, *Tetrahedron*, 1999, **55**, 1427–1440.

67 S. Kim, H. Ko, T. Lee and D. Kim, *J. Org. Chem.*, 2005, **70**, 5756–5759.

68 D. L. Comins, Y.-M. Zhang and X. Zheng, *Chem. Commun.*, 1998, 2509–2510.

69 J. D. Winkler and P. M. Hershberger, *J. Am. Chem. Soc.*, 1989, **111**, 4852–4856.

70 X. Q. Ma and D. R. Gang, *Nat. Prod. Rep.*, 2004, **21**, 752–772.

71 K. Bödeker, *Liebigs Ann. Chem.*, 1881, **208**, 363–367.

72 G. Stork, R. A. Kretchmer and R. H. Schlessinger, *J. Am. Chem. Soc.*, 1968, **90**, 1647–1648.

73 W. A. Ayer, W. R. Bowman, G. A. Cooke and A. C. Soper, *Tetrahedron Lett.*, 1966, **7**, 2021–2026.

74 W. A. Ayer, W. R. Bowman, T. C. Joseph and P. Smith, *J. Am. Chem. Soc.*, 1968, **90**, 1648–1650.

75 A. Padwa, M. A. Brodney, J. P. Marino and S. M. Sheehan, *J. Org. Chem.*, 1997, **62**, 78–87.

76 K. Wiesner and L. Poon, *Tetrahedron Lett.*, 1967, **8**, 4937–4940.

77 H. Dugas, M. E. Hazenberg, Z. Valenta and K. Wiesner, *Tetrahedron Lett.*, 1967, **8**, 4931–4936.

78 K. Wiesner, V. Musil and K. J. Wesner, *Tetrahedron Lett.*, 1968, **9**, 5643–5646.

79 K. Wiesner, L. Poon, I. Jirkovský and M. Fishman, *Can. J. Chem.*, 1969, **47**, 433–444.

80 C. H. Heathcock, E. Kleinman and E. S. Binkley, *J. Am. Chem. Soc.*, 1978, **100**, 8036–8037.

81 E. Kleinman and C. H. Heathcock, *Tetrahedron Lett.*, 1979, **20**, 4125–4128.

82 C. H. Heathcock, E. F. Kleinman and E. S. Binkley, *J. Am. Chem. Soc.*, 1982, **104**, 1054–1068.

83 K. Nakahara, K. Hirano, R. Maehata, Y. Kita and H. Fujioka, *Org. Lett.*, 2011, **13**, 2015–2017.

84 D. A. Evans and J. R. Scheerer, *Angew. Chem., Int. Ed.*, 2005, **44**, 6038–6042.

85 H. Yang, R. G. Carter and L. N. Zakharov, *J. Am. Chem. Soc.*, 2008, **130**, 9238–9239.

86 H. Yang and R. G. Carter, *J. Org. Chem.*, 2010, **75**, 4929–4938.

87 G. A. Kraus and Y. S. Hon, *J. Am. Chem. Soc.*, 1985, **107**, 4341–4342.

88 P. A. Grieco and Y. Dai, *J. Am. Chem. Soc.*, 1998, **120**, 5128–5129.

89 E. Wenkert, B. Chauncy, K. G. Dave, A. R. Jeffcoat, F. M. Schell and H. P. Schenk, *J. Am. Chem. Soc.*, 1973, **95**, 8427–8436.

90 E. Wenkert and C. A. Broka, *J. Chem. Soc., Chem. Commun.*, 1984, 714–715.

91 G. A. Kraus and Y. S. Hon, *Heterocycles*, 1987, **25**, 377–386.

92 W. A. Ayer and L. S. Trifonov, in *The Alkaloids*, ed. G. A. Cordell and A. Brossi, Academic, New York, 1994, vol. 45, pp. 233–266.

93 F. A. L. Anet and C. R. Eves, *Can. J. Chem.*, 1958, **36**, 902–909.

94 D. F. Fischer and R. Sarpong, *J. Am. Chem. Soc.*, 2010, **132**, 5926–5927.

95 C. Tsukano, L. Zhao, Y. Takemoto and M. Hirama, *Eur. J. Org. Chem.*, 2010, 4198–4200.

96 C. Yuan, C.-T. Chang and D. Siegel, *J. Org. Chem.*, 2013, **78**, 5647–5668.

97 M. Azuma, T. Yoshikawa, N. Kogure, M. Kitajima and H. Takayama, *J. Am. Chem. Soc.*, 2014, **136**, 11618–11621.

98 D. Schumann, H.-J. Müller and A. Naumann, *Liebigs Ann. Chem.*, 1982, 2057–2061.

99 D. Schumann, H. J. Müller and A. Naumann, *Liebigs Ann. Chem.*, 1982, 1700–1705.

100 D. Schumann and A. Naumann, *Liebigs Ann. Chem.*, 1983, 220–225.

101 H. Morita, K. I. Ishiuchi, A. Haganuma, T. Hoshino, Y. Obara, N. Nakahata and J. I. Kobayashi, *Tetrahedron*, 2005, **61**, 1955–1960.

102 B. B. Liau and M. D. Shair, *J. Am. Chem. Soc.*, 2010, **132**, 9594–9595.

103 A. S. Lee, B. B. Liau and M. D. Shair, *J. Am. Chem. Soc.*, 2014, **136**, 13442–13452.

104 H. Li, X. Wang and X. Lei, *Angew. Chem., Int. Ed.*, 2012, **51**, 491–495.

105 J. Zhang, J. Wu, B. Hong, W. Ai, X. Wang, H. Li and X. Lei, *Nat. Commun.*, 2014, **5**, 4614.

106 Y.-R. Yang, Z.-W. Lai, L. Shen, J.-Z. Huang, X.-D. Wu, J.-L. Yin and K. Wei, *Org. Lett.*, 2010, **12**, 3430–3433.

107 Y.-R. Yang, L. Shen, J.-Z. Huang, T. Xu and K. Wei, *J. Org. Chem.*, 2011, **76**, 3684–3690.

108 S.-H. Hou, Y.-Q. Tu, L. Liu, F.-M. Zhang, S.-H. Wang and X.-M. Zhang, *Angew. Chem., Int. Ed.*, 2013, **52**, 11373–11376.

109 Y. Xia and A. P. Kozikowski, *J. Am. Chem. Soc.*, 1989, **111**, 4116–4117.

110 L. Qian and R. Ji, *Tetrahedron Lett.*, 1989, **30**, 2089–2090.

111 A. P. Kozikowski, Y. Xia, E. R. Reddy, W. Tuckmantel, I. Hanin and X. C. Tang, *J. Org. Chem.*, 1991, **56**, 4636–4645.

112 A. P. Kozikowski, G. Campiani, P. Aagaard and M. McKinney, *J. Chem. Soc., Chem. Commun.*, 1993, 860–862.

113 I. Y. C. Lee, M. H. Jung, H. W. Lee and J. Y. Yang, *Tetrahedron Lett.*, 2002, **43**, 2407–2409.

114 T. Koshiba, S. Yokoshima and T. Fukuyama, *Org. Lett.*, 2009, **11**, 5354–5356.

115 M. K. M. Tun, D.-J. Wüstmann and S. B. Herzon, *Chem. Sci.*, 2011, **2**, 2251–2253.

116 S. M. A. Sohel and T. Opatz, *ARKIVOC*, 2014, **i**, 92–108.

117 Y. Hirasawa, J. Kobayashi and H. Morita, *Heterocycles*, 2009, **77**, 679–729.

118 G. T. Ha, R. K. Wong and Y. Zhang, *Chem. Biodiversity*, 2011, **8**, 1189–1204.

119 S. R. Tudhope, J. A. Bellamy, A. Ball, D. Rajasekar, M. Azadi-Ardakani, H. S. Meera, J. M. Gnanadeepam, R. Saiganesh, F. Gibson, L. He, C. H. Behrens, G. Underiner, J. Marfurt and N. Favre, *Org. Process Res. Dev.*, 2012, **16**, 635–642.

120 R. Ding, B.-F. Sun and G.-Q. Lin, *Org. Lett.*, 2012, **14**, 4446–4449.

121 J. D. White, Y. Li, J. Kim and M. Terinek, *Org. Lett.*, 2013, **15**, 882–885.

122 Y. Hirasawa, H. Morita and J. Kobayashi, *Org. Lett.*, 2004, **6**, 3389–3391.

123 Y. Hirasawa, J. Kobayashi, Y. Obara, N. Nakahata, N. Kawahara, Y. Goda and H. Morita, *Heterocycles*, 2006, **68**, 2357–2364.

124 B. L. Nilsson, L. E. Overman, J. R. de Alaniz and J. M. Rohde, *J. Am. Chem. Soc.*, 2008, **130**, 11297–11299.

125 R. A. Altman, B. L. Nilsson, L. E. Overman, J. R. de Alaniz, J. M. Rohde and V. Taupin, *J. Org. Chem.*, 2010, **75**, 7519–7534.

126 X. Cheng and S. P. Waters, *Org. Lett.*, 2010, **12**, 205–207.

127 W. A. Denne, S. R. Johns, J. A. Lamberton and A. M. Mathieson, *Tetrahedron Lett.*, 1971, **12**, 3107–3108.

128 W. A. Denne, S. R. Johns, J. A. Lamberton and A. M. Mathieson, *Tetrahedron Lett.*, 1973, **14**, 794.

129 E. J. Corey and R. D. Balanson, *J. Am. Chem. Soc.*, 1974, **96**, 6516–6517.

130 D. M. Ryckman and R. V. Stevens, *J. Am. Chem. Soc.*, 1987, **109**, 4940–4948.

131 M. Matsui, in *The Alkaloids: Chemistry and Pharmacology*, ed. B. Arnold, Elsevier Academic Press, 1988, vol. 33, pp. 307–347.

132 Y. Inubushi, T. Ibuka and M. Kitano, *Tetrahedron Lett.*, 1969, **10**, 1611–1614.

133 D. A. Evans, C. A. Bryan and G. M. Wahl, *J. Org. Chem.*, 1970, **35**, 4122–4127.

134 T. Ibuka, K. Tanaka and Y. Inubushi, *Tetrahedron Lett.*, 1970, **11**, 4811–4814.

135 Y. Inubushi, M. Kitano and T. Ibuka, *Chem. Pharm. Bull.*, 1971, **19**, 1820–1841.

136 T. Ibuka, K. Tanaka and Y. Inubushi, *Tetrahedron Lett.*, 1972, **13**, 1393–1396.

137 T. Ibuka, K. Tanaka and Y. Inubushi, *Chem. Pharm. Bull.*, 1974, **22**, 782–798.

138 T. Ibuka, K. Tanaka and Y. Inubushi, *Chem. Pharm. Bull.*, 1974, **22**, 907–921.

139 A. G. Schultz and A. Wang, *J. Am. Chem. Soc.*, 1998, **120**, 8259–8260.

140 S. B. Jones, L. He and S. L. Castle, *Org. Lett.*, 2006, **8**, 3757–3760.

141 F. Li, S. S. Tartakoff and S. L. Castle, *J. Am. Chem. Soc.*, 2009, **131**, 6674–6675.

142 F. Li, S. S. Tartakoff and S. L. Castle, *J. Org. Chem.*, 2009, **74**, 9082–9093.

143 S. B. Herzon, N. A. Calandra and S. M. King, *Angew. Chem., Int. Ed.*, 2011, **50**, 8863–8866.

144 K. V. Chuang, R. Navarro and S. E. Reisman, *Angew. Chem., Int. Ed.*, 2011, **50**, 9447–9451.

145 S. M. King, N. A. Calandra and S. B. Herzon, *Angew. Chem., Int. Ed.*, 2013, **52**, 3642–3645.

146 N. A. Calandra, S. M. King and S. B. Herzon, *J. Org. Chem.*, 2013, **78**, 10031–10057.

147 D. K. Nielsen, L. L. Nielsen, S. B. Jones, L. Toll, M. C. Asplund and S. L. Castle, *J. Org. Chem.*, 2009, **74**, 1187–1199.

148 G.-W. Qin and R.-S. Xu, *Med. Res. Rev.*, 1998, **18**, 375–382.

149 R.-S. Xu, in *Studies in Natural Products Chemistry*, ed. R. Attaur, Elsevier, 2000, vol. vol. 21, Part B, pp. 729–772.

150 H. S. Chung, P. M. Hon, G. Lin, P. P. H. But and H. Dong, *Planta Med.*, 2003, **69**, 914–920.

151 R. Aloise Pilli and M. da Conceição Ferreira de Oliveira, *Nat. Prod. Rep.*, 2000, **17**, 117–127.

152 R. A. Pilli, G. B. Rosso and M. da Conceição Ferreira de Oliveira, *Nat. Prod. Rep.*, 2010, **27**, 1908–1937.

153 A. S. Kende, J. I. Martin Hernando and J. B. J. Milbank, *Org. Lett.*, 2001, **3**, 2505–2508.

154 A. S. Kende, J. I. Martin Hernando and J. B. J. Milbank, *Tetrahedron*, 2002, **58**, 61–74.

155 T. Taniguchi and H. Ishibashi, *Tetrahedron*, 2008, **64**, 8773–8779.



156 T. Taniguchi, G. Tanabe, O. Muraoka and H. Ishibashi, *Org. Lett.*, 2008, **10**, 197–199.

157 P.-Q. Huang, S.-Y. Huang, L.-H. Gao, Z.-Y. Mao, Z. Chang and A.-E. Wang, *Chem. Commun.*, 2015, **51**, 4576–4578.

158 Y.-M. Zhao, P. Gu, Y.-Q. Tu, C.-A. Fan and Q. Zhang, *Org. Lett.*, 2008, **10**, 1763–1766.

159 Z.-H. Chen, Z.-M. Chen, Y.-Q. Zhang, Y.-Q. Tu and F.-M. Zhang, *J. Org. Chem.*, 2011, **76**, 10173–10186.

160 Z.-H. Chen, Y.-Q. Zhang, Z.-M. Chen, Y.-Q. Tu and F.-M. Zhang, *Chem. Commun.*, 2011, **47**, 1836–1838.

161 M. Brüggemann, A. I. McDonald, L. E. Overman, M. D. Rosen, L. Schwink and J. P. Scott, *J. Am. Chem. Soc.*, 2003, **125**, 15284–15285.

162 A. S. Kende, T. L. Smalley and H. Huang, *J. Am. Chem. Soc.*, 1999, **121**, 7431–7432.

163 C. Fang, C. S. Shanahan, D. H. Paull and S. F. Martin, *Angew. Chem., Int. Ed.*, 2012, **51**, 10596–10599.

164 P. Gu, Y.-M. Zhao, Y. Q. Tu, Y. Ma and F. Zhang, *Org. Lett.*, 2006, **8**, 5271–5273.

165 M. Greshoff, *Ber. Dtsch. Chem. Ges.*, 1890, **23**, 3537–3550.

166 A. R. Battersby and H. Gregory, *J. Chem. Soc.*, 1963, 22–32.

167 C. Djerassi, H. Schmid, W. G. Kump, A. R. Battersby, J. M. Wilson, R. J. Owellen, D. J. Lecount and H. Budzikiewicz, *Helv. Chim. Acta*, 1963, **46**, 742–751.

168 B. W. Bycroft, D. Schumann, M. B. Patel and H. Schmid, *Helv. Chim. Acta*, 1964, **47**, 1147–1152.

169 D. Schumann, B. W. Bycroft and H. Schmid, *Experientia*, 1964, **20**, 202–203.

170 H. Achenbach and K. Biemann, *J. Am. Chem. Soc.*, 1965, **87**, 4944–4950.

171 J. M. F. Filho, B. Gilbert, M. Kitagawa, L. A. P. Leme and L. J. Durham, *J. Chem. Soc. C*, 1966, 1260–1266.

172 B. M. Craven, B. Gilbert and L. A. P. Leme, *Chem. Commun.*, 1968, 955–956.

173 B. Craven, *Acta Crystallogr., Sect. B: Struct. Sci.*, 1969, **25**, 2131–2139.

174 H. Kinoshita, T. Ohnuma, T. Oishi and Y. Ban, *Chem. Lett.*, 1986, **15**, 927–930.

175 D. Cartier, M. Ouahrani and J. Lévy, *Tetrahedron Lett.*, 1989, **30**, 1951–1954.

176 J. Xie, A. L. Wolfe and D. L. Boger, *Org. Lett.*, 2013, **15**, 868–870.

177 Y. Ban, Y. Honma and T. Oishi, *Tetrahedron Lett.*, 1976, **17**, 1111–1114.

178 M. E. Kuehne and P. J. Seaton, *J. Org. Chem.*, 1985, **50**, 4790–4796.

179 E. Wenkert and S. Liu, *J. Org. Chem.*, 1994, **59**, 7677–7682.

180 D. Gagnon and C. Spino, *J. Org. Chem.*, 2009, **74**, 6035–6041.

181 T. Gallagher and P. Magnus, *J. Am. Chem. Soc.*, 1983, **105**, 2086–2087.

182 P. Magnus, T. Gallagher, P. Brown and J. C. Huffman, *J. Am. Chem. Soc.*, 1984, **106**, 2105–2114.

183 P. Magnus and P. Brown, *J. Chem. Soc., Chem. Commun.*, 1985, 184–186.

184 P. Magnus, I. R. Matthews, J. Schultz, R. Waditschatka and J. C. Huffman, *J. Org. Chem.*, 1988, **53**, 5772–5776.

185 P. Magnus, T. Katoh, I. R. Matthews and J. C. Huffman, *J. Am. Chem. Soc.*, 1989, **111**, 6707–6711.

186 K. Awang, T. Sévenet, A. Hamid, A. Hadi, B. David and M. Païs, *Tetrahedron Lett.*, 1992, **33**, 2493–2496.

187 K. Awang, T. Sévenet, M. Païs and A. H. A. Hadi, *J. Nat. Prod.*, 1993, **56**, 1134–1139.

188 T.-S. Kam, K. Yoganathan and C.-H. Chuah, *Tetrahedron Lett.*, 1995, **36**, 759–762.

189 T.-S. Kam, K.-H. Lim, K. Yoganathan, M. Hayashi and K. Komiyama, *Tetrahedron*, 2004, **60**, 10739–10745.

190 S. Jin, J. Gong and Y. Qin, *Angew. Chem., Int. Ed.*, 2015, **54**, 2228–2231.

191 S. Arai, M. Nakajima and A. Nishida, *Angew. Chem., Int. Ed.*, 2014, **53**, 5569–5572.

192 S. Arai, M. Nakajima and A. Nishida, *Angew. Chem., Int. Ed.*, 2014, **53**, 14295.

193 M. Hoshi, O. Kaneko, M. Nakajima, S. Arai and A. Nishida, *Org. Lett.*, 2014, **16**, 768–771.

194 W. H. Pearson, I. Y. Lee, Y. Mi and P. Stoy, *J. Org. Chem.*, 2004, **69**, 9109–9122.

195 W. H. Pearson, Y. Mi, I. Y. Lee and P. Stoy, *J. Am. Chem. Soc.*, 2001, **123**, 6724–6725.

196 T. J. Greshock, A. W. Grubbs, S. Tsukamoto and R. M. Williams, *Angew. Chem., Int. Ed.*, 2007, **46**, 2262–2265.

197 G. D. Artman III, A. W. Grubbs and R. M. Williams, *J. Am. Chem. Soc.*, 2007, **129**, 6336–6342.

198 D. Seebach, M. Boes, R. Naef and W. B. Schweizer, *J. Am. Chem. Soc.*, 1983, **105**, 5390–5398.

199 P. S. Baran, C. A. Guerrero, N. B. Ambhaikar and B. D. Hafenstein, *Angew. Chem., Int. Ed.*, 2005, **44**, 606–609.

200 P. S. Baran, C. A. Guerrero, B. D. Hafenstein and N. B. Ambhaikar, *Angew. Chem., Int. Ed.*, 2005, **44**, 3892–3895.

201 S. B. Herzon and A. G. Myers, *J. Am. Chem. Soc.*, 2005, **127**, 5342–5344.

202 R. F. Bond, J. C. A. Boeyens, C. W. Holzapfel and P. S. Steyn, *J. Chem. Soc., Perkin Trans. 1*, 1979, 1751–1761.

203 M. Tsuda, Y. Kasai, K. Komatsu, T. Sone, M. Tanaka, Y. Mikami and J. I. Kobayashi, *Org. Lett.*, 2004, **6**, 3087–3089.

204 T. Mugishima, M. Tsuda, Y. Kasai, H. Ishiyama, E. Fukushi, J. Kawabata, M. Watanabe, K. Akao and J. Kobayashi, *J. Org. Chem.*, 2005, **70**, 9430–9435.

205 Z. Bian, C. C. Marvin and S. F. Martin, *J. Am. Chem. Soc.*, 2013, **135**, 10886–10889.

206 K. Kong, J. A. Enquist, M. E. McCallum, G. M. Smith, T. Matsumaru, E. Menhaji-Klotz and J. L. Wood, *J. Am. Chem. Soc.*, 2013, **135**, 10890–10893.

207 E. V. Mercado-Marin, P. Garcia-Reynaga, S. Romminger, E. F. Pimenta, D. K. Romney, M. W. Lodewyk, D. E. Williams, R. J. Andersen, S. J. Miller, D. J. Tantillo, R. G. S. Berlinck and R. Sarpong, *Nature*, 2014, **509**, 318–324.

208 E. V. Mercado-Marin and R. Sarpong, *Chem. Sci.*, 2015, **6**, 5048–5052.



209 B. M. Trost, D. A. Bringley, T. Zhang and N. Cramer, *J. Am. Chem. Soc.*, 2013, **135**, 16720–16735.

210 B. M. Trost, N. Cramer and H. Bernsmann, *J. Am. Chem. Soc.*, 2007, **129**, 3086–3087.

211 W. G. Beyersbergen van Henegouwen, R. M. Fieseler, F. P. J. T. Rutjes and H. Hiemstra, *Angew. Chem., Int. Ed.*, 1999, **38**, 2214–2217.

212 H. Lin and S. J. Danishefsky, *Angew. Chem., Int. Ed.*, 2003, **42**, 36–51.

213 X. Zhou, T. Xiao, Y. Iwama and Y. Qin, *Angew. Chem., Int. Ed.*, 2012, **51**, 4909–4912.

214 J. Shimokawa, T. Harada, S. Yokoshima and T. Fukuyama, *J. Am. Chem. Soc.*, 2011, **133**, 17634–17637.

215 S. Diethelm and E. M. Carreira, *J. Am. Chem. Soc.*, 2013, **135**, 8500–8503.

216 K. Stratmann, R. E. Moore, R. Bonjouklian, J. B. Deeter, G. M. L. Patterson, S. Shaffer, C. D. Smith and T. A. Smitka, *J. Am. Chem. Soc.*, 1994, **116**, 9935–9942.

217 J. I. Jimenez, U. Huber, R. E. Moore and G. M. L. Patterson, *J. Nat. Prod.*, 1999, **62**, 569–572.

218 J. C. Menéndez, in *Bioactive Heterocycles V*, ed. M. Khan, Springer, Berlin Heidelberg, 2007, vol. 11, ch. 63, pp. 63–101.

219 J. L. Wood, *Nat. Chem.*, 2012, **4**, 341–343.

220 J. A. MacKay, R. L. Bishop and V. H. Rawal, *Org. Lett.*, 2005, **7**, 3421–3424.

221 V. Bhat, K. M. Allan and V. H. Rawal, *J. Am. Chem. Soc.*, 2011, **133**, 5798–5801.

222 K. M. Allan, K. Kobayashi and V. H. Rawal, *J. Am. Chem. Soc.*, 2012, **134**, 1392–1395.

223 K. Jae Nyoung and E. K. Ryu, *Tetrahedron Lett.*, 1993, **34**, 8283–8284.

224 J. N. Kim, K. S. Jung, H. J. Lee and J. S. Son, *Tetrahedron Lett.*, 1997, **38**, 1597–1598.

225 T.-H. Fu, W. T. McElroy, M. Shamszad and S. F. Martin, *Org. Lett.*, 2012, **14**, 3834–3837.

226 X. Tian, A. D. Huters, C. J. Douglas and N. K. Garg, *Org. Lett.*, 2009, **11**, 2349–2351.

227 A. D. Huters, K. W. Quasdorf, E. D. Styduhar and N. K. Garg, *J. Am. Chem. Soc.*, 2011, **133**, 15797–15799.

228 K. W. Quasdorf, A. D. Huters, M. W. Lodewyk, D. J. Tantillo and N. K. Garg, *J. Am. Chem. Soc.*, 2012, **134**, 1396–1399.

229 N. A. Weires, E. D. Styduhar, E. L. Baker and N. K. Garg, *J. Am. Chem. Soc.*, 2014, **136**, 14710–14713.

230 E. D. Styduhar, A. D. Huters, N. A. Weires and N. K. Garg, *Angew. Chem., Int. Ed.*, 2013, **52**, 12422–12425.

231 K. Komine, Y. Nomura, J. Ishihara and S. Hatakeyama, *Org. Lett.*, 2015, **17**, 3918–3921.

232 P. S. Baran, R. A. Shenvi and C. A. Mitsos, *Angew. Chem., Int. Ed.*, 2005, **44**, 3714–3717.

233 P. S. Baran and R. A. Shenvi, *J. Am. Chem. Soc.*, 2006, **128**, 14028–14029.

234 T. Newhouse and P. S. Baran, *J. Am. Chem. Soc.*, 2008, **130**, 10886–10887.

235 K. Foo, T. Newhouse, I. Mori, H. Takayama and P. S. Baran, *Angew. Chem., Int. Ed.*, 2011, **50**, 2716–2719.

236 T. Newhouse, C. A. Lewis and P. S. Baran, *J. Am. Chem. Soc.*, 2009, **131**, 6360–6361.

237 T. Newhouse, C. A. Lewis, K. J. Eastman and P. S. Baran, *J. Am. Chem. Soc.*, 2010, **132**, 7119–7137.

238 C. Li, C. Chan, A. C. Heimann and S. J. Danishefsky, *Angew. Chem., Int. Ed.*, 2007, **46**, 1448–1450.

239 H. Abdelkafi and B. Nay, *Nat. Prod. Rep.*, 2012, **29**, 845–869.

240 S. M. Weinreb and J. Auerbach, *J. Am. Chem. Soc.*, 1975, **97**, 2503–2506.

241 T. P. Burkholder and P. L. Fuchs, *J. Am. Chem. Soc.*, 1988, **110**, 2341–2342.

242 L. F. Tietze and H. Schirok, *J. Am. Chem. Soc.*, 1999, **121**, 10264–10269.

243 Q. Liu, E. M. Ferreira and B. M. Stoltz, *J. Org. Chem.*, 2007, **72**, 7352–7358.

244 W.-D. Z. Li and Y.-Q. Wang, *Org. Lett.*, 2003, **5**, 2931–2934.

245 L. Planas, J. Pérard-Viret and J. Royer, *J. Org. Chem.*, 2004, **69**, 3087–3092.

246 J. D. Eckelbarger, J. T. Wilmot and D. Y. Gin, *J. Am. Chem. Soc.*, 2006, **128**, 10370–10371.

247 J. D. Eckelbarger, J. T. Wilmot, M. T. Epperson, C. S. Thakur, D. Shum, C. Antczak, L. Tarassishin, H. Djaballah and D. Y. Gin, *Chem.-Eur. J.*, 2008, **14**, 4293–4306.

248 Z. Zhao and P. S. Mariano, *Tetrahedron*, 2006, **62**, 7266–7273.

249 A. Hameed, A. J. Blake and C. J. Hayes, *J. Org. Chem.*, 2008, **73**, 8045–8048.

250 T. Taniguchi and H. Ishibashi, *Org. Lett.*, 2008, **10**, 4129–4131.

251 M. E. Kuehne, W. G. Bornmann, W. H. Parsons, T. D. Spitzer, J. F. Blount and J. Zubieta, *J. Org. Chem.*, 1988, **53**, 3439–3450.

252 M. Ikeda, S. A. A. El Bialy, K. Hirose, M. Kotake, T. Sato, S. M. M. Bayomi, I. A. Shehata, A. M. Abdelal, L. M. Gad and T. Yakura, *Chem. Pharm. Bull.*, 1999, **47**, 983–987.

253 S. Suga, M. Watanabe and J. Yoshida, *J. Am. Chem. Soc.*, 2002, **124**, 14824–14825.

254 S. Yasuda, T. Yamada and M. Hanaoka, *Tetrahedron Lett.*, 1986, **27**, 2023–2026.

255 H. Ishibashi, M. Okano, H. Tamaki, K. Maruyama, T. Yakura and M. Ikeda, *J. Chem. Soc., Chem. Commun.*, 1990, 1436–1437.

256 L. F. Tietze, H. Braun, P. L. Steck, S. A. A. El Bialy, N. Tölle and A. Düfert, *Tetrahedron*, 2007, **63**, 6437–6445.

257 Y.-M. Zhao, P. Gu, H.-J. Zhang, Q.-W. Zhang, C.-A. Fan, Y.-Q. Tu and F.-M. Zhang, *J. Org. Chem.*, 2009, **74**, 3211–3213.

258 Y. Koseki, H. Sato, Y. Watanabe and T. Nagasaka, *Org. Lett.*, 2002, **4**, 885–888.

259 X. Lin, R. W. Kavash and P. S. Mariano, *J. Am. Chem. Soc.*, 1994, **116**, 9791–9792.

260 K. Folkers and R. T. Major, *J. Am. Chem. Soc.*, 1937, **59**, 1580–1581.

261 E. Reimann, in *Progress in the Chemistry of Organic Natural Products*, ed. W. Herz, H. Falk and G. W. Kirby, Springer, Vienna, 2007, vol. 88, ch. 1, pp. 1–62.

262 H. Ishibashi, T. Sato, M. Takahashi, M. Hayashi, K. Ishikawa and M. Ikeda, *Chem. Pharm. Bull.*, 1990, **38**, 907–911.

263 S. Chikaoka, A. Toyao, M. Ogasawara, O. Tamura and H. Ishibashi, *J. Org. Chem.*, 2003, **68**, 312–318.

264 T. Sano, J. Toda, T. Ohshima and Y. Tsuda, *Chem. Pharm. Bull.*, 1992, **40**, 873–878.

265 R. L. Funk and J. Belmar, *Tetrahedron Lett.*, 2012, **53**, 176–178.

266 S. T. Heller, T. Kiho, A. R. H. Narayan and R. Sarpong, *Angew. Chem., Int. Ed.*, 2013, **52**, 11129–11133.

267 H.-T. Luu, S. Wiesler, G. Frey and J. Streuff, *Org. Lett.*, 2015, **17**, 2478–2481.

268 N. A. Braun, M. Ousmer, J. D. Bray, D. Bouchu, K. Peters, E.-M. Peters and M. A. Ciufolini, *J. Org. Chem.*, 2000, **65**, 4397–4408.

269 M. Paladino, J. Zaifman and M. A. Ciufolini, *Org. Lett.*, 2015, **17**, 3422–3425.

270 T. Kawasaki, N. Onoda, H. Watanabe and T. Kitahara, *Tetrahedron Lett.*, 2001, **42**, 8003–8006.

271 J. M. Joo, R. A. David, Y. Yuan and C. Lee, *Org. Lett.*, 2010, **12**, 5704–5707.

272 K. V. Chuang, R. Navarro and S. E. Reisman, *Chem. Sci.*, 2011, **2**, 1086–1089.

273 M. A. le Dreau, D. Desmaele, F. Dumas and J. d'Angelo, *J. Org. Chem.*, 1993, **58**, 2933–2935.

274 Y. Yoshida, K. Mohri, K. Isobe, T. Itoh and K. Yamamoto, *J. Org. Chem.*, 2009, **74**, 6010–6015.

275 K. Shimizu, M. Takimoto, Y. Sato and M. Mori, *J. Organomet. Chem.*, 2006, **691**, 5466–5475.

276 H. Fukumoto, T. Esumi, J. Ishihara and S. Hatakeyama, *Tetrahedron Lett.*, 2003, **44**, 8047–8049.

277 Q. Wang and A. Padwa, *Org. Lett.*, 2006, **8**, 601–604.

278 F. Zhang, N. S. Simpkins and A. J. Blake, *Org. Biomol. Chem.*, 2009, **7**, 1963–1979.

279 J. H. Rigby, C. Deur and M. J. Heeg, *Tetrahedron Lett.*, 1999, **40**, 6887–6890.

280 J. W. Daly, T. F. Spande and H. M. Garraffo, *J. Nat. Prod.*, 2005, **68**, 1556–1575.

281 S. M. Weinreb, *Chem. Rev.*, 2006, **106**, 2531–2549.

282 K. Sakamoto, E. Tsujii, F. Abe, T. Nakanishi, M. Yamashita, N. Shigematsu, S. Izumi and M. Okuhara, *J. Antibiot.*, 1996, **49**, 37–44.

283 J. F. Biard, S. Guyot, C. Roussakis, J. F. Verbist, J. Vercauteren, J. F. Weber and K. Boukef, *Tetrahedron Lett.*, 1994, **35**, 2691–2694.

284 D. L. J. Clive, M. L. Yu, J. Wang, V. S. C. Yeh and S. Z. Kang, *Chem. Rev.*, 2005, **105**, 4483–4514.

285 B. B. Snider and T. Liu, *J. Org. Chem.*, 1997, **62**, 5630–5633.

286 B. M. Trost and M. T. Rudd, *Org. Lett.*, 2003, **5**, 4599–4602.

287 A. C. Flick, M. J. A. Caballero and A. Padwa, *Org. Lett.*, 2008, **10**, 1871–1874.

288 J. Liu, R. P. Hsung and S. D. Peters, *Org. Lett.*, 2004, **6**, 3989–3992.

289 J. Liu, J. J. Swidorski, S. D. Peters and R. P. Hsung, *J. Org. Chem.*, 2005, **70**, 3898–3902.

290 H. Abe, S. Aoyagi and C. Kibayashi, *J. Am. Chem. Soc.*, 2005, **127**, 1473–1480.

291 J. F. Liu and C. H. Heathcock, *J. Org. Chem.*, 1999, **64**, 8263–8266.

292 G. A. Molander and M. Rönn, *J. Org. Chem.*, 1999, **64**, 5183–5187.

293 T. Arai, H. Abe, S. Aoyagi and C. Kibayashi, *Tetrahedron Lett.*, 2004, **45**, 5921–5924.

294 G. Pandey and V. Janakiram, *Chem.-Eur. J.*, 2015, **21**, 13120–13126.

295 G. Lapointe, K. Schenk and P. Renaud, *Org. Lett.*, 2011, **13**, 4774–4777.

296 G. Scheffler, H. Seike and E. J. Sorensen, *Angew. Chem., Int. Ed.*, 2000, **39**, 4593–4596.

297 M. Ousmer, N. A. Braun and M. A. Ciufolini, *Org. Lett.*, 2001, **3**, 765–767.

298 A.-J. Ma, Y.-Q. Tu, J.-B. Peng, Q.-Y. Dou, S.-H. Hou, F.-M. Zhang and S.-H. Wang, *Org. Lett.*, 2012, **14**, 3604–3607.

299 S. Ieda, T. Kan and T. Fukuyama, *Tetrahedron Lett.*, 2010, **51**, 4027–4029.

300 H.-H. Huo, H.-K. Zhang, X.-E. Xia and P.-Q. Huang, *Org. Lett.*, 2012, **14**, 4834–4837.

301 H.-H. Huo, X.-E. Xia, H.-K. Zhang and P.-Q. Huang, *J. Org. Chem.*, 2013, **78**, 455–465.

302 K. M. Brummond and J. L. Lu, *Org. Lett.*, 2001, **3**, 1347–1349.

303 K. M. Brummond and S.-P. Hong, *J. Org. Chem.*, 2005, **70**, 907–916.

304 M. Movassaghi, M. Tjandra and J. Qi, *J. Am. Chem. Soc.*, 2009, **131**, 9648–9650.

305 D. A. Evans, D. J. Adams and E. E. Kwan, *J. Am. Chem. Soc.*, 2012, **134**, 8162–8170.

306 M. Kuramoto, C. Tong, K. Yamada, T. Chiba, Y. Hayashi and D. Uemura, *Tetrahedron Lett.*, 1996, **37**, 3867–3870.

307 T. Chou, M. Kuramoto, Y. Otani, M. Shikano, K. Yazawa and D. Uemura, *Tetrahedron Lett.*, 1996, **37**, 3871–3874.

308 J. Kobayashi and M. Ishibashi, *Chem. Rev.*, 1993, **93**, 1753–1769.

309 D. Trauner, J. B. Schwarz and S. J. Danishefsky, *Angew. Chem., Int. Ed.*, 1999, **38**, 3542–3545.

310 M. W. Carson, G. Kim and S. J. Danishefsky, *Angew. Chem., Int. Ed.*, 2001, **40**, 4453–4456.

311 M. W. Carson, G. Kim, M. F. Hentemann, D. Trauner and S. J. Danishefsky, *Angew. Chem., Int. Ed.*, 2001, **40**, 4450–4452.

312 H. S. Christie and C. H. Heathcock, *Proc. Natl. Acad. Sci. U. S. A.*, 2004, **101**, 12079–12084.

313 S. Xu, H. Arimoto and D. Uemura, *Angew. Chem., Int. Ed.*, 2007, **46**, 5746–5749.

314 S. Omura, K. Matsuzaki, T. Fujimoto, K. Kosuge, T. Furuya, S. Fujita and A. Nakagawa, *J. Antibiot.*, 1991, **44**, 117–118.

315 S. Omura, T. Fujimoto, K. Otoguro, K. Matsuzaki, R. Moriguchi, H. Tanaka and Y. Sasaki, *J. Antibiot.*, 1991, **44**, 113–116.



316 R. H. Feling, G. O. Buchanan, T. J. Mincer, C. A. Kauffman, P. R. Jensen and W. Fenical, *Angew. Chem., Int. Ed.*, 2003, **42**, 355–357.

317 M. Shibasaki, M. Kanai and N. Fukuda, *Chem.-Asian J.*, 2007, **2**, 20–38.

318 E. J. Corey and G. A. Reichard, *J. Am. Chem. Soc.*, 1992, **114**, 10677–10678.

319 E. J. Corey and G. A. Reichard, *Tetrahedron Lett.*, 1993, **34**, 6973–6976.

320 E. J. Corey and S. Choi, *Tetrahedron Lett.*, 1993, **34**, 6969–6972.

321 T. Sunazuka, T. Nagamitsu, K. Matsuzaki, H. Tanaka, S. Omura and A. B. Smith, *J. Am. Chem. Soc.*, 1993, **115**, 5302.

322 H. Uno, J. E. Baldwin and A. T. Russell, *J. Am. Chem. Soc.*, 1994, **116**, 2139–2140.

323 N. Chida, J. Takeoka, N. Tsutsumi and S. Ogawa, *J. Chem. Soc., Chem. Commun.*, 1995, 793–794.

324 T. Nagamitsu, T. Sunazuka, H. Tanaka, S. Ōmura, P. A. Sprengeler and A. B. Smith, *J. Am. Chem. Soc.*, 1996, **118**, 3584–3590.

325 N. Chida, J. Takeoka, K. Ando, N. Tsutsumi and S. Ogawa, *Tetrahedron*, 1997, **53**, 16287–16298.

326 E. J. Corey, W. Li and T. Nagamitsu, *Angew. Chem., Int. Ed.*, 1998, **37**, 1676–1679.

327 E. J. Corey, W. Li and G. A. Reichard, *J. Am. Chem. Soc.*, 1998, **120**, 2330–2336.

328 J. S. Panek and C. E. Masse, *Angew. Chem., Int. Ed.*, 1999, **38**, 1093–1095.

329 H. Ooi, N. Ishibashi, Y. Iwabuchi, J. Ishihara and S. Hatakeyama, *J. Org. Chem.*, 2004, **69**, 7765–7768.

330 L. R. Reddy, P. Saravanan and E. J. Corey, *J. Am. Chem. Soc.*, 2004, **126**, 6230–6231.

331 L. R. Reddy, J.-F. Fournier, B. V. S. Reddy and E. J. Corey, *Org. Lett.*, 2005, **7**, 2699–2701.

332 A. Endo and S. J. Danishefsky, *J. Am. Chem. Soc.*, 2005, **127**, 8298–8299.

333 E. P. Balskus and E. N. Jacobsen, *J. Am. Chem. Soc.*, 2006, **128**, 6810–6812.

334 N. Fukuda, K. Sasaki, T. V. R. S. Sastry, M. Kanai and M. Shibasaki, *J. Org. Chem.*, 2006, **71**, 1220–1225.

335 N. P. Mulholland, G. Pattenden and I. A. S. Walters, *Org. Biomol. Chem.*, 2006, **4**, 2845–2846.

336 G. Ma, H. Nguyen and D. Romo, *Org. Lett.*, 2007, **9**, 2143–2146.

337 T. Ling, V. R. Macherla, R. R. Manam, K. A. McArthur and B. C. M. Potts, *Org. Lett.*, 2007, **9**, 2289–2292.

338 C. J. Hayes, A. E. Sherlock, M. P. Green, C. Wilson, A. J. Blake, M. D. Selby and J. C. Prodger, *J. Org. Chem.*, 2008, **73**, 2041–2051.

339 N. P. Mulholland, G. Pattenden and I. A. S. Walters, *Org. Biomol. Chem.*, 2008, **6**, 2782–2789.

340 T. Fukuda, K. Sugiyama, S. Arima, Y. Harigaya, T. Nagamitsu and S. Ōmura, *Org. Lett.*, 2008, **10**, 4239–4242.

341 K. Takahashi, M. Midori, K. Kawano, J. Ishihara and S. Hatakeyama, *Angew. Chem., Int. Ed.*, 2008, **47**, 6244–6246.

342 Y. Sato, H. Fukuda, M. Tomizawa, T. Masaki, M. Shibuya, N. Kanoh and Y. Iwabuchi, *Heterocycles*, 2010, **81**, 2239–2246.

343 Y. Kaiya, J.-I. Hasegawa, T. Momose, T. Sato and N. Chida, *Chem.-Asian J.*, 2011, **6**, 209–219.

344 N. Satoh, S. Yokoshima and T. Fukuyama, *Org. Lett.*, 2011, **13**, 3028–3031.

345 H. Nguyen, G. Ma, T. Gladysheva, T. Fremgen and D. Romo, *J. Org. Chem.*, 2011, **76**, 2–12.

346 D. J. Wardrop and E. G. Bowen, *Chem. Commun.*, 2005, 5106–5108.

347 S. Yoshioka, M. Nagatomo and M. Inoue, *Org. Lett.*, 2015, **17**, 90–93.

348 J. Kobayashi, F. Kanda, M. Ishibashi and H. Shigemori, *J. Org. Chem.*, 1991, **56**, 4574–4576.

349 T. Jahn, G. M. König, A. D. Wright, G. Wörheide and J. Reitner, *Tetrahedron Lett.*, 1997, **38**, 3883–3884.

350 S. Tsukamoto, K. Tane, T. Ohta, S. Matsunaga, N. Fusetani and R. W. M. van Soest, *J. Nat. Prod.*, 2001, **64**, 1576–1578.

351 K. Namba, T. Shinada, T. Teramoto and Y. Ohfune, *J. Am. Chem. Soc.*, 2000, **122**, 10708–10709.

352 P. M. Wehn and J. Du Bois, *J. Am. Chem. Soc.*, 2002, **124**, 12950–12951.

353 A. Hinman and J. Du Bois, *J. Am. Chem. Soc.*, 2003, **125**, 11510–11511.

354 K. Tran, P. J. Lombardi and J. L. Leighton, *Org. Lett.*, 2008, **10**, 3165–3167.

355 Y. Ichikawa, K. Okumura, Y. Matsuda, T. Hasegawa, M. Nakamura, A. Fujimoto, T. Masuda, K. Nakano and H. Kotsuki, *Org. Biomol. Chem.*, 2012, **10**, 614–622.

356 T. Kano, T. Hashimoto and K. Maruoka, *J. Am. Chem. Soc.*, 2006, **128**, 2174–2175.

357 M. P. Sibi, L. M. Stanley and T. Soeta, *Org. Lett.*, 2007, **9**, 1553–1556.

358 Y. Wang, X. Liu and L. Deng, *J. Am. Chem. Soc.*, 2006, **128**, 3928–3930.

359 J. C. S. Woo and D. B. MacKay, *Tetrahedron Lett.*, 2003, **44**, 2881–2883.

360 C. Drouin, J. C. S. Woo, D. B. MacKay and R. M. A. Lavigne, *Tetrahedron Lett.*, 2004, **45**, 7197–7199.

361 J. C. Lanter, H. F. Chen, X. Q. Zhang and Z. H. Sui, *Org. Lett.*, 2005, **7**, 5905–5907.

362 T. Shinada, E. Ikebe, K. Oe, K. Namba, M. Kawasaki and Y. Ohfune, *Org. Lett.*, 2007, **9**, 1765–1767.

363 T. Shinada, E. Ikebe, K. Oe, K. Namba, M. Kawasaki and Y. Ohfune, *Org. Lett.*, 2010, **12**, 2170.

364 K. Oe, T. Shinada and Y. Ohfune, *Tetrahedron Lett.*, 2008, **49**, 7426–7429.

365 K. Sankar, H. Rahman, P. P. Das, E. Bhimireddy, B. Sridhar and D. K. Mohapatra, *Org. Lett.*, 2012, **14**, 1082–1085.

366 T. Yoshimura, T. Kinoshita, H. Yoshioka and T. Kawabata, *Org. Lett.*, 2013, **15**, 864–867.

367 M. Nagatomo, H. Nishiyama, H. Fujino and M. Inoue, *Angew. Chem., Int. Ed.*, 2015, **54**, 1537–1541.

368 Y. Tahara, *J. Pharm. Soc. Jpn.*, 1909, **29**, 587–625.

369 Y. Tahara, *Biochem. Z.*, 1911, **30**, 255–275.



370 T. Goto, Y. Kishi, S. Takahashi and Y. Hirata, *Tetrahedron*, 1965, **21**, 2059–2088.

371 K. Tsuda, O. Amakasu, S. Ikuma, M. Kawamura, C. Tamura, R. Tachikawa and K. Sakai, *Chem. Pharm. Bull.*, 1964, **12**, 1357–1374.

372 R. B. Woodward and J. Z. Gougoutas, *J. Am. Chem. Soc.*, 1964, **86**, 5030.

373 A. Furusaki, Y. Tomiie and I. Nitta, *Bull. Chem. Soc. Jpn.*, 1970, **43**, 3325–3331.

374 A. Furusaki, Y. Tomiie and I. Nitta, *Bull. Chem. Soc. Jpn.*, 1970, **43**, 3332–3341.

375 F. A. Fuhrman, *Ann. N. Y. Acad. Sci.*, 1986, **479**, 1–14.

376 C. Y. Kao, *Ann. N. Y. Acad. Sci.*, 1986, **479**, 52–67.

377 W. Ulbricht, H. H. Wagner and J. Schmidtmayer, *Ann. N. Y. Acad. Sci.*, 1986, **479**, 68–83.

378 R. E. Weiss and R. Horn, *Ann. N. Y. Acad. Sci.*, 1986, **479**, 152–161.

379 B. K. Krueger, J. F. Worley and R. J. French, *Ann. N. Y. Acad. Sci.*, 1986, **479**, 257–268.

380 J. Chau and M. A. Ciufolini, *Mar. Drugs*, 2011, **9**, 2046–2074.

381 Y. Kishi, F. Nakatsubo, M. Aratani, T. Goto, S. Inoue and H. Kakoi, *Tetrahedron Lett.*, 1970, **11**, 5129–5132.

382 Y. Kishi, F. Nakatsubo, M. Aratani, T. Goto, S. Inoue, H. Kakoi and S. Sugiura, *Tetrahedron Lett.*, 1970, **11**, 5127–5128.

383 Y. Kishi, M. Aratani, T. Fukuyama, F. Nakatsubo, T. Goto, S. Inoue, H. Tanino, S. Sugiura and H. Kakoi, *J. Am. Chem. Soc.*, 1972, **94**, 9217–9219.

384 Y. Kishi, T. Fukuyama, M. Aratani, F. Nakatsubo, T. Goto, S. Inoue, H. Tanino, S. Sugiura and H. Kakoi, *J. Am. Chem. Soc.*, 1972, **94**, 9219–9221.

385 T. Nishikawa, M. Asai, N. Ohyabu, N. Yamamoto, Y. Fukuda and M. Isobe, *Tetrahedron*, 2001, **57**, 3875–3883.

386 N. Ohyabu, T. Nishikawa and M. Isobe, *J. Am. Chem. Soc.*, 2003, **125**, 8798–8805.

387 T. Nishikawa, D. Urabe and M. Isobe, *Angew. Chem., Int. Ed.*, 2004, **43**, 4782–4785.

388 N. Ünver and G. İ. Kaya, *Turk. J. Chem.*, 2005, **29**, 547–553.

389 S. Tian, W. Zi and D. Ma, *Angew. Chem., Int. Ed.*, 2012, **51**, 10141–10144.

390 Y. Shi, B. Yang, S. Cai and S. Gao, *Angew. Chem., Int. Ed.*, 2014, **53**, 9539–9543.

391 B. D. Morris and M. R. Prinsep, *J. Nat. Prod.*, 1999, **62**, 688–693.

392 C. C. Hughes and D. Trauner, *Angew. Chem., Int. Ed.*, 2002, **41**, 4556–4559.

393 K. Sakaguchi, M. Ayabe, Y. Watanabe, T. Okada, K. Kawamura, T. Shinada and Y. Ohfune, *Tetrahedron*, 2009, **65**, 10355–10364.

394 K. Chiyoda, J. Shimokawa and T. Fukuyama, *Angew. Chem., Int. Ed.*, 2012, **51**, 2505–2508.

395 A. Soheili and U. K. Tambar, *Org. Lett.*, 2013, **15**, 5138–5141.

396 M. O'Connor, C. Sun and D. Lee, *Angew. Chem., Int. Ed.*, 2015, **127**, 10101–10104.

