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On the enthalpic preference of fragment binding^{†‡}

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Detailed thermodynamic analysis of fragment binding revealed that unlike drug-like compounds, fragments bind with significant enthalpic preference. This observation is in line with the size dependency of binding enthalpy contributions and is also supported by a large body of experimental data from direct binding thermodynamic measurements. The enthalpy-driven binding of fragments represents a thermodynamic rationale for fragment-based drug discovery programs and suggests guidelines for fragment-based optimization programs.

Introduction

The thermodynamics of ligand–protein binding has recently gained increased attention due to the recognition that enthalpy and entropy changes accompanying binding can carry significant information on the quality of interactions. In the context of drug discovery, the thermodynamic quantities of binding have been suggested to be related to the drug-like properties of ligands and ultimately to the developability of drug candidates.^{1,2} The thermodynamic profile of ligand binding, *i.e.* the enthalpic and entropic contributions to the binding free energy, reflects the structural features and the interactions of the ligand and the protein. However, a detailed understanding of how structural changes affect thermodynamic profiles is still missing. It appears that the complex process of ligand binding which includes conformational changes, desolvation and formation of new interactions between the ligand and the protein gives various and, to a large extent, compensating contributions to both enthalpy and entropy. Nevertheless, it has been recognized that enthalpic contributions are often dominated by optimal polar interactions while entropic contributions come at a large extent from the desolvation that releases water molecules from the solvation shell and allows them to join bulk water. These trends manifest themselves in the ligand size dependence of thermodynamic profiles, that is, smaller ligands tend to bind with favourable enthalpy while favourable entropy changes become increasingly important in the binding of larger ligands.³ Since fragments usually bind in the high micromolar to the low

millimolar range and endothermic binding for small molecule–protein interactions is relatively rare, (around 20% for all interactions in the SCORPIO and BINDING databases), the enthalpy gain of fragment binding is not unexpected. In the current contribution, however, we show that – in contrast to drug-like molecules – the contribution of favourable fragment binding enthalpy tends to dominate over entropic terms. We analyse this trend by investigating the theoretical background and experimental data of the binding of small compounds. We focus our analyses on fragment-sized compounds whose heavy atom number does not exceed 22. It is worth mentioning that fragments have emerged as a key tool in drug discovery as they provide highly suitable chemical starting points for medicinal chemistry optimizations. In the forthcoming analysis, we show that in contrast to other opinions which are more ambivalent about the enthalpic signature of fragments^{4,5} they do have some particular features that lead to specific thermodynamic profiles and this discriminates them from larger compounds.

Theoretical considerations

Fragments used in drug discovery are small polar compounds. Owing to their small size, they typically exhibit low affinity towards protein targets. In order to detect their binding, a high screening concentration (typically >100 μM) is required and thus sufficient water solubility is needed. Thus, the fragments used in drug discovery must have a high polar surface area and a limited lipophilic character. Indeed, fragments have been shown to bind to protein hot spots through a limited number of optimal geometry H-bonds.⁶ It has to be noted that not all the H-bonds identified in ligand–protein complexes are expected to contribute to the binding free energy. Owing to the dependence of the H-bond energy on geometrical parameters, most notably on the donor–acceptor distance and the donor–hydrogen–acceptor angle,⁷ H-bonds

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with non-optimal geometrical parameters give little contribution to the binding free energy as they fail to overcome unfavourable desolvation effects. The estimated free energy gain for the formation of a single H-bond in water does not to exceed 6 kJ mol^{-1} in either peptide-peptide^{8,9} or small molecule interactions.¹⁰ Taking into account the fact that the average number of optimal geometry H-bonds in fragment-protein interactions was found to be 2, the free energy gain of H-bond formation would be 12 kJ mol^{-1} on average, based on these numbers. However, the above estimates for the free-energy of H-bonds are not directly related to fragment-protein binding whose thermodynamics may be different as protein hot spots are involved, as discussed below.

Fragments bind to protein hot spots^{6,11,12} which are good binding sites in general.¹³ No universal characterization of hot spots is available but there are certain specific features attributed to them. It was shown¹⁴ that water molecules displaced by fragments in the hot spot have high unfavourable excess entropies as calculated by the WaterMap method.¹⁵ Various fragments binding to the same protein hot spot appear to displace the same set of water molecules without affecting distal water networks.¹⁶ This is in contrast to larger compounds which disturb the water network in the binding pocket to a larger extent and therefore need to expel water molecules at a larger free energy cost. Another feature of fragment-hot spot binding was pointed out by Vajda and co-workers showing that pose conservation upon fragment expansion is associated with binding to the hot spot.¹² These observations make it reasonable to assume that the cited 6 kJ mol^{-1} estimation for the free energy gain of H-bonds represents a lower limit to that available for fragments at protein hot spots.

A clue to the enthalpic and entropic components of the free energy gain attributed to H-bonds is provided by an analysis of the polar surface area burial upon binding. Plots of enthalpy and entropy against buried polar surface area for an extensive data set show that $50\text{--}100 \text{ \AA}^2$ polar surface burial associated with the formation of two H-bonds corresponds to several tens of kJ mol^{-1} enthalpic gain and a smaller entropic penalty (Fig. 4c and d in ref. 17). It has to be noted that this high favourable enthalpy change is not proportional to the number of H-bonds; instead, it diminishes with increasing polar surface burial.¹⁷

Ligand-protein binding is accompanied by a rigid body entropy loss. Various estimates for its magnitude come from indirect experimental measurements and from computations (see *e.g.* ref 18). Specific analyses of fragment-protein binding suggest a $15\text{--}20 \text{ kJ mol}^{-1}$ entropic loss.^{19,20} This value only slightly increases for larger ligands, and thus a rigid body entropy loss represents a relatively more important barrier to fragment binding owing to the limited number of interactions available to compensate for it.

The desolvation of the ligand and the protein typically contributes favourably to binding. The magnitude of this effect correlates with the apolar surface buried in the complex formation. An analysis of the buried apolar surface area and

binding free energy data for a large number of protein-ligand complexes showed that the burial of approximately 20 \AA^2 apolar Connolly surface corresponds to 1 kJ mol^{-1} free energy gain.¹⁷ The enthalpy and entropy components of the binding free energy show weaker correlation with apolar surface area burial, but a trend of unfavourable enthalpy change and a twice as large favourable entropy change was identified.¹⁷ This is in line with the interpretation that these effects are dominated by the transition of water molecules from the solvation shell into the bulk water. The small size and polarity of the fragments represent limitations for the magnitude of their apolar surface area and for the corresponding thermodynamic effects upon binding. The apolar part of the Connolly surface of fragment-sized compounds in drug discovery is typically below 250 \AA^2 and in most cases it is much lower owing to the small size and the polar nature of the fragments.²¹ Taking into account the fact that the burial of the ligand surface is accompanied by the burial of the protein surface with a similar magnitude, the binding free energy gain corresponding to the burial of 250 \AA^2 of ligand apolar surface is 25 kJ mol^{-1} . This is an upper limit, and the binding free energy gain due to apolar desolvation is significantly lower in most cases. This means that apolar desolvation alone typically does not give a high enough contribution to compensate for the rigid body entropy loss and to produce productive binding for the fragments.

The above analysis shows that fragment-protein binding has to be additionally driven by the enthalpy gain of few optimal geometry polar interactions, most notably by H-bonds that are able to overcome the rigid body entropy loss in spite of the typically less important entropy gain coming from desolvation. Consequently, theoretical considerations suggest that fragment binding should be enthalpic in nature.

Experimental fragment thermodynamics data

There are relatively few studies that are aimed at the experimental investigation of fragment binding thermodynamics. In a previous study, we analysed ITC data of fragment-protein complexes and found that the majority of the fragments bind enthalpically.³ The distribution of thermodynamic signatures showed that except for charged ions (7 out of 159) all other fragments bind enthalpically. In line with this observation, the large ITC fragment dataset of Astex measured under standardized conditions shows that 621 out of the 624 compounds bind with favourable enthalpy.^{22,23} This trend also is reflected the present dataset of 138 neutral fragments and druglike compounds^{24,26} on 17 targets where the pronounced tendency of fragments to bind with more favourable enthalpy and less favourable entropy with respect to drug-like compounds is apparent in the ΔH vs. $-T\Delta S$ scatterplot (Fig. 1a). Furthermore, a statistical analysis of ΔH and $-T\Delta S$ values showed a statistically significant difference between fragments and drug-like compounds (Mann-Whitney *U*-test, $p < 0.005$). On the other hand, charged compounds fail to show such a trend owing to the increased role of the desolvation penalty accompanying binding



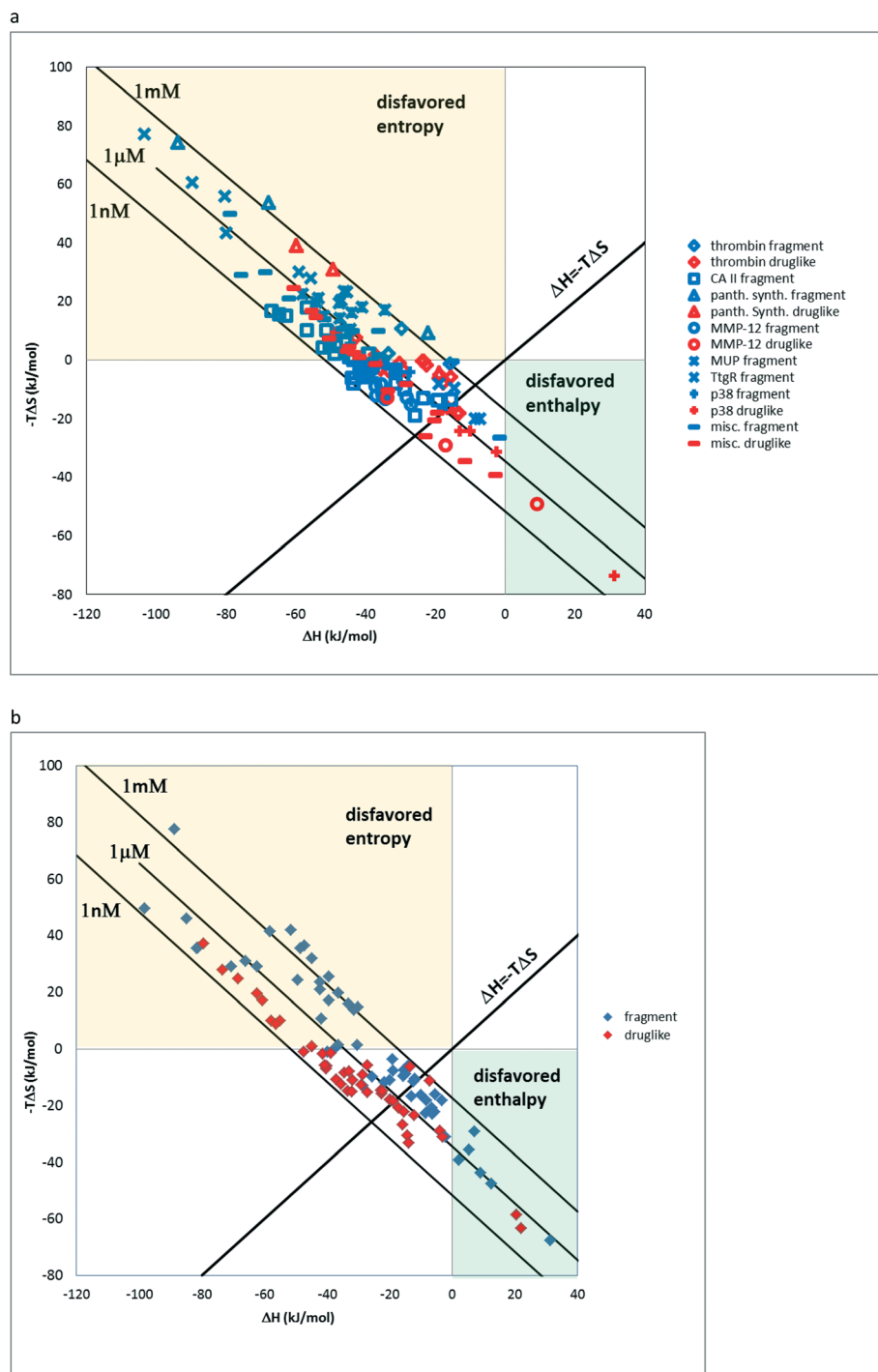


Fig. 1 a) Enthalpic and entropic components of binding for complexes of neutral fragments and drug-like molecules. b) Enthalpic and entropic components of binding for complexes of charged fragments and drug-like molecules.

(Fig. 1b). Interestingly enough, for thrombin fragments the authors compared the thermodynamic profiles obtained by direct and displacement titration.²⁵ This analysis revealed that only direct ITC data can be used to compare the thermodynamic profiles since these are not further affected by superimposed implications of the displacement ligand.¹⁶ The

authors suggested that the surrounding water network has a significant impact on the thermodynamic profiles; however, we argue that these effects in fact do not override the enthalpic preference of the fragments as suggested by the size dependency of the binding enthalpy^{3,27} and the above analysis.



Impact on fragment optimization

Investigating the enthalpic preference of fragment binding relative to optimized compounds, we performed a comparative structural and thermodynamic analysis of thrombin,²⁵ carbonic anhydrase II (ref. 28) and MMP12 (ref. 29) ligands (– Table 1). These examples were selected as both thermodynamic data and high resolution X-ray structures are available both for the fragments and for the optimized compounds. Owing to the selection of examples, our analysis focuses on true enzymatic hot spots and may not be valid for protein–protein interfaces. First we investigated the number of optimal H-bonds that contribute most significantly to the binding enthalpy gain upon binding. In the case of thrombin, we found that the benzamidine fragment forms four H-bonds within the S1 pocket of thrombin out of which two H-bonds fulfil the geometric criteria of optimal interactions.⁷ Although the corresponding optimized compound forms more H-bonds (6 optimized out of the 9 as reflected in the corresponding binding enthalpy contribution), the number of optimal H-bonds (2/4) does not change within the S1 pocket. FMAP calculations¹² revealed that all the fragments bind to the hot spot of protein targets. In the case of thrombin, the S1 pocket was identified by FMAP as the hot spot. The benzamidine fragment displaced 5 water molecules from the S1 pocket and other thrombin fragments displaced the very same set of water molecules.¹⁶ The optimized ligand displaced basically the same set of water molecules from the S1 pocket; however, we found 3 more water molecules displaced from other thrombin pockets. This indicates that in the case of thrombin, the larger ligand disturbs the water network more significantly than the fragment. Apolar desolvation, as approximated by the change of the apolar contact surface area, was found to be also more significant for the optimized ligand relative to

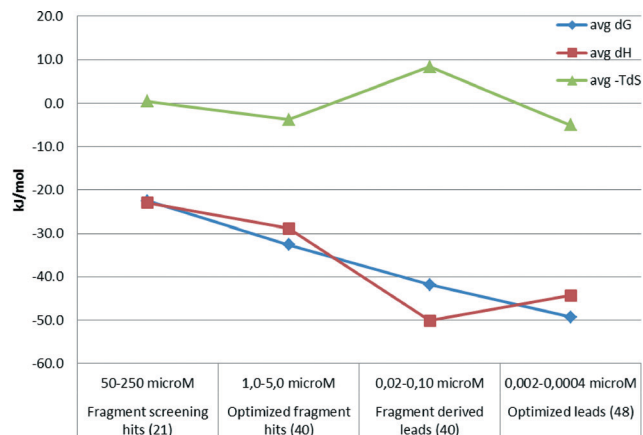


Fig. 2 Fragment optimization data from Astex.

the fragment (217 Å² versus 172 Å²). Both the larger number of water molecules replaced and the larger extent of apolar desolvation contributed to the increased entropy gain observed for the optimized compound. Considering the larger improvement found in the binding enthalpy, this optimization is enthalpy driven which improved the potency with the simultaneous increase in MW but controlled the lipophilicity effectively.

Benzenesulphonic amide, the carbonic anhydrase II fragment, forms only one H-bond within the binding cavity which is optimal. The most important interactions, however, are heteroatom–metal contacts formed between the Zn ion and sulphonic amide O and N atoms. The number of optimal H-bonds increased to 2 out of the total 3 for the optimized compound, which is in line with the enthalpy gain realized in the corresponding thermodynamic profile. The

Table 1 Structural, physicochemical and thermodynamic data of fragment and optimized ligand complexes of thrombin, carbonic anhydrase II (CA-II) and MMP12

	Thrombin		CA-II		MMP12	
	Frag	Opt	Frag	Opt	Frag	Opt
Ligand structure						
H-bonds (opt/all)	2/4	6/9	1/1	2/3	2/2	2/4
ΔCSA _{apo} (Å ²)	172	217	98	326	59	196
# of H ₂ O replaced	4	8	5	5	2	5
K _d (μm)	355	0.11	0.839	0.027	6180	0.0611
ΔG (kcal mol ⁻¹)	-4.73	-9.48	-8.3	-10.3	-3.01	-9.83
ΔH (kcal mol ⁻¹)	-3.32	-6.88	-7.5	-9.7	-3.18	-9.09
-TΔS (kcal mol ⁻¹)	-1.41	-2.60	-0.8	-0.6	0.17	-0.74
N _{heavy}	9	24	10	20	5	15
MW	120.2	331.4	157.2	290.3	75.1	230.2
log P	0.89	-0.26	0.58	1.38	-1.03	-0.53
Reference	25		28		29	
PDB	4UEH	2ZGX	4JSZ (H94C)	1G4O (F131V)	1Y93	3F1A
PDB (apo)	2UUF		1G3Z		4IJO	



benzenesulphonic amide moiety binds to the hot spot of the protein in both cases and displaces the very same number of water molecules. The apolar contact surface area is much larger for the optimized compounds. However, only a small part of the contact surface is used upon binding since the phenyl group goes out from the binding pocket. Consequently, only a minor improvement in the binding entropy was detected. Comparing the thermodynamic profiles of the initial fragment and the optimized compound, this optimization is also enthalpy driven and improved the potency at the expense of some limited increase in both MW and lipophilicity.

The starting fragment of the MMP12 optimization was hydroxamic acid that forms two H-bonds, both optimal, with the protein and also heteroatom-metal contacts with the active site Zn ion. The number of optimal H-bonds remained constant during the optimization; however, two new less than optimal H-bonds were detected. The fragment binds to the protein hot spot and displaces two water molecules. Although the position of the hydroxamic acid moiety does not change too much, the optimized compound displaces 3 more water molecules. The larger compound therefore disturbs the water network more significantly relative to the fragment. The apolar contact surface area is much increased and thus improved the contribution of the binding entropy to some extent. More importantly however, the introduced phenyl group forms van der Waals contacts with His218 that in addition to extra H-bonds make the binding of the optimized compound enthalpy dominant. Optimization of the initial fragment is again enthalpy driven which improves the potency with the simultaneous control of physicochemical parameters.

The above examples demonstrate that optimized fragment hits can be further developed with a more favourable binding enthalpy than the fragment screening hits they are derived from. In line with this finding, a thermodynamic analysis of 150 compounds from Astex drug discovery programs revealed that favorable binding enthalpy of fragment screening hits can be further improved by optimizing the compounds to leads (Fig. 2). However, lead optimization seems to be then associated with reduced favourable enthalpy, although with more favourable binding free energy. This is supported by the observation that late phase optimizations are usually entropy driven.³⁰ In fact, Astex published a thermodynamic optimization scheme of HSP90 inhibitors indicating that both the fragment starting points and the optimized compounds are clearly enthalpic.³¹

These observations together with supporting theoretical considerations suggest that:

- the higher the enthalpy content of binding for a fragment hit, the higher the chance that it can be optimized to a clinical candidate with advantageous thermodynamic properties and associated beneficial physicochemical and pharmacokinetic profiles;
- early phase optimization offers more opportunity for increasing the enthalpic character of binding;
- although the measured binding enthalpy is multifactorial and the individual contributions of intermolecular

interactions cannot be measured directly, the net enthalpy gain reflects the quality of interactions;³²

- the optimization of fragment-protein interactions must be explored without looking for overall free energy improvement before lead optimization really starts.

Conclusions

In summary, theoretical considerations and detailed examples support the hypothesis that fragments form an optimal geometry *via* polar interactions with the protein hot spot without any important apolar desolvation contribution and without significantly disturbing the water network in the binding site. These findings together with the large amount of raw thermodynamic data all demonstrate that fragments bind enthalpically. Analyses of fragment optimizations support that appropriate screening hit selection and early phase optimization offer the best opportunity to maximize the favourable enthalpy content of binding.

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Notes and references

- 1 J. E. Ladbury, G. Klebe and E. Freire, *Nat. Rev. Drug Discovery*, 2009, 9, 23.
- 2 G. G. Ferenczy and G. M. Keserű, *Drug Discovery Today*, 2010, 15, 919.
- 3 G. G. Ferenczy and G. M. Keserű, *J. Chem. Inf. Model.*, 2010, 50, 1536.
- 4 G. Klebe, *Nat. Rev. Drug Discovery*, 2015, 14, 95.
- 5 E. Ruhmann, M. Betz, A. Heine and G. Klebe, *J. Med. Chem.*, 2015, 58, 6960.
- 6 G. G. Ferenczy and G. M. Keserű, *J. Chem. Inf. Model.*, 2012, 52, 1039.
- 7 A. V. Morozov, T. Kortemme, K. Tsemekhman and D. Baker, *Proc. Natl. Acad. Sci. U. S. A.*, 2004, 101, 6946.
- 8 A. R. Fersht, *Trends Biochem. Sci.*, 1987, 12, 301.
- 9 S.-Y. Sheu, D.-Y. Yang, H. L. Selzle and E. W. Schlag, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, 100, 12683.
- 10 C. A. Hunter, *Angew. Chem., Int. Ed.*, 2004, 43, 5310.
- 11 S. Barelrier, J. Pons, O. Marcillat, J. M. Lancelin and I. Krimm, *J. Med. Chem.*, 2010, 53, 2577.
- 12 D. Kozakov, D. R. Hall, S. Jehle, L. Luo, S. O. Ochiana, E. V. Jones, M. Pollastri, K. N. Allen, A. Whitty and S. Vajda, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, 112, E2585.
- 13 W. L. DeLano, *Curr. Opin. Struct. Biol.*, 2002, 12, 14.
- 14 Y. Yang, F. C. Lightstone and S. E. Wong, *Expert Opin. Drug Discovery*, 2013, 1.
- 15 O. Ichiara, Y. Shimada and D. Yoshidome, *ChemMedChem*, 2014, 9, 2708.
- 16 E. Rühmann, M. Betz, A. Heine and G. Klebe, *J. Med. Chem.*, 2015, 58, 6960.



- 17 T. S. Olsson, M. A. Williams, W. R. Pitt and J. E. Ladbury, *J. Mol. Biol.*, 2008, **384**, 1002.
- 18 S. J. Irudayam and R. H. Henchman, *J. Phys. Chem. B*, 2009, **113**, 5871.
- 19 C. W. Murray and M. L. Verdonk, *J. Comput.-Aided Mol. Des.*, 2002, **16**, 741.
- 20 V. Borsi, V. Calderone, M. Fragai, C. Luchinat and N. Sart, *J. Med. Chem.*, 2010, **53**, 4285.
- 21 The apolar Connolly surfaces for the 91 fragments included in the analysis of thermodynamic profiles are available in the Supplementary Information.
- 22 E. Chiarparin, N. Amin and G. Williams, The behaviour of physico-chemical properties during fragment progression. FRAGMENTS III - Third RSC-BMCS Fragment-based Drug Discovery meeting, March 7–8; Poster 31, GlaxoSmithKline, Stevenage, UK, 2011.
- 23 F. E. Torres, M. I. Recht, J. E. Coyle, R. H. Bruce and G. Williams, *Curr. Opin. Struct. Biol.*, 2007, **20**, 598.
- 24 C. A. Lipinski, F. Lombardo, B. W. Dominy and P. J. Feeney, *Adv. Drug Delivery Rev.*, 2001, **46**, 3.
- 25 E. Rühmann, M. Betz, M. Fricke, A. Heine, M. Schäfer and G. Klebe, *Biochim. Biophys. Acta*, 2015, **1850**, 647.
- 26 Compounds with either pK_{basic} higher than 7 or pK_{acidic} lower than 7 were considered as charged. pK values were calculated with JChem for Excel, version 15.8, Chemaxon, Budapest, Hungary, 2014.
- 27 C. H. Reynolds and M. K. Holloway, *ACS Med. Chem. Lett.*, 2011, **2**, 433.
- 28 A. D. Scott, C. Phillips, A. Alex, M. Flocco, A. Bent, A. Randall, R. O'Brien, L. Damian and L. H. Jones, *ChemMedChem*, 2009, **4**, 1985.
- 29 I. Bertini, V. Calderone, M. Fragai, A. Giachetti, M. Loconte, C. Luchinat, M. Maletta, C. Nativi and K. J. Yeo, *J. Am. Chem. Soc.*, 2007, **129**, 2466.
- 30 M. M. Hann and G. M. Keserű, *Nat. Rev. Drug Discovery*, 2012, **11**, 355.
- 31 G. Williams, The PyramidTM Approach to Fragment-Based Biophysical Screening, in *Label-Free Technologies For Drug Discovery*, ed. M. Cooper and L. M. Mayr, John Wiley & Sons, New York, USA, 2011, p. 241.
- 32 E. Freire, *Drug Discovery Today*, 2008, **13**, 869.

