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Biological processing of dinuclear ruthenium complexes in eukaryotic cells

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The biological processing – mechanism of cellular uptake, effects on the cytoplasmic and mitochondrial membranes, intracellular sites of localisation and induction of reactive oxygen species – of two dinuclear polypyridylruthenium(II) complexes has been examined in three eukaryotic cells lines. Flow cytometry was used to determine the uptake of $[\{\text{Ru}(\text{phen})_2\}_2\{\mu\text{-bb}_{12}\}]^{4+}$ (Rubb₁₂) and $[\text{Ru}(\text{phen})_2(\mu\text{-bb}_7)\text{Ru}(\text{tpy})\text{Cl}]^{3+}$ (Rubb₇-Cl, where phen = 1,10-phenanthroline, tpy = 2,2':6',2''-terpyridine and bb_n = bis[4(4'-methyl-2,2'-bipyridyl)]-1,*n*-alkane) in baby hamster kidney (BHK), human embryonic kidney (HEK-293) and liver carcinoma (HepG2) cell lines. The results demonstrated that the major uptake mechanism for Rubb₁₂ and Rubb₇-Cl was active transport, although with a significant contribution from carrier-assisted diffusion for Rubb₁₂ and passive diffusion for Rubb₇-Cl. Flow cytometry coupled with Annexin V/TO-PRO-3 double-staining was used to compare cell death by membrane damage or apoptosis. Rubb₁₂ induced significant direct membrane damage, particularly with HepG2 cells, while Rubb₇-Cl caused considerably less membrane damage but induced greater levels of apoptosis. Confocal microscopy, coupled with JC-1 assays, demonstrated that Rubb₁₂ depolarises the mitochondrial membrane, whereas Rubb₇-Cl had a much smaller affect. Cellular localisation experiments indicated that Rubb₁₂ did not accumulate in the mitochondria, whereas significant mitochondrial accumulation was observed for Rubb₇-Cl. The effect of Rubb₁₂ and Rubb₇-Cl on intracellular superoxide dismutase activity showed that the ruthenium complexes could induce cell death *via* a reactive oxygen species-mediated pathway. The results of this study demonstrate that Rubb₁₂ predominantly kills eukaryotic cells by damaging the cytoplasmic membrane. As this dinuclear ruthenium complex has been previously shown to exhibit greater toxicity towards bacteria than eukaryotic cells, the results of the present study suggest that metal-based cationic oligomers can achieve selective toxicity against bacteria, despite exhibiting a non-specific membrane damage mechanism of action.

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Introduction

There is considerable interest in developing ruthenium complexes as therapeutic agents, and a variety of coordinatively-labile mononuclear ruthenium(II) and ruthenium(III) complexes have shown significant promise as anticancer drugs.^{1–6} Furthermore, a range

of inert mononuclear ruthenium(II) complexes have shown promising *in vitro* anticancer and/or antimicrobial properties.^{7–15} More recently, there has been increasing interest in “tuning” the structure of the ruthenium complexes to increase their activity and selectivity for bacterial or cancer cells. One such strategy, the synthesis of cationic oligomers,^{16–19} takes advantage of the greater presence of negatively-charged components in the bacterial membrane compared to eukaryotic cells.^{20,21} The cell wall and outer membrane of bacteria contain a higher proportion of negatively-charged phospholipids – *e.g.* phosphatidylglycerol, as well as a variety of negatively-charged macromolecules such as teichoic acids and lipopolysaccharides – than do eukaryotic cells.^{20,21}

We have synthesised and examined the antibacterial activities of a series of dinuclear polypyridylruthenium(II) complexes $[\{\text{Ru}(\text{phen})_2\}_2\{\mu\text{-bb}_n\}]^{4+}$ (Rubb_n, see Fig. 1) where (phen = 1,10-phenanthroline and bb_n = bis[4(4'-methyl-2,2'-bipyridyl)]-1,*n*-alkane).^{22–25} The Rubb_n complexes with a short linking chain

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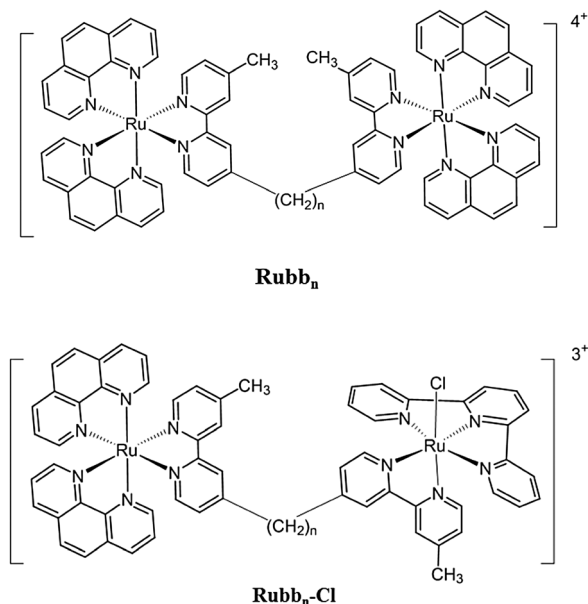


Fig. 1 Structure of the Rubb_n and $\text{Rubb}_n\text{-Cl}$ complexes ($n = 7, 12$ and 16).

(bb_2 , bb_5 and bb_7) showed little or no antibacterial activity, whereas Rubb_{12} showed excellent activity and was 40–100 fold less toxic towards eukaryotic cells.^{22,25} However, it was noted that Rubb_{12} showed very similar 24-hour IC_{50} values towards three significantly different eukaryotic cell lines – a baby hamster kidney (BHK), a human embryonic kidney (HEK-293) and a human liver carcinoma (HepG2).²⁵ The similar cytotoxicity data between the eukaryotic cell lines suggest a common, non-specific, mechanism of action. Previous studies with bacteria demonstrated that although Rubb_{12} preferentially localises at ribosomes within a bacterial cell,²⁶ it also significantly permeabilises the cytoplasmic membrane.²⁷ Non-specific membrane damage would likely result in the ruthenium complex exhibiting very similar IC_{50} values against the three eukaryotic cell lines. However, a previous study suggested that the Rubb_n complexes enter L1210 murine leukaemia cells largely by passive diffusion, accumulate in the mitochondria and induce cell death by apoptosis.²⁸ By contrast, the results of a confocal microscopy study with the BHK, HEK-293 and HepG2 cell lines demonstrated that Rubb_{12} did not accumulate in the mitochondria, but predominantly localised in the nucleolus of the cell.²⁵ Taken together, the combined results suggest that the biological processing of the Rubb_n complexes in white blood cells (L1210) is significantly different to that in kidney and liver cells (BHK, HEK-293 and HepG2). In order to gain a deeper understanding of the effect of the Rubb_n complexes on organ cells, and in particular the organs responsible for processing drugs, it was of interest to determine the cellular uptake mechanism and mechanism of cell death of the Rubb_{12} complex in the BHK, HEK-293 and HepG2 cells.

Although this study was aimed at understanding the biological processing of the Rubb_n complexes in eukaryotic cells, it also provides important information for the development of cationic antimicrobial or anticancer oligomers in general.

As previously noted, there is considerable current interest in the use of cationic oligomers/polymers for combating bacterial resistance to current drugs.^{16,19} However, because many of these potential drugs disrupt the bacterial membrane, their clinical application could be limited as they would also interact with eukaryotic membranes in a similar manner. As alkyl chains are commonly used to link the monomeric units of antimicrobial (and anticancer) cationic oligomers/polymers,^{16–19} the results of this study could well be applicable to a wider range of drugs than the Rubb_n complexes.

In addition to the Rubb_n complexes, we have also synthesised and examined the biological activities of a series of dinuclear complexes that contain one inert metal centre and one coordinatively-labile metal centre (the $\text{Rubb}_n\text{-Cl}$ complexes, see Fig. 1).²⁹ Interestingly, while each of the $\text{Rubb}_{12}\text{-Cl}$ and $\text{Rubb}_{16}\text{-Cl}$ complexes showed similar toxicities across the BHK, HEK-293 and HepG2 cell lines, $\text{Rubb}_7\text{-Cl}$ showed surprising differences in cytotoxicities.²⁹ Against the BHK cells, $\text{Rubb}_7\text{-Cl}$ was by far the least toxic of the $\text{Rubb}_n\text{-Cl}$ and Rubb_n complexes (24 h- $\text{IC}_{50} = 238 \mu\text{M}$); whereas by contrast, against the HepG2 cell line it was by far the most toxic (24 h- $\text{IC}_{50} = 3.7 \mu\text{M}$).²⁹ The significantly different response exhibited by the BHK, HEK-293 and HepG2 cells to $\text{Rubb}_7\text{-Cl}$ suggests that the ruthenium complex does not act in a non-specific manner (*e.g.* membrane damage), but through a specific pathway that leads to apoptosis. Consequently, the cellular uptake mechanism and mechanism of cell death for $\text{Rubb}_7\text{-Cl}$ against the BHK, HEK-293 and HepG2 cells was also examined and compared to that obtained for Rubb_{12} .

Results

Cellular uptake mechanism of the ruthenium complexes

Cellular uptake studies {described in (a)–(d) below} were carried out using flow cytometry for the Rubb_{12} and $\text{Rubb}_7\text{-Cl}$ complexes in the kidney and liver cell lines. These studies aimed to differentiate between three major uptake pathways: (1) active transport, which is energy-dependent; (2) passive diffusion, an energy-independent form of molecular transport; and (3) carrier-assisted diffusion, an energy-independent pathway, facilitated by membrane protein carriers such as organic cation transporters (OCT).

(a) **Effect of cell membrane permeability on uptake of Rubb_{12} and $\text{Rubb}_7\text{-Cl}$.** To differentiate between passive uptake and active transport/carrier-facilitated diffusion, the cellular uptake of Rubb_{12} and $\text{Rubb}_7\text{-Cl}$ were initially examined in fully permeabilised (heat treated, representing the maximum cellular uptake possible) and PFA-fixed (active transport/carrier-impaired) cells (Fig. 2). PFA-fixation causes cross-linking of primary amino groups in proteins that results in protein related uptake of the ruthenium complex being blocked. Consequently, the cellular uptake of the ruthenium complexes after heat treatment or PFA-fixation provides useful comparative data to that obtained under control conditions (37 °C in the absence of metabolic inhibitors or activators).



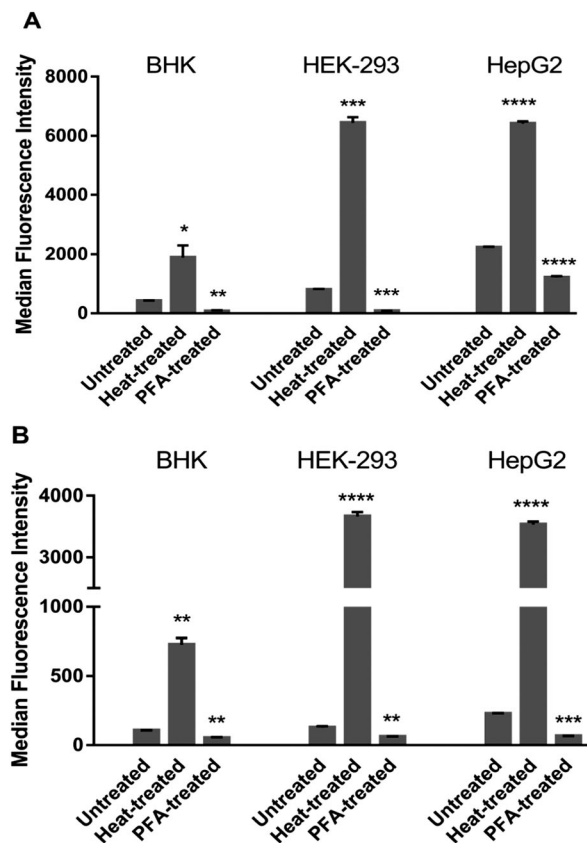


Fig. 2 Cellular uptake as median fluorescence intensity (MFI) values of 20 μ M ruthenium complexes (A, Rubb_{12} and B, $\text{Rubb}_{7\text{-Cl}}$) after a 4-hour incubation at 37 $^{\circ}\text{C}$ in untreated, heat-treated and PFA-fixed cells for the three cell lines BHK, HEK-293 and HepG2.

The Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$ complexes have different luminescence signal strengths. Therefore, the median fluorescence intensity (MFI) values cannot be used to compare cellular uptake of different complexes, but only for analysing changes in the uptake of one complex under different conditions in different cells.

Both complexes accumulated significantly more in heat-treated cells than in untreated and PFA-fixed cells irrespective of the cell line. In control and PFA-fixed samples, the uptake generally followed $\text{HepG2} \geq \text{HEK-293} > \text{BHK}$ for both complexes. However, in heat-treated cells, MFIs in HEK-293 cells were slightly higher than HepG2, and the MFIs in both human cell lines were more than 3-fold higher than for the baby hamster kidney BHK cells. After PFA-fixation, the cellular uptake of Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$ was strongly impeded by 50–90% compared with the untreated controls in the three cell lines.

(b) Temperature-dependent cellular uptake of Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$. To differentiate between temperature-dependent and -independent transport routes, the cellular uptake of Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$ were investigated at 4, 24 and 37 $^{\circ}\text{C}$ by flow cytometry in the three cell lines. Relative MFI values (percentage of MFI at each temperature *versus* MFI at 37 $^{\circ}\text{C}$) are summarised in Fig. 3. Uptake increased with temperature for both ruthenium complexes in the three cell lines. At 4 $^{\circ}\text{C}$, almost no uptake was observed in BHK and HEK-293 cells, with a slightly higher

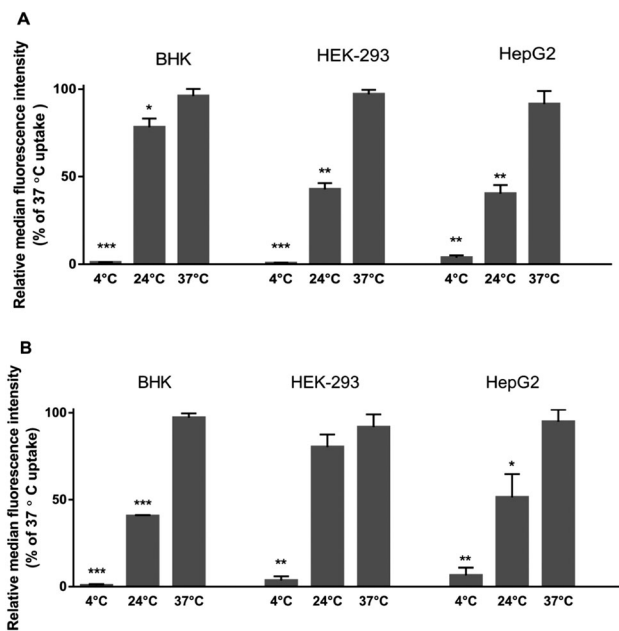


Fig. 3 Relative cellular uptake (MFIs at each temperature to MFIs at 37 $^{\circ}\text{C}$) of (A) Rubb_{12} and (B) $\text{Rubb}_{7\text{-Cl}}$ at 4, 24 and 37 $^{\circ}\text{C}$ in BHK, HEK-293 and HepG2 cells after a 4-hour incubation. MFIs: medium fluorescence intensities.

uptake in HepG2. At 24 $^{\circ}\text{C}$, the uptake of Rubb_{12} increased sharply in BHK cells, as it did for $\text{Rubb}_{7\text{-Cl}}$ in HEK-293, and moderately increased (40–50%) in the other cell lines. These data suggest a larger contribution to uptake of Rubb_{12} by an energy-independent process in BHK, but predominantly an active uptake mechanism in HEK-293. However, the opposite was observed for $\text{Rubb}_{7\text{-Cl}}$, where a larger contribution to uptake by an energy-independent process was seen in HEK-293 cells. By contrast, the uptake of both ruthenium complexes in HepG2 cells appeared to be predominantly by an active process.

(c) Dependence of cellular uptake of Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$ on ATP production. Flow cytometry experiments were conducted to confirm the ATP requirement for the cellular uptake of the ruthenium complexes (see Fig. 4). Cells were pretreated with the glycolysis inhibitor 2-deoxy-glucose³⁰ and the mitochondria inhibitor oligomycin to inhibit ATP production,³¹ and with D-glucose to stimulate ATP production.

The ATP-production stimulator D-glucose did not enhance uptake of the ruthenium complexes in BHK and HEK-293 cells, with uptake of Rubb_{12} in BHK even slightly lower. This suggested that the cellular ATP quota was optimal, preventing further accumulation by the treatment. However, for HepG2 cells, D-glucose treatment did induce slightly greater accumulation for Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$. The inhibition of ATP production caused a significant decrease in the uptake of both ruthenium complexes in all three cell lines, indicating that uptake was predominantly through an energy-dependent (active) process.

(d) Effect of organic cation transport (OCT) inhibitors on the uptake of Rubb_{12} and $\text{Rubb}_{7\text{-Cl}}$. Organisms use polyspecific organic cation (OCT) transporters for facilitating the diffusion of endogenous organic cations, cationic drugs and toxins across the cell membrane.³² The possibility of energy-independent



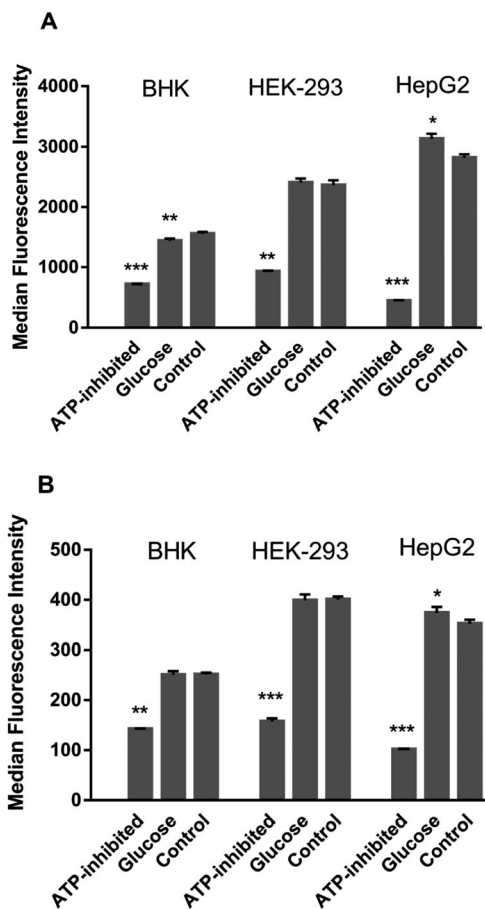


Fig. 4 Cellular uptake (MFIs) of (A), Rubb₁₂ and (B), Rubb_{7-Cl} (4-hour incubation) in ATP-inhibitor-treated, glucose-treated and untreated BHK, HEK-293 and HepG2 cells. MFI: medium fluorescence intensity.

carrier-assisted diffusion *via* OCTs was investigated by flow cytometry (see Fig. 5). To determine if the ruthenium complexes were transported by OCTs, cells were pre-treated with tetraalkylammonium salts or procainamide to inhibit the function of organic cation transporters then incubated with the ruthenium complexes. In BHK cells, uptake of Rubb₁₂ was inhibited slightly but significantly by the tetra-*n*-butylammonium, tetramethylammonium and tetraethylammonium salts, suggesting OCT1, OCT2 and OCTN1 could be involved in transportation. Interestingly, uptake appeared to be stimulated for Rubb_{7-Cl}, suggesting that these OCTs were not involved in the uptake of Rubb_{7-Cl}. A significant decrease in uptake of both ruthenium complexes was observed in OCT-inhibited HEK-293 cells. This observation suggests that HEK-293 cells can transport both ruthenium complexes by OCT-facilitated diffusion as an additional pathway to active transport mechanism. Rubb₁₂ uptake was significantly lower (by 32–42%) in the OCT-inhibited HepG2 cells, suggesting a significant contribution of OCTs in the transportation process. By contrast, OCTs do not appear to be involved in the uptake of Rubb_{7-Cl} in HepG2 cells.

Detection of apoptosis and membrane damage

Annexin V/TO-PRO-3 double-staining assays were carried out with BHK and HepG2 cells to compare cell death by membrane

damage or apoptosis for Rubb₁₂ and Rubb_{7-Cl}. Early stages of apoptosis are characterised by the translocation of phosphatidylserine from the inner leaflet of the plasma membrane to the outer leaflet, which can be visualised by binding to the Annexin V fluorescent conjugate.³³ As cell death progresses, TO-PRO-3 (a cell-impermeant dye) diffuses into the cells with compromised membrane integrity and binds to DNA.³³ Alternatively, if the cytoplasmic membrane is directly damaged by the ruthenium complex, TO-PRO-3 will enter the cell and bind to DNA without any translocation of phosphatidylserine from the inner leaflet of the plasma membrane to the outer leaflet. Accordingly, in the flow cytometry assays, apoptosis is demonstrated by cells “moving” from Q4 (bottom-left quadrant) to Q3 (bottom-right) and then finally to Q2 (top-right). On the other hand, direct membrane damage is detected by cells moving from Q4 to Q1 (top-left), and then to Q2 as the cell becomes totally permeabilised (allowing the large Annexin V fluorescent conjugate into the cell) and then possibly to Q3 as TO-PRO-3 is displaced from the DNA by the ruthenium complex.

Time-course (4-, 24-, 48- and 72-hours) flow cytometry was conducted using BHK cells with Rubb₁₂ and Rubb_{7-Cl} at half of their 24 h-IC₅₀ concentrations (50 and 100 μM, respectively). A small percentage of the BHK cells (5–10%) were always damaged (observed in Q1) during the trypsin treatment required for cell harvesting; however, little membrane damage was observed for HepG2 cells. After incubation with Rubb₁₂, viable BHK cells (bottom-left quadrant, Q4) decreased gradually with incubation time (see Fig. 6). Noticeably, there was a sharp increase in the top-left quadrant, indicating that half the cells had significant membrane damage, while the early apoptotic cells (bottom-right quadrant, Q3) only increased slowly and to a lesser extent. BHK cells treated with Rubb_{7-Cl} showed a different cell population distribution to that observed with Rubb₁₂. The percentage of cells that exhibited direct membrane damage was considerably smaller (Q1), but the percentage of cells exhibiting apoptosis (Q2 and Q3) was significantly higher than was observed with Rubb₁₂.

Time-course Annexin V/TO-PRO-3 double-staining assays were also carried out with HepG2 cells treated with either Rubb₁₂ or Rubb_{7-Cl} (see Fig. 7). For Rubb₁₂, extensive membrane damage was observed after a 4-hour incubation, with over 50% of the cell population observed in Q1 after 24 hours. Interestingly, after a 72-hour incubation a significant percentage of the cells were Annexin positive and TO-PRO-3 negative (Q3). Given that most of the cells were non-viable due to membrane damage at 4 and 24 hours, the significant percentage of cells in Q3 after a 72-hour incubation suggests the DNA was saturated with Rubb₁₂, rather being indicative of early apoptosis. Saturation with Rubb₁₂ would inhibit TO-PRO-3 from binding to DNA, leading to the observed low levels of TO-PRO-3 fluorescence.

HepG2 cells treated with 10 μM Rubb_{7-Cl} appeared to induce relatively weak responses, with few cells exhibiting significant membrane damage (Q1) and even fewer cells being early-apoptotic (Q3). To further amplify cell responses, the assays were repeated with a Rubb_{7-Cl} concentration of 20 μM. The increased concentration of Rubb_{7-Cl} did slightly enhance the responses, with the



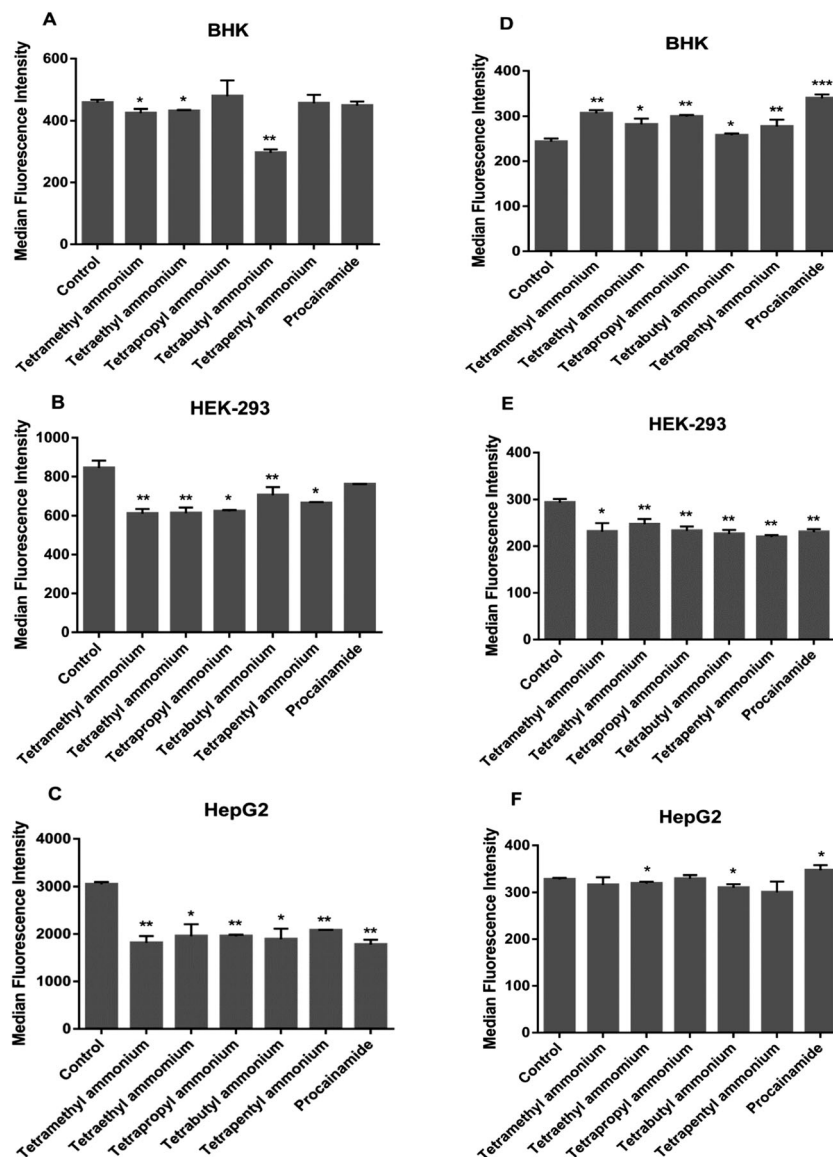


Fig. 5 Effect of OCT inhibition on Rubb₁₂ (A–C) and Rubb_{7-Cl} (D–F) uptake in BHK, HEK-293 and HepG2 cells after a 4-hour incubation.

percentage of cells exhibiting direct membrane damage doubling (Q1); however there was only a very slight increase in the number of early apoptotic cells (Q3). As the 24 h-IC₅₀ is 3.7 μ M, the flow cytometry results suggest that Rubb_{7-Cl} is not cytotoxic against HepG2 cells due to direct membrane damage or any process leading to apoptosis.

Mitochondrial membrane potential

Mitochondrial dysfunction plays an important role in different cell death mechanisms, and the depolarisation of the mitochondrial membrane potential (MMP) due to changes in the mitochondrial permeability has been demonstrated to be an early stage of mitochondrial dysfunction.³⁴ To evaluate the disruption of mitochondrial function induced by Rubb₁₂ and Rubb_{7-Cl}, the change in the mitochondrial membrane potential was determined by confocal microscopy using JC-1 assays.³⁵ Functional polarised mitochondria have a high MMP promoting

the formation of red fluorescent JC-1 aggregates, while depolarised mitochondria with low MMP show a diffuse green fluorescence from JC-1 monomers.

BHK cells incubated with 50 μ M of Rubb₁₂ and Rubb_{7-Cl} showed different changing patterns in MMP (Fig. 8). After a 4-hour incubation, Rubb₁₂ treatment affected the distribution of mitochondria in BHK cells, with low MMP mitochondria spreading throughout the cytoplasm. Mitochondria with high MMP appeared to be confined to the cell periphery. Moreover, some polarised mitochondria appeared elongated. By contrast, in Rubb_{7-Cl} treated BHK cells very few low MMP mitochondria were observed, with the high MMP mitochondria red fluorescence being maintained.

HepG2 cells incubated with 20 μ M of Rubb₁₂ and Rubb_{7-Cl} displayed some differences in the staining pattern (Fig. 9). Treatment with Rubb₁₂ led to the localisation of depolarised mitochondria in the perinuclear region. The presence of several



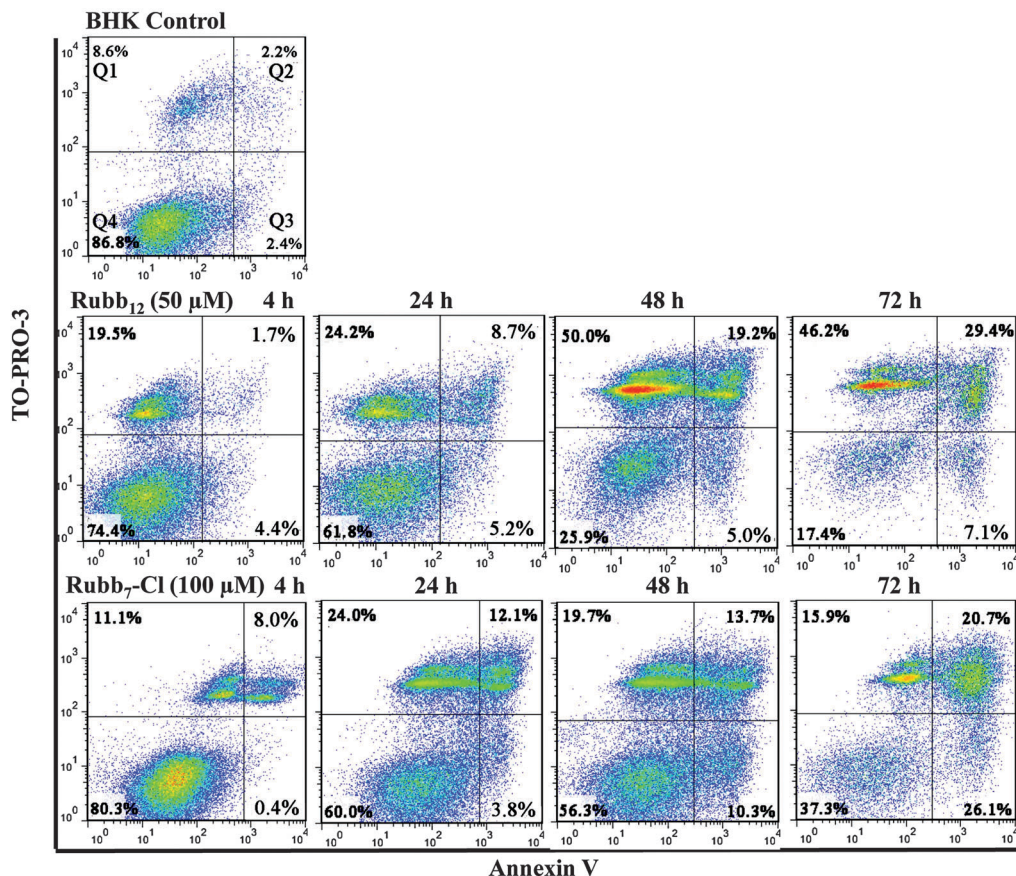


Fig. 6 Time-course Annexin V/TO-PRO-3 staining of BHK cells (control, top) and with treatment by the ruthenium complexes (middle: Rubb₁₂ and bottom: Rubb_{7-Cl}). Cells plotted in each quadrant represent the following: bottom-left (Annexin V-negative/TO-PRO-3-negative), viable cells; bottom-right (Annexin V-positive/TO-PRO-3-negative), early apoptotic cells; top-right (Annexin V-positive/TO-PRO-3-positive), late apoptotic/necrotic cells; top-left (Annexin V-negative/TO-PRO-3-positive), membrane-damaged dead cells.

condensed green hotspots could be indicative of condensed mitochondria with low MMP in response to Rubb₁₂ treatment. On the other hand, mitochondria with high MMP displayed condensed red fluorescence. After incubation with Rubb_{7-Cl} at the concentration of 20 μM, HepG2 cells displayed only slightly decreased red fluorescence of high MMP mitochondria, with only a few low MMP mitochondria observed in the perinuclear region.

Additionally, time-course MMP changes (0-, 4- and 20-hour incubations) were monitored in BHK cells with Rubb₁₂ and Rubb_{7-Cl}. As expected, longer incubation time led to increased depolarisation of mitochondria (data not shown).

Cellular localisation

As the JC-1 assays demonstrated that Rubb₁₂ and Rubb_{7-Cl} affected the mitochondrial membrane potential, it was of interest to determine if the ruthenium complexes localised to any extent in the mitochondria. The cellular localisation of high and low concentrations (relative to 24 h-IC₅₀ values) of Rubb₁₂ and Rubb_{7-Cl} in BHK cells has been previously reported.²⁹ Both Rubb₁₂ and Rubb_{7-Cl} preferentially accumulated in the nucleolus at low concentrations, but exhibited significant DNA binding at higher concentrations. Furthermore, mitochondrial localisation was observed for Rubb_{7-Cl} at high concentration,

but not for Rubb₁₂.²⁹ The localisation of both ruthenium complexes at low concentrations (5 μM) in HepG2 cells has also been reported.²⁹ However, the localisation of higher concentrations of Rubb₁₂ and Rubb_{7-Cl} in HepG2 cells has not been reported. Although considerable Rubb₁₂ phosphorescence was observed in the cytoplasm of HepG2 cells, no co-localisation was observed in the mitochondria (see Fig. 10). Co-localisation experiments with ERtracker indicated that a significant proportion of the cytoplasmic Rubb₁₂ phosphorescence was due to accumulation in the ER (Fig. 10). By contrast, Rubb_{7-Cl} showed intense co-localisation with mitochondria (yellow overlay), but little in the ER (Fig. 10).

Reactive oxygen superoxide dismutase assays

The primary superoxide scavenger, superoxide dismutase (SOD), is an attractive target for pharmacological intervention.^{36–38} To investigate whether the treatment with ruthenium complexes affected intracellular SOD activity, SOD assays were conducted on BHK, HEK-293 and HepG2 cells after a 20-hour incubation with Rubb₁₂ and Rubb_{7-Cl} (Fig. 11). Compared with the untreated controls, BHK cells appeared to slightly increase SOD activity in response to treatment with the ruthenium complexes, with a non-significant increase for Rubb₁₂ and Rubb_{7-Cl} being observed.



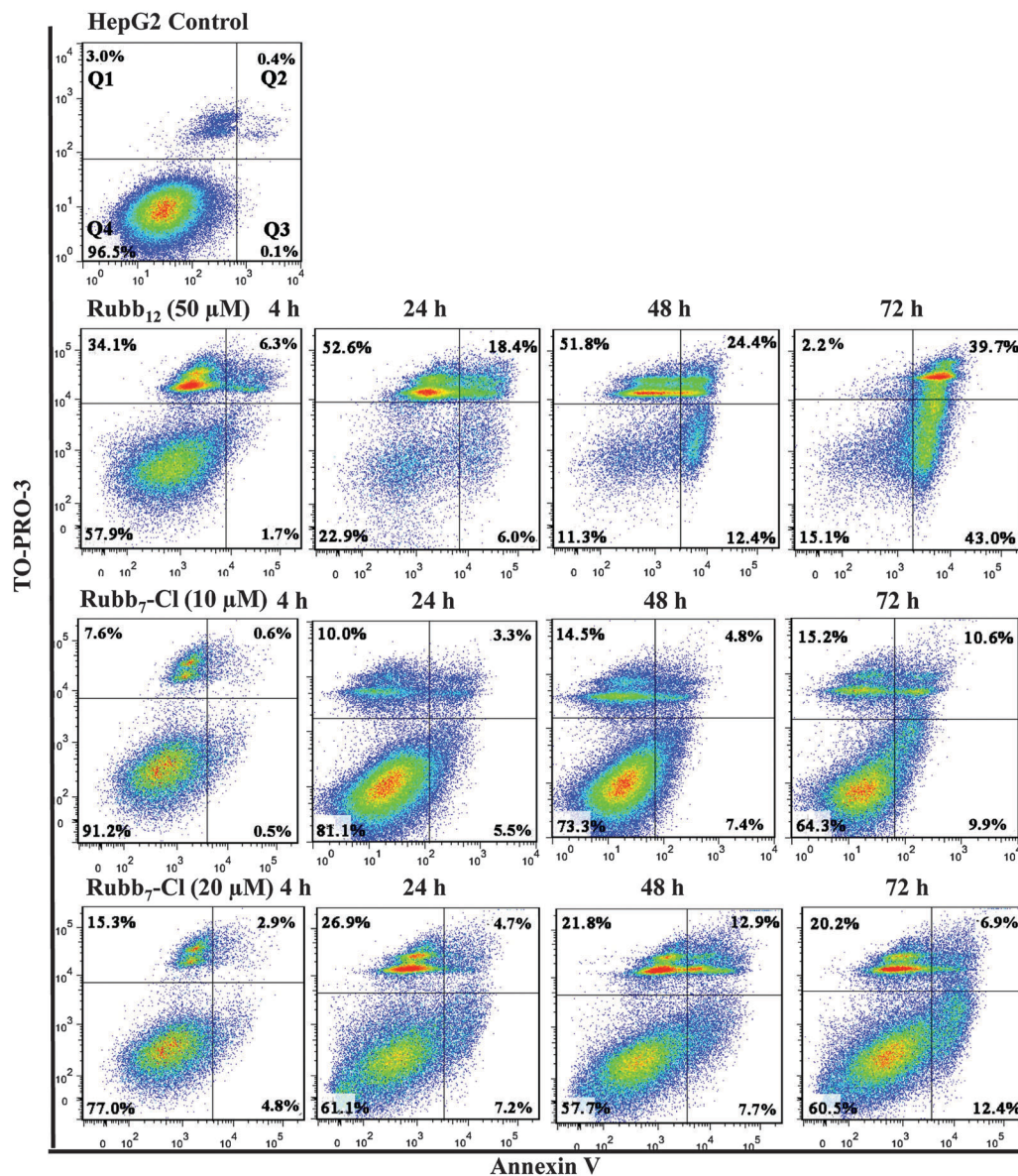


Fig. 7 Annexin V/TO-PRO-3 staining of HepG2 cells (control) and with treatment by the ruthenium complexes treatment (from top to bottom: 50 μM Rubb₁₂, 10 μM Rubb₇-Cl and 20 μM Rubb₇-Cl).

Similarly, SOD activity in HEK-293 cells also showed a slight increase for both ruthenium complexes. In contrast, the cancer cell line HepG2 showed a reverse trend in SOD activity, with a significant down-regulation for Rubb₁₂, but no obvious change for Rubb₇-Cl.

Discussion

Cationic oligomers, such as cationic peptides and di- and oligo-nuclear ruthenium complexes, have considerable potential as selective antibacterial agents due to their preferential interactions with the negatively-charged bacterial membrane and cell wall. One approach to the preparation of di- and oligo-nuclear ruthenium complexes is to link the individual metal centres

with a ligand that contains an alkyl chain. The length of the alkyl chain could affect cellular uptake, membrane damage and intracellular localisation in bacterial and eukaryotic cells. It has been previously demonstrated that the alkyl chain length controls the uptake in bacterial and L1210 cells.^{23,28} However, for both bacterial and L1210 cells, the dominant mechanism of cellular uptake was passive diffusion.^{23,28} The aims of this study were to determine if the Rubb₁₂ and Rubb₇-Cl complexes were taken up by passive diffusion in organ cells, and to examine the effect of the ruthenium complexes on the cytoplasmic and mitochondrial membranes.

Heat treatment at 60 °C destroys membrane integrity completely³⁹ and compounds freely pass into the cells and associate with all available biological ligands. Following 60 °C treatment, both Rubb₁₂ and Rubb₇-Cl uptake was greater in the human cell



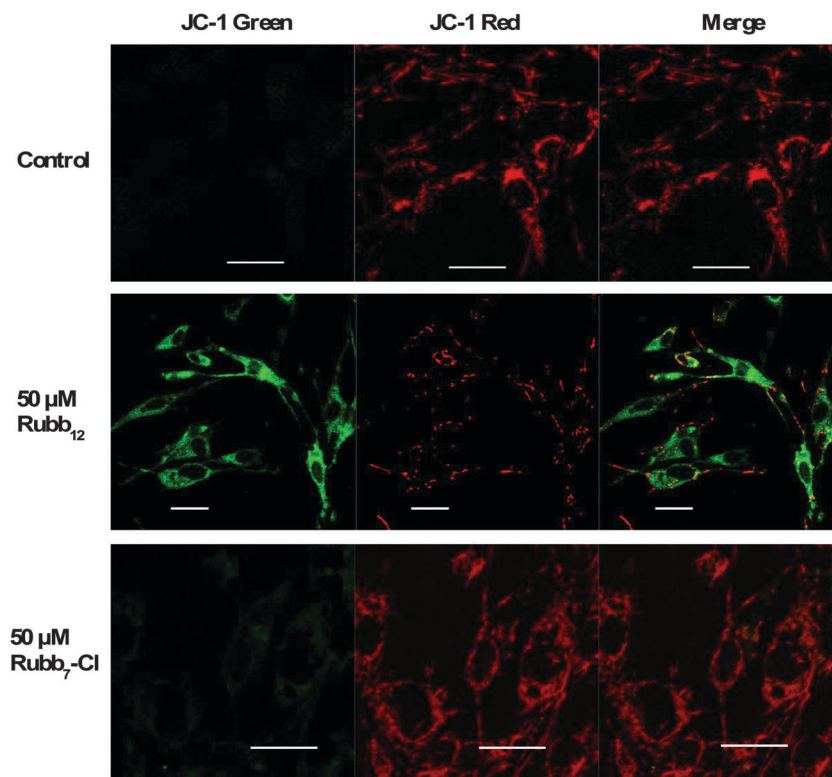


Fig. 8 JC-1 fluorescence imaging of mitochondria in BHK cells after a 4-hour incubation with 50 μM of Rubb₁₂ (middle) and Rubb_{7-Cl} (bottom). Red fluorescence is the sign of high MMP, whereas the green fluorescent signal indicates mitochondrial membrane depolarisation. Scale bar = 20 μm .

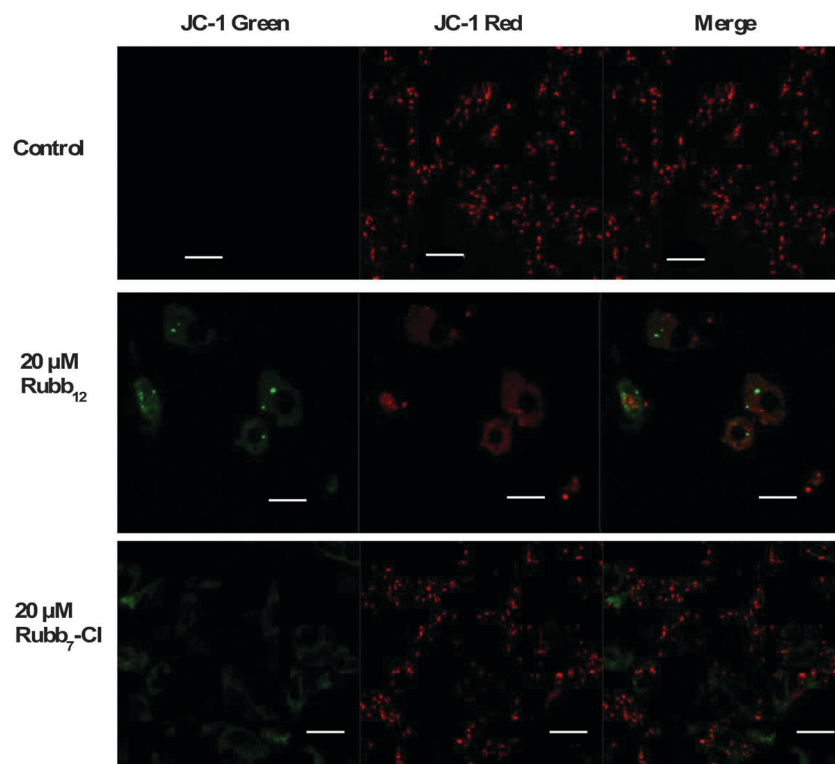


Fig. 9 JC-1 fluorescence imaging of mitochondria in HepG2 cells after a 4-hour incubation with 20 μM of Rubb₁₂ (middle) and Rubb_{7-Cl} (bottom). Scale bar = 20 μm .



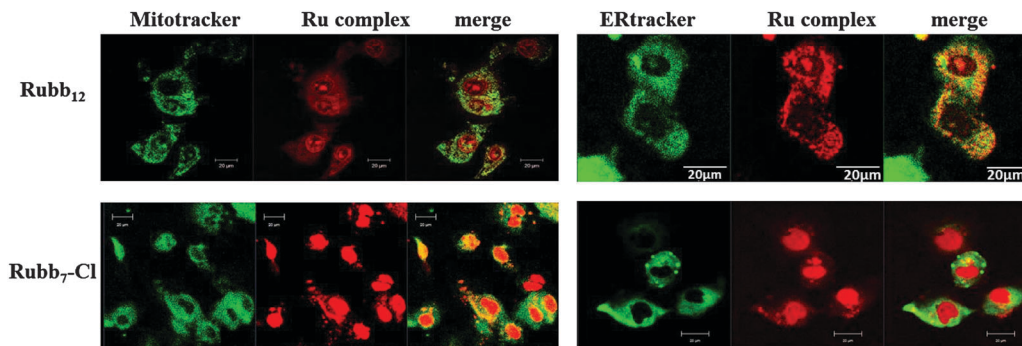


Fig. 10 Rubb₁₂ (top) and Rubb_{7-Cl} (20 hour incubation) localisation in HepG2 cells at 20 μM, co-stained with Mitotracker Green (left) or ERTracker Green (right), where the yellow coloration arises from co-localisation of Mitotracker Green/ERTracker Green and Rubb_n-Cl. Scale bar = 20 μm.

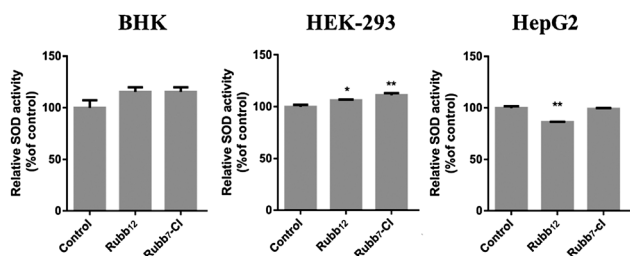


Fig. 11 Effect of the ruthenium complexes on SOD activity in BHK, HEK-293 and HepG2 cells.

lines than in the baby hamster cells (HEK-293 ≥ HepG2 > BHK), suggesting more intracellular binding sites in the two human cell lines than in BHK cells. As chromosomal DNA was one of the main intracellular targets, these results could be partly explained by the different chromosome numbers in HEK-293, HepG2 and BHK cells – with 64, 55 and 44 chromosomes, respectively.⁴⁰ PFA-fixation causes cross-linking of membrane proteins (including protein carriers),⁴¹ leading to diminished uptake, with compounds predominantly entering cells *via* passive diffusion. The observed large loss of Rubb₁₂ and Rubb_{7-Cl} uptake in PFA-fixed cells may suggest that the inhibited active/carrier-assisted transportation routes contribute more than 50% of the total uptake in all cases.

As carrier proteins are inactivated and ATP-production is limited at 4 °C,⁴² the small uptake of Rubb₁₂ and Rubb_{7-Cl} at 4 °C compared with the greater uptake at higher temperatures demonstrated the strong temperature-dependence of the uptake in the eukaryotic cells. This is consistent with the previously reported energy-dependent uptake of the rigidly linked dinuclear complex $[\{Ru(phen)_2\}_2\{\mu\text{-tpphz}\}]^{4+}$ in MCF-7 cells.⁴³ However, other ruthenium complexes are transported *via* passive diffusion; for instance the mononuclear complex $[Ru(DIP)_2dppz]^{2+}$ showed much higher uptake at 4 °C.⁴⁴ Due to the decreased membrane fluidity at 4 °C (increased rigidity of the phospholipids chains), the cell membrane would be less permeable than it would be at 37 °C.^{45,46} Consequently, and particularly as the interactions between the phospholipid alkyl chains and the alkyl chains of the bb_n ligand are likely to be important, it is possible that the ruthenium complexes might not be able to

enter the cells by passive diffusion at 4 °C. On the other hand, at 24 °C, ATP-dependent transport is mostly inhibited, while passive transport and carrier-facilitated diffusion function normally.^{47,48} Other studies have demonstrated that strictly ATP-required uptake of peptides was reduced by ~75% at 24 °C compared to 37 °C,⁴⁹ and Na-K-ATPase activity is inhibited by ~70% in mammalian erythrocytes at 24 °C.⁵⁰ These findings indicate that ~25–30% of active uptake occurs at 24 °C. Accordingly, the actual active uptake can be estimated. Corrected estimations suggest active transport generally appears to be the dominant cellular uptake mechanism, contributing to more than half of the total uptake. However, for Rubb₁₂ in BHK cells and Rubb_{7-Cl} in HEK-293 cells slightly lower contributions of active uptake (~30%) are estimated, indicating carrier-assisted diffusion could also be involved in the uptake.

To further ascertain the contribution of active uptake, the energy dependence of cellular uptake of Rubb₁₂ and Rubb_{7-Cl} was determined by pretreating cells with the ATP-production stimulator D-glucose or the metabolic inhibitors 2-deoxy-glucose and oligomycin. The unchanged uptake in the D-glucose-treated BHK and HEK-293 cells indicate an optimal energy condition, which could not be further improved. However, the increased uptake in D-glucose-treated HepG2 cells was indicative of glucose deprivation in HepG2 cells. This may be a result of a heavy reliance on glycolysis for ATP production in cancer cells instead of mitochondrial oxidative phosphorylation,⁵¹ the latter being the primary pathway to produce ATP in healthy cells.⁵² Therefore, it is possible that the external supply of glucose enhanced energy production in HepG2 cells, consequently leading to greater active uptake of ruthenium complexes. In contrast, the significant decrease in uptake in ATP-inhibited cells for both Rubb₁₂ and Rubb_{7-Cl} confirmed a significantly large contribution of active uptake of the ruthenium complexes.

The actual contribution of active uptake could be even larger than the observed proportions, considering that ATP-production cannot be completely blocked by the metabolic inhibitors.^{53,54} These findings, in conjunction with the lower temperature study results, strongly support the conclusion that active uptake is the major mechanism of cell entry for the ruthenium complexes studied in kidney and liver cell lines.

Based on the 24 °C uptake and the ATP-inhibited uptake results, the possible role of polyspecific organic cation transporters



(OCT)-assisted diffusion of Rubb₁₂ and Rubb_{7-Cl} was explored. The lower uptake of Rubb₁₂ following OCTs inhibition in the three cell lines, supports the conclusion that the uptake of Rubb₁₂ was partially facilitated by OCTs. In contrast, Rubb_{7-Cl} displayed no OCTs-facilitated diffusion in BHK and HepG2 cells, but a considerable contribution of OCTs-facilitated uptake was noted in HEK-293 cells. It is possible that the labile complex Rubb_{7-Cl} interacts with the carrier proteins in a covalent mode, making dissociation harder from the membrane protein, and transport by carriers like OCTs less likely. On the other hand, a correlation between OCT binding with molecules that contain both a positive charge and high lipophilicity has been observed.⁵⁵ This may explain the greater OCT-assisted uptake of the more positively-charged and possibly more lipophilic Rubb₁₂.

In summary, the major uptake mechanism for Rubb₁₂ and Rubb_{7-Cl} is active transport in kidney and liver cells – with a significant contribution from carrier-assisted diffusion for Rubb₁₂ but mainly passive diffusion for Rubb_{7-Cl}. However, previous studies from our research group on Rubb_n complexes demonstrated uptake by passive diffusion in L1210 leukaemia cells and bacteria cells.^{23,28} As the function of kidney and liver are filtering and detoxification,⁵⁶ uptake of molecules can be expected to be more selective and finely managed, which is only possible by active uptake.

To determine the extent of the damage to the cytoplasmic membrane, and to compare the cellular consequences of membrane damage with cell death through an apoptotic pathway, Annexin V/TO-PRO-3 assays were carried out. The results clearly demonstrated that Rubb₁₂ caused significant direct membrane damage to both BHK and HepG2 cells; however, membrane damage was considerably more rapid and extensive with the HepG2 cells. By contrast, Rubb_{7-Cl} caused significantly less membrane damage than Rubb₁₂ in both cell lines. Of note however, was the relatively low level of apoptotic cells up to 48 hours for both cell lines.

As an indicator of mitochondrial health, mitochondrial membrane potential (MMP) was monitored using the JC-1 assays. The results clearly demonstrate that Rubb₁₂ depolarises the mitochondrial membrane in both BHK and HepG2 cells; whereas, Rubb_{7-Cl} had a much smaller affect. The cellular localisation experiments indicated that although Rubb₁₂ did not accumulate in the mitochondria in either cell line, significant mitochondrial accumulation was observed for Rubb_{7-Cl} in both BHK and HepG2 lines. While DNA damage and ER stress beyond a certain repair-threshold could reduce MMP,^{55,57,58} the combined results (lack of localisation and reduction in MMP) are consistent with Rubb₁₂ damaging the mitochondrial membrane. Given the previously-reported membrane damage induced by Rubb_n in bacterial cells,²⁷ the lipophilic ruthenium complexes could lead to depolarisation of membranes in eukaryotic cells, including the cell membrane and intracellular membrane compartments (e.g. ER, mitochondria, Golgi, endosome and lysosome), resulting in irreparable membrane damage.

Reactive oxygen species (ROS) have long been known as inducers of programmed cell death.⁵⁹ ER/mitochondrial localisation of the ruthenium complexes makes generation of ROS likely,

as these two organelles are intricately involved in the overall ROS regulation.⁶⁰ Many ruthenium complexes have been demonstrated to increase ROS generation and induce oxidative stress and apoptosis.^{61–64} To estimate the oxidative perturbation caused by the ruthenium complexes Rubb₁₂ and Rubb_{7-Cl}, their effects on the intracellular activity of superoxide dismutase (SOD), the essential enzyme eliminating ROS, was studied. The observed enhanced SOD activity in BHK and HEK-293 cells can be indicative of increased ROS stress induced by the ruthenium complexes. This hypothesis is supported by the reported increased SOD activity in response to oxidative stress in plants⁶⁵ and other animal cells.⁶⁶ However, in HepG2 cells, SOD activity decreased with treatment by Rubb₁₂ but remained unchanged for Rubb_{7-Cl}. Since HepG2 cells are more susceptible to the ruthenium complexes than the other two cell lines, a decrease in SOD activity rather than increased activity was to be expected, due to the direct oxidative damage of SOD enzymes and/or ROS-altered SOD gene expression.^{67–69} Similar reduction of SOD activity has been reported in cisplatin treated rat ovarian cells⁷⁰ and in vitamin-treated breast cancer cells.⁷¹ The unchanged SOD activity in Rubb_{7-Cl} treated HepG2 could be explained by a balance between up-regulation and the inhibition of the enzyme activity/expression. These results suggest that the ruthenium complexes could induce cell death *via* a ROS-mediated pathway.

The present study demonstrates that the dinuclear ruthenium complex linked by the bb₁₂ ligand (Rubb₁₂) extensively damages the cytoplasmic membrane and probably the mitochondrial membrane. Although it has been established that Rubb₁₂ selectively localises with RNA in eukaryotic cells,²⁵ it is likely that membrane damage is the dominant mechanism of cell death. By contrast, Rubb_{7-Cl} has a considerably smaller affect on the cytoplasmic and mitochondrial membranes. As a consequence, Rubb_{7-Cl} shows greater variability in its 24 h-IC₅₀s towards eukaryotic cells. After activation by aquation, Rubb_{7-Cl} can form covalent bonds with biomolecules. Although Rubb_{7-Cl} only requires 3.5 hours to be 50% aquated in aqueous solution at 25 °C,²⁹ covalent bond formation with nucleotides and amino acids is very slow.²⁹ This suggests that Rubb_{7-Cl} predominantly induces its biological effect through non-covalent interactions within eukaryotic cells. HepG2 cells treated with Rubb_{7-Cl} at a concentration more than six-fold the 72 h-IC₅₀ exhibited only a slight cell response, with weak apoptosis and weak membrane damage. These weak cellular responses could be partly due to the low incidence of drug-induced apoptosis reported for some cancer cells.^{72,73} However, Rubb_{7-Cl} targets nucleic DNA in HepG2 cells,²⁹ which may suggest another predominant cell death pathway. The DNA binding caused by Rubb_{7-Cl} may induce DNA damage (for example condensation), therefore halting the cell cycle.

The results of this study indicate that Rubb₁₂ causes significant membrane damage – both cytoplasmic and mitochondrial – whereas little membrane damage was observed for Rubb_{7-Cl}. These conclusions are consistent with the IC₅₀ values observed for Rubb₁₂ and Rubb_{7-Cl} against the three eukaryotic cell lines – very similar values for Rubb₁₂, and highly divergent values for Rubb_{7-Cl}. Furthermore, the results of this study suggest that



cationic oligomers joined by alkyl chain lengths of twelve carbons or longer will likely cause significant membrane damage to eukaryotic cells. Alternatively, cationic oligomers joined by alkyl chain lengths of seven carbons are unlikely to cause significant membrane damage, and hence may provide a better scaffold for oligomeric or polymeric antibacterial agents. The mechanism by which Rubb₁₂ damages membranes is unknown, but it could be related to the lipophilicity of the dinuclear ruthenium complex or the actual chain length. Given that the log *P* for Rubb₁₂ is -2.7 ,²² it seems unlikely that differences in lipophilicity between Rubb₁₂ and Rubb₇-Cl is the dominant determinant of membrane damage. It is noted that the highly non-polar section of a membrane is 25–30 Å in width,⁷⁴ which is approximately the same distance as that between the ruthenium centres (26 Å) in Rubb₁₂. As the ruthenium metal centres can favourably interact (through π - π interactions of the phenanthroline ligands) and the distance between the ruthenium centres is compatible with the non-polar region of a membrane, it is possible that membrane damage is caused by aggregated Rubb₁₂ molecules inserting perpendicularly into the membrane to form a pore. This could then lead to greater levels of membrane disruption, eventually leading to the complete permeabilisation of the cell.

Conclusions

Dinuclear ruthenium complexes have shown considerable potential as antimicrobial agents; however, in order to rationally design more active and more selective compounds, it is necessary to gain an understanding of the biological processing of the dinuclear complexes in bacterial and eukaryotic cells. This study provides a detailed analysis of the biological processing of the dinuclear complexes in eukaryotic cells, and in particular, an analysis of the effects of these compounds on the membranes of eukaryotic cells. The results indicate that Rubb₁₂ damages the cytoplasmic membrane, and probably the membranes of internal organelles like the mitochondria. However, despite the fact that Rubb₁₂ kills eukaryotic cells in a non-specific manner, it can still be 40–100 times more active against bacteria than eukaryotic cells.²⁹ By contrast, Rubb₇-Cl does not significantly damage membranes and is cytotoxic through other specific mechanism(s). As a consequence of this, Rubb₇-Cl exhibits very variable cytotoxicities against various cell lines. It is commonly believed that potential new antimicrobial drugs should not be active through non-specific mechanisms (as generally evidenced by similar IC₅₀ values against eukaryotic cells), and in particular, should not just damage cell membranes. The results of this study suggest there are sufficient differences, particularly in the charge density, between bacterial and eukaryotic membranes that cationic oligomers (like Rubb₁₂) could form the basis of new types of antimicrobial agents.

Experimental procedures

Material

The ruthenium complexes Rubb₁₂ and Rubb₇-Cl were synthesised as previously described.^{29,75}

Cell culture

Three mammalian cell lines BHK (baby hamster kidney), HEK-293 (embryonic kidney) and HepG2 (liver carcinoma) were used in this study. All cell lines were generously supplied by the Australian Army Malaria Institute (AMI, Enoggera, QLD, Australia), and originated from the American Type Culture Collection (ATCC, Manassas, USA). Cells were cultured as previously described.²⁵ Cells used in the study were in the logarithmic growth phase and were grown to 70% confluence, and then trypsinised with 0.25% trypsin–0.02% EDTA (Sigma-Aldrich) for detachment and used in the assays described below.

Cellular localisation of the ruthenium complexes

The trypsinised cells were seeded in Lumox[®] multiwell plates (Sarstedt, Numbrecht, Germany), and incubated with 20 to 50 μ M ruthenium complexes at 37 °C with 5% CO₂ for 4 or 20 h as described. Following incubation, cells were stained with 100 nM Mitotracker[®] Green FM (Invitrogen, Carlsbad, USA) for mitochondrial staining and 1 μ M ER-Tracker[™] Green (Invitrogen) for endoplasmic reticulum (ER) staining. Staining was carried out in RPMI-1640 medium under standard cultivation conditions as per the manufacturer's instructions. Following staining, cells were gently rinsed with phosphate buffer solution (PBS; pH = 7.1) prior to confocal laser scanning microscopy.

The cellular localisation of the ruthenium complexes was determined using a Zeiss laser scanning confocal microscope (LSM 700, Carl Zeiss, Jena, Germany). Samples were viewed under 63 \times oil immersion using the following excitation (λ_{ex}) and emission (λ_{em}) wavelengths. Metal complexes (λ_{ex} = 450 nm, λ_{em} = 610 nm), Mitotracker Green FM (λ_{ex} = 490 nm, λ_{em} = 516 nm) and ER-Tracker[™] Green (λ_{ex} = 504 nm, λ_{em} = 511 nm) were excited using a blue argon laser (λ_{ex} = 488 nm), and emissions were collected at λ_{em} = 570–650 nm for the ruthenium complexes, λ_{em} = 470–550 nm for Mitotracker and ER-tracker. Image data acquisition and processing was performed using Zen software 2009 (Carl Zeiss).

Cellular uptake of ruthenium complexes

Flow cytometry. Cultures at ~85% confluency were treated with ruthenium complexes at the desired concentrations in 6-well plates, for various periods of time. Cells were gently trypsinised and harvested by centrifuging at 1000g for 3 min, and re-suspended and washed in PBS to remove ruthenium complexes from the culture medium and the cell surface. Cellular uptake of the ruthenium complexes was measured by monitoring the median fluorescence intensity (MFI) using a Guava EasyCyte[™] 5HT flow cytometer (Merck Milipore, Darmstadt, Germany). Experiments were performed in triplicate. Ruthenium complexes were excited with a λ_{ex} = 488 nm blue argon laser and emissions collected over the range λ_{em} = 570–615 nm. All flow cytometry data were processed with GuavaSoft 2.5 (Merck Milipore).

Effect of cell membrane permeability on the uptake of ruthenium complexes

Cell membrane permeability was modified by heat treatment or fixation with paraformaldehyde (PFA, Sigma-Aldrich, St. Louis, USA).



The cells were cultured in three aliquots. Given the upper temperature limit for eukaryotic cells near 60 °C (due to their inability to form membranes that are both thermostable and functional),³⁹ one aliquot was pre-incubated at 60 °C for 30 min before the ruthenium complexes were added. The heat treatment permeabilised and killed the cells, allowing for the maximal interaction between the ruthenium complexes and their intracellular targets. The second aliquot was incubated in fresh 4% PFA solution for 10 min at room temperature. The PFA solution was removed and the cell layer was rinsed with PBS before incubation with the ruthenium complexes. The third aliquot served as the control group following standard staining procedures. Ruthenium complexes (20 μM) were added to pretreated and control cells, followed by incubation at 37 °C for 4 h for all samples. After the incubation, cells were gently trypsinised, centrifuged and washed twice with PBS, and re-suspended in PBS for flow cytometry. Statistical analysis was performed using a *t*-test in GraphPad Prism 6.0 (GraphPad Software, San Diego, USA). A *P* value of <0.05 was considered significant.

Dependence of cellular uptake of the ruthenium complexes on temperature

Cells were incubated with the ruthenium complexes (50 μM for BHK, 20 μM for HEK-293 and 10 μM for HepG2, reflecting the trend in the IC₅₀ values for Rubb₇-Cl) for 4 h at 4, 24 or 37 °C. Following incubation, the cells were trypsinised, centrifuged and rinsed with PBS before analysis by flow cytometry. *t*-Tests were performed to evaluate the significance of temperature on the uptake with 37 °C treatments serving as controls.

Dependence of cellular uptake of the ruthenium complexes on ATP production

Cells were pre-incubated with 2-deoxy-glucose (30 mM, Sigma-Aldrich) and oligomycin (10 μM, Sigma-Aldrich), or the ATP stimulator D-glucose (30 mM; Sigma-Aldrich) for 2 h at 37 °C. Ruthenium complexes (20 μM) were added to pretreated cells and untreated cells (controls) and incubated at 37 °C for 4 h. After incubation, the cells were trypsinised, centrifuged and rinsed twice with PBS for flow cytometry analysis. Results were statistically analysed using a *t*-test.

Effect of organic cation transport inhibitors on the uptake of the ruthenium complexes

Cells were pre-incubated for 1 h at 37 °C with six organic cation transport inhibitors³² (Table 1; tetramethylammonium chloride, tetraethylammonium chloride monohydrate, tetrapropylammonium

chloride, tetra-*n*-butylammonium chloride, tetra-*n*-pentylammonium chloride and procainamide hydrochloride; Sigma-Aldrich) at 2 mM in RPMI-1640 complete medium. Thereafter, the ruthenium complexes (20 μM) were added and incubations continued for 4 h at 37 °C. The cells were trypsinised, centrifuged and washed for analysis by flow cytometry. To compare the treated groups with controls, the data were analyzed by multiple *t*-tests.

Determination of membrane damage and apoptosis induced by the ruthenium complexes

Apoptosis induced by the ruthenium complexes was assessed using the Alexa Fluor[®] 488 Annexin V assay (Sigma-Aldrich) for detecting phosphatidyl-serine in the external plasma membrane leaflet and TO-PRO-3 (Molecular Probes[®], Invitrogen) for assessing cell membrane permeability. BHK and HepG2 cells were incubated with the ruthenium complexes for the desired times. Cells were gently trypsinised, washed and collected by centrifugation. Cell pellets were resuspended to 1 × 10⁵ to 1 × 10⁶ cells mL⁻¹ in 100 μL Annexin V binding buffer. Alexa Fluor[®] 488 Annexin V (5 μL) was added and the samples were incubated at room temperature for 15 min in darkness. Annexin V binding buffer (400 μL) and TO-PRO-3 (5 μL) were added to each sample immediately prior to analysis on a BD LSR Fortessa flow cytometer (Franklin Lakes, USA), with a 100 000 events per sample recorded for each experiment. Data were analysed using FlowJo 7.6.1 (FlowJo Software, Ashland, USA).

Detection of mitochondrial membrane potential

Detection of mitochondrial membrane potential (MMP) was carried out using a Zeiss laser scanning confocal microscope (LSM 700, Carl Zeiss, Göttingen, Germany). BHK and HepG2 cells were treated with Rubb₁₂ and Rubb₇-Cl for 4 h in Lumox[®] multiwell plates (Sarstedt). After rinsing, treated cells were incubated with 5 μM JC-1 dye (mitochondrial membrane potential probe, Invitrogen) in complete RPMI-1640 medium for 30 min at 37 °C. Cells were washed with PBS before confocal microscopy. JC-1 monomers were excited with a λ_{ex} = 488 nm argon-ion laser and emissions collected over the range of λ_{em} = 515–545 nm. J-aggregate forms were excited with a 568 nm argon-krypton laser and the emission signals collected over the range of λ_{em} = 575–625 nm. Time-course MMP tests were conducted using the BHK cell line. The cells were incubated with 20 μM of Rubb₁₂ or 50 μM of Rubb₇-Cl (approximately 25% of the respective 24 h-IC₅₀ values) for 4 or 20 h, and the treated cells were processed as above.

Reactive oxygen superoxide dismutase (SOD) assays

The SOD activities in the cell lines BHK, HEK-293 and HepG2 after treatment with Rubb₁₂ and Rubb₇-Cl were determined using the SOD assay kit (Sigma-Aldrich). The cell lines were incubated with the ruthenium complexes for 20 h at different concentrations (50 μM for BHK, 20 μM for HEK-293 and 10 μM for HepG2, respectively). Cells were detached from the tissue culture flask by trypsinisation and collected by centrifugation. The cell pellets were resuspended in PBS buffer solution to 5 × 10⁶ cells mL⁻¹. Glass beads (150–212 μm; Sigma-Aldrich)

Table 1 Inhibitors of polyspecific organic cation transporters (OCTs)

OCT inhibitors	OCTs inhibited
Tetramethylammonium chloride	OCT1, OCT2
Tetraethylammonium chloride	OCT1, OCT2, OCT3, OCTN1, OCTN2
Tetrapropylammonium chloride	OCT1, OCT2
Tetra- <i>n</i> -butylammonium chloride	OCT1, OCT2, OCTN1
Tetra- <i>n</i> -pentylammonium chloride	OCT1, OCT2, OCT3, OCTN1
Procainamide	OCT1, OCT2, OCT3, OCTN1, OCTN2



were added to the cell suspension and agitated using a bead beater (BBY24M-CE; Next Advance, Averill Park, USA) for 10 min at 4 °C to break the cell membrane. Supernatants were collected after centrifugation, and the SOD activity was determined as per the procedure provided by the manufacturer of the assay kit. Data were statistically analysed using *t*-tests, with the SOD activity in untreated cells serving as controls.

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References

- 1 S. Kapitza, M. A. Jakupiec, M. Uhla, B. K. Keppler and B. Marian, *Cancer Lett.*, 2005, **226**, 115.
- 2 E. Alessio, G. Mestroni, A. Bergamo and G. Sava, in *Metal Ions Biological Systems*, ed. A. Sigel, H. Sigel and M. Dekker, CRC Press, New York, 2004, pp. 323–351.
- 3 L. Ronconi and P. J. Sadler, *Coord. Chem. Rev.*, 2007, **251**, 1633.
- 4 Y. K. Yan, M. Melchart, A. Habtemariam and P. J. Sadler, *Chem. Commun.*, 2005, 4764.
- 5 C. A. Vock, W. H. Ang, C. Scolaro, A. D. Phillips, L. Lagopoulos, L. Juillerat-Jeanneret, G. Sava, R. Scopelliti and P. J. Dyson, *J. Med. Chem.*, 2007, **50**, 2166.
- 6 G. Sava, E. Alessio, A. Bergamo and G. Mestroni, in *Biological Inorganic Chemistry*, ed. M. J. Clarke and P. J. Sadler, Springer, Berlin, 1999, vol. 1, pp. 143–169.
- 7 M. R. Gill and J. A. Thomas, *Chem. Soc. Rev.*, 2012, **41**, 3179.
- 8 C. S. Devi, D. A. Kumar, S. S. Singh, N. Gabra, N. Deepika, Y. P. Kumar and S. Satyanarayana, *Eur. J. Med. Chem.*, 2013, **64**, 410.
- 9 F. P. Dwyer, E. C. Gyrfas, W. P. Rogers and J. H. Koch, *Nature*, 1952, **170**, 190.
- 10 A. Bolhuis, L. Hand, J. E. Marshall, A. D. Richards, A. Rodger and J. Aldrich-Wright, *Eur. J. Pharm. Sci.*, 2011, **42**, 313.
- 11 S. V. Kumar, W. K. C. Lo, H. J. L. Brooks and J. D. Crowley, *Inorg. Chim. Acta*, 2015, **425**, 1.
- 12 V. Pierroz, T. Joshi, A. Leonidova, C. Mari, J. Schur, I. Ott, L. Spiccia, S. Ferrari and G. Gasser, *J. Am. Chem. Soc.*, 2012, **134**, 20376.
- 13 H. Huang, P. Zhang, B. Yu, Y. Chen, J. Wang, L. Ji and H. Chao, *J. Med. Chem.*, 2014, **57**, 8971.
- 14 H. Huang, B. Yu, P. Zhang, J. Huang, Y. Chen, G. Gasser, L. Ji and H. Chao, *Angew. Chem., Int. Ed.*, 2015, **54**, 14049.
- 15 H. Huang, P. Zhang, H. Chen, L. Ji and H. Chao, *Chem. – Eur. J.*, 2015, **21**, 715.
- 16 A. S. Abd-El-Aziz, C. Agatemor, N. Etkin, D. P. Overy, M. Lanteigne, K. McQuillan and R. G. Kerr, *Biomacromolecules*, 2015, **16**, 3694.
- 17 A. Martin, A. Byrne, C. S. Burke, R. J. Forster and T. E. Keyes, *J. Am. Chem. Soc.*, 2014, **136**, 15300.
- 18 P. Liu, B.-Y. Wu, J. Liu, Y.-C. Dai, Y.-J. Wang and K.-Z. Wang, *Inorg. Chem.*, 2016, **55**, 1412.
- 19 F. Li, J. G. Collins and F. R. Keene, *Chem. Soc. Rev.*, 2015, **44**, 2529.
- 20 H. Jenssen, P. Hamill and R. E. W. Hancock, *Clin. Microbiol. Rev.*, 2006, **19**, 491.
- 21 S. K. Vooturi, C. M. Cheung, M. J. Rybak and S. M. Firestone, *J. Med. Chem.*, 2009, **52**, 5020.
- 22 F. Li, Y. Mulyana, M. Feterl, J. M. Warner, J. G. Collins and F. R. Keene, *Dalton Trans.*, 2011, **40**, 5032.
- 23 F. Li, M. Feterl, Y. Mulyana, J. M. Warner, J. G. Collins and F. R. Keene, *J. Antimicrob. Chemother.*, 2012, **67**, 2686.
- 24 F. Li, M. Feterl, J. M. Warner, A. I. Day, F. R. Keene and J. G. Collins, *Dalton Trans.*, 2013, **42**, 8868.
- 25 X. Li, A. K. Gorle, T. D. Ainsworth, K. Heimann, C. E. Woodward, J. G. Collins and F. R. Keene, *Dalton Trans.*, 2015, **44**, 3594.
- 26 F. Li, E. J. Harry, A. L. Bottomley, M. D. Edstein, G. W. Birrell, C. E. Woodward, F. R. Keene and J. G. Collins, *Chem. Sci.*, 2014, **5**, 685.
- 27 F. Li, M. Feterl, J. M. Warner, F. R. Keene and J. G. Collins, *J. Antimicrob. Chemother.*, 2013, **68**, 2825.
- 28 M. J. Pisani, P. D. Fromm, Y. Mulyana, R. J. Clarke, H. Körner, K. Heimann, J. G. Collins and F. R. Keene, *ChemMedChem*, 2011, **6**, 848.
- 29 X. Li, K. Heimann, F. Li, J. M. Warner, F. R. Keene and J. G. Collins, *Dalton Trans.*, 2016, **45**, 4017.
- 30 E. C. Slater, *Methods Enzymol.*, 1967, **10**, 48.
- 31 H. Koepsell and H. Endou, *Eur. J. Appl. Physiol.*, 2004, **447**, 666.
- 32 S. Barban and H. O. Schulze, *J. Biol. Chem.*, 1961, **236**, 1887.
- 33 I. Vermes, C. Haanen, H. Steffens-Nakken and C. Reutelingsperger, *J. Immunol. Methods*, 1995, **184**, 39.
- 34 V. Borutaite, *Environ. Mol. Mutagen.*, 2010, **51**, 406.
- 35 J. D. Ly, D. R. Grubb and A. Lawen, *Apoptosis*, 2003, **8**, 115.
- 36 B. Halliwell and J. M. C. Gutteridge, *Free radicals in biology and medicine*, Oxford University Press, New York, 2007, pp. 449–450.
- 37 L. W. Oberley, G. R. Buettner and G. R. Buettner, *Cancer Res.*, 1979, **39**, 1141.
- 38 I. Fridovich, *Acc. Chem. Res.*, 1972, **5**, 321.
- 39 M. R. Tansey and T. D. Brock, *Proc. Natl. Acad. Sci. U. S. A.*, 1972, **69**, 2426.
- 40 American Type Culture Collection, <http://www.atcc.org/>.
- 41 J. A. Kiernan, *Microsc. Today*, 2000, **1**, 8.
- 42 H. Ginsburg and S. Yeroushalmy, *J. Appl. Physiol.*, 1978, **282**, 399.
- 43 M. R. Gill, J. Garcia-Lara, S. J. Foster, C. Smythe, G. Battaglia and J. A. Thomas, *Nat. Chem.*, 2009, **1**, 662.
- 44 C. A. Puckett and J. K. Barton, *Biochemistry*, 2008, **47**, 11711.
- 45 M. Rossignol, T. Uso and P. Thomas, *J. Membr. Biol.*, 1985, **87**, 269.
- 46 M. B. Lande, J. M. Donovan and M. L. Zeidel, *J. Gen. Physiol.*, 1995, **106**, 67.
- 47 U. D. Kulkarni, R. Mahalingam, X. Li, I. Pather and B. Jasti, *AAPS PharmSciTech*, 2011, **12**, 579.
- 48 C. A. Lesch, C. A. Squier, A. Cruchley, D. M. Williams and P. Speight, *J. Dent. Res.*, 1989, **68**, 1345.



- 49 J. C. Wolters, R. Abele and R. Tampé, *J. Biol. Chem.*, 2005, **280**, 23631.
- 50 S. L. Kimzey and J. S. Willis, *J. Gen. Physiol.*, 1971, **58**, 634.
- 51 O. Warburg, *Science*, 1956, **123**, 309.
- 52 I. M. Ahmad and M. Y. Abdalla, in *Oxidative stress in cancer biology and therapy*, ed. D. R. Spitz, K. J. Dornfeld, K. Krishnan and D. Gius, Humana Press, New York, 2012, pp. 1–19.
- 53 A. N. Wick, D. R. Drury, H. I. Nakada and J. B. Wolfe, *J. Biol. Chem.*, 1957, **224**, 963.
- 54 M. Jastroch, A. S. Divakaruni, S. Mookerjee, J. R. Treberg and M. D. Brand, *Essays Biochem.*, 2010, **47**, 53.
- 55 G. Kroemer, L. Galluzzi and C. Brenner, *Physiol. Rev.*, 2007, **87**, 99.
- 56 N. Ahmed, D. Maureen, C. Smith and E. Wood, *Biology of Disease*, Taylor & Francis, 2006, p. 600.
- 57 W. X. Zong, C. Li, G. Hatzivassiliou, T. Lindsten, Q. C. Yu, J. Yuan and C. B. Thompson, *J. Cell Biol.*, 2003, **162**, 59.
- 58 L. Filippin, P. J. Magalhães, G. Di Benedetto, M. Colella and T. Pozzan, *J. Biol. Chem.*, 2003, **278**, 39224.
- 59 S. C. Gupta, D. Hevia, S. Patchva, B. Park, W. Koh and B. B. Aggarwal, *Antioxid. Redox Signaling*, 2012, **16**, 1295.
- 60 A. A. Alfadda and R. M. Sallam, *J. Biomed. Biotechnol.*, 2012, 936486.
- 61 W. Li, B.-J. Han, J.-H. Yao, G.-B. Jiang and Y.-J. Liu, *RSC Adv.*, 2015, **5**, 24534.
- 62 Z. Zhao, Z. Luo, Q. Wu, W. Zheng, Y. Feng and T. Chen, *Dalton Trans.*, 2014, **43**, 17017.
- 63 Z. Luo, L. Yu, F. Yang, Z. Zhao, B. Yu, H. Lai, K.-H. Wong, S.-M. Ngai, W. Zheng and T. Chen, *Metallomics*, 2014, **6**, 1480.
- 64 C. Tan, S. Lai, S. Wu, S. Hu, L. Zhou, Y. Chen, M. Wang, Y. Zhu, W. Lian, W. Peng, J. Liangnian and A. Xu, *J. Med. Chem.*, 2010, **53**, 7613.
- 65 A. S. Gupta, J. L. Heinen, A. S. Holaday, J. J. Burke and R. D. Allen, *Proc. Natl. Acad. Sci. U. S. A.*, 1993, **90**, 1629.
- 66 P. States, T. Promotion and N. Series, *Science*, 1985, **227**, 375.
- 67 M. S. Cooke, M. D. Evans, M. Dizdaroglu and J. Lunec, *FASEB J.*, 2003, **17**, 1195.
- 68 P. V. Limaye, N. Raghuram and S. Sivakami, *Mol. Cell. Biochem.*, 2003, **243**, 147.
- 69 C. Gemma, J. Vila, A. Bachstetter and P. Bickford, in *Brain aging: models, methods and mechanisms*, ed. D. R. Riddle, CRC Press/Taylor & Francis, 2007, pp. 353–374.
- 70 X. Li, S. Yang, X. Lv, H. Sun, J. Weng, Y. Liang and D. Zhou, *J. Gynecol. Oncol.*, 2013, **24**, 177.
- 71 D. Feldman, J. W. Pike and F. H. Glorieux, *Vitamin D*, Elsevier Academic Press, 2005, p. 1952.
- 72 D. Hanahan and R. A. Weinberg, *Cell*, 2000, **100**, 57.
- 73 S. Fulda, *Int. J. Cancer*, 2009, **124**, 511.
- 74 B. A. Lewis and D. M. Engelman, *J. Mol. Biol.*, 1983, **166**, 211.
- 75 Y. Mulyana, D. K. Weber, D. P. Buck, C. A. Motti, J. G. Collins and F. R. Keene, *Dalton Trans.*, 2011, **40**, 1510.

