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## Compounds from *Caesalpinia sappan* with anti-inflammatory properties in macrophages and chondrocytes

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The heartwood of *Caesalpinia sappan* is a traditional ingredient of food and beverages in South East Asia and has been used in traditional medicine as an analgesic and anti-inflammatory drug or to promote blood circulation. Scientific studies have confirmed different bioactivities associated with its use. Here, five fractions were isolated from the ethanolic extract of *C. sappan* heartwood, including episappanol (1), protosappanin C (2), brazilin (3), (iso-)protosappanin B (4) and sappanol (5) using high-performance liquid chromatography (HPLC). All compounds were tested for their anti-inflammatory effects in two different cell lines. Cytokine concentrations in the cell supernatant were determined using enzyme-linked immunosorbent assay (ELISA), and mRNA levels were measured using reverse-transcription quantitative polymerase chain reaction (RT-qPCR). In lipopolysaccharide-stimulated macrophages, all compounds significantly inhibited the secretion of the pro-inflammatory cytokines interleukin (IL-6) and tumor necrosis factor-alpha (TNF- $\alpha$ ). Sappanol (5) increased the secretion of the anti-inflammatory IL-10. In IL-1 $\beta$ -stimulated chondrocytes, all fractions reduced the mRNA expression and the secretion of the pro-inflammatory cytokines IL-6 and TNF- $\alpha$ . The highest anti-inflammatory effect was found for brazilin (3) in both cell lines. Of note, this is the first study which shows the anti-inflammatory effect of sappanol and episappanol. This study provides evidence for the efficacy of the traditional use of *C. sappan* as an anti-inflammatory remedy. Given the high prevalence of inflammation-related pathologies including arthritis, and the urgent need to clinically intervene with these diseases, the anti-inflammatory activity of diverse compounds from *C. sappan* may be of interest for the development of complementary and alternative treatment strategies.

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### 1. Introduction

The heartwood of *Caesalpinia sappan* L. (Leguminosae), a plant native to South East Asia and South India, has been used as a natural red dye, as a traditional ingredient of food and beverages and in traditional Chinese medicine as an analgesic and anti-inflammatory drug or to promote menstruation and blood circulation.<sup>1</sup> Scientific studies have confirmed different bioactivities of this medicinal plant including vasorelaxation,<sup>2</sup> anti-atherosclerotic,<sup>3</sup> antioxidant,<sup>4</sup> antibacterial,<sup>5</sup> anti-inflammatory<sup>6</sup> or anti-arthritis effects.<sup>7</sup>

In addition to the major compound brazilin, several compounds have been isolated and identified including protosappanins, chalcones and homoisoflavones.<sup>8–10</sup> Most studies have been published on the bioactivity of brazilin showing vasorelaxation in endothelial cells *in vitro*,<sup>2</sup> hypoglycemic activity in diabetic mice,<sup>11</sup> and antibacterial activity.<sup>12</sup> More recently, it has been shown that brazilin reduces high glucose-induced vascular inflammation,<sup>13</sup> and exhibits anti-thrombotic properties,<sup>14</sup> anti-cancer activities as shown in liver, breast, lung and gingival cancer cells<sup>15</sup> and anti-allergic activities in a murine asthma model.<sup>16</sup>

A limited number of studies have been conducted on other compounds from *C. sappan*. As such, protosappanin A, B and brazilein exhibited antioxidative activity.<sup>4,17</sup> Brazilein, sappan-chalcone, protosappanin C, D and E showed anti-inflammatory effects in lipopolysaccharide (LPS)-induced macrophages (J774.1) as indicated by a reduction of nitric oxide (NO) or prostaglandin E2 (PGE<sub>2</sub>) production.<sup>10,17</sup> Sappanone A or protosappanin E inhibited IL-6 secretion in LPS-stimulated macrophages (RAW 264.7).<sup>18</sup> Brazilein, protosappanin A, and

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sappanchalcone exerted inhibitory effects on a drug target for influenza treatment.<sup>19</sup> Protosappanin A was found to be an immunosuppressive agent in heart-transplanted rats and exhibited anti-rejection activity.<sup>20</sup>

Inflammation plays a major role in a broad range of diseases including asthma, atherosclerosis, cancer, and arthritis. IL-1 $\beta$  stimulated chondrocytes serve as a joint inflammation model for arthritis. Previously, we have shown the anti-inflammatory activity of the ethanolic extract of *C. sappan* (CSE) in chondrocytes as indicated by the inhibited expression of the pro-inflammatory cytokines IL-1 $\beta$  and TNF- $\alpha$  and the inhibition of the synthesis of NO and COX-2 expression in primary chondrocytes.<sup>21</sup> Furthermore, CSE inhibited IL-1 $\beta$ -induced overexpression of matrix metalloproteinases in human chondrocytes, and thus may attenuate the progression of osteoarthritis.<sup>22</sup> Brazilin was suggested to be the major active compound of CSE, however, the contribution of other chemical entities to the suggested anti-arthritis activity of *C. sappan* is currently unknown. Furthermore, the anti-inflammatory effect of only a few compounds from *C. sappan* has been elucidated so far, while the effect of most compounds is still not clear. Additionally, the literature is not consistent on the biological activities of several of the isolated compounds.

The present study aimed to assess the anti-inflammatory activity of episappanol, protosappanin C, brazilin, (iso-)protosappanin B and sappanol isolated from CSE in macrophages and chondrocytes *in vitro*. Three cytokines indicating the inflammatory response were chosen, two cytokines which enhance inflammation (IL-6 and TNF- $\alpha$ ) and one which counteracts inflammation (IL-10).

## 2. Materials and methods

### 2.1. Chemicals and reagents

Lipopolysaccharides from *E. coli* and thiazolyl blue tetrazolium bromide (MTT) were obtained from Sigma-Aldrich (St. Louis, MO, USA). Acetonitrile (HPLC grade) was purchased from Promocore (Wesel, Germany). Dulbecco's modified Eagle's medium (DMEM), foetal bovine serum (FBS), L-glutamine, gentamicin, penicillin and streptomycin were purchased from Life Technologies (Carlsbad, CA, USA). Fetal calf serum (FCS) was obtained from Biochrom (Cambridge, UK). 12-Well microplates were obtained from Iwaki (Asahi glass, Chiyoda, Japan). The kits for enzyme-linked immunosorbent assays (ELISA, Ready-SET-Go!) of mouse and human TNF- $\alpha$ , IL-6, and IL-10 were purchased from eBioscience (San Diego, CA, USA). All other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA) and Merck (Darmstadt, Germany).

### 2.2. Plant material and extraction

The heartwood of *Caesalpinia sappan* was collected in Chiang Mai province, Thailand in 2013, and identified by comparison with the voucher specimen (no. 87-1631) at the Herbarium Section, Faculty of Pharmacy, Chiang Mai University. Powdered heartwood was extracted with ethanol for 24 h under stirring

and the resulting extract was filtered and concentrated under vacuum to yield a solid extract.

### 2.3. Characterization of the extract

The extract was dissolved in 20% ethanol and characterized using HPLC with ultraviolet detection (HPLC-UV) on an UltiMate 3000 HPLC system (Thermo Fisher Scientific, Waltham, MA, USA) connected to a security guard cartridge followed by a Kinetex C-18 column (5  $\mu$ m C18, 4.6  $\times$  150 mm, Phenomenex, Torrance, CA, USA). The mobile phase consisted of solvent A (water-acetonitrile 95:5, 0.1% trifluoroacetic acid, TFA in water) and solvent B (acetonitrile, 0.1% TFA). The gradient profile was as follows: from 0–5 min, 100% A; 5–30 min, from 0 to 17.5% B; 30–33 min from 17.5 to 50% B; 33–34 min, 50 to 100% B; 34–35 min, 100% B; 36–38 min, 100% A. A flow rate of 0.5 ml min<sup>-1</sup> was used. The elution profile was recorded using a photodiode array detector PDA-100 (Dionex) set at 280 nm.

### 2.4. Preparative scale separation of the extract

The extract was separated using UltiMate 3000 HPLC system with a Hypersil Gold preparative guard column and HPLC column (5  $\mu$ m C18, 21  $\times$  250 mm, Thermo Scientific). The same gradient profile and eluents as for analytical HPLC were used with a flow rate of 20 ml min<sup>-1</sup>. Isolated fractions were lyophilized using a Heto Power Dry LL3000 freeze-dryer (Thermo Scientific).

Due to the instability of fractions 1 and 5 under acidic conditions, the preparative separation of these fractions was performed using solvent A (5% acetonitrile in water) and solvent B (acetonitrile) without TFA, but with the same gradient as described before.

### 2.5. Identification of isolated fractions using NMR

NMR spectra were recorded from acetone- $d_6$  solutions on a Bruker Avance III 400 instrument (Bruker, Germany) at 25 °C using a directly detecting BBFO probe (400 MHz for <sup>1</sup>H, 100 MHz for <sup>13</sup>C). The solvent (residual) signals served as an internal standard which were related to TMS with  $\delta$  2.05 ppm (<sup>1</sup>H) and  $\delta$  29.84 ppm (<sup>13</sup>C). The unambiguous assignment of NMR resonances was achieved by the combined application of standard NMR spectroscopic techniques such as APT, COSY, TOCSY, NOESY, gs-HSQC and gs-HMBC using standard Bruker software.

### 2.6. Identification of the compounds by high resolution mass spectrometry (HR-MS)

HR-MS was performed using a MaXis ESI (electrospray ionization) Q-TOF (quadrupole time of flight) mass spectrometer (Bruker Daltonics, Bremen, Germany). All measurements were performed in positive mode, with a capillary voltage of 4500 V, nebulizer pressure of 0.4 bar, dry gas flow of 4 L min<sup>-1</sup>, dry temperature of 180 °C. The quadrupole ion energy was set at 4 eV and the collision cell energy at 6 eV and a mass range between 50 and 2500 *m/z* was screened. Data were analyzed using a Bruker Compass Data Analysis 4.0.

**Table 1** RT-qPCR: primers used

Gene	Species	Accession number	Forward primer	Reverse primer	Efficiency (%)
IL6	Human	NM_000600	ATAGGACTGGAGATGTCTGAGG	AGGCAACTGGACCCGAAGG	93.5
SDHA	Human	NM_004168	TGGGACAAGAGGGCATCTG	CCACCACTGCATCAAATTATG	94.6
TNFA	Human	NM_000594	TCAGCAAGGACAGCAGAGG	CAGTATGTGAGAGGAAGAGAAC	100
IL6	Mouse	NM_031168	CTGTCTATACCACTTCAC	CATCATCGTTGTTCATAC	90.6
SDHA	Mouse	NM_023281	CCAGGACTTACAATTGT	TTGACTGTTGATGAGAAT	94.5
TNFA	Mouse	NM_013693	TTCTGTCTACTGAACTTC	CCATAGAACTGATGAGAG	83.8

## 2.7. Analysis of the anti-inflammatory activity in macrophages

The analysis of the anti-inflammatory activity in LPS-stimulated murine macrophages (RAW 264.7, American Type Culture Collection, ATCC-TIB-71) was performed as previously described by Mueller *et al.* (2010).<sup>23</sup> In brief, cells were seeded at a density of  $2 \times 10^6$  cells per well in 12-well plates. After 24 h incubation at 37 °C, cells were pretreated with test substances for 3 h before LPS was added at a final concentration of 1 µg ml<sup>-1</sup>. After further 24 h of incubation, the medium was removed, centrifuged and stored at -20 °C prior to analysis by ELISA.

The secretion of TNF-α, IL-6, and IL-10 in the cell supernatant was analyzed using ELISA kits according to the manufacturer's protocol. The optical density at 450 nm, corrected by the reference wavelength 570 nm, was measured using an Infinite M200 microplate reader (Tecan, Crailsheim, Germany).

The viability of the cells was tested using a MTT assay as previously described.<sup>23</sup> If the viability was 25% below the control, a compound was considered cytotoxic. The ELISA results indicating the cytokine secretion were normalized to the MTT values to reduce any variation arising from the differences in the cell density. Cells treated with only LPS served as a positive control and the amount of secreted cytokines was defined as 100%. All the results from the tested compounds were then calculated as a percent of this value. Dexamethasone was used as the reference drug. The assay was performed in triplicate on separate days. The mean and standard deviation are presented in the figures.

Total RNA extraction was carried out using the NucleoSpin RNA II Kit (Macherey-Nagel, Duren, Germany) according to the manufacturer's instructions. Each sample was run on the Agilent 2100 Bioanalyzer Nano LabChip (Santa Clara, CA, USA) for quality control and quantification of total RNA prior to reverse transcription into cDNA using the high capacity cDNA reverse transcription kit (Life Technologies). RNA integrity numbers were between 9.3 and 9.9.

SYBR-green-based RT-qPCR assays for IL6, TNFA and IL10 expression were used as previously described<sup>21,22</sup> and with respect to the MIQE guidelines.<sup>24</sup> The mRNA levels of the target genes were calculated as quantities relative to the untreated control group, considering both amplification efficiencies and normalization to succinate dehydrogenase complex, subunit A (SDHA). Details on primers are given in Table 1.

## 2.8. Analysis of the anti-inflammatory activity in chondrocytes

As an established *in vitro* model for human chondrocytes, SW1353 cells (ATCC-HTB-94) were cultured as previously described.<sup>22</sup> In brief, cells were seeded at a density of  $2 \times 10^4$  cm<sup>-2</sup> in 6-well plates and cultured in DMEM containing 10% FCS and 100 µg per ml gentamicin at 37 °C.

Prior to treatment, SW1353 cells were starved overnight in serum-free DMEM supplemented with gentamicin. Then, cells were pretreated for 1 h with the isolated fractions in serum-free medium followed by the addition of 10 ng per ml IL-1β and further incubation for 24 h. As a negative control, cells were left untreated, and as a positive control, cells were treated only with IL-1β. Afterwards, cell supernatants were collected, centrifuged and stored at -80 °C. All treatments were performed in duplicate and were repeated three times. ELISA assays were conducted according to the manufacturer's protocol (Ready-SET-Go! ELISA, eBioscience). SYBR-green-based RT-qPCR assays for IL6 and TNFA expression were conducted as described above (section 2.7). Details on primers are given in Table 1. Concomitantly, MTT assays were performed as indicated above (section 2.7) to determine cell viability.

## 2.9. Statistics

Statistics were performed using one way analysis of variance using Tukey's test comparing all study groups (95% confidence interval). *P*-Values <0.05 were considered as significant. The IC<sub>50</sub> values were determined using Table Curve 2D (Systat Software, San Jose, CA, USA).

## 3. Results

### 3.1. Separation of the plant extract

Five fractions were separated from CSE as shown in the chromatogram of the preparative separation (Fig. 1). After lyophilization, the fractions were dissolved in 20% ethanol and analyzed by analytical HPLC. Fractions 2, 3 and 4 showed single peak purity. Fractions 1 and 5 were not stable when isolated under acidic conditions and showed 2–3 peaks. We changed the solvent system for preparative HPLC to solvent A (5% acetonitrile in water) and solvent B (acetonitrile) and applied the same gradient as described in the Materials and methods section. Finally, fractions 1 and 5 were obtained with one peak purity.



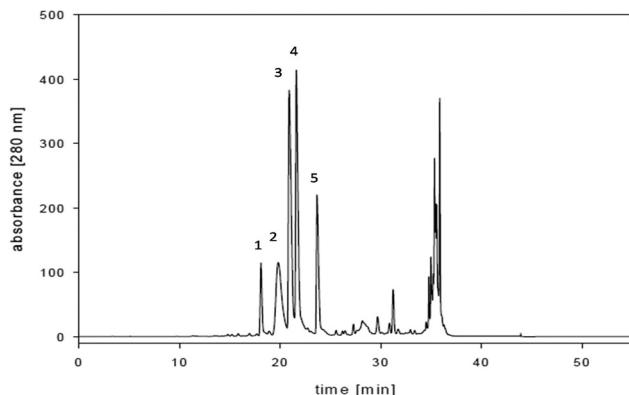


Fig. 1 Chromatogram of the preparative HPLC used for separation of the 20% ethanolic extract of *C. sappan*.

### 3.2. Identification of the compounds using NMR

The isolated compounds were identified as episappanol (1), protosappanin C (2), brazilin (3), a mixture of protosappanin B and isoprotosappanin B (4) and sappanol (5) using NMR. The structures are shown in Fig. 2.

**Episappanol (1).**  $^1\text{H-NMR}$  (400 MHz, acetone- $d_6$ ):  $\delta$  8.32 (br s, 0.42 H, OH), 7.73 (br s, 1 H, OH), 7.06 (d, 1 H,  $^3J_{5,6} = 8.4$  Hz, H-5), 6.89 (d, 1 H,  $^4J_{2,6'} = 1.8$  Hz, H-2'), 6.73 (d, 1 H,  $^3J_{5',6'} = 8.0$  Hz, H-5'), 6.69 (dd, 1 H,  $^3J_{6,5'} = 8.0$  Hz,  $^4J_{6',2'} = 1.8$  Hz, H-6'), 6.36 (dd, 1 H,  $^3J_{6,5} = 8.4$  Hz,  $^4J_{6,8} = 2.3$  Hz, H-6), 6.23 (d, 1 H,  $^4J_{8,6} = 2.3$  Hz, H-8), 4.31 (br d, 0.48 H, OH), 4.22 (s, 1 H, H-4), 4.05 (d, 1 H,  $^2J_{2b,2a} = 11.0$  Hz, H-2b), 3.75 (d, 1 H,  $^2J_{2a,2b} = 11.0$  Hz, H-2a), 2.91 (d, 1 H,  $^2J_{9b,9a} = 13.8$  Hz, H-9b), 2.64 (d, 1 H,  $^2J_{9a,9b} = 13.8$  Hz, H-9a), 2.95 (s) and 2.92 (s) (13H, OH and H<sub>2</sub>O).

$^{13}\text{C-NMR}$  (100 MHz, acetone- $d_6$ ):  $\delta$  158.9 (C-7), 155.7 (C-8a), 145.2 (C-3'), 144.3 (C-4'), 132.2 (C-5), 129.0 (C-1'), 123.2 (C-6'), 119.0 (C-2'), 117.0 (C-4a), 115.5 (C-5'), 108.9 (C-6), 103.0 (C-8), 71.2 (C-3), 70.0 (C-4), 69.2 (C-2), 39.0 (C-9).

The  $^1\text{H-NMR}$  data are in agreement with Namikoshi *et al.* (1987).<sup>8</sup>

**Protosappanin C (2).**  $^1\text{H-NMR}$  (400 MHz, acetone- $d_6$ ):  $\delta$  9.78 (s, 1 H, 7-CHO), 7.03 (d, 1 H,  $^3J_{1,2} = 8.4$  Hz, H-1), 6.79 (s, 1 H, H-12), 6.78 (s, 1 H, H-9), 6.61 (m, 1 H, H-2), 6.51 (broad s, 1 H, H-4), 4.25 (A-part of an AB-system, 1 H,  $^2J_{6,6'} = 12.5$  Hz, H-6), 4.02 (B-part of an AB-system, 1 H,  $^2J_{6',6} = 12.5$  Hz, H-6'), 2.80 (A-part of an AB-system, 1 H,  $^2J_{8,8'} = 13.8$  Hz, H-8), 2.57 (B-part of an AB-system, 1 H,  $^2J_{8',8} = 13.8$  Hz, H-8').

$^{13}\text{C-NMR}$  (100 MHz, acetone- $d_6$ ):  $\delta$  204.0 (7-CHO), 158.9 (C-4a), 158.6 (C-3), 145.0 (C-10), 144.7 (C-11), 133.4 (C-1), 132.4 (C-12a), 125.3 (C-8a), 123.0 (C-12b), 119.9 (C-9), 117.2 (C-12), 111.5 (C-2), 108.0 (C-4), 75.1 (C-7), 74.4 (C-6), 37.1 (C-8).

**Brazilin (3).**  $^1\text{H-NMR}$  (400 MHz, acetone- $d_6$ ):  $\delta$  7.20 (d, 1 H,  $^3J_{5,6} = 8.3$  Hz, H-5), 6.76 (s, 1 H, H-5'), 6.65 (s, 1 H, H-2'), 6.49 (dd, 1 H,  $^3J_{6,5} = 8.3$  Hz,  $^4J_{6,8} = 2.5$  Hz, H-6), 6.29 (d, 1 H,  $^4J_{8,6} = 2.5$  Hz, H-8), 3.97 (broad s, 1 H, H-4), 3.94 (dd, 1 H,  $^2J_{2a,2b} = 11.2$  Hz,  $^4J_{2a,4} = 1.4$  Hz, H-2a), 3.71 (d, 1 H,  $^2J_{2b,2a} = 11.2$  Hz, H-2b), 3.01 (d, 1 H,  $^2J_{9a,9b} = 15.7$  Hz, H-9a), 2.82 (d, 1 H,  $^2J_{2b,2a} = 15.7$  Hz, H-9b).

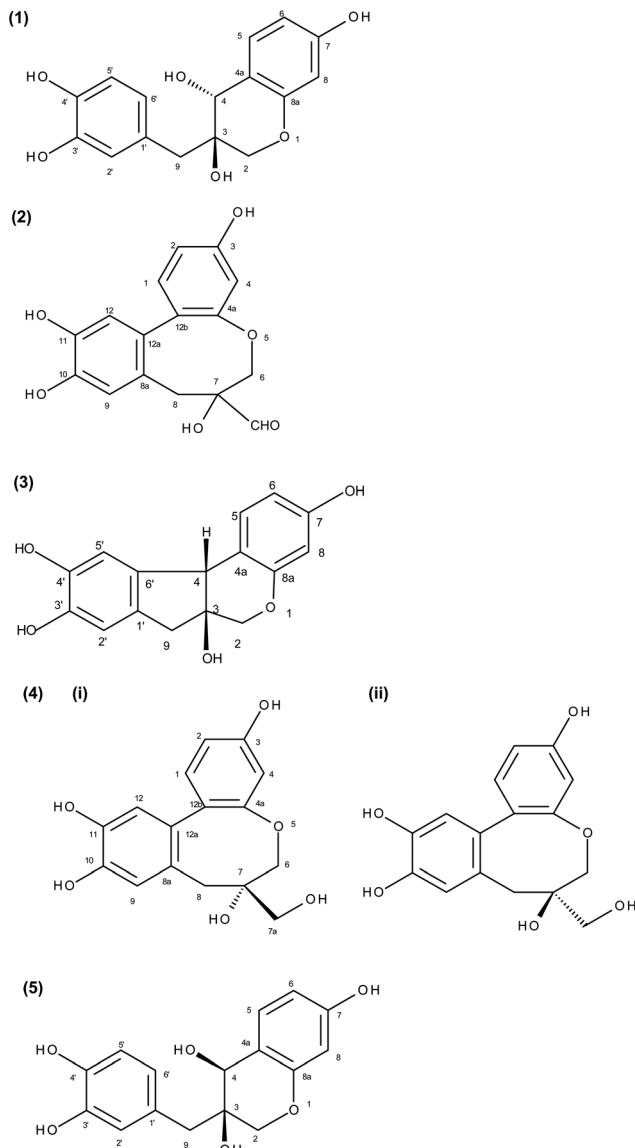


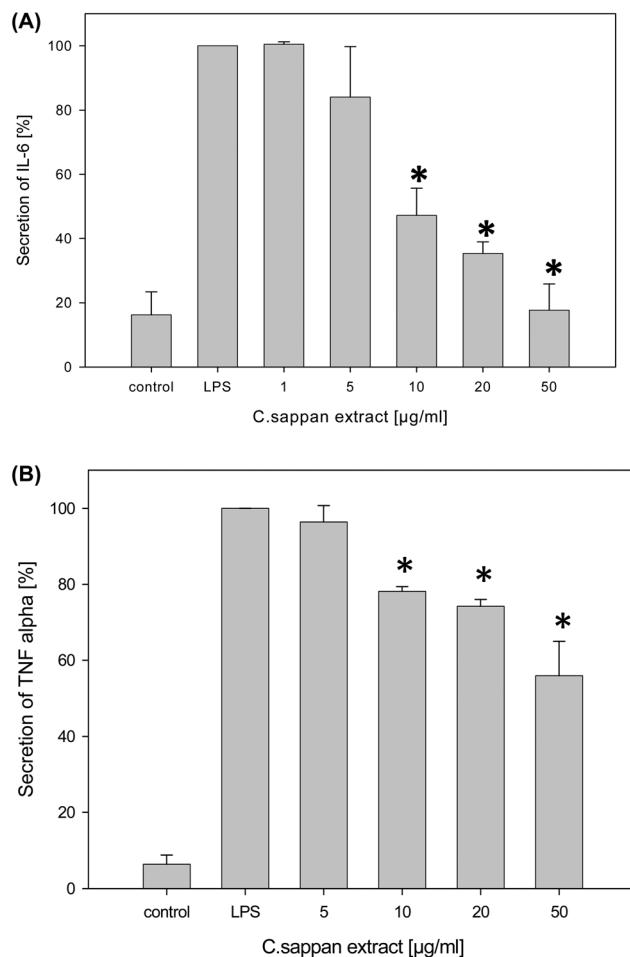
Fig. 2 Structure of the five isolated fractions: (1) episappanol, (2) protosappanin C, (3) brazilin, (4) (i) protosappanin B, (ii) isoprotosappanin B and (5) sappanol.

$^{13}\text{C-NMR}$  (100 MHz, acetone- $d_6$ ):  $\delta$  157.6 (C-7), 155.5 (C-8a), 145.1 (C-3'), 144.8 (C-4'), 137.4 (C-6'), 132.0 (C-5), 131.5 (C-1'), 115.6 (C-4a), 112.6 (C-2'), 112.3 (C-5'), 109.6 (C-6), 103.9 (C-8), 77.7 (C-3), 70.8 (C-2), 51.1 (C-4), 42.9 (C-9).

The numbering of the brazilin ring system used here (Fig. 3C) is the one introduced by Fu *et al.* (2008).<sup>9</sup> Fukue *et al.* and Kim *et al.* used a different numbering system.<sup>25,26</sup>

**(Iso-)protosappanin B (4). Major isomer (isoprotosappanin B):**  $^1\text{H-NMR}$  (400 MHz, acetone- $d_6$ ):  $\delta$  6.99 (d, 1 H,  $^3J_{1,2} = 8.3$  Hz, H-1), 6.77 (br s, 1 H, H-12), 6.75 (br s, 1 H, H-9), 6.57 (dd, 1 H,  $^3J_{2,1} = 8.3$  Hz,  $^4J_{2,4} = 2.4$  Hz, H-2), 6.48 (d, 1 H,  $^4J_{4,2} = 2.4$  Hz, H-4), 4.12 (d, 1 H,  $^2J_{6,6'} = 12.0$  Hz, H-6), 3.86 (d, 1 H,  $^2J_{6',6} = 12.0$  Hz, H-6'), 3.58 (A-part of an AB-system, 1 H,  $^2J_{7a,7a'} = 10.9$  Hz, H-7a), 3.48 (B-part of an AB-system, 1 H,





**Fig. 3** Influence of CSE on the secretion of cytokines in macrophages: (A) IL-6, (B) TNF-α.

$^2J_{7a',7a} = 10.9$  Hz, H-7a'), 2.58 (A-part of an AB-system, 1 H,  $^2J_{8,8'} = 13.4$  Hz, H-8), 2.50 (B-part of an AB-system, 1 H,  $^2J_{8',8} = 13.4$  Hz, 1 H, H-8').

$^{13}\text{C-NMR}$  (100 MHz, acetone- $d_6$ ):  $\delta$  159.2 (C-4a), 158.7 (C-3), 144.4 (C-10 and C-11), 133.3 (C-1), 132.3 (C-12a), 127.3 (C-8a), 123.3 (C-12b), 119.6 (C-9), 117.2 (C-12), 111.0 (C-2), 108.0 (C-4), 75.8 (C-6), 72.1 (C-7), 67.9 (C-7a), 39.8 (C-8).

**Minor isomer (protosappanin B):**  $^1\text{H-NMR}$  (400 MHz, acetone- $d_6$ ):  $\delta$  7.00 (d, 1 H,  $^3J_{1,2} = 8.1$  Hz, H-1), 6.85 (br s, 1 H, H-9), 6.71 (br s, 1 H, H-12), 6.62 (m, 1 H, H-2), 6.56 (m, 1 H, H-4), 4.35 (d, 1 H,  $^2J_{6,6'} = 11.7$  Hz, H-6), 3.57 (m, 2 H, H-6' and H-7a), 3.38 (d, 1 H,  $^2J_{7a',7a} = 10.9$  Hz, H-7a'), 2.74 (A-part of an AB-system, 1 H,  $^2J_{8,8'} = 13.5$  Hz, H-8), 2.67 (B-part of an AB-system, 1 H,  $^2J_{8',8} = 13.5$  Hz, 1 H, H-8').

$^{13}\text{C-NMR}$  (100 MHz, acetone- $d_6$ ):  $\delta$  160.6 (C-4a), 158.7 (C-3), 144.4 (C-10 and C-11), 132.3 (C-1), 131.7 (C-12a), 128.3 (C-8a), 125.2 (C-12b), 118.9 (C-9), 117.4 (C-12), 111.7 (C-2), 108.8 (C-4), 77.2 (C-6), 72.6 (C-7), 65.6 (C-7a), 42.5 (C-8).

The  $^1\text{H}$  and  $^{13}\text{C-NMR}$  data are in agreement with Fu *et al.* (2008).<sup>9</sup>

**Sappanol (5).**  $^1\text{H-NMR}$  (400 MHz, acetone- $d_6$ ):  $\delta$  8.47 (br s, 0.54 H, OH), 7.74 (br s, 1 H, OH), 7.11 (d, 1 H,  $^3J_{5,6} = 8.3$  Hz,

H-5), 6.76 (d, 1 H,  $^4J_{2',6'} = 2.0$  Hz, H-2'), 6.70 (d, 1 H,  $^3J_{5',6'} = 8.0$  Hz, H-5'), 6.52 (dd, 1 H,  $^3J_{6',5'} = 8.0$  Hz,  $^4J_{6',2'} = 2.0$  Hz, H-6'), 6.43 (dd, 1 H,  $^3J_{6,5} = 8.3$  Hz,  $^4J_{6,8} = 2.4$  Hz, H-6), 6.28 (d, 1 H,  $^4J_{8,6} = 2.4$  Hz, H-8), 4.45 (br d, 0.61 H, OH), 4.22 (s, 1 H, H-4), 3.86 (d, 1 H,  $^2J_{2,2'} = 10.6$  Hz, H-2), 3.67 (dd, 1 H,  $^2J_{2',2} = 10.6$  Hz,  $^4J_{2',4} = 1.2$  Hz, H-2'), 2.63 (A-part of an AB-system, 1 H,  $^2J_{9,9'} = 14.1$  Hz, H-9), 2.59 (B-part of an AB-system, 1 H,  $^2J_{9',9} = 14.1$  Hz, 1 H, H-9').

$^{13}\text{C-NMR}$  (100 MHz, acetone- $d_6$ ):  $\delta$  159.3 (C-7), 155.6 (C-8a), 145.2 (C-3'), 144.5 (C-4'), 132.7 (C-5), 129.0 (C-1'), 122.9 (C-6'), 118.7 (C-2'), 116.4 (C-4a), 115.4 (C-5'), 109.4 (C-6), 103.1 (C-8), 70.2 (C-3), 69.4 (C-4), 67.8 (C-2), 40.7 (C-9).

The  $^1\text{H-NMR}$  data are in agreement with Namikoshi *et al.* (1987).<sup>8</sup>

### 3.3. Identification of the compounds using HR-MS

As shown in Table 2, the results of the HR-MS were consistent with the NMR data.

### 3.4. Anti-inflammatory activity in LPS-stimulated macrophages

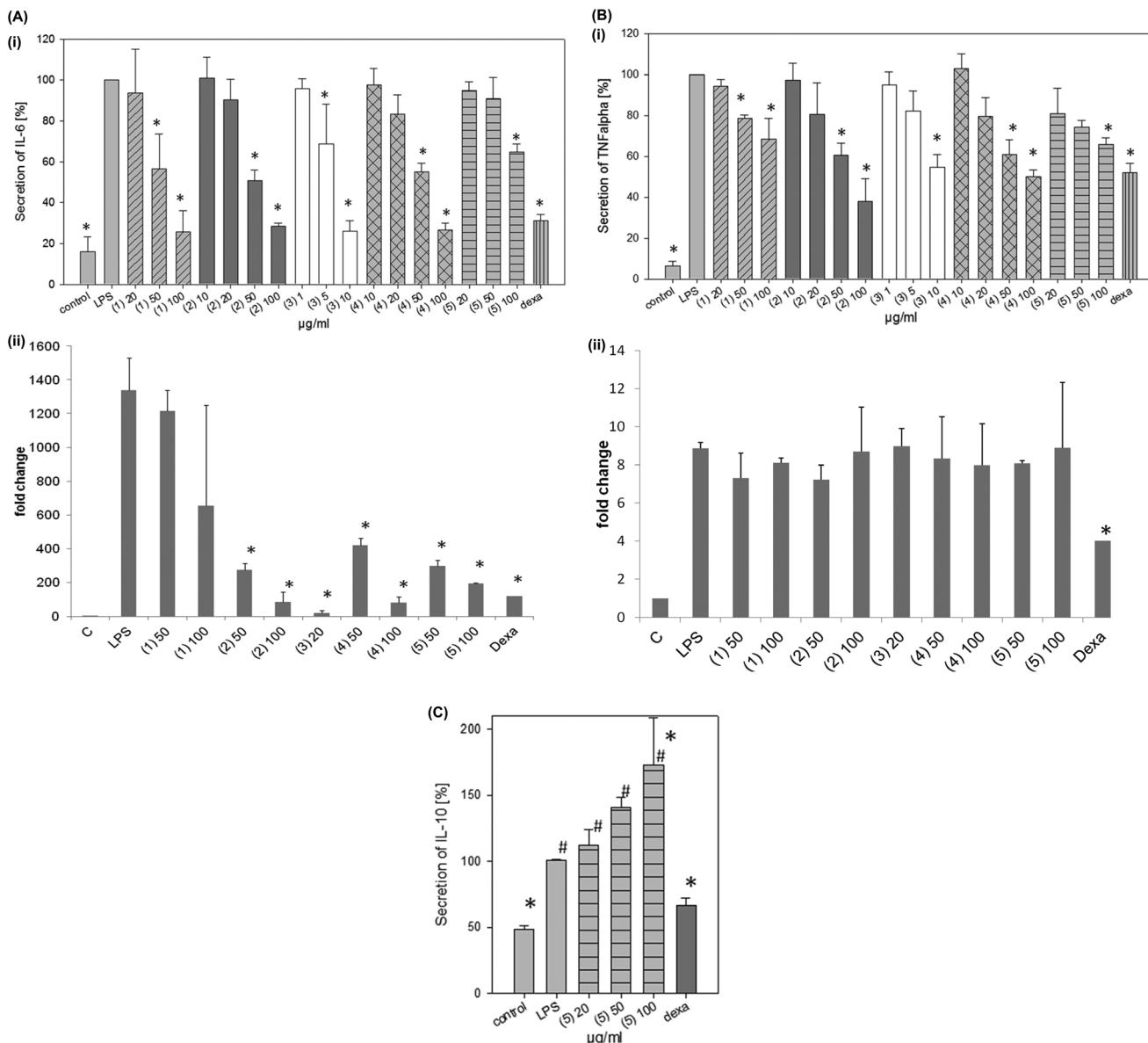
Brazolin (3) was significantly cytotoxic at a concentration of  $\geq 20$   $\mu\text{g ml}^{-1}$  as determined by the MTT assay and could therefore only be analyzed at lower concentrations. The remaining fractions were not cytotoxic up to 100  $\mu\text{g ml}^{-1}$  and tested up to this concentration. The entire extract was cytotoxic at a concentration of 100  $\mu\text{g ml}^{-1}$ , but not at a concentration of 50  $\mu\text{g ml}^{-1}$  and thus it was only applied up to 50  $\mu\text{g ml}^{-1}$  in subsequent bioactivity assays.

The absolute level of IL-6 secretion was up to 5000  $\text{pg ml}^{-1}$  for the positive control and up to 100  $\text{pg ml}^{-1}$  for the negative control. The values for the TNF- $\alpha$  secretion were up to 2500  $\text{pg ml}^{-1}$  for the positive control and up to 300  $\text{pg ml}^{-1}$  for the negative control, respectively. CSE significantly exerted anti-inflammatory effects down to a concentration of 10  $\mu\text{g ml}^{-1}$  as indicated by a reduction of the IL-6 and TNF- $\alpha$  secretion with an  $IC_{50}$  of 8 and 36  $\mu\text{g ml}^{-1}$ , respectively (Fig. 3). No significant effect on IL-10 was observed.

All the isolated fractions (1–5) induced a significant and dose-dependent change in the cytokine secretion profile as indicated by a decrease of the pro-inflammatory cytokines IL-6 (Fig. 4Ai) and TNF- $\alpha$  (Fig. 4Bi) (Table 3) at the cytokine level. At the mRNA level, the decrease of IL6 was significant whereas the decrease of TNFA was not significant (Fig. 4Aii and 4Bii). Fraction 5 increased the secretion of the anti-inflammatory IL-10 (Fig. 4C), whereas the other fractions had no significant

**Table 2** Results of HR-MS

Fraction	Compound	[M + H]	MW [g]
1	Episappanol	305.1020	304.0941
2	Protosappanol C	303.0863	302.0784
3	Brazolin	287.0914	286.0835
4	Protosappanol B + isoprotosappanol B	305.1020	304.0941
5	Sappanol	305.1020	304.0941



**Fig. 4** Influence of the 5 isolated fractions (i) on the secretion of cytokines and (ii) on the mRNA expression in macrophages: (A) IL-6, (B) TNF- $\alpha$ ; (C) influence of sappanol on the secretion of IL-10. Untreated cells (control), cells treated with 1  $\mu$ g per ml LPS only (LPS) or treated with LPS and 10  $\mu$ M dexamethasone (dexa) were included. \*Significant differences compared to positive control ( $p < 0.05$ ). #Significant differences compared to untreated cells ( $p < 0.05$ ).

**Table 3** IC<sub>50</sub> values of the 5 fractions in macrophages and chondrocytes, as calculated from ELISA experiments

Sample	Macrophages		Chondrocytes	
	IL-6	TNF- $\alpha$	IL-6	TNF- $\alpha$
1	155 $\mu$ M	>164 $\mu$ M	>33 $\mu$ M	24 $\mu$ M
2	123 $\mu$ M	121 $\mu$ M	21 $\mu$ M	16 $\mu$ M
3	18 $\mu$ M	29 $\mu$ M	5 $\mu$ M	3 $\mu$ M
4	128 $\mu$ M	>164 $\mu$ M	>33 $\mu$ M	22 $\mu$ M
5	>164 $\mu$ M	>164 $\mu$ M	>33 $\mu$ M	>33 $\mu$ M
CSE	8 $\mu$ g ml <sup>-1</sup>	36 $\mu$ g ml <sup>-1</sup>	>10 $\mu$ g ml <sup>-1</sup>	1.6 $\mu$ g ml <sup>-1</sup>
Dexamethasone	2 nM	1 $\mu$ M	65 nM	66 nM

effect on this cytokine. Brazilin (3) exerted the highest anti-inflammatory effect with a significant inhibition of the IL-6 secretion down to 5  $\mu$ g ml<sup>-1</sup> and a significant inhibition of the TNF- $\alpha$  secretion down to 10  $\mu$ g ml<sup>-1</sup>. The IC<sub>50</sub> values were 18  $\mu$ M for IL-6 and 29  $\mu$ M for TNF- $\alpha$ , respectively. Episappanol (1), protosappanol C (2), and (iso-)protosappanol B (4) significantly inhibited the IL-6 and TNF- $\alpha$  secretion at 50  $\mu$ g ml<sup>-1</sup>. Sappanol (5) significantly reduced the IL-6 and TNF- $\alpha$  secretion at the highest concentration tested (100  $\mu$ g ml<sup>-1</sup>). Besides brazilin, protosappanol C (2) and (iso-)protosappanol B (4) were the most effective fractions with IC<sub>50</sub> values of 123  $\mu$ M and 128  $\mu$ M as indicated by IL-6 secretion (Table 3).

### 3.5. Anti-inflammatory activity in IL-1 $\beta$ -stimulated chondrocytes

Based on the results of preceding MTT experiments (data not shown), bразилин and CSE were tested up to a concentration of 10  $\mu\text{g ml}^{-1}$  and fractions 1, 2, 4 and 5 up to 20  $\mu\text{g ml}^{-1}$  to avoid cytotoxic effects. The absolute level of IL-6 secretion was up to 110  $\text{pg ml}^{-1}$  for the positive control and up to 10  $\text{pg ml}^{-1}$  for the negative control. The values for the TNF- $\alpha$  secretion were up to 150  $\text{pg ml}^{-1}$  for the positive control and up to 12  $\text{pg ml}^{-1}$  for the negative control, respectively. CSE significantly inhibited the IL-6 secretion at 10  $\mu\text{g ml}^{-1}$  and the TNF- $\alpha$  secretion at 2  $\mu\text{g ml}^{-1}$  (Fig. 5). All the isolated fractions (1–5) significantly inhibited the IL-6 secretion in chondrocytes (Fig. 6A, Table 3). Brazilin (3) was most effective in this cell line also and significantly inhibited the IL-6 secretion at 5  $\mu\text{g ml}^{-1}$  and an IC<sub>50</sub> of 3  $\mu\text{M}$ . Fractions 2 and 4 showed a significant effect on the IL-6 secretion at 10  $\mu\text{g ml}^{-1}$  and fractions 1 and 5 at 20  $\mu\text{g ml}^{-1}$ . The IC<sub>50</sub> values for IL-6 ranged between 5  $\mu\text{M}$  for bразилин and >33  $\mu\text{M}$  for sappanol (Table 3). IL-10 was not secreted by chondrocytes at detectable levels and was therefore not analyzed in this cell line.

All the fractions (1–5) exerted a significant reduction of the TNF- $\alpha$  secretion at a concentration of 10  $\mu\text{g ml}^{-1}$  or lower

(Fig. 6B, Table 3). Brazilin (3) was effective at 1  $\mu\text{g ml}^{-1}$  also. The IC<sub>50</sub> values for TNF- $\alpha$  ranged between 3  $\mu\text{M}$  for bразилин and >33  $\mu\text{M}$  for sappanol (Table 3). These findings are in agreement with the qPCR results, showing that the fractions (1–5) significantly inhibited IL6 and TNFA mRNA expression levels at a concentration of 10  $\mu\text{g ml}^{-1}$  (Fig. 6ii) with the highest effect observed for bразилин.

## 4. Discussion

The anti-inflammatory and anti-arthritic effects of the extract of *C. sappan* have been shown previously.<sup>6,21,22</sup> However, the anti-inflammatory effect of several compounds from CSE has not been fully elucidated and the anti-arthritic effect of isolated compounds from *C. sappan* besides bразилин has not been previously shown.

In this study, five fractions were isolated from the ethanolic (20%) extract of *C. sappan*, namely episappanol (1), protosappanin C (2), bразилин (3), (iso-)protosappanin B (4) and sappanol (5). The occurrence of the isolated compounds in *C. sappan* has already been shown previously.<sup>8,9,27</sup>

To the best of our knowledge, this is the first study showing the anti-inflammatory effect of sappanol and episappanol. Interestingly, sappanol even increases the secretion of the anti-inflammatory, IL-10. The anti-inflammatory effect of protosappanin C and protosappanin B (2, 4) in macrophages has been described previously. Our result showing a moderate effect of protosappanin C is in agreement with Washiyama *et al.* (2009)<sup>10</sup> and Sasaki *et al.* (2007)<sup>17</sup> who showed the effect on PGE<sub>2</sub> or NO reduction in J774.1 macrophage cells. Our results on the anti-inflammatory effects of protosappanin B are in contrast to those of Washiyama *et al.* (2009)<sup>10</sup> who found no activity of protosappanin B in J774.1 macrophage cells. Of note, the literature is not consistent on the anti-inflammatory activity of different protosappanins. Sasaki *et al.*<sup>17</sup> found a slight anti-inflammatory activity of protosappanin B and in J774.1 cells as indicated by the inhibition of NO production and iNOS expression. The compounds isolated from *C. sappan* are unstable which may contribute to the inconsistency of the literature.

The anti-inflammatory effect of CSE and bразилин in RAW 264.7 macrophages in this study is consistent with the literature. Hong *et al.* showed an inhibition of COX-2 and iNOS expression of the methanolic extract in RAW 264.7 macrophages.<sup>6</sup> Brazilin exhibits anti-inflammatory activity *via* various mechanisms such as inhibition of NO production and regulation of nuclear factor kappa-B (NF)- $\kappa$ B and activator protein-1 in RAW 264.7 cells<sup>28–30</sup> or *via* inducing heme oxygenase-1 expression and inhibition of the production of PGE<sub>2</sub>, TNF- $\alpha$  and IL1- $\beta$ .<sup>29</sup>

Here, we also show the bioactivity of 1–5 in human SW1353 chondrocytes. To the best of our knowledge, this is the first report that demonstrates the anti-inflammatory effect of protosappanin B and C, sappanol and episappanol in chondrocytes. The *in vitro* and *in vivo* anti-inflammatory activity of CSE was shown previously.<sup>7,21</sup> We also showed the anti-arthritic effect

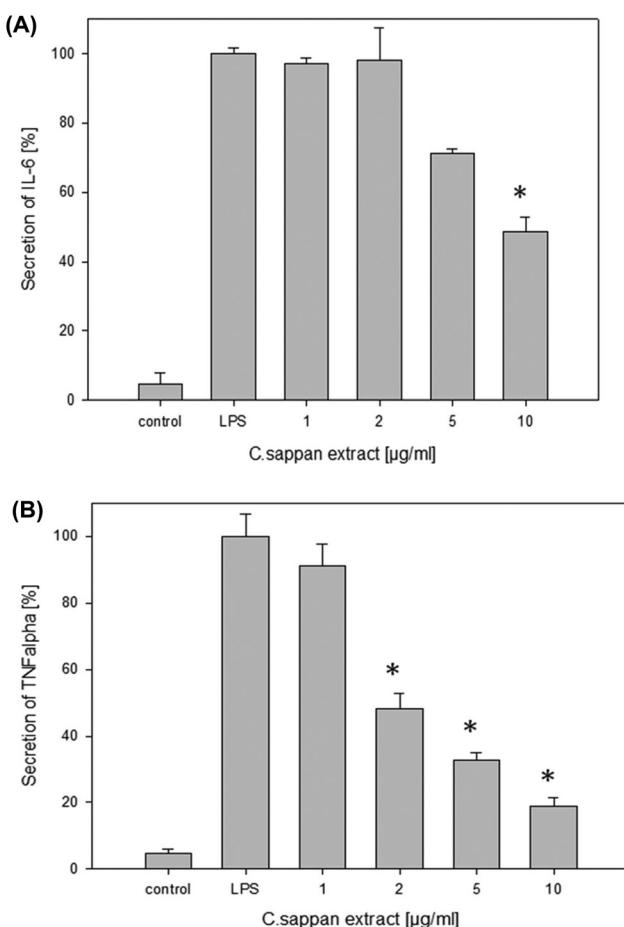
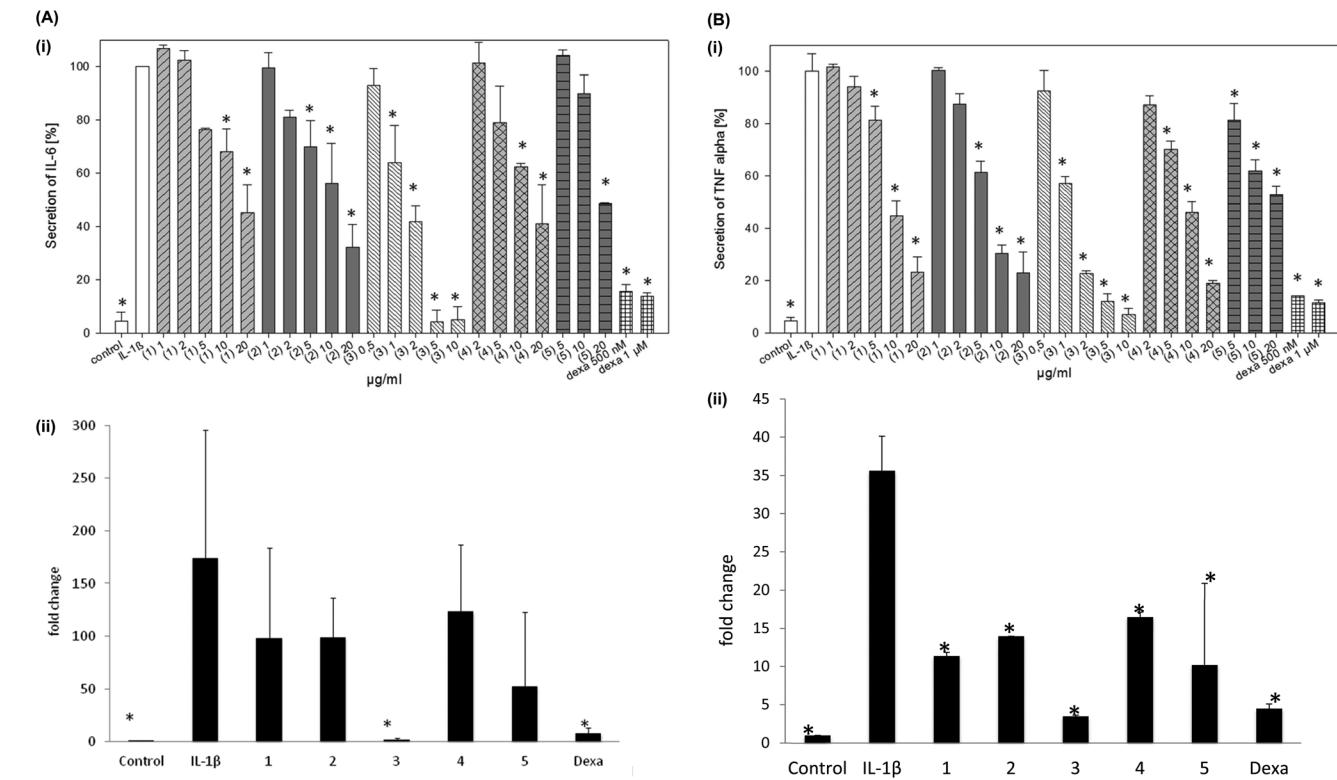


Fig. 5 Influence of CSE on the secretion of cytokines in chondrocytes: (A) IL-6, (B) TNF- $\alpha$ .





**Fig. 6** Influence of the 5 isolated fractions (i) on the secretion of cytokines and (ii) on the mRNA expression in chondrocytes: (A) IL-6, (B) TNF- $\alpha$ . Untreated cells (control), cells treated with 10 ng per ml IL-1 $\beta$  only or with IL-1 $\beta$  and dexamethasone (dexam) were included. For mRNA analyses, all compounds were tested at a concentration of 10  $\mu$ g ml $^{-1}$  and dexamethasone at 10  $\mu$ M. \* Significant differences compared to positive control ( $p < 0.05$ ).

of brazilin previously.<sup>21,22</sup> Thus, this study elucidates the anti-arthritis effect of other constituents besides brazilin.

Furthermore, this study confirms that *C. sappan* exerts an anti-inflammatory effect and might therefore provide an explanation for the analgesic effect *via* the reduction of swelling and inflammation according to its traditional use.<sup>1</sup>

## 5. Conclusion

CSE and the five isolated fractions, episappanol (1), protosappanin C (2), brazilin (3), (iso-)protosappanin B (4) and sappanol (5) showed significant anti-inflammatory properties in two different cell lines, macrophages and chondrocytes. Thus, CSE and its fractions may be potent candidates for complementary and alternative treatment of inflammation and associated diseases such as arthritis. The study confirms the traditional use of *C. sappan* to reduce inflammation, swelling and pain.

## References

- 1 *Chinese Pharmacopoeia Commission*, Medical Science Press, Beijing, 2010.
- 2 C. M. Hu, J. J. Kang, C. C. Lee, C. H. Li, J. W. Liao and Y. W. Cheng, *Eur. J. Pharmacol.*, 2003, **468**, 37–45.
- 3 G. Taeg Oh, J. Hoon Choi, J. Joo Hong, D.-Y. Kim, S.-B. Lee, J.-R. Kim, C.-H. Lee, B.-H. Hyun, S. Ryang Oh, S.-H. Bok and T.-S. Jeong, *Atherosclerosis*, 2001, **159**, 17–26.
- 4 J. Hu, X. Yan, W. Wang, H. Wu, L. Hua and L. Du, *Tsinghua Sci. Technol.*, 2008, **13**, 474–479.
- 5 K.-J. Kim, H.-H. Yu, S.-I. Jeong, J.-D. Cha, S.-M. Kim and Y.-O. You, *J. Ethnopharmacol.*, 2004, **91**, 81–87.
- 6 C. H. Hong, S. K. Hur, O. J. Oh, S. S. Kim, K. A. Nam and S. K. Lee, *J. Ethnopharmacol.*, 2002, **83**, 153–159.
- 7 Y.-Z. Wang, S.-Q. Sun and Y.-B. Zhou, *J. Ethnopharmacol.*, 2011, **136**, 271–278.
- 8 M. Namikoshi, H. Nakata, H. Yamada, M. Nagai and T. Saitoh, *Chem. Pharm. Bull.*, 1987, **35**, 2761–2773.
- 9 L. Fu, X. Huang, Z. Lai, Y. Hu, H. Liu and X. Cai, *Molecules*, 2008, **13**, 1923–1930.
- 10 M. Washiyama, Y. Sasaki, T. Hosokawa and S. Nagumo, *Biol. Pharm. Bull.*, 2009, **32**, 941–944.
- 11 C. Moon, S. Lee, M. Lee and S. Kim, *Life Sci.*, 1993, **53**, 1291–1297.
- 12 H. Xu and S. Lee, *Phytother. Res.*, 2004, **18**, 647–651.
- 13 T. Jayakumar, C. Chang, S. Lin, Y. Huang, C. Hu, A. Elizabeth, S. Lin and C. Choy, *Biomed. Res. Int.*, 2014, **2014**, 403703.
- 14 Y. Chang, S. Huang, W. Lu, C. Chung, W. Chen, S. Lu, K. Lin and J. Sheu, *J. Biomed. Sci.*, 2013, **20**.
- 15 C.-T. Yen, K. Nakagawa-Goto, T.-L. Hwang, P.-C. Wu, S. L. Morris-Natschke, W.-C. Lai, K. F. Bastow, F.-R. Chang,



Y.-C. Wu and K.-H. Lee, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 1037–1039.

16 C. Lee, C. Wang, J. Kang, J. Liao, B. Chiang, H. Chen, C. Hu, C. Lin, S. Huang and Y. Lai, *J. Agric. Food Chem.*, 2012, **60**, 9405–9414.

17 Y. Sasaki, T. Hosokawa, M. Nagai and S. Nagumo, *Biol. Pharm. Bull.*, 2007, **30**, 193–196.

18 M.-J. Chu, Y.-Z. Wang, K. Itagaki, H.-X. Ma, P. Xin, X.-G. Zhou, G.-Y. Chen, S. Li and S.-Q. Sun, *J. Ethnopharmacol.*, 2013, **148**, 37–44.

19 A. Liu, S. Shu, H. Qin, S. Lee, Y. Wang and G. Du, *Planta Med.*, 2009, **75**, 337–339.

20 J. Wu, M. Zhang, H. Jia, X. Huang, Q. Zhang, J. Hou and Y. Bo, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 2010, **381**, 83–92.

21 S. Q. Wu, M. Otero, F. M. Unger, M. B. Goldring, A. Phrutivorapongkul, C. Chiari, A. Kolb, H. Viernstein and S. Toegel, *J. Ethnopharmacol.*, 2011, **138**, 364–372.

22 S. Toegel, S. Wu, M. Otero, M. Goldring, P. Leelaporntpisid, C. Chiari, A. Kolb, F. Unger, R. Windhager and H. Viernstein, *Genes Nutr.*, 2012, **7**, 307–318.

23 M. Mueller, S. Hobinger and A. Jungbauer, *Food Chem.*, 2010, 987–996.

24 S. A. Bustin, J. F. Beaulieu, J. Huggett, R. Jaggi, F. S. Kibenge, P. A. Olsvik, L. C. Penning and T. S. Toegel, *BMC Mol. Biol.*, 2010, **74**.

25 C. Fuke, J. Yamahara, T. Shimokawa, J.-E. Kinjo, T. Tomimatsu and T. Nohara, *Phytochemistry*, 1985, **24**, 2403–2405.

26 D. S. Kim, N.-I. Baek, S. R. Oh, K. Y. Jung, I. S. Lee and H.-K. Lee, *Phytochemistry*, 1997, **46**, 177–178.

27 K. Mitani, F. Takano, T. Kawabata, A. Allam, M. Ota, T. Takahashi, N. Yahagi, C. Sakurada, S. Fushiya and T. Ohta, *Planta Med.*, 2013, **79**, 37–44.

28 I.-K. Bae, H.-Y. Min, A.-R. Han, E.-K. Seo and S. K. Lee, *Eur. J. Pharmacol.*, 2005, **513**, 237–242.

29 C. M. Hu, Y. H. Liu, K. P. Cheah, J. S. Li, C. S. K. Lam, W. Y. Yu and C. S. Choy, *J. Ethnopharmacol.*, 2009, **121**, 79–85.

30 B. S. Min, T. D. Cuong, T. M. Hung, B. K. Min, B. S. Shin and M. H. Woo, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 7436–7439.

