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Nickel(II) complexes of *N*-CH₂CF₃ cyclam derivatives as contrast agents for ¹⁹F magnetic resonance imaging†

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Kinetically inert Ni(II) complexes of *N*¹,*N*⁸-bis(2,2,2-trifluoroethyl) cyclams with hydrogen atoms or phosphonic acid groups in the *N*⁴,*N*¹¹-positions show significant ¹⁹F NMR relaxation rate enhancement useful for 19-fluorine MRI imaging.

Magnetic resonance imaging (MRI) is one of the most common techniques in molecular imaging. It is based on the detection of the NMR signal originating from water protons in a tissue. To increase its sensitivity, paramagnetic contrast agents (CAs) are often applied.^{1,2} They affect mainly the longitudinal (*T*₁) relaxation time of the ¹H signal, which leads to an increase in the intensity of the water proton MRI signal. However, essentially all tissues contain water and, thus, the background signal compromises the detection accuracy. This problem can be solved by using non-proton MRI and the ¹⁹F nucleus seems to be the most promising candidate.^{3–6} Natural monoisotopic ¹⁹F has an NMR resonance frequency close to that of ¹H (40.08 MHz T^{–1} for ¹⁹F compared to 42.58 MHz T^{–1} for ¹H) and exhibits sensitivity comparable to ¹H (83%). Fluorine concentration in organisms is virtually zero and, therefore, the lack of background in fluorine-based images enables “hot-spot” imaging. The wider spectral range of the ¹⁹F nucleus (~350 ppm) compared to ¹H (~10 ppm) is also beneficial for some applications. Moreover, only small hardware and software adjustments of standard ¹H scanners are needed for ¹⁹F detection.⁶ This makes the nucleus very potent for *e.g.* cellular tracking of labelled cell cultures.^{7–11}

However, the ¹⁹F nucleus present in organic molecules has usually a very long *T*₁ relaxation time requiring a long delay between excitation pulses; this prolongs the total duration of imaging experiments to unrealistic lengths. Shortening of the *T*₁ relaxation time can result in significant shortening of the experimental time. However, it is necessary to take into account the concomitant shortening of the transversal (*T*₂ or *T*₂^{*}) relaxation time, which leads to signal broadening and can result in very fast loss of signal intensity. It has been shown that the introduction of highly paramagnetic lanthanide(III) ions to the close vicinity of the fluorine atom(s) leads to significant shortening of the relaxation times,¹² and the *T*₂^{*}/*T*₁ ratio is in the range of 0.3–0.9, which is suitable for MRI measurements.^{12c,13}

It is known that, despite the low overall electronic spin (*S* = 1) and magnetic momentum (*μ*_{eff} ~ 3 B.M.) of Ni(II), this ion can induce a large paramagnetic chemical shift and relaxation enhancement comparable to that of lanthanide(III) ions with higher *S* and *μ*.¹⁴ Therefore, some Ni(II) complexes have been studied as MRI agents employable in the Chemical Exchange Saturation Transfer method.¹⁵ Here, we decided to study the ¹⁹F NMR relaxation properties of Ni(II) complexes. The Ni(II) ion fits perfectly in the cavity of 1,4,8,11-tetraazacyclotetradecane (cyclam) and cyclam derivatives are well-known to form Ni(II) complexes with high thermodynamic stability, especially with ligands having coordinating pendant arms enabling octahedral binding to the metal. The 2,2,2-trifluoroethyl side arm was chosen as a group containing a high number of equivalent fluorine atoms. Therefore, ligands **1** and H₄te2p-tfe₂ (Fig. 1)

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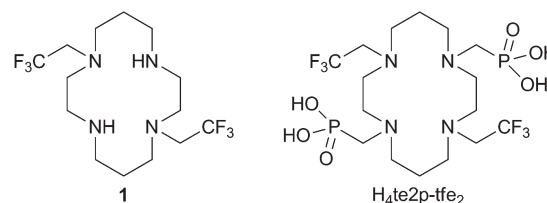


Fig. 1 Ligands studied in this work.

were suggested for testing the ^{19}F NMR parameters of their complexes.

Acylation of 1,8-dibenzylcyclam with ethyl trifluoroacetate or trifluoroacetic anhydride yielded the corresponding bis(amide). It was followed by BH_3 reduction¹⁶ in diglyme at elevated temperature and the benzyl protecting groups were removed by Pd/C hydrogenolysis to obtain ligand **1**. The reaction¹⁷ of amine **1** in neat $\text{P}(\text{OEt})_3$ with CH_2O led to the tetraethyl bis(methylenephosphonate) cyclam derivative. The ethylester groups were removed by transesterification with trimethylsilylbromide¹⁸ followed by the silylester hydrolysis to yield $\text{H}_4\text{te2p-tfe}_2$, which was isolated in a zwitterionic form after ion exchange chromatography. Synthetic details and results of a single-crystal X-ray diffraction study of $1\cdot 2\text{HCl}\cdot 2\text{H}_2\text{O}$ and $\text{H}_4\text{te2p-tfe}_2\cdot 4\text{HBr}\cdot 0.5\text{H}_2\text{O}$ are given in the ESI (Fig. S5 and S6†).

Ligand **1** (in the form of a hydrochloride) reacts with $\text{Ni}(\text{II})$ salts in aqueous solutions to give a light greenish-blue precipitate. The structure of this compound was determined by a single-crystal X-ray study as $\text{cis-}[\text{Ni}(\text{1})(\text{Cl})_2]$ (see ESI Fig. S7†); the central ion is surrounded by four cyclam nitrogen atoms in the *cis-V* configuration¹⁹ with two-fold symmetry ($d_{\text{Ni-N}} = 2.10$ and 2.26 Å for secondary and tertiary amines, respectively) and *cis*-chloride anions coordinated with $d_{\text{Ni-Cl}} = 2.42$ Å.

However, the $\text{cis-}[\text{Ni}(\text{1})(\text{Cl})_2]$ complex shows extremely low solubility in all solvents. Thus, the presence of chloride ions had to be avoided during the preparation of a water-soluble complex. Therefore, ligand **1** in the form of a free base and $\text{Ni}(\text{ClO}_4)_2$ was used for further complex preparation. The course of the reaction in the $\text{H}_2\text{O}:\text{DMSO}$ 1:6.5 mixture at 50°C was followed by ^{19}F NMR spectroscopy (Fig. S2†). Such a solvent mixture was used to keep the reaction mixture fully homogeneous right from the beginning as compound **1** is poorly soluble in water. The reaction proceeds (at 50°C) through an intermediate ($\delta_{\text{F}} = -22.9$ ppm) and is completed during 90 min to give the final complex with $\delta_{\text{F}} = -29.3$ ppm (at 50°C). On cooling to 25°C , the signal shifts to -26.5 ppm. No further ^{19}F NMR spectral changes were observed upon heating the solution at 100°C for several days.

To obtain an aqueous stock solution, prolonged heating (80°C) of the suspension of ligand **1** with $\text{Ni}(\text{ClO}_4)_2$ in $\text{H}_2\text{O}:\text{MeOH}$ 1:1 (with subsequent evaporation of MeOH) was used. It led to the formation of a light blue aqueous solution of a single product with $\delta_{\text{F}} = -26.2$ ppm. When the aqueous solution of the complex prepared in $\text{H}_2\text{O}:\text{MeOH}$ was mixed with the sample prepared in $\text{H}_2\text{O}:\text{DMSO}$, only one symmetric signal in ^{19}F NMR was observed revealing that the species formed in both experiments are identical complexes.

Despite a number of attempts, we were not able to crystallize this light blue product. However, red single-crystals of $\text{trans-}[\text{Ni}(\text{1})](\text{ClO}_4)_2$ (Fig. 2) were obtained when the blue aqueous solution of the complex was saturated with NaClO_4 and was left standing for a few weeks. In this complex, the cyclam ring is coordinated in the centrosymmetric *trans*-III configuration¹⁹ ($d_{\text{Ni-N}} = 1.95$ and 1.99 Å for secondary and ter-

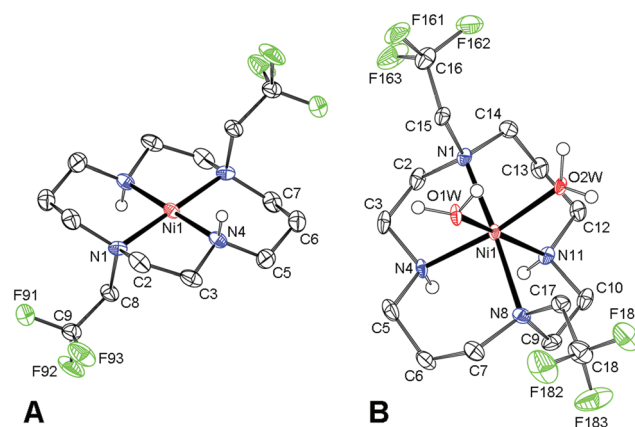


Fig. 2 Structures of (A): $\text{trans-}[\text{Ni}(\text{1})]^{2+}$ and (B): $\text{cis-}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2]^{2+}$ complex cations found in the solid state structures of $\text{trans-}[\text{Ni}(\text{1})](\text{ClO}_4)_2$ and $\text{cis-}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2](\text{TsO})_2$, respectively. Carbon-bound hydrogen atoms are omitted for clarity. Thermal ellipsoids are shown at the 60% probability level.

tiary amino groups, respectively). Consistent with the red colour, only very weak axial interaction with perchlorate anions located in distant positions ($d_{\text{Ni-O}} = 2.83$ Å) was observed.

Dissolution of the red $\text{trans-}[\text{Ni}(\text{1})](\text{ClO}_4)_2$ complex in water produced a light blue solution with $\delta_{\text{F}} = -19.3$ ppm and the species slowly isomerizes with first-order kinetics ($\tau_{1/2} \sim 3.5$ h, 25°C) to a species with $\delta_{\text{F}} = -26.3$ ppm (Fig. S3†). The final species is identical to the original $\text{Ni}(\text{II})$ -**1** complex, as it was confirmed by ^{19}F NMR after the addition of a standard. Taking into account the isomerism of metal ion-cyclam complexes,¹⁹ red $\text{trans-}[\text{Ni}(\text{1})](\text{ClO}_4)_2$ with the *trans*-III configuration probably forms a blue hexacoordinated $\text{trans-}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2]^{2+}$ species upon dissolution, and this complex is rearranged in solution to the $\text{cis-}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2]^{2+}$ species with the *cis*-V cyclam conformation.

Typically, the *trans*-III isomer of cyclam complexes is considered to be the thermodynamically most stable one,²⁰ and the preference of the *cis*-V cyclam conformation for the $\text{Ni}(\text{II})$ -**1** complex is rather surprising. However, the higher stability of the *cis*-V isomer over the *trans*-III one was supported also by the isolation of $\text{cis-}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2](\text{TsO})_2$ (Fig. 2). The structure shows Ni-N distances of 2.07 and 2.09 Å for secondary amino groups, and 2.22 and 2.25 Å for tertiary ones, respectively, with two water molecules coordinated with $d_{\text{Ni-O}} = 2.07$ and 2.10 Å.

Based on the data presented above, one can conclude that the reaction of ligand **1** with $\text{Ni}(\text{ClO}_4)_2$ leads to the formation of the $\text{cis-}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2](\text{ClO}_4)_2$ complex. The geometries of $\text{Ni}(\text{II})$ coordination polyhedra found in the solid state structures are compiled in Table S2† and are comparable to other $[\text{Ni}(\text{L})(\text{H}_2\text{O})_2]$ complexes of cyclam derivatives.^{21,22} The Ni-F distances found in all solid-state structures were in the range of 4.8–5.4 Å (Table S3†).

The complex of the second studied ligand, $\text{H}_4\text{te2p-tfe}_2$, was prepared by heating the ligand together with a slight excess of



NiCl₂ in aq. ammonia (pH 10) at 75 °C for 24 h; the excess of Ni(II) ions was removed by column chromatography. As mentioned above, the course of the reaction was followed by ¹⁹F NMR (Fig. S4†), which showed a fast drop in the concentration of the free ligand ($\delta_F = -68.3$ ppm), and the formation of an intermediate ($\delta_F = -41.1$ ppm) and its slower rearrangement to the final product ($\delta_F = -26.4$ ppm). The time-dependence of intensities of all three signals could be satisfactorily fitted using a monoexponential function (Fig. S4†) and showed comparable rate constants for all three processes (see the ESI†). Such behaviour points to the presence of an equilibrium between the free ligand and the intermediate with an irreversible (rate-determining) reaction step leading to the formation of the final complex.

The final product was isolated in the form of light blue crystals, which were identified as (NH₄){*trans*-[Ni(Hte2p-tfe₂)]}·3.25H₂O by single-crystal X-ray diffraction. Therefore, the complex species present in solution are expected to be *trans*-[Ni(H_{*n*}te2p-tfe₂)]^{*n*-2} (*n* = 0, 1) depending on pH. The molecular structure of the complex anion is shown in Fig. 3, and geometric parameters of the coordination sphere of Ni(II) and Ni–F distances are listed in Tables S2 and S3.† The cyclam ring exhibits the *trans*-III configuration¹⁹ (*d*_{Ni–N} are ~2.10 and 2.11 Å for amino groups bearing the methylenephosphonate pendant arms, and 2.22 and 2.23 Å for those substituted by trifluoroethyl groups) with the oxygen atoms of phosphonate groups occupying apical positions (Ni–O distances are 2.06 and 2.10 Å, respectively). The molecules of the complex are connected *via* short hydrogen bonds between the oxygen atoms of protonated and unprotonated phosphonate pendants (*d*_{O...O} = 2.47 Å), forming infinite chains, similar to what was found for analogous complexes of cyclam-methylenephosphonate derivatives.^{23,24} Bonding distances and the

overall molecular structure are very similar to those of Ni(II) complexes of analogous derivatives.^{22,24}

The thermodynamics of complexing properties of the phosphonate H₄te2p-tfe₂ ligand was studied by potentiometry (see the ESI and Table S4†). The comparison of ligand stepwise protonation constants (log *K*_{1–4} 10.86, 10.09, 5.60 and 4.73) with those of the *N*¹,*N*⁸-dimethyl-*N*⁴,*N*¹¹-bis(methylenephosphonate) analogue²⁵ (log *K*_{1–4} 11.47, 12.17, 7.20 and 6.33, Table S5†) points to significantly decreased ligand basicity caused by the presence of electron-withdrawing –CH₂CF₃ groups. Surprisingly, it affects not only the first two protonation constants corresponding to the ring amino groups but also those of the phosphonate moieties, probably as a result of a strong electron-withdrawing effect transferred through intramolecular hydrogen bonds, which are expected to have a geometry analogous to that found for related cyclam derivatives.²⁵ Equilibration of the Ni(II)–H₄te2p-tfe₂ system is relatively slow and, therefore, the out-of-cell titration method had to be used. As the complexation mechanism is not fully straightforward (see above), the samples used for the out-of-cell titration were heated at 50 °C for 2 weeks to ensure quantitative rearrangement of the intermediate to the final *trans* isomer. The time required for equilibration was checked by ¹⁹F NMR of separate samples. The stability constant, log *K*_{NiL} = 13.28, is about 2–7 orders of magnitude lower than the constants of complexes with related ligands (Table S5†),^{26,27} mainly as a consequence of the lower ligand basicity. The distribution diagram of the system (Fig. S8†) shows that full Ni(II) complexation by H₄te2p-tfe₂ is completed at pH 7 and the complex is present at this pH almost entirely in a fully deprotonated form.

For possible *in vitro/in vivo* utilization, kinetic inertness is a more important parameter than thermodynamic stability. Kinetic inertness is often tested in acidic solutions as acid-assisted complex dissociation. Thus, the decomposition of both studied complexes was examined in 1 M aq. HCl at 37 and 80 °C. Both complexes are decomposed relatively slowly by HCl at 37 °C ($\tau_{1/2}$ ~8 and ~10 h for *cis*-[Ni(1)(H₂O)₂]²⁺ and *trans*-[Ni(H_{*n*}te2p-tfe₂)]^{*n*-2}, respectively) but the decomplexation of the *cis*-[Ni(1)(H₂O)₂]²⁺ complex is substantially accelerated at the higher temperature (80 °C, $\tau_{1/2}$ ~3 min compared to ~5 h for *trans*-[Ni(H_{*n*}te2p-tfe₂)]^{*n*-2}, Table S6†), as the presence of apically coordinated pendant arms in the H₄te2p-tfe₂ complex enhances kinetic inertness. High inertness has been observed for several Cu(II) complexes with analogous cyclam-based ligands,²⁸ and highly protonated species of several Ni(II)^{22,24} complexes of phosphonated cyclam derivatives have been isolated even in the solid state. These results suggest sufficient complex stability under physiological conditions and warrant the possible use of the Ni(II)–H₄te2p-tfe₂ complex in *in vitro/in vivo* applications.

As the *trans*-[Ni(te2p-tfe₂)]²⁻ complex is kinetically inert and promises reasonable stability *in vivo*, its ¹⁹F MRI-related parameters were investigated. Although the *cis*-[Ni(1)(H₂O)₂]²⁺ complex is not suitable for any *in vivo* application due to its low solubility in chloride-containing media, it was studied as well for comparative purposes as there are no related data

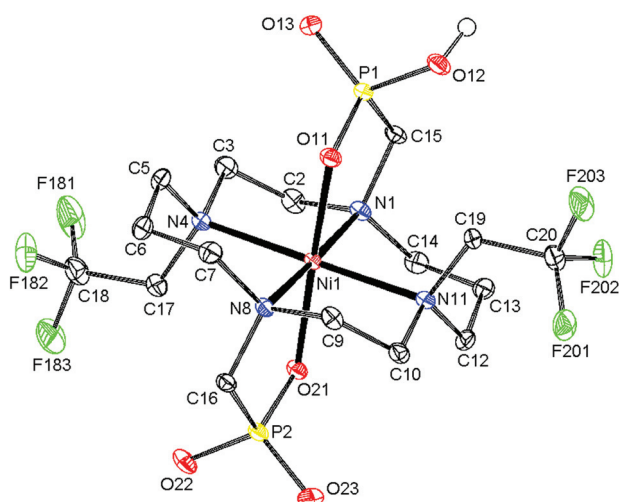


Fig. 3 Molecular structure of the *trans*-[Ni(Hte2p-tfe₂)]⁻ anion found in the crystal structure of *trans*-(NH₄)[Ni(Hte2p-tfe₂)]·3.25H₂O. Carbon-bound hydrogen atoms are omitted for clarity. Thermal ellipsoids are shown at the 60% probability level.



reported in the literature at all. The ^{19}F NMR relaxation measurement of both Ni(II) complexes at $B_0 = 7.05$ T showed extreme shortening of ^{19}F NMR T_1 relaxation times by 2–3 orders of magnitude compared with the values observed for the free ligands, **1** and $\text{H}_4\text{te2p-tfe}_2$ (Table 1).

The suitability of the $\text{trans}[\text{Ni}(\text{te2p-tfe}_2)]^{2-}$ complex for ^{19}F MRI was tested by phantom visualization at $B = 4.7$ T. At this magnetic field, the T_1 of the complex is very short in the millisecond range with a still convenient T_2^*/T_1 relaxation times ratio (Table 1). The observed relaxation times are even slightly shorter than those reported for studied Ln(III) complexes – in the cases of highly paramagnetic Tb(III), Dy(III) and Ho(III) complexes with the estimated Ln(III)–F distance lying in the range of 5–7 Å, the reported T_1 is typically in the range of 7–11 ms at 4.7 T and room temperature.^{12c,13} The very short relaxation time of the Ni(II) complex required optimization of the fast pulse sequence – for the visualization of the complex, a fast gradient echo sequence with TE = 1.3 ms and TR = 3 ms was used. The slowly relaxing samples (containing the free ligand and trifluoroethanol used as a standard) were best measured using a long turbospin echo sequence employing TE = 40 ms and TR = 2000 ms. For localization of the samples, the ^1H MRI scan (Fig. 4A) was also acquired. Fig. 4 shows the results of the MRI visualization. The brightness of aq. solution of the Ni(II) complex compared to aq. solutions of the free ligand and trifluoroethanol is caused by its paramagnetism, which shortens the T_1 relaxation time of water protons ($r_1(\text{complex}) = 0.12 \text{ mm}^{-1} \text{ s}^{-1}$, 4.7 T, 25 °C). As each sample has a different ^{19}F NMR chemical shift (δ_{F} –26 ppm, –68 ppm and –77 ppm for the complex, free ligand and trifluoroethanol, respectively), each signal can be excited separately. In the case of the fast sequence, only a negligible signal of the free ligand was detected, as virtually no diamagnetic sample relaxation occurs during the sequence time-scale. On the contrary, in the experiment employing the long sequence, no signal of the paramagnetic sample was found as its magnetization relaxes before the start of acquisition.

The samples of free ligand **1** and $\text{cis}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2](\text{ClO}_4)_2$ show fully concordant behaviour (Fig. S9†).

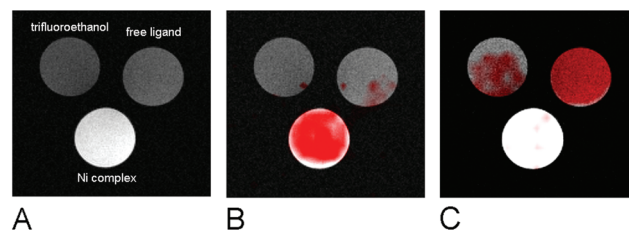


Fig. 4 MRI study of phantoms containing trifluoroethanol, free $\text{H}_4\text{te2p-tfe}_2$ and the $\text{trans}[\text{Ni}(\text{te2p-tfe}_2)]^{2-}$ complex ($c_{\text{F}} = 0.004$ M in all samples), $B = 4.7$ T, 25 °C, home-made $^1\text{H}/^{19}\text{F}$ surface single loop coil. (A) ^1H MRI scan, gradient echo sequence, flip angle 30°, TE = 3.7 ms, TR = 100 ms, matrix 256×256 . (B) Overlay of ^1H MRI with ^{19}F MRI; ^{19}F MRI was optimized for the complex; acquired at $\delta = -26$ ppm, gradient echo sequence, TE = 1.3 ms, TR = 3 ms, matrix 32×32 interpolated to 256×256 . (C) Overlay of ^1H MRI with ^{19}F MRI; ^{19}F MRI was optimized for the ligand; acquired at $\delta = -77$ ppm, turbospin echo sequence, TE = 40 ms, TR = 2000 ms, matrix 32×32 interpolated to 256×256 .

In conclusion, transition metal ion complexes of fluorine-containing ligands can be considered a new class of ^{19}F MRI contrast agents, as shown in the case of Ni(II). The presence of strongly complexing and electron donating phosphonates enhances the kinetic inertness of the studied complexes and compensates the disadvantageous coordination properties of fluorine-containing ligands. Relaxation parameters of the $\text{trans}[\text{Ni}(\text{te2p-tfe}_2)]^{2-}$ complex with fluorine atoms located about 5 Å from the Ni(II) centre are highly suitable for ^{19}F MRI hot-spot imaging employing fast pulse sequences. As the Ni(II) complexes with coordinated water molecules exhibit useful water proton T_1 -relaxivity, properly designed compounds could be potentially used as dual $^1\text{H}/^{19}\text{F}$ MRI contrast agents.²⁹

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Table 1 ^{19}F NMR relaxation times^a and ^1H relaxivity of the studied compounds (pH 7, 25 °C)

Parameter	1	$\text{cis}[\text{Ni}(\text{1})(\text{H}_2\text{O})_2]^{2+}$	$\text{H}_4\text{te2p-tfe}_2$	$\text{trans}[\text{Ni}(\text{te2p-tfe}_2)]^{2-}$
$B_0 = 7.05$ T (300 MHz for ^1H , 282 MHz for ^{19}F)				
$T_1(^{19}\text{F})$	0.8(3) s	1.72(1) ms	0.5(1) s	2.8(7) ms
$T_2^*(^{19}\text{F})$	≈76 ms	≈0.82 ms	≈50 ms	≈0.90 ms
$T_2^*/T_1(^{19}\text{F})$	0.1	0.48	0.1	0.32
$r_1(^1\text{H})$	—	0.83(3)	—	0.18(1)
$B_0 = 4.70$ T (200 MHz for ^1H , 188 MHz for ^{19}F)				
$T_1(^{19}\text{F})$	0.82(1) s	1.2(1) ms	1.1(2) s	4.2(1.1) ms
$T_2^*(^{19}\text{F})$	3.1(1) ms	0.62(1) ms	3.1(2) ms	1.1(1) ms
T_2^*/T_1	0.0038	0.52	0.0028	0.26
$r_1(^1\text{H})$	—	0.66(4) s ^{−1} mM ^{−1}	—	0.12(2) s ^{−1} mM ^{−1}

^a T_1 was determined using inversion recovery pulse sequence; T_2^* was determined from line-width using Lorentzian-shape fitting of the signal.



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