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Systems biocatalysis: *para*-alkenylation of unprotected phenols†

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Commercially available phenol derivatives were transformed with pyruvate to form a new C–C bond leading to the corresponding *para*-coumaric acids and only one molecule of water as an innocent side product in buffer. The reaction was catalysed by a biocatalytic system consisting of two enzymatic steps, which were run simultaneously: (i) in the first step catalysed by a tyrosine phenol lyase the C–C coupling of phenol derivatives with pyruvate and ammonia yielded L-tyrosine derivatives, (ii) which were transformed in the second step to the final product *via* ammonia elimination catalysed by a tyrosine ammonia lyase. The reactions proceeded with exquisite regio- and stereoselectivity yielding just the *para*-products with the (*E*)-configuration. The method represents an efficient approach for the direct alkenylation of phenols on a preparative scale (up to 0.6 mmol) affording (*E*)-*para*-coumaric acids in excellent isolated yields without requiring chromatographic purification. Co-expression of the involved enzymes in a single host gave access to single catalyst preparation.

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1. Introduction

α,β -unsaturated carboxylic acids are central compounds in organic chemistry which are not only widely present in a broad range of biologically active molecules but also exhibit outstanding potential¹ to be used as building blocks for organic synthesis.² Among them, coumaric acid represents a privileged scaffold being part of a broad range of pharmacologically active compounds widely distributed in the plant kingdom. For instance, caffeic acid and specially its phenethyl ester have demonstrated “*in vitro*” activities such as anti-inflammatory and anti-tumour effects or provided protection against UV light.³ Furthermore, *p*-coumaric acid, the most abundant isomer of coumaric acid, is known to reduce the risk of stomach cancer by reducing the formation of carcinogenic nitrosamines.⁴ Related compounds such as ferulic acid have shown remarkable activities against breast and liver cancers.⁵ Additionally, functionalized derivatives such as chlorogenic acid and cichoric acid have been shown to slow the release of glucose to the bloodstream and inhibit the HIV integrase, respectively.⁶

Traditional methods to prepare *p*-coumaric acid derivatives are generally based on Knoevenagel⁷ or Heck⁸ reactions, thus requiring a functional group (aldehyde or halogen, respectively) in the *para*-position to the phenolic hydroxyl. On the other hand, biocatalytic approaches are limited to the carboxylation of *para*-vinylphenol derivatives using carboxylases,⁹ which require the corresponding *p*-hydroxy styrene precursors to be prepared using Wittig reactions of the corresponding aldehydes.¹⁰ Direct C–H alkenylations of phenols have been reported with (i) Ru-catalysts,¹¹ (ii) Pd-catalysis to access *ortho*-coumaric acid derivatives,¹² and (iii) Fe-containing mesoporous aluminosilicates.¹³ All direct alkenylation methods lead to *ortho* products.

2. Results and discussion

We have recently reported a biocatalytic system for the *para*-vinylation of phenols (Scheme 1a).¹⁴ In the present work, the biocatalytic system was adapted for the preparation of *p*-coumaric acid derivatives from unsubstituted phenols using only pyruvate as a stoichiometric reagent (Scheme 1b), which represents a *para*-selective alkenylation. This contribution represents the first example of direct *para*-selective alkenylation, without requiring an auxiliary group at the *p*-position (Br, CHO, *etc.*) with respect to the phenolic OH moiety.

The cascade¹⁵ investigated was designed by combining two enzymatic reaction steps. In the first step of the cascade, the C–C coupling between pyruvate, ammonia and different phenol derivatives **1** should give access to substituted L-tyrosine derivatives **3** (Scheme 2).¹⁶ Subsequent elimination

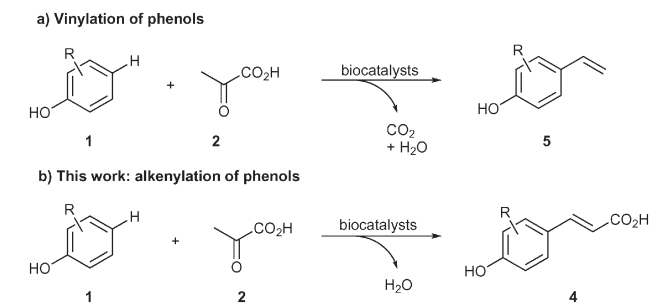
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Scheme 1 a) Vinylation of phenols. b) C–H *para*-alkenylation of phenols leading to *p*-coumaric acid derivatives.

of ammonia by a tyrosine ammonia lyase (TAL)¹⁷ should give the (*E*)-*p*-coumaric acids **4** in the second step. The overall reaction requires only pyruvate and phenols as stoichiometric reagents since ammonia is internally recycled. For the first step the M379V variant of the recombinant tyrosine phenol lyase from *Citrobacter freundii* was used as a cell-free extract due to its broad scope towards non-natural substrates.¹⁶ For the elimination step the recombinant TAL from *Rhodobacter sphaeroides* gave the best results when employed as an *E. coli* whole cell catalyst.¹⁴

To identify the optimal pH value for the overall biotransformation, the biocatalytic alkenylation of phenol **1a** was studied at pH values between pH 8–10 (Fig. 1) since the TPL as well as the TAL gave the highest conversions at alkaline pHs.¹⁴ At pH 8 the formation of (*E*)-**4a** was observed albeit at a low rate due to the accumulation of *L*-**3a** in the reaction (see the ESI† for a detailed profile). This is consistent with the lower activity of the TAL. The highest concentration of (*E*)-**4a** was obtained at pH 10 where TPL and TAL also gave the highest conversions in the individual reactions. This is in contrast to the vinylation cascade (Scheme 1A),¹⁴ where pH 8 turned out to be optimum due to a compromise with the last additional step, the decarboxylation, which proceeded best at pH 6.

Monitoring the alkenylation of **1a** over time at pH 10 and 30 °C revealed that under the conditions employed the two steps were well coupled since the concentration of *L*-**3a** never exceeded 15% of the total amount of products (Fig. 2). Accumulation of **3a** would indicate that elimination is the rate-

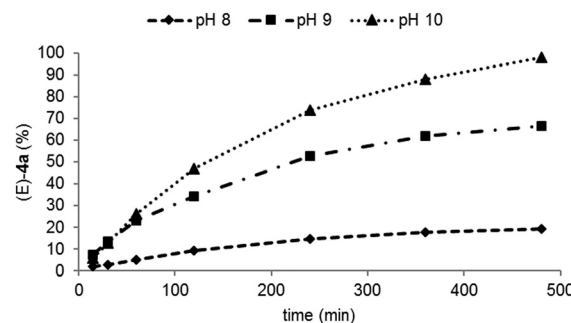
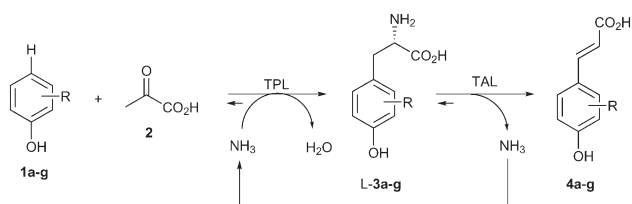


Fig. 1 Conversion of **1a** to *p*-coumaric acid (*E*)-**4a** at different pH values. Reaction conditions: **1a** (23 mM), pyruvate (46 mM), NH₄Cl (180 mM), TPL M379V (5 mg cell-free extract, 1.5 U), TAL (20 mg freeze-dried *E. coli*/TAL, 0.18 U), buffer (50 mM, potassium phosphate, pH 8, CHES, pH 9–10), PLP (0.04 mM), Et₂O (5% v/v), 30 °C, 850 rpm.

limiting step of the cascade. The formation of (*E*)-**4a** followed a linear profile for the first two hours, then slowed down reaching completion after 24 h when the starting materials and intermediates were below the detection limit. Although all reactions are reversible, the high conversion resulted from the thermodynamic preference for the formation of (*E*)-**4a** (−24.0 kcal mol^{−1} for the transformation of the *L*-tyrosine binding state in the enzyme to (*E*)-coumarate + H₂N-MIO).^{17a}

Next, the scope of the cascade was investigated using a set of 2- and 3-substituted phenols. Phenols bearing halogens (fluoro, chloro and bromo, **1a–c**) in the 2-position were efficiently transformed with >99% conversion and perfect selectivity towards the *E*-isomer (Table 1, entries 1–4). 2-Methylphenol (**1d**) was also transformed achieving in this case good conversion of the alkenylated product (entry 5). Higher amounts of (*E*)-**4d** were formed at longer reaction times (48 h, entry 6).

The bioalkenylation protocol was also successfully applied for 3-substituted phenols (entries 7–8), observing quantitative conversion of 3-fluorophenol (**1e**) and high conversion of 3-chlorophenol (**1f**). Interestingly, a 2,3-disubstituted phenol (**1g**) was also efficiently transformed into the corresponding *p*-hydroxycinnamic acid (entry 9). However, phenols bearing



1: a) R = 2-F; b) R = 2-Cl; c) R = 2-Br; d) R = 2-Me; e) R = 3-F; f) R = 3-Cl; g) R = 2,3-diF

Scheme 2 Two-step cascade for the biocatalytic alkenylation of phenols. TAL: recombinant tyrosine ammonia lyase from *Rhodobacter sphaeroides* employed as an *E. coli* whole cell catalyst; TPL: recombinant tyrosine phenol lyase from *Citrobacter freundii* M379V used as a cell-free extract.

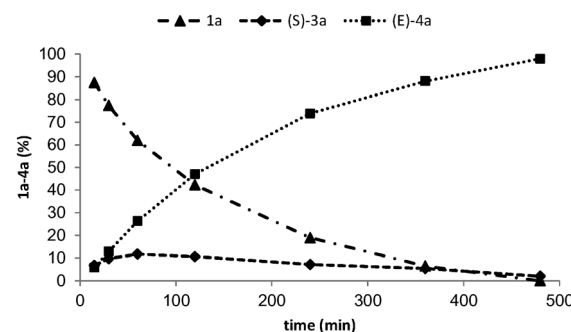
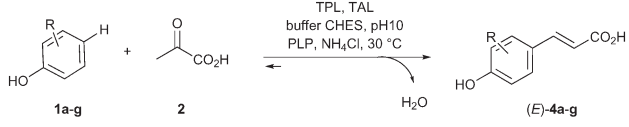


Fig. 2 Time course for the alkenylation of 2-fluorophenol (**1a**) into 3-fluoro-4-hydroxycinnamic acid (*E*)-**4a**. Reaction conditions: **1a** (23 mM), pyruvate (46 mM), NH₄Cl (180 mM), buffer (50 mM, CHES, pH 10), PLP (0.04 mM), TPL M379V (5 mg cell-free extract, 1.5 U in 1 mL total volume), TAL (20 mg freeze-dried *E. coli*/TAL, 0.18 U in 1 mL total volume), Et₂O (5% v/v), 30 °C, 850 rpm.



Table 1 Enzymatic alkenylation of phenols to yield *p*-hydroxycinnamic acids **4a–g**^a


1a-g + 2 $\xrightarrow[\text{PLP, NH}_4\text{Cl, 30 } ^\circ\text{C}]{\text{TPL, TAL, buffer CHES, pH10}}$ (E)-4a-g + H₂O

1: a) R = 2-F; b) R = 2-Cl; c) R = 2-Br; d) 2-Me; e) R = 3-F; f) R = 3-Cl; g) R = 2,3-diF

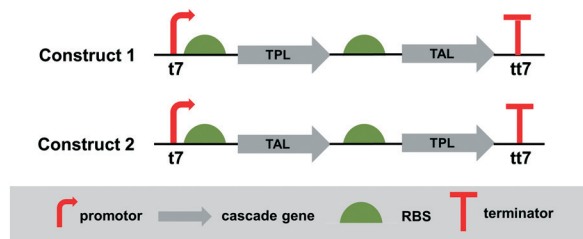
Entry	1a–g	1 [mM]	1 [%] ^b	3 [%] ^b	4 [%] ^{b,c}
1	a	23	<1	<1	>95
2	a ^e	46	<1	<1	>95(96)
3	b	23	<1	<1	>95(85)
4	c	23	<1	<1	>95(96)
5	d	23	10	20	71 ^d
6	d ^f	23	2	15	83 ^d
7	e	23	<1	<1	>95(88)
8	f	23	26	5	69 ^d
9	g	23	<1	<1	>95(80)

^a Reaction conditions: phenol **1**, pyruvate (2 equiv.), NH₄Cl (180 mM), TPL (50 mg cell-free extract, 15 U), TAL (200 mg freeze-dried *E. coli*/TAL, 1.8 U), CHES buffer (pH 10, 10 mL, 50 mM), PLP (0.04 mM), Et₂O (5% v/v), 30 °C, 120 rpm, 24 h. ^b Determined by reverse-phase HPLC analysis. ^c Isolated yields are shown in brackets. ^d Reactions performed on an analytical scale only. ^e Double amounts of the two biocatalysts were used. ^f Reaction time: 48 h.

polar *ortho* groups such as in catechol or *o*-aminophenol were not transformed at all.

Subsequently, *para*-alkenylation was successfully shown on a semi-preparative scale (0.6 mmol, 46 mM substrate concentration), isolating for instance the fluorinated derivative (E)-4a as a single isomer in 96% isolated yield after a simple extraction protocol without requiring chromatographic purification (entry 2). Semi-preparative biotransformations were also performed for 2-substituted phenols (**1b–c**), the 3-substituted phenol (**1e**) as well as for the 2,3-substituted phenol (**1g**), demonstrating the potential of the present methodology for the preparation of the corresponding alkenylated products in excellent isolated yields (80–96%) and complete stereoselectivity without requiring chromatographic purification steps.

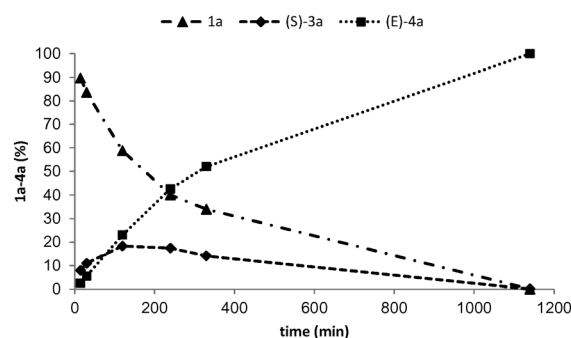
Since the cascade requires two enzymes, the next step was to co-express both enzymes in a single host, which simplifies enzyme production and handling of the catalysts. Co-expression of enzymes in a single cell enables the generation of efficient whole-cell catalysts.¹⁸ To identify the most suitable construct for the expression of the two enzymes in *E. coli*, two bicistronic constructs were designed, wherein in one case the TPL was first in line followed by TAL (construct 1), while in the other case the order is reversed, thus the TAL enzyme comes first (construct 2) (Fig. 3). The TPL and TAL genes were assembled under the control of a strong inducible promoter (T7/*lacO*). The translation of TAL was driven by the RBS of pET28a(+) and the gene carried an N-terminal hexahistidine-tag. TPL was driven by the RBS of the pET22b(+) vector. The constructs were generated using Gibson isothermal assembly.

**Fig. 3** Design of constructs for co-expression of TPL and TAL. t7: IPTG inducible T7/*lacO* promoter; tt7: T7 terminator.

Initial rates for the overall cascade were measured for freeze-dried cells of *E. coli* containing the overexpressed enzymes encoded by constructs 1–2. A higher activity was observed for the transformation of **1a** to (E)-4a with *E. coli*(TPL–TAL) (1.2 mU mg^{−1}) compared to *E. coli*(TAL–TPL) (0.23 mU mg^{−1}). The TAL activities for the single step transformation of **3a** into **4a** were 1.35 mU mg^{−1} for *E. coli*(TPL–TAL) and 0.31 mU mg^{−1} for *E. coli*(TAL–TPL). Consequently, the transformation of **1a** into (E)-4a was monitored over time for the best performing catalyst, that is, *E. coli*(TPL–TAL) (Fig. 4). The reaction reached completion after about 19 hours. This may indicate a positive influence of the co-expression in a single cell, because the TAL activity of the *E. coli*(TPL–TAL) catalyst was limiting (intermediate **3a** reached up to 23%). However, only 22% of the units used in the experiment were employed for the profile in Fig. 2 and the reaction only took roughly double the amount of time. Nevertheless, the expression level may be improved for efficient catalyst preparation.

3. Conclusions

In conclusion, phenols have been efficiently *para*-alkenyated using a biocatalytic system involving two steps, namely C–C bond formation¹⁹ and NH₃-elimination. Only pyruvate is required as the stoichiometric reagent. Since only one molecule of water is released in the transformation, the overall

**Fig. 4** Time course for the alkenylation of 2-fluorophenol (**1a**) into 3-fluoro-4-hydroxycinnamic acid (E)-4a employing *E. coli*(TPL–TAL). Reaction conditions: total volume: 1 mL, **1a** (23 mM), pyruvate (46 mM), NH₄Cl (180 mM), buffer (50 mM, CHES, pH 10), PLP (0.04 mM), *E. coli*(TPL–TAL) (30 mg freeze-dried cells, 36 mU), Et₂O (5% v/v), 30 °C, 850 rpm.

reaction possesses an excellent atom economy (82% for (*E*)-4a-e, 83% for (*E*)-4b, 86% for (*E*)-4c and 83% for (*E*)-4g). *p*-Alkenylation was successfully performed on a semi-preparative scale at substrate concentrations of 23–46 mM obtaining the alkenylated *p*-coumaric acid products as a single (*E*)-isomer in excellent isolated yields (80–96%) without requiring chromatographic purification. By co-expression of the two enzymes in a single host, only a single catalyst preparation may be required for the reaction.

4. Experimental section

4.1 General

Chemical reagents were purchased from different commercial sources and used without further purification. Melting points were measured on samples in an open capillary tube and were uncorrected. ^1H and ^{13}C -NMR spectra were obtained using a Bruker spectrometer (^1H , 300.13 MHz; ^{13}C 75.5 MHz). The chemical shifts are given in delta (δ) and the coupling constants in Hertz (Hz). Mass spectra experiments (MS) were carried out by ESI⁺ using an Agilent HPLC coupled to a MS.

4.2 Single enzyme preparations

Single biocatalysts were prepared as previously described.¹⁴ Tyrosine phenol lyase M379V (TPL) from *Citrobacter freundii* (0.30 U mg⁻¹) was used as a freeze-dried cell-free extract and prepared as previously described.¹⁴ Tyrosine ammonia lyase (TAL) from *Rhodobacter sphaeroides* (9.2 mU mg⁻¹) was used as a whole-cell freeze-dried catalyst.¹⁴

4.3 Co-expression of TPL and TAL

The coding sequences for tyrosine phenol lyase (TPL) from *Citrobacter freundii* M379V and tyrosine ammonia lyase (TAL) from *Rhodobacter sphaeroides* were PCR amplified from pET22b(+)-TPL and pET28a(+)-TAL¹⁴ using the following primers: TPL with pBP856 (5'-gataacaattccggaattgtgagcggataa-caattccggaattcaaggagatatacatatgatgaactatccggc-3') and pBP857 (5'-gctgcccattgtatatctcttatttaaattattagatataatcgaagcgcgcgg-taaaaaac-3') for construct 1; with pBP939 (5'-gctgcaaca gactcgggttaagctagcaaggagatatacatatgatgaactatccggc-3') and pBP943 (5'-gggggttatgctaggatcgatccttattagatata atcgaagcgcgcg-gtaaaaaaac-3') for construct 2. TAL was amplified with pBP858 (5'-cgcgctctcgattatat ctaataaatttaaataaggagatataaccatg-ggcagcagcc-3') and pBP866 (5'-caagggttatgctaggatcgatccttaa-accggactctgttcagcaga tg-3') for construct 1; and with pBP937 (5'-gataacaattccggaattgtgagcggataaacaattccggaattcaaggagat-ataccatgggcagcagccatc-3') and pBP938 (5'-atagttcatcatatgtatat-ctcttgctagcttaaacggactctgttcagcagatg-3') for construct 2. The PCR fragments included the RBSs of pET28a(+) (TAL) and pET22b(+) (TPL) and TAL carried an N-terminal hexahistidine-tag.¹⁴ pCAS1 (ref. 18c) was linearized with EcoRI and BamHI and dephosphorylated using alkaline phosphatase (Thermo Fisher Scientific, Waltham, MA). All PCR and DNA fragments were gel purified (Promega, Fitchburg, WI) before the TPL and TAL genes were inserted into pCAS1

by Gibson isothermal assembly.²⁰ The resulting constructs 1 and 2 were sequence verified.

For the co-expression of the two cascade enzymes, *E. coli* BL21-Gold(DE3) (Agilent Technologies, Santa Clara, CA) was transformed with constructs 1 and 2. The cells were grown in LB medium (Carl Roth GmbH, Karlsruhe, Germany) until they reached a density of OD₆₀₀ ~ 0.6–0.8. The expression of the cascade enzymes was induced with 1 mM IPTG (Sigma-Aldrich, St. Louis, MO) and the expression was performed at 28 °C overnight. The cells were then harvested by low speed centrifugation (2800 × *g*, 4 °C, 20 min), washed once in PBS (137 mM NaCl, 2.7 mM KCl, 8 mM Na₂HPO₄, 2 mM KH₂PO₄) and the pellets were stored at -20 °C until lyophilisation in 15 mL of PBS overnight.

4.4 Overall activities for *E. coli*(TPL-TAL) and *E. coli*(TAL-TPL)

For better comparison of constructs 1 and 2, the activity for the transformation of the corresponding phenol 1a into the cinnamic acid derivative (*E*)-4a was determined by measuring the initial rate by HPLC. One unit of activity was defined as the amount of catalyst prepared that catalysed the formation of 1 μmol of 3-fluoro-4-hydroxycinnamic acid (4a) per minute under the following conditions: 30 °C, 850 rpm, pH 10. The assay mixture contained 2-fluorophenol (1a, 23 mM), pyruvate (46 mM), NH₄Cl (180 mM) and *E. coli* whole cells overexpressing TPL and TAL (30 mg). The reaction activities were determined corresponding to 1.2 mU mg⁻¹ *E. coli* whole cells for construct 1 and 0.23 mU mg⁻¹ *E. coli* whole cells for construct 2.

4.5 Biotransformations on analytical scale

Recombinant TPL M379V from *C. freundii* (1.5 U, 5 mg freeze-dried cell-free extract) and TAL from *R. sphaeroides* (0.18 U, 20 mg freeze-dried *E. coli*/TAL cells) were rehydrated in a CHES buffer (50 mM, 180 mM NH₄Cl, 0.04 mM PLP, pH 1, 1 mL) for 10 min at 30 °C and 120 rpm. The corresponding phenols 1a–g (0.023 mmol, 23 mM) and pyruvate (0.046 mmol, 46 mM; 11.5 μL of 4 M stock solution in CHES buffer pH 10) were added to the mixture and the reaction was incubated at 21 °C and 800 rpm for 24 h. Reactions were quenched with a MeCN/H₂O 1:1 solution containing 0.1% TFA (1 mL), centrifuged to remove the cells and the supernatant was injected in the HPLC system for conversion measurement.

4.6 Biotransformations on preparative scale

Recombinant TPL M379V from *C. freundii* (15 U, 50 mg freeze-dried cell-free extract) and TAL from *R. sphaeroides* (3.68 U, 400 mg freeze-dried *E. coli*/TAL cells) were rehydrated in a CHES buffer (50 mM, 180 mM NH₄Cl, 0.04 mM PLP, pH 10, 10 mL) for 10 min at 30 °C and 120 rpm. Then the phenol (0.23 mmol, 23 mM) and pyruvate (101.2 mg, 0.46 mmol, 46 mM) were added to the mixture and the reaction was incubated for 24 h at 21 °C and 170 rpm. The reaction was stopped with an aqueous saturated NH₄Cl solution (5 mL), acidified with 2 M HCl (1 mL) and extracted with EtOAc (3 ×



15 mL). The combined organic phases were dried (Na_2SO_4) and the solvent was evaporated under reduced pressure. The crude product was washed with *n*-heptane (3×5 mL) obtaining the corresponding *p*-hydroxycinnamic acids as white solids (80–96%).

4.7 Reaction scale and isolated yields

1a: (51 mg, 0.46 mmol, 46 mM); isolated yield: (*E*)-**4a**: 80.6 mg, 96%. **1b:** (30 mg, 0.23 mmol, 23 mM); isolated yield: (*E*)-**4b**: (39 mg, 85%). **1c:** (40 mg, 0.23 mmol, 23 mM); isolated yield: (*E*)-**4c**: (54 mg, 96%). **1e:** (26 mg, 0.23 mmol, 23 mM); isolated yield: (*E*)-**4e**: (37 mg, 88%). **1g:** (30 mg, 0.23 mmol, 0.23 mM); (*E*)-**4g**: isolated yield: (36 mg, 80%).

(*E*)-3-(3-Fluoro-4-hydroxyphenyl)acrylic acid (**4a**). White solid. Mp: 215 (dec.). ^1H NMR (CD_3OD , 300.13 MHz): δ 6.33 (d, $^3J_{\text{HH}} = 17.5$ Hz, 1H), 6.94 (at, $J = 8.7$ Hz, 1H), 7.23–7.27 (m, 1H), 7.37 (dd, $^3J_{\text{HH}} = 12.2$ Hz, $^3J_{\text{HH}} = 2.2$ Hz, 1H), 7.57 (d, $^3J_{\text{HH}} = 17.5$ Hz, 1H). ^{13}C NMR (CDCl_3 , 75.5 MHz): δ 116.6, 116.8, 127.0, 128.5, 145.9, 149.1, 151.9, 155.1, 171.0. ^{19}F NMR (CD_3OD , 282 MHz): δ -140.0. MS (ESI^+): 183 [(M + H) $^+$, 100%].

(*E*)-3-(3-Chloro-4-hydroxyphenyl)acrylic acid (**4b**). White solid. Mp: 220 °C (dec.). ^1H NMR (CD_3OD , 300.13 MHz): δ 6.33 (d, $^3J_{\text{HH}} = 15.0$ Hz, 1H), 6.94 (d, $^3J_{\text{HH}} = 6.0$ Hz, 1H), 7.40 (dd, $^3J_{\text{HH}} = 8.2$ Hz, $^3J_{\text{HH}} = 2.1$ Hz, 1H), 7.54–7.60 (m, 2H). ^{13}C NMR (CDCl_3 , 75.5 MHz): δ 117.5, 118.0, 122.5, 128.7, 129.9, 131.1, 145.3, 156.7, 170.8. MS (ESI^+): 199 [($^{35}\text{ClM} + \text{H}$) $^+$, 100%], 199 [($^{35}\text{ClM} + \text{H}$) $^+$, 100%].

(*E*)-3-(3-Bromo-4-hydroxyphenyl)acrylic acid (**4c**). White solid. Mp: 225 (dec.). ^1H NMR (CD_3OD , 300.13 MHz): δ 6.33 (d, $^3J_{\text{HH}} = 14.8$ Hz, 1H), 6.92 (d, $^3J_{\text{HH}} = 8.2$ Hz, 1H), 7.45 (dd, $^3J_{\text{HH}} = 8.0$ Hz, $^3J_{\text{HH}} = 2.0$ Hz, 1H), 7.55 (d, 1H, $^3J_{\text{HH}} = 14.8$ Hz), 7.74 (d, 1H, $^4J_{\text{HH}} = 2.1$ Hz). ^{13}C NMR (CDCl_3 , 75.5 MHz): δ 111.9, 117.7, 129.3, 130.3, 134.7, 145.5, 158.1, 171.0. MS (ESI^+) 243 [($^{79}\text{BrM} + \text{H}$) $^+$, 100%], 245 [($^{81}\text{BrM} + \text{H}$) $^+$, 100%].

(*E*)-3-(2-Fluoro-4-hydroxyphenyl)acrylic acid (**4e**). White solid. Mp: 210 °C (dec.). ^1H NMR (CD_3OD , 300.13 MHz): δ 6.38 (d, $^3J_{\text{HH}} = 17.5$ Hz, 1H), 6.54–6.59 (m, 1H), 6.64–6.68 (m, 1H), 7.51 (at, $J = 8.7$ Hz, 1H), 7.72 (d, $^3J_{\text{HH}} = 17.5$ Hz, 1H). ^{13}C NMR (CDCl_3 , 75.5 MHz): δ 104.3, 113.8, 115.2, 118.4, 132.7, 139.4, 162.7, 164.1, 171.2. ^{19}F NMR (CD_3OD , 282 MHz): δ -115.6. MS (ESI^+): 183 [(M + H) $^+$, 100%].

(*E*)-3-(2,3-Difluoro-4-hydroxyphenyl)acrylic acid (**4g**). White solid. Mp: 225 °C (dec.). ^1H NMR (CD_3OD , 300.13 MHz): δ 6.34 (d, $^3J_{\text{HH}} = 18.0$ Hz, 1H), 6.77–6.81 (m, 1H), 7.28–7.33 (m, 1H), 7.67 (d, $^3J = 17.3$ Hz, 1H). ^{13}C NMR (CDCl_3 , 75.5 MHz): δ 114.8, 116.5, 120.1, 125.0, 125.1, 138.4, 141.8, 151.5, 170.7. ^{19}F NMR (CD_3OD , 282 MHz): δ -142.3, -164.3. MS (ESI^+): 183 [(M + H) $^+$, 100%].

4.8 Analytics

Conversions for substrates were determined with an Agilent chromatograph UV detector at different wavelengths using a Luna C18 column (25 cm \times 4.6 mm I.D.). *Sample preparation*: a MeCN/ H_2O solution (1 mL, 1 : 1) containing 0.1% TFA was added to an aliquot of the reaction solution (1 mL). The

protein was removed by centrifugation and the solution was filtered through VIVAspin membrane polyethersulfone filters. The resulting solution was analysed by HPLC under the conditions stated below (see the ESI^+ for retention times). *Column*: Luna C18 5 μm ; *flow*: 1 mL min $^{-1}$; *temperature*: 30 °C; *gradient*: from 100% H_2O (0.1% TFA) to 100% MeCN (0.1% TFA) in 22 min. Wavelength: 280 nm. The relative amount of the species was determined by correcting the value with the response factor of each compound.

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Notes and references

- (a) H. Röller, K. H. Dahm, C. C. Sweely and B. M. Trost, *Angew. Chem., Int. Ed. Engl.*, 1967, 6, 179–180; (b) A. T. Fuller, G. Mellows, M. Woolford, G. T. Banks, K. D. Barrow and E. B. Chain, *Nature*, 1971, 234, 416–417.
- (a) G. Li, T. Wang, F. Fei, Y.-M. Su, Y. Li, Q. Lan and X.-S. Wang, *Angew. Chem., Int. Ed.*, 2016, 55, 3491–3495; (b) X.-Y. Chen, Z.-H. Gao, C.-Y. Song, C.-L. Zhang, Z.-X. Wang and S. Ye, *Angew. Chem., Int. Ed.*, 2014, 53, 11611–11615.
- (a) M. Demestre, S. M. Messerli, N. Celli, M. Shahossini, L. Kluwe, V. Mautner and H. Maruta, *Phytother. Res.*, 2009, 23, 226–230; (b) Z. Orban, M. Mitsiades, T. R. Burke Jr., M. Tsokos and G. P. Chouros, *NeuroImmunoModulation*, 2000, 7, 99–105; (c) T.-W. Chung, S.-K. Moon, Y.-C. Chang, J.-H. Ko, Y.-C. Lee, G. Cho, S.-H. Kim, J.-G. Kim and C.-H. Kim, *FASEB J.*, 2004, 18, 1670–1681; (d) V. Staniforth, L.-T. Chung and N.-S. Yang, *Carcinogenesis*, 2006, 27, 1803–1811; (e) K. Natajaran, S. Singh, T. R. Burke Jr., D. Bruberger and B. B. Aggarwal, *Proc. Natl. Acad. Sci. U. S. A.*, 1996, 93, 9090–9095.
- L. R. Ferguson, S.-T. Zhu and P. J. Harris, *Mol. Nutr. Food Res.*, 2005, 49, 585–593.
- (a) Y. S. Lee, *Arch. Pharmacol. Res.*, 2005, 28, 1183–1189; (b) M. Kampa, V. I. Alexaki, G. Notas, A. P. Nifli, A. Nistikaki, A. Hatzoglou, E. Bakogeorgou, E. Kowimtzoglou, G. Blekces and D. Boskou, *Breast Cancer Res.*, 2004, 6, 63–74.
- (a) K. L. Johnston, M. N. Clifford and L. M. Morgan, *Am. J. Clin. Nutr.*, 2003, 78, 728–733; (b) J. Y. Lee, K. J. Yoon and Y. S. Lee, *Bioorg. Med. Chem. Lett.*, 2003, 13, 4331–4334.
- (a) N. Sharma, U. K. Sharma, R. Kumar, N. Katoch, R. Kumar and A. K. Sinha, *Adv. Synth. Catal.*, 2011, 353, 871–878; (b) V. Percec, M. Peterca, M. J. Sienkowska, M. A. Ilies, E. Aqad, J. Smidkral and P. A. Heiney, *J. Am. Chem. Soc.*, 2006, 128, 3324–3334.



- 8 A. Shard, N. Sharma, B. Bharti, S. Dadhwal, R. Kumar and A. K. Sinha, *Angew. Chem., Int. Ed.*, 2012, **51**, 12250–12253.
- 9 (a) C. Wuensch, T. Pavkov-Keller, G. Steinkellner, J. Gross, M. Fuchs, A. Hromic, A. Lyskowski, K. Fauland, K. Gruber, S. M. Glueck and K. Faber, *Adv. Synth. Catal.*, 2015, **357**, 1909–1918; (b) C. Wuensch, S. M. Glueck, J. Gross, D. Koszelewski, M. Schober and K. Faber, *Org. Lett.*, 2012, **14**, 1974–1977.
- 10 C. Wuensch, J. Gross, G. Steinkellner, K. Gruber, S. M. Glueck and K. Faber, *Angew. Chem., Int. Ed.*, 2013, **52**, 2293–2297.
- 11 (a) D.-H. Lee, K.-H. Kwon and C. S. Yi, *J. Am. Chem. Soc.*, 2012, **134**, 7325–7328; (b) M. C. Reddy and M. Jeganmohan, *Eur. J. Org. Chem.*, 2013, 1150–1157.
- 12 X.-S. Zhang, Z.-W. Li and Z.-J. Shi, *Org. Chem. Front.*, 2014, **1**, 44–49.
- 13 S. Halder and S. Koner, *Beilstein J. Org. Chem.*, 2013, **9**, 49–55.
- 14 E. Busto, R. C. Simon and W. Kroutil, *Angew. Chem., Int. Ed.*, 2015, **54**, 10899–10902.
- 15 (a) W.-D. Fessner, *New Biotechnol.*, 2015, **32**, 658–664; (b) R. Sigrist, B. Z. d. Costa, A. J. Marsaioli and L. G. de Oliveira, *Biotechnol. Adv.*, 2015, **33**, 394–411; (c) V. Köhler and N. J. Turner, *Chem. Commun.*, 2015, **51**, 450–464; (d) J. Muschiol, C. Peters, N. Oberleitner, M. D. Mihovilovic, U. T. Bornscheuer and F. Rudroff, *Chem. Commun.*, 2015, **51**, 5798–5811; (e) J.-K. Guterl and V. Sieber, *Eng. Life Sci.*, 2013, **13**, 4–18; (f) E. Ricca, B. Brucher and J. H. Schrittwieser, *Adv. Synth. Catal.*, 2011, **353**, 2239–2262; (g) J. H. Schrittwieser, J. Sattler, V. Resch, F. G. Mutti and W. Kroutil, *Curr. Opin. Chem. Biol.*, 2011, **15**, 249–256.
- 16 (a) B. Seisser, R. Zinkl, K. Gruber, F. Kaufmann, A. Hafner and W. Kroutil, *Adv. Synth. Catal.*, 2010, **352**, 731–736; (b) K. Kim and P. A. Code, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 1205–1208; (c) R. Stephen-Phillips, K. Ravichandran and R. L. Von Tersch, *Enzyme Microb. Technol.*, 1989, **11**, 80–83.
- 17 (a) S. Pilbák, O. Farkas and L. Poppe, *Chem. – Eur. J.*, 2012, **18**, 7793–7802; (b) J. A. Kyndt, T. E. Meyer, M. A. Cusanovic and J. J. Von Beeumen, *FEBS Lett.*, 2002, **512**, 240–244.
- 18 (a) P. Both, H. Busch, P. P. Kelly, F. G. Mutti, N. J. Turner and S. L. Flitsch, *Angew. Chem., Int. Ed.*, 2016, **55**, 1511–1513; (b) Z. Xu, Z. Xu, B. Tang, S. Li, J. Gao, B. Chi and H. Xu, *Biochem. Eng. J.*, 2016, **109**, 28–34; (c) G. Gourinchas, E. Busto, M. Killinger, N. Richter, B. Wiltschi and W. Kroutil, *Chem. Commun.*, 2015, **51**, 2828–2831; (d) S.-Y. Chen, C.-X. Yang, J.-P. Wu, G. Xu and L.-R. Yang, *Adv. Synth. Catal.*, 2013, **355**, 3179–3190; (e) E. Lorenz, S. Klatte and V. F. Wendisch, *J. Biotechnol.*, 2013, **168**, 289–294; (f) M. Schrewe, N. Ladkau, B. Bühler and A. Schmid, *Adv. Synth. Catal.*, 2013, **355**, 1693–1697.
- 19 (a) For selected reviews on biocatalytic C–C bond formation see: N. G. Schmidt, E. Eger and W. Kroutil, *ACS Catal.*, 2016, **6**, 4286–4311; (b) *Science of Synthesis: Biocatalysis in Organic Synthesis*, ed. K. Faber, W.-D. Fessner and N. J. Turner, Thieme, Stuttgart, 2015, ch. 2.1 and 2.2, pp. 1–212; (c) Y. Miao, M. Rahimi, E. M. Geertsema and G. J. Poelarends, *Curr. Opin. Chem. Biol.*, 2015, **25**, 115–123; (d) B. M. Nestl, S. C. Hammer, B. A. Nebel and B. Hauer, *Angew. Chem., Int. Ed.*, 2014, **53**, 3070–3095; (e) M. Müller, *Adv. Synth. Catal.*, 2014, **354**, 3161–3174; (f) V. Resch, J. H. Schrittwieser, E. Siirola and W. Kroutil, *Curr. Opin. Biotechnol.*, 2011, **22**, 793–799; (g) M. Pohl and A. Liese, *Industrial Processes Using Lyases for C–C, C–N and C–O Bond Formation*, in *Biocatalysis in the Pharmaceutical and Biotechnology Industry*, ed. R. N. Patel, CRC Press, 2007, pp. 661–676.
- 20 D. G. Gibson, L. Young, R.-Y. Chuang, J. C. Venter, C. A. Hutchison III and H. O. Smith, *Nat. Methods*, 2009, **6**, 343–345.

