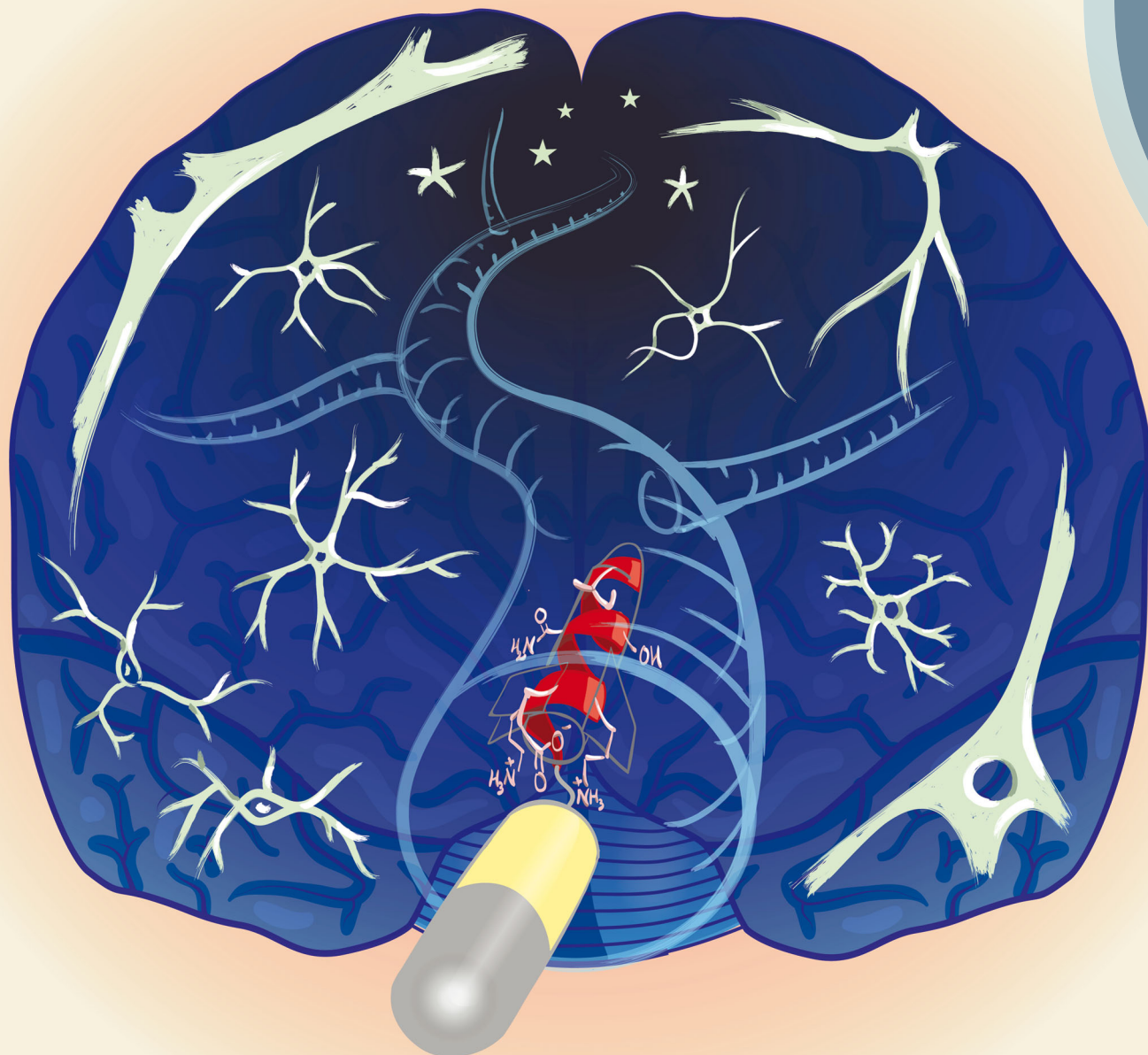


Chem Soc Rev

Chemical Society Reviews

www.rsc.org/chemsocrev



ISSN 0306-0012



ROYAL SOCIETY
OF CHEMISTRY

REVIEW ARTICLE

Benjamí Oller-Salvia *et al.*

Blood–brain barrier shuttle peptides: an emerging paradigm for brain delivery

175
YEARS



Cite this: *Chem. Soc. Rev.*, 2016, 45, 4690

Received 28th January 2016

DOI: 10.1039/c6cs00076b

www.rsc.org/chemsocrev

Blood–brain barrier shuttle peptides: an emerging paradigm for brain delivery

Benjami Oller-Salvia,^a Macarena Sánchez-Navarro,^a Ernest Giralt^{ab} and Meritxell Teixidó^a

Brain delivery is one of the major challenges in drug development because of the high number of patients suffering from neural diseases and the low efficiency of the treatments available. Although the blood–brain barrier (BBB) prevents most drugs from reaching their targets, molecular vectors – known as BBB shuttles – offer great promise to safely overcome this formidable obstacle. In recent years, peptide shuttles have received growing attention because of their lower cost, reduced immunogenicity, and higher chemical versatility than traditional Trojan horse antibodies and other proteins.

1. Introduction

Delivery to the brain is a major challenge in drug development because an ageing population and the growing prevalence of brain cancers are increasing the incidence of central nervous system (CNS) diseases.¹ Moreover, the lack of efficient treatments generates high direct and indirect costs, which together correspond to 1/4th of the burden of all diseases in Europe and

high-income countries.² Therefore, improving CNS drugs would not only enhance the well-being of many people but also considerably reduce health costs. However, a formidable obstacle must be overcome to enable active compounds to reach their targets in therapeutically relevant amounts: the blood–brain barrier (BBB).³

Although many strategies to circumvent the BBB have been proposed, to date none has shown a satisfactory efficiency–safety balance. At one end of the spectrum, direct drug administration into the brain has a high risk and is very local and, at the other end, the modification of molecules to enhance their diffusion through the barrier is applicable only for some small drugs. Among the non-invasive approaches, molecular vectors – also known as BBB shuttles – (Fig. 1) have proved their

^a *Institute for Research in Biomedicine (IRB Barcelona), The Barcelona Institute of Science and Technology (BIST), 08028 Barcelona, Spain. E-mail: meritxell.teixido@irbbarcelona.org, ernest.giralt@irbbarcelona.org*

^b *Department of Organic Chemistry, University of Barcelona, 08028 Barcelona, Spain*



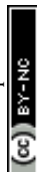
Benjami Oller-Salvia

Benjami Oller-Salvia obtained his PhD from the University of Barcelona, conducting research at the Institute for Research in Biomedicine (IRB Barcelona). His PhD thesis focussed on venom-inspired peptides for brain delivery and was mentored by Prof. Ernest Giralt and Dr. Meritxell Teixidó. He has performed research stages in several institutes, working under the supervision of Prof. Salvador Borrós group at IQS, Prof. Elazer Edelman and Prof. Mercedes Balcells at MIT and Prof. Kai Johnsson at EPFL. Since January 2016 he has been working on genetic code expansion applied to therapeutic proteins in Prof. Jason Chin's laboratory at the MRC Laboratory of Molecular Biology in Cambridge.



Macarena Sánchez-Navarro

Macarena Sánchez-Navarro obtained her PhD from the University of Seville under the supervision of Dr Javier Rojo where she worked on the synthesis of glyco-dendrimers. In a first postdoctoral stay at Nazario's Martin Lab (Universidad Complutense de Madrid, Spain), she worked on the preparation and evaluation of two families of water soluble fullerenes. In 2010 she joined the Prof. Ben Davis group at University of Oxford where she was involved in the site-selective modification of proteins. Since 2012 she has been a part of the group of Prof. Ernest Giralt at IRB Barcelona where her research is focused on understanding the main mechanisms of brain transport.



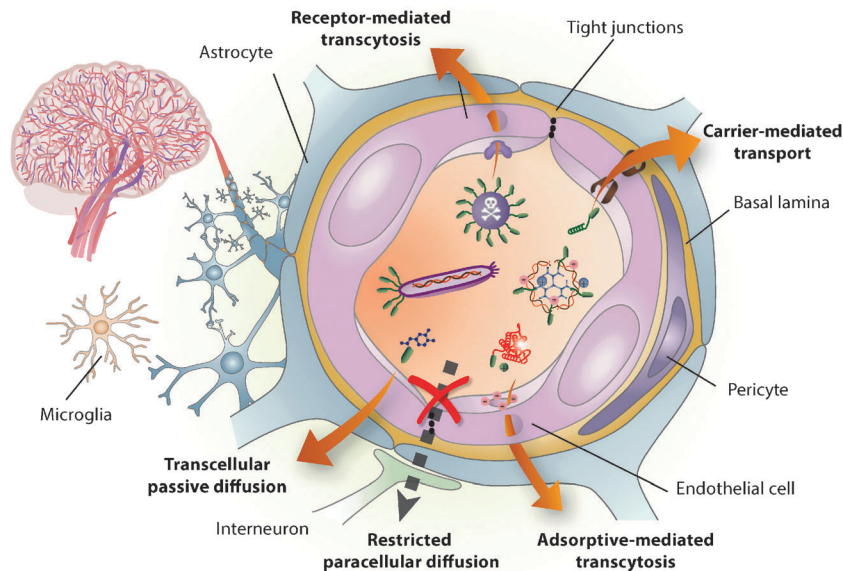


Fig. 1 BBB-shuttle mediated transport across the BBB. The BBB is formed mainly by the tight brain endothelium, which is surrounded by the basal lamina and regulated by the other cells in the neurovascular unit, including pericytes, glial cells and neurons.⁶ BBB shuttles mediate drug delivery across this barrier by taking advantage of endogenous transport pathways. Small molecules and peptides can be delivered using shuttles that undergo passive diffusion and carrier-mediated transport. However, passive diffusion is altered by the physicochemical characteristics of the cargo, including $\log P$, pK_a , molecular weight, topological polar surface area and hydrogen bonding.¹⁸⁶ Carrier proteins usually undergo precise rearrangements to translocate constructs into the cytosol,⁹ thus they are also highly sensitive to the modification of substrate properties. Conversely, adsorptive- or receptor-mediated transcytosis allow the transport of a wider variety of cargoes, including proteins, viruses and nanocarriers. Despite the high efficiency of the former mechanism in mediating tissue uptake, the latter has focussed most attention on brain delivery due to its potential targeting capacity.

potential in preclinical research over the last two decades, and some of these compounds are in clinical trials. The BBB shuttle⁴ concept includes Trojan horse antibodies⁵ and any other molecule capable of transporting a cargo into the brain parenchyma without affecting the BBB integrity. Over the last five years, research into peptide shuttles has thrived because they overcome some of the weaknesses of classical protein shuttles,

including complex derivatization and characterization, high immunogenicity, and costly production. Of note, here we will use the word peptide to refer to small proteins (with or without structure) containing up to 50 amino acid residues.

In this review, crossing the BBB is presented as a tremendous challenge but also as an excellent opportunity for drug delivery into the brain. We first provide an overview of BBB shuttle



Ernest Giralt

Ernest Giralt is Professor of Organic Chemistry at the University of Barcelona and Group Leader at the Institute for Research in Biomedicine (IRB Barcelona) where he is the Head of the Chemistry and Molecular Pharmacology Programme. He has published around 420 papers, 16 patents and several books. He has received several awards including, recently, the Josep Rüdinger Memorial Lecture Award and the Max Bergmann Medal.

His major interests lie in the fields of peptide synthesis and molecular recognition, in particular using NMR, with emphasis on the design of specific ligands for interaction with protein surfaces, related to possible therapeutic uses. This includes studies concerning new brain delivery systems, and modulators of protein-protein interactions.



Meritxell Teixidó

Meritxell Teixidó has been a Research Associate in the Design, Synthesis and Structure of Peptides and Proteins Group at the IRB Barcelona since 2006. Her major interest lies in the field of peptide synthesis and the discovery of blood-brain barrier peptidic shuttles, in particular using these shuttles to deliver drugs, diagnostic agents and nanoparticles that otherwise cannot reach their target site inside the CNS. In this regard, her research combines

protease-resistant peptides, mass spectrometry techniques, and transport evaluation tools to achieve delivery systems of this nature. She has published around 45 papers and review articles and participated in 7 patents.



peptides and subsequently present the most advanced ones, which are included in drug formulations that have reached clinical trials. Then, the focus is given to three representative case studies to illustrate some of the main achievements of shuttle transport of diverse cargoes. Additionally, some strategies that rescue unspecific shuttles are reported and the relevance of protease-resistance is highlighted. Finally, we point out the main trends in the field and the challenges to be addressed.

2. Toward minimized brain delivery vectors

2.1. The blood–brain barrier

The BBB is a physical, metabolic and transport barrier that tightly controls the transfer of substances from blood to neural tissues and *vice versa*, thereby contributing to brain homeostasis.^{6–8} Endothelial cells on brain capillary walls are the main constituents of this barrier and they form tight junctions that hinder paracellular passage. Additionally, these cells have many cytosolic and extracellular-membrane enzymes, down-regulated vesicular transport and efflux pumps. The permeability of the brain endothelium is influenced by the other cells belonging to the neurovascular unit (Fig. 1) and is affected by most CNS pathologies; however, BBB dysfunction is usually only significant in advanced stages of disease and in the most affected sites.

Despite its efficient role as a barrier, the BBB is the main gateway to the brain as it grants access to necessary ions, nutrients and hormones – the maximal cell-capillary distance is 20 μm , which can be permeated by small molecules in half a second.⁹ Therefore, taking advantage of the endogenous transport mechanisms present on the BBB is potentially the most efficient way to deliver substances to any part of the brain.⁶ Many small hydrophobic compounds (<500 Da) diffuse across the endothelium membrane, while polar molecules such as glucose, amino acids, and several peptides have specific carriers; these highly selective transporters mediate transport into the endothelium cytosol and from there to the brain extracellular space. Indeed, even some macromolecules and also certain peptides cross the BBB through endocytic mechanisms involving receptor-mediated transcytosis (RMT) and/or adsorptive-mediated transcytosis (AMT). In the process of transcytosis, the vesicle formed circulates across the cell, bypassing the degradation pathway, and eventually releases its content into the parenchyma by exocytosis. AMT is considered non-specific and comprises all vesicular transport mechanisms that do not involve protein receptors; in AMT, endocytosis is generally promoted by the interaction of the often positively charged molecule with membrane phospholipids and the glycocalyx.

2.2. Brain delivery approaches

Nowadays most strategies for drug delivery into the brain that circumvents the BBB are invasive,¹⁰ involving the highest risk of brain damage or infection and requiring demanding set-ups.^{11,12} In addition, administration is often excessively localized and

diffusion in the brain is very limited, especially for large molecules.¹³ Although alternative routes such as nasal delivery are under investigation, most attention is focused on achieving efficient distribution of the drug through the extensive brain vasculature.

Temporal disruption of the tight junctions of brain endothelium by chemical or physical stimuli entails toxicity and the risk of neuronal dysfunction.¹⁴ Hence, much effort has been channelled into improving transport across endothelial cells. The most common strategies rely on enhancing lipophilicity and positive charge, in order to increase passive diffusion and mediate interaction with the anionic glycocalyx, respectively. However, these modifications lead to higher unspecific uptake in many tissues often resulting in off-target effects and, in addition, they enhance recognition by efflux pumps.^{12,15} A more selective way to boost the permeation of certain small molecules into the brain is to modify them by mimicking endogenous substrates of BBB carriers.¹⁶ However, all these approaches require a high degree of tailoring and are rarely applicable to large drugs such as biotherapeutics.

A more general approach for drug delivery to the CNS focuses on delivery vectors. Although biological vectors such as viruses^{17,18} and modified cells¹⁹ have been used to increase BBB transport, their safety, permeability across an intact barrier, and brain selectivity are still limited.^{18–20} Conversely, molecular vectors, dubbed BBB shuttles,^{4,21–33} aim to provide broadly applicable, selective and safer delivery systems.

2.3. BBB shuttle peptides

BBB shuttles allow the transport of a wide range of cargoes, comprising small molecules, proteins, nanoparticles and genetic material across the BBB. Substrates of natural carriers such as glucose and neutral amino acids have been applied to transport small molecules through their natural carriers on the BBB, while for nanoparticles and biomolecules the focus has been set on receptor ligand proteins since vesicular mechanisms tolerate a wide range of cargo sizes.³ Remarkably, peptides have bridged the gap between these two worlds.

The BBB shuttle concept was conceived by William M. Pardridge in the mid-1980s,³⁴ inspired by chimeric proteins targeting cell receptors. The first successful attempts relied first on cationized albumin,³⁵ which lacked brain selectivity, and then on IgGs directed against insulin and transferrin receptors.³⁶ However, the success of these initial antibodies was limited by their high affinity, which hampered an efficient release into the brain parenchyma.^{15,37} Therefore, a variety of protein shuttles have been investigated; most of them are ligands of receptors on the brain endothelium and include the following: apolipoproteins (Apo) A and E,³⁸ receptor-associated protein (RAP),³⁹ transferrin (Tf),⁴⁰ lactotransferrin,⁴¹ melanotransferrin (p97),⁴² and leptin.⁴³ However, these proteins compete with their endogenous counterparts. Although a few non-endogenous proteins, such as wheat germ agglutinin⁴⁴ and a non-toxic mutant of diphtheria toxin (CRM197),⁴⁵ have been used, they also have shown moderate efficacy and selectivity. In recent years, research on antibodies has been relaunched focussing on lower-affinity IgG derivatives.^{21,37,46,47}



Despite the relative success of antibodies and other large proteins, their production is expensive and high immunogenicity is an issue. This is why research in this field in the last decade has focussed to a great extent on peptides. These molecules combine the low cost of small drugs with the high specificity of biologics.^{37,48} Peptides are easier to obtain and characterize than the latter and have very low immunogenicity, especially those without a rigid structure.⁴⁹ Moreover, they display medium to low affinities, a property that has been critical to the development of anti-TfR shuttles.³⁷ In addition, peptides are amenable to chemical synthesis. This feature opens up the possibility of applying a wide range of non-natural modifications and of introducing a plethora of functional groups for site-specific conjugation to proteins and nanocarriers. Furthermore, reduced functional alteration of the cargo and enhanced shelf-life are also important advantages over most proteins. Although peptides have often been undervalued in pharmaceutical chemistry because of their low resistance to proteolytic degradation, this limitation can now be overcome by means of various strategies that will be described in section 7.⁵⁰

Although some peptides had long been shown to cross the BBB,⁵¹ the field of peptide shuttles was pioneered in 1999 by Stephen Dowdy and coworkers.⁵² In this seminal paper, the authors demonstrated the capacity of a fragment from the HIV TAT protein to deliver β -galactosidase into the brain and other organs. However, it was in 2007 with RVG29⁵³ that a peptide was proven capable of transporting cargoes into the brain in a selective fashion. Soon after, the great potential of Angiopep-2⁵⁴ and glutathione (GSH)^{55,56} as BBB shuttles was unravelled – formulations including these peptides are currently in clinical trials. Remarkably, in the last 5 years over 30 BBB shuttle peptides with increasing efficiency and versatility have been reported (Table 1).

3. Aiming for selectivity

The discovery of TAT peptide as a brain delivery vector, directed initial research efforts into finding BBB shuttles with high permeability across cell membranes. Passive diffusion and AMT provide the highest transport since the first is considered unsaturable and saturation concentrations for AMT are 3 orders of magnitude higher than for RMT.⁵⁷ Nevertheless, the need for safer therapeutics has pushed research towards targeted strategies in an attempt to achieve Paul Ehrlich's "magic bullet".

3.1. Unspecific uptake

BBB shuttle peptides capable of increasing the brain uptake of large cargoes in a non-selective way belong mainly to the cell-penetrating peptide (CPP) family. CPPs comprise short amphipathic and/or cationic sequences with a high capacity to cross cell membranes without the need of a receptor.^{58,59} Most peptides with this property have been derived from protein transduction domains (e.g. TAT⁶⁰ and penetratin⁶¹), by hybridizing these domains with antibiotic peptides (e.g. SynB1^{62,63} and transportan⁶⁴), or through biomimetic design (e.g. oligoarginine⁶⁵).

Although CPPs can enter cells through different mechanisms, when linked to large cargoes they mostly undergo endocytosis. Therefore, it is generally assumed that they undergo AMT across the BBB.⁵⁷ However, exocytosis of the entire BBB shuttle constructs from the endothelium is more controversial than in RMT and they may accumulate in the endothelium as has been reported for positively charged proteins such as lectins.⁴⁴ By contrast, a recent study using CPPs of four different classes suggests that trapping in brain capillaries of peptides alone may be relatively low but indicates that brain parenchymal accumulation does not correlate with their cell internalization capacity.⁶⁶

For small drugs (<300 Da), peptide shuttles formed by 2–4 amino acid residues that cross the BBB through passive diffusion are more attractive since they may minimize the loss of activity upon conjugation. In peptides diffusing across the BBB, hydrogen bonding and water desolvation have a better correlation with permeability than $\log P$.⁶⁷ Based on this criterion, three families of BBB shuttle peptides, namely diketopiperazines,³² *N*-methylphenylalanines^{29–31} and phenylprolines,⁶⁸ have been developed.

3.2. Targeting transporters

Most BBB shuttle peptides that interact with transporters have been obtained from either neurotropic biomolecules or phage display biopanning (Fig. 2). Natural peptides or proteins targeting the brain can be endogenous, like hormones and apolipoproteins, or exogenous, such as certain viruses and neurotoxins.^{33,69} Regarding phage display, although it has been extensively applied for the last three decades,⁷⁰ it has not been exploited to find BBB shuttles until the last few years. Filamentous phages, which are the most commonly used ones, measure 6.5 nm in diameter and 900 nm in length and are generally engineered to display only 5 copies of peptide per viral particle.⁷¹ These features, together with the high number of sequences that can be displayed in a phage library, explain why this screening technique has provided peptides capable of transporting large cargoes such as nanoparticles.

Ideally, BBB shuttles should target receptors with the following attributes: high expression in the luminal side of brain vasculature with respect to other tissues; capacity to mediate transcytosis; high turnover and broad substrate recognition.⁷² Moreover, the physiological role of the transporter should not be easily altered. Unfortunately, quantitative physiological information of the wide variety of brain endothelium receptors reported is very limited, thus screening together with trial and error rather than design has mainly driven the discovery of new shuttle peptides. Of note, although many sequences have been reported to target a particular receptor, the contribution of other mechanisms cannot be excluded.¹⁵ Determining whether the molecule has reached the brain parenchyma is already a challenge and studying the delivery route requires a multifocal and integrated approach (Table 2).

The receptors with high expression on the BBB for which transcytosis has been best characterized include: transferrin (TfR1),⁴⁰ low-density lipoproteins (LDLRs),⁷³ insulin⁷⁴ and leptin.⁷⁵





Table 1 Selected features of representative peptide BBB shuttles

Peptide	Typical sequence	Proposed transporter	Origin	Main cargoes	BBB passage evidence	Ref.
Angiopoep-2	TFYGGSRGRKRNFKTEEY-OH	LRP1	Neurotropic endog. Protein	Small drugs, proteins, nanopart., and DNA/RNA	<ul style="list-style-type: none"> • BBBCM • Capillary depletion • Fluorescence microscopy • TEM 	54 and 113–135
ApoB (3371–3409)	SSVIDALQYKLEGGITRLTRK-RGLKATALSLSNKFVEGS (LRKLRKRL) ₂	LRP2 LDLR	Neurotropic endog. Protein	Proteins	<ul style="list-style-type: none"> • Effect on glioma, epilepsy and Parkinson's mouse models • Capillary depletion • Effect on MPS mouse model 	81 and 156
ApoE (159–167) ₂	LRP2 LDLR	LRP2 LDLR	Neurotropic endog. protein	Proteins and nanopart.	<ul style="list-style-type: none"> • BBBCM • Capillary depletion • Fluorescence microscopy • Effect on the MLD mouse model 	82, 157 and 160
Peptide-22	Ac-C(&)MPRLRG(&)-NH ₂	LDLR	Phage display (receptor)	Nanopart.	<ul style="list-style-type: none"> • BBBCM • Live fluorescence microscopy 	27
THR	THRPPMWSVPVWP-NH ₂	TR1	Phage display (cells)	RNA and nanopart.	<ul style="list-style-type: none"> • BBBCM • TEM 	88 and 183
THR <i>retro-entatio</i>	pwpswmprrht-NH ₂	TR1	Phage display-derived	Small drugs and nanopart.	<ul style="list-style-type: none"> • BBBCM • Live fluorescence microscopy 	182
CRT	C(&)RTGPSVC(&)	TR1	Phage display (mice)	Virus	<ul style="list-style-type: none"> • Capillary depletion 	89
Leptin30	YQQLTSMPSRNVIQISND-LENLRDLLHLV	Leptin receptors	Neurotropic endog. protein	DNA	<ul style="list-style-type: none"> • Capillary depletion 	76 and 77
RVG29	YTWMPENPRPGTPCDIFT-NSRGKRASNG-OH	nAChR	Neurotropic exog. protein	Nanopart. and RNA/DNA	<ul style="list-style-type: none"> • Capillary depletion • BBBCM • Effect in viral encephalitis and mouse models of Parkinson's disease 	53 and 146–154
P ₁ CDX	GreitGraetwsekf-OH	nAChR	Neurotoxin-derived	Nanopart.	<ul style="list-style-type: none"> • BBBCM • Effect on glioblastoma 	94
Apamin	C(&)NC(&)KAPETALC(&)-AR-RC(&)QQH-NH ₂	KCa channel?	Neurotoxin	Proteins and nanopart.	<ul style="list-style-type: none"> • Human BBBCM • Fluorescence microscopy 	95 and 96
MiniAp-4	[Dap](&)KAPETALD(&)	KCa channel?	Neurotoxin-derived	Small drugs, proteins and nanopart.	<ul style="list-style-type: none"> • Intracerebral microdialysis 	97
GSH	γ -L-glutamyl-CG-OH	GSH transporter	Endog. peptide	Nanopart.	<ul style="list-style-type: none"> • Effect on glioma and MS mouse models 	55, 56 and 141–145
G23	HLNILSTLWKYRC	GM1	Phage display (receptor)	Nanopart.	<ul style="list-style-type: none"> • BBBCM 	107
g7	GFtGFLS(O- β -Glc)-NH ₂	Unknown receptor	Endog. peptide-derived	Nanopart.	<ul style="list-style-type: none"> • Fluorescence microscopy • Fluorescence microscopy 	100 and 164–166
TGN	TGNYKALHPHG	Unknown receptor	Phage display (<i>in vivo</i>)	Nanopart.	<ul style="list-style-type: none"> • TEM 	109–111
TAT (47–57)	YGRKKRRQRRR-NH ₂	AMT	Exog. protein	Proteins and nanopart.	<ul style="list-style-type: none"> • Effect on glioma and AD mouse models • BBBCM • Capillary depletion • Fluorescence microscopy • TEM 	52 and 167–174
SynB1	RGGRLSYRRRFFSTSTGR	AMT	Toxin	Small drugs	<ul style="list-style-type: none"> • Brain perfusion • Capillary depletion 	62 and 63
Diketopiperazines	&(N-MePhe)-(N-MePhe)Diketopiperazines	Passive diffusion	Design (+serendipity)	Small drugs	<ul style="list-style-type: none"> • PAMPA 	32 and 180
PhPro	(Phenylproline) ₄ -NH ₂	Passive diffusion	Design	Small drug	<ul style="list-style-type: none"> • BBBCM • PAMPA 	68

"BBB passage evidence" includes the main strategies used to assess the presence of the compound targeted by the BBB-shuttle in the brain parenchyma or the effects derived from it – these approaches do not provide information about brain selectivity and do not prove that the whole cargo-shuttle construct crosses the BBB. Abbreviations: Endog.: endogenous; Exog.: exogenous; nanopart.: nanoparticles; BBBCM: a cell-based BBB model; MPS: mucopolysaccharidosis; MLD: metachromatic leukodystrophy; AD: Alzheimer's disease. Nomenclature for cyclic peptides (&|) is adapted to the 3-letter amino acid code from the one described by Spengler *et al.*¹⁸⁸ [Dap] stands for diamino propionic acid. Only selected references relevant to the study of these peptides as BBB-shuttles are cited here.

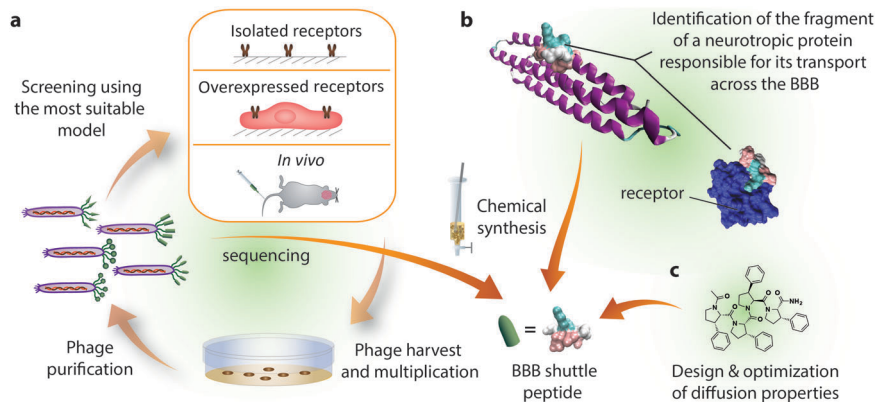


Fig. 2 Sources of BBB shuttle peptides. (a) Phage display biopanning may be performed on isolated receptors, cells overexpressing a particular receptor or *in vivo*. The phages that bind the receptors or accumulate in the brain are recovered, amplified through bacterial infection, titrated and sequenced. The biopanning cycle is generally repeated 2–3 times and the most abundant sequences are chemically synthesized for further study. (b) For peptides coming from (neurotropic) proteins, the moiety responsible for transport is identified through one or more of the following techniques: sequence alignment, screening of synthesized fragments or structural studies involving X-ray and NMR. Here we highlight a sequence of ApoE (PDB: 1LPE) that has been shown to interact with LRP1 (PDB: 2KNY).¹⁸⁷ (c) Passive diffusion shuttles have been designed taking into account the parameters involved in this permeation mechanism. The structure of phenylproline is presented here as the BBB peptide shuttle with the highest solubility and transport capacity.

However, while the first two have been widely used for brain delivery, very few BBB shuttle peptides derive from the others.^{76,77}

LDLRs have been extensively studied for their roles in transport and signalling⁷⁸ and they are the most exploited receptors for delivery across the BBB using peptides. Moreover, some of them (particularly LRP1) are overexpressed in the brain⁷⁹ and in tumours.⁸⁰ Peptides targeting this family of receptors either are based on natural protein ligands, namely ApoB and ApoE fragments^{81,82} and Angiopep-2, or they are found by phage display biopanning against LDLR, like Peptide-22.²⁷

TfR1 is also well characterized among BBB transport receptors. Furthermore, it has even higher expression than LRP1 on the brain endothelium^{83–85} and is widely present in tumours.⁸⁶ Peptides interacting with TfR1 were discovered by applying phage display biopanning in various ways. B6 was identified in a nonamer library screened against the extracellular domain of human TfR,⁸⁷ whereas THR and T7 were found through panning against a human receptor expressed in chicken fibroblasts (chicken TfR does not bind human Tf),⁸⁸ and the CRT peptide was found to selectively target mouse brain parenchyma *in vivo*.⁸⁹

In addition to the aforementioned receptors, many other pathways have been explored in an attempt to increase brain delivery efficiency and selectivity. The widespread occurrence of some ion channels in the CNS, as well as their intracellular traffic and recycling ability, have inspired several shuttles, comprising RVG29, RDPs,^{90,91} KC2S,⁹² L¹CDX,⁹³ P¹CDX,⁹⁴ apamin,^{95,96} and MiniAp-4.⁹⁷ By contrast, the endogenous peptide GSH was identified as a BBB shuttle following from its reported capacity to reach the brain through a saturable and specific mechanism. Many transporters, some of which are preferentially expressed in the CNS,⁹⁸ mediate the influx and efflux of GSH and its endogenous conjugates;⁹⁹ however, further research is required to elucidate the putative transcytotic mechanism of GSH. Another shuttle derived from an endogenous peptide is g7,¹⁰⁰ which is discussed later in this review.

Although integrin receptors do not display a particularly high expression in the brain microvasculature, they have been extensively used for targeting brain tumours and inflamed regions of the CNS.¹⁰¹ Cyclic RGD (cRGD)¹⁰² is a peptide derived from a sequence present in many proteins that recognise these receptors. Because integrin $\alpha_v\beta_3$ is overexpressed in the neovasculature, cRGD has been extensively used to target nanoparticles into gliomas.¹⁰³ However, it can mediate transcytosis only indirectly through internalization into leukocytes and other immune cells, which are recruited into the brain in response to inflammation.¹⁹

Protein transporters are not the sole means to achieve a certain degree of selectivity in the transcytosis across brain endothelium. Gangliosides have a heterogeneous tissue distribution and can also mediate transport across polarized cells.^{104,105} This particular selective AMT mechanism has been exploited by G23 peptide, which was found by phage display biopanning against gangliosides G_{T1b}¹⁰⁶ and GM1,¹⁰⁷ the latter of which is present in caveolae. G23 has been shown to promote the transport of nanoparticles across the BBB¹⁰⁷ and provide a targeting effect.¹⁰⁸

Finally, several peptide shuttles have been found through *in vivo* phage display biopanning without aiming for a particular receptor. The most prominent example is that of TGN.¹⁰⁹ This sequence is actively transported across brain endothelial cells and its brain selectivity suggests that the mechanism is receptor-mediated. The brain delivery capacity of this shuttle is supported by enhanced therapeutic effect in glioblastoma and Alzheimer's mouse models.^{110,111}

4. On the way to clinical application

The first generation of BBB shuttle peptides has reached clinical trials in the last few years. Here we will describe the two best-documented examples, Angiopep-2 and GSH. However, others are



Table 2 Techniques used to study the transport of BBB shuttles

Model	Advantages	Limitations	Observations & examples	Ref.
Passive diffusion BBB models	<ul style="list-style-type: none"> Suitable for high-throughput screening 	<ul style="list-style-type: none"> Only useful for compounds that cross mainly through diffusion Cannot predict the effect of efflux pumps 	<ul style="list-style-type: none"> <i>E.g.</i> diketopiperazine, <i>N</i>-methyl-Phe and phenylproline shuttles have been optimized using the parallel artificial membrane permeability assay (PAMPA) 	29–32, 68 and 180
Cell uptake	<ul style="list-style-type: none"> High throughput Mechanistic understanding of the transport process using simple settings: active transport (temperature, sodium azide), endocytic mechanism (saturation, inhibition), and receptor type (competition with substrates) 	<ul style="list-style-type: none"> Cannot be used to predict BBB permeability Expression of transporters may differ from physiological human BBB Inhibitors may affect different pathways in diverse cell-lines No evidence of whether the construct has been degraded 	<ul style="list-style-type: none"> Immortalized cell-lines are preferred to primary endothelial cells since the latter rapidly lose their natural phenotype; bEnd3 is the most used cell-line <i>E.g.</i> mechanisms of Angiopoep-2 and TGN have been partly elucidated through cell uptake 	25, 109, 113 and 132
Cell-based BBB models	<ul style="list-style-type: none"> Good correlation with <i>in vivo</i> permeability values of small molecules Compromise between costs, throughput and predictive value Paracellular transport restricted by tight junction proteins Functional expression of many transporters Can be used to study transport mechanisms when paracellular contribution is low 	<ul style="list-style-type: none"> Difficult to compare permeability values between different models Only certain trends and certain permeability values can be predicted for macromolecules & NPs Trans-endothelial resistance is still low in robust models compared to <i>in vivo</i> (0.05–1 vs. > 2–8 kΩ cm²) To minimize paracellular contribution some authors perform pulse-chase assays Expression of transporters may differ from physiological human BBB High cost of Transwell® and media Very low throughput Requires highly skilled personnel Tissue damage and glial activation produced by the probe Interaction with receptors cannot always be extrapolated to humans (like all techniques applied to animals/tissues derived from them) 	<ul style="list-style-type: none"> Various cell alternatives: non-cerebral cells or brain capillary endothelial cells (BCEC). BCEC can be from different species and primary or immortalized, generally in co-culture with glial cells or pericytes. hCMEC/D3 is the most used cell-line, although it is considered leaky <i>E.g.</i> Angiopoep-2, THRre and MiniAp-4 were discovered in a bovine BBB cell-based model 	21, 54, 82, 94, 95, 97, 135, 160, 182 and 183
<i>In vivo</i> intracerebral microdialysis	<ul style="list-style-type: none"> Measures unbound drug in the interstitial fluid site-specifically with a putatively intact BBB Fast sampling and possibility to measure during extended periods (days) Current probes preclude sampling large constructs such as nanopart. 	<ul style="list-style-type: none"> High cost of Transwell® and media Requires highly skilled personnel Tissue damage and glial activation produced by the probe Interaction with receptors cannot always be extrapolated to humans (like all techniques applied to animals/tissues derived from them) 	<ul style="list-style-type: none"> <i>E.g.</i> drugs carried by PEGylated liposomes targeted with GSH have been measured using microdialysis 	98 and 144
<i>In vivo</i> non-invasive imaging	<ul style="list-style-type: none"> General: rough assessment of PK with a limited number of animals Fluorescence and luminescence are available without restrictions and are the fastest noninvasive methods (< 1 min vs. rest > 30 min); wide choice of fluorescent labels; luminescence is more sensitive PET and SPECT: high sensitivity and penetration MRI: high penetration and resolution (0.1–1 mm); no regulatory restrictions High resolution (< 10 μm) Only technique that provides live visual evidence of BBB permeation 	<ul style="list-style-type: none"> General: lower resolution than <i>ex vivo</i> (generally > 1 mm); no possibility to verify the integrity of the construct Fluorescence and luminescence: low-penetrating and semi-quantitative PET and SPECT: regulatory restrictions for radioactivity; PET requires short-lived radiotracers and cyclotron MRI: difficult to quantify Need for craniotomy Low throughput; requires skilled personnel 	<ul style="list-style-type: none"> Fluorescence: most widely used Luminescence: luciferase gene delivery PET: herpes simple virus thymidine kinase (<i>e.g.</i> CRT) SPECT: ¹²⁵I-labelled proteins (<i>e.g.</i> 9Lys-ApoE (151–170)) MRI: iron oxide NPs and DTTPA or DOTA-Ga³⁺ (<i>e.g.</i> Angiopoep-2, cRGD) 	23, 89, 92, 93, 97, 126–134, 136, 154 and 161
<i>In vivo</i> two-photon microscopy	<ul style="list-style-type: none"> General: removal of blood and external tissues; minimal alteration of the sample 	<ul style="list-style-type: none"> General: requires the use of labels and does not allow the identification of the entities 	<ul style="list-style-type: none"> Quantum dots targeted with THR <i>retro-entatio</i> 	27 and 182
<i>Ex vivo</i> quantification of entire organs	<ul style="list-style-type: none"> Fluorescence: no special regulation, only the amount of BBB-shuttle-cargo vs. cargo in brain) High-energy radiotracers: accurate measure of the total amount of constructs; autoradiography of brain slices provides low-resolution brain distribution 	<ul style="list-style-type: none"> Fluorescence: semi-quantitative → only the same organ in different animals can be compared High-energy radiotracers: specific regulations 	<ul style="list-style-type: none"> Fluorescence is the most extensively used ¹²⁵I and ¹³¹I have also been used to label BBB shuttle constructs, especially with Angiopoep-2 	23, 111, 126, 127, 132 and 134





Table 2 (continued)

Model	Advantages	Limitations	Observations & examples	Ref.
<i>Ex vivo</i> quantification in organ homogenates	<ul style="list-style-type: none"> Capillary depletion allows measurement of the amount of compound in brain parenchyma Identity of the compound can be verified If extraction is quantitative, most techniques provide absolute amounts that can be compared between different organs A wide variety of quantification techniques can be applied 	<ul style="list-style-type: none"> Capillary depletion is not quantitative and results should be interpreted with great care Many steps before the final measure increase the chances of introducing artefacts 	<ul style="list-style-type: none"> ^3H and ^{14}C have been used to label drugs encapsulated in GSH-coated PEGylated liposomes and Angiopep-2 constructs ICP-MS is the reference technique for metal NPs (e.g. THR, Angiopep-2) Absorbance, fluorescence and luminescence have been used to quantify delivered molecules/enzymes/genes encoding them (e.g. RVG29, RDPs) Flow cytometry (e.g. TAT, RVG29) Optical microscopy: β-galactosidase (e.g. TAT and RDPs) TEM (e.g. THR) Fluorescence microscopy (e.g. ApoB and ApoE peptides, g7) 	36, 37, 39, 44, 53, 54, 82, 83, 91, 111, 127, 157, 162 and 176
<i>Ex vivo</i> microscopy	<ul style="list-style-type: none"> Optical microscopy gives rough localization of the cargo (i.e. enzyme) TEM is the most reliable, sensitive and precise technique for metal nanoparticles Fluorescence microscopy can reveal the real position of cargo (staining the capillaries or colocalization with certain cell types, such as neurons) 	<ul style="list-style-type: none"> Fixation efficiency of the constructs may be deficient Processing of the sample may introduce artefacts Punctuated or diffuse patterns without a clear localization may be difficult to distinguish from noise 	<ul style="list-style-type: none"> Flow cytometry (e.g. TAT, RVG29) Optical microscopy: β-galactosidase (e.g. TAT and RDPs) TEM (e.g. THR) Fluorescence microscopy (e.g. ApoB and ApoE peptides, g7) 	21, 44, 45, 81, 82, 161 and 183

Despite the plethora of BBB models currently available,^{189–192} evaluating the capacity of BBB shuttles to deliver a cargo across the BBB into the brain parenchyma remains challenging. Thus, an integrated approach is required. Cell-based BBB models¹⁹³ have often been applied to screen BBB shuttles mainly due to their balance between high-throughput and predictive values. Moreover, these models allow focusing on transcellular transport without any interference of other physiological factors, which is particularly useful in mechanistic studies provided paracellular diffusion is low enough; direct internalization assays can also deliver valuable information in this regard. In particular, experiments with human endothelial cells are essential to complement transport studies in animals. Notwithstanding, further refinement and validation of transporters in cell-based BBB models would boost their use. Regarding *in vivo* studies, the capacity of the best-validated shuttles has been evaluated in healthy mice and in animal models of disease. *In situ* brain perfusion in tandem with microdialysis of the brain interstitial fluid provides the most reliable quantification of the unbound drug capable of crossing the BBB. However, these techniques are seldom used due to their complexity, low throughput and limitations of probes for large cargoes. Conversely, a capillary depletion step is more common to minimize to a certain extent the quantification of the compound adsorbed onto or internalized in brain endothelium. This method, which tends to overestimate the parenchymal concentration,¹⁹⁴ is often complemented with an image showing the construct outside the capillaries. Since it is difficult to unambiguously assign spots or a background increase in fluorescence, establishing co-localization with a localized target (e.g. amyloid plaques) is more reliable. Additionally, some studies take advantage of an intrinsic property of the cargo to show that it remains functional upon reaching the brain parenchyma and that a label or a metabolite is not quantified instead. The use of BBB integrity markers and controls including scrambled peptides gives further strength to these studies. Finally, an increase in the therapeutic effect of a drug in an animal model can also be an indirect proof that the drug-shuttle conjugate has reached the brain parenchyma providing the target is only in the CNS and the BBB is intact.

in advanced pre-clinical stages, and MTfp,¹¹² a dodecapeptide derived from melanotransferrin, has been announced to be ready to enter clinical development.

4.1. Angiopep-2: an example of versatility

Angiopep-2 was identified by sequence alignment of aprotinin with other human proteins having a Kunitz domain, which interacts with LRP1 (Fig. 3).^{54,113} This BBB shuttle was initially exploited to transport small molecules such as doxorubicin,¹¹⁴ etoposide,¹¹⁴ paclitaxel,¹¹⁵ and also peptides.¹¹⁶ Its conjugate with paclitaxel (ANG1005 or GRN1005)^{117,118} showed good tolerance in Phase I clinical studies^{119,120} and reached Phase II for the treatment of recurrent high-grade glioma in combination with bevacizumab (ClinicalTrials.gov identifier: NCT01480583). ANG1005 is currently also in Phase II clinical trials for breast cancer (ClinicalTrials.gov identifier: NCT02048059), and preliminary results show that this compound reduces tumours up to 60% in patients.¹²¹

Angiopep-2 has been used to transport a wide variety of nanocarriers loaded with small molecules, proteins or genetic material into the CNS. These carriers include liposomes,^{122,123} nanotubes,¹²⁴ dendrimers made of polyamidoamine^{125–128} and poly-L-lysine,¹²⁹ and also nanoparticles made of PEG-polycaprolactam,^{130–133} PEG-poly(lactic-co-glycolate) (PEG-PLGA),¹³⁴ thermoresponsive hydrogels,¹³⁵ upconversion nanocrystals¹³⁶ and gold.¹³⁷ The diameter of these particles ranged from 7 to 200 nm thus further confirming the versatility of this shuttle. Moreover, the number of peptides required for efficient delivery is relatively low; four peptides are considered optimal for 7–8 nm dendrons¹²⁷ and 53 peptides on the surface of a 90 nm nanoparticle provided efficient transport.²⁵ The increase in brain delivery for most constructs is in the range of 1.5- to 3-fold in mice.

The vast majority of studies with Angiopep-2 describe about conjugates for the diagnosis^{125,136} or the treatment^{124,131,137} of brain tumours; in the second case a significant increase in survival with respect to the free drug or untargeted nanocarriers has often been reported. Although this peptide has been used mainly to transport small molecules and nanoparticles, conjugation of the shuttle to trastuzumab has recently been shown to enhance the therapeutic effect of this antibody in mice bearing HER2+ brain tumours.¹³⁸ Angiopep-2 also increases the anti-fungal activity of amphotericin B in meningoencephalitis^{139,140} and the therapeutic index of phenytoin sodium against epilepsy in rats.¹³⁵ Additionally, delivery of Angiopep-2-coated nanoparticles loaded with hGDNF boosts the neuroprotective effect of this protein in a Parkinsonian rat model, improving the locomotor activity and recovery of dopaminergic neurons.¹²⁹ Although in some of these models, especially in those involving tumours, the BBB may be compromised, permeation has also been assessed in healthy mice and cell-based BBB models.¹³⁵

4.2. GSH: a highly specialized shuttle

Together with Angiopep-2, GSH is the BBB shuttle peptide that has reached most advanced stages in the route towards clinical application. GSH has been mainly applied to target PEGylated nanoliposomes loaded with drugs, which are thereby protected

from degradation and clearance. This formulation, known as G-Technology[®], has been applied to a wide range of compounds, encompassing small molecules,^{141–143} peptides¹⁴⁴ and, very recently, biologics.¹⁴⁵

G-Technology[®] for doxorubicin delivery (2B3-101) has reached Phase I/IIa clinical trials for brain cancer treatment (ClinicalTrials.gov identifier: NCT01386580). This nanoplatform has also been exploited for the delivery of methylprednisolone (2B3-201), enhancing its transport up to 6.5-fold.¹⁴² 2B3-201 is capable of reducing neuroinflammation in rats with encephalomyelitis, and reached Phase I clinical trials for multiple sclerosis⁵⁵ (ClinicalTrials.gov identifier: NCT02048358). Even more remarkable is the selective increase in brain delivery of single domain antibodies against amyloid plaques in APP/PS1 mice.¹⁴⁵

5. One shuttle, one cargo

The large change in physicochemical properties induced by therapeutic cargoes and the distinct location of the targets of these drugs inside the brain has limited the universal aspiration of most BBB shuttles. Hence, in general, each peptide shuttle is prominent in the delivery of a particular family of cargoes. Although many peptides have been reported for each type of cargo, here we will focus on three well-documented case studies.

5.1. Gene delivery with RVG29

RVG29 was found when studying the neurotropism of the rabies virus, which is mediated by its glycoprotein (RVG).⁵³ Although a tail with 9 arginines was introduced to bind siRNA, the unmodified peptide was shown to reach the brain parenchyma. This observation suggested that the increase in brain delivery of the oligoarginine-RVG29 construct was due to the targeting peptide and not to the potential opening of tight junctions promoted by the polycationic sequence. This construct was first used to transport oligonucleotides into healthy mouse brains to silence GFP in GFP-transgenic mice as well as endogenous SOD1 in the CNS. As a further demonstration of its value, this delivery strategy was successfully applied to protect mice with JEV-induced encephalitis. The authors reasoned that transport across the BBB could take place by RMT through interaction with the $\alpha 7$ subunit of the nicotinic acetylcholine receptor (nAChR), as shown by the selective binding of the peptide to neurons and its competition with α -bungarotoxin. In a subsequent study,¹⁴⁶ RVG29 was intravenously injected into mice and was found inside cells that overexpress nAChR, unlike a scrambled version of the sequence. Moreover, RVG29 was not detected when administered to knockout animals devoid of this receptor.

The high potential of this sequence for gene delivery has been confirmed using either the oligoarginine tail,^{147,148} a polylysine dendrigraft,¹⁴⁹ polyethylenimine,^{150,151} polyasparthydrizide¹⁵¹ or polyamidoamide dendrimers¹⁵² to bind the oligonucleotide chains. RVG29 linked to exosomes is particularly efficient as it mediates higher protein knockdown than when linked to oligoarginine with five-fold less RNA.¹⁵³ As an example of therapeutic effect, the polylysine dendrigraft targeted delivery of caspase-3 RNAi reduced



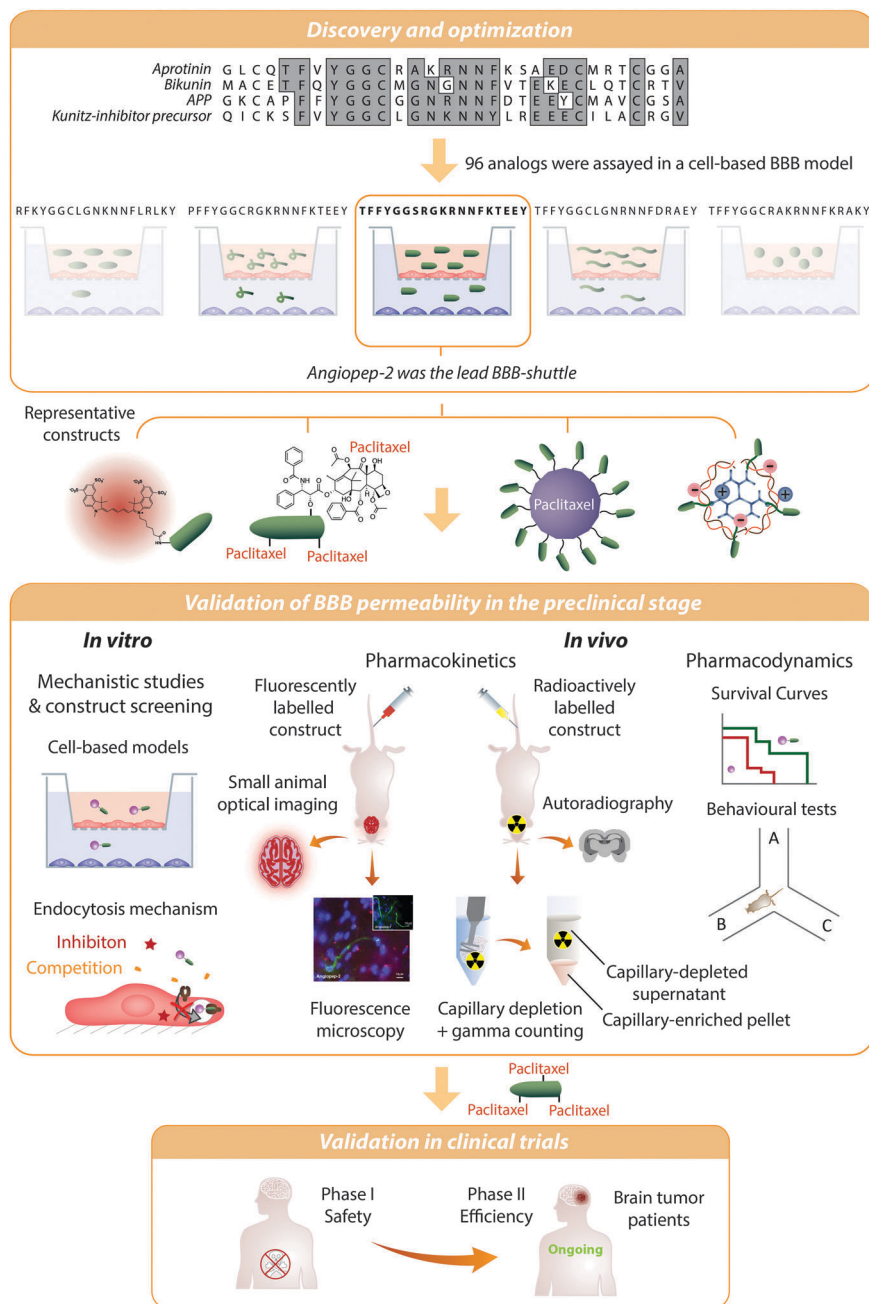


Fig. 3 Angiopep-2 discovery and validation as a BBB shuttle. Angiopep-2 was discovered by alignment of Kunitz protein domains followed by a screening of 96 analogues in a bovine cell-based blood–brain barrier model. The transport of the selected peptide was inhibited by low temperature, RAP protein, and α_2 -macroglobulin, thereby indicating that the transcytosis was active and probably mediated by LRP-1.¹¹³ However, several uptake mechanisms may contribute to the transport of conjugates.²⁵ This peptide has proven its efficiency as a BBB shuttle *in vitro* and *in vivo*. Whole-animal fluorescence imaging and autoradiography among other techniques have shown that Angiopep-2 constructs accumulate in the brain. Additionally, there is qualitative (e.g. microscopy. Image reproduced with permission from ref. 195. Copyright 2010 Elsevier) as well as quantitative (e.g. radioisotopic labelling and capillary depletion) evidence proving that this peptide reaches the brain parenchyma. Angiopep-2 conjugated to paclitaxel is currently in Phase II clinical trials.

caspase-3 levels, improving locomotor activity and rescuing dopaminergic neuronal loss in a rat model of Parkinson's disease.¹⁵⁴

5.2. Apolipoprotein-derived peptides for enzyme delivery

Apolipoproteins have been applied for brain delivery taking advantage of their roles in lipid transport. Although the whole

proteins have been used to transport nanoparticles,¹⁵⁵ the peptides derived from them have mainly been applied to enzymes, probably because of the lower effect of the targeting moiety on the enzymatic activity.

Apolipoprotein B100 (ApoB) is the primary component of low-density lipoprotein and it interacts with LDLR and LRP2.⁷⁹



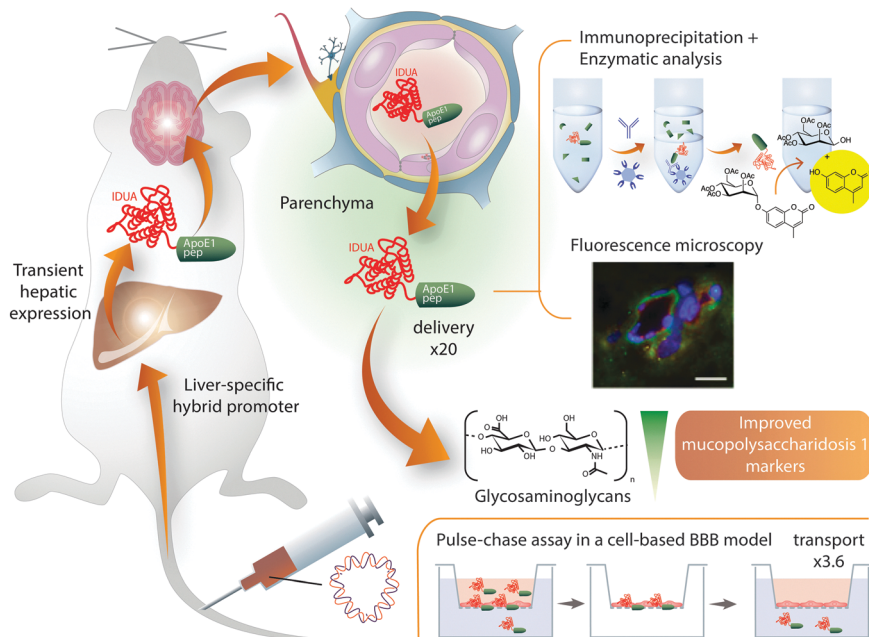


Fig. 4 Brain delivery of IDUA–ApoE (159–167)₂ expressed in the liver. Increased delivery of IDUA into the mouse brain parenchyma was achieved by expressing it as a fusion protein with ApoE (159–167)₂ in mouse liver.⁸² The fusion of ApoE peptide increased 3.6-fold the transport of IDUA in a pulse-chase assay performed on a cell-based BBB model – in this experiment the protein is incubated and, after washing, the transport of the protein adsorbed onto or internalized by endothelial cells is measured; this strategy intends to minimize the paracellular contribution to the transport, which is still a concern in cell-based BBB models.²¹ Remarkably, the amount of IDUA found in the brain when expressed in mice livers fused to ApoE (159–167)₂ was 20-fold higher than when expressed without the BBB shuttle. Quantification was performed after capillary-depletion. In addition, these authors provided fluorescence microscopy images, which indicate colocalization of the construct with neurons and astrocytes and show a clear pattern of diffusion from the capillary into the parenchyma. With this approach, 23% of normal brain IDUA activity was restored, which was sufficient to decrease brain glycosaminoglycan and β -hexosaminidase concentrations to normal levels in mucopolysaccharidosis type I mice.

Spencer and Verma⁸¹ achieved hepatic expression of proteins in the liver fused to the binding domain of ApoB (3371–3409, P04114 UniProt), which resulted in sustained brain delivery. The cDNA was introduced into the liver and spleen using a lentivirus vector. Through this strategy, GFP and glucocerebrosidase were delivered into the brain. Fluorescence microscopy revealed the co-localization of these molecules in neurons and astrocytes in the regions of the brain with a high expression of LDLR. Also using ApoB (3371–3409) to target iduronate-2-sulphatase, the overall brain pathology was improved in a mouse model of mucopolysaccharidosis type IIIA.¹⁵⁶

Despite the successful cases of ApoB BBB shuttles, peptides derived from ApoE showed higher efficiency when applied to transport other proteins.^{82,157} This observation may be related to ApoE binding a variety of LDLRs, including LRP1.⁷⁹ Of note, altering the homeostasis of this protein entails many secondary effects,⁹ which could be minimized using only the moiety involved in transport. With the aim of finding the most suitable fragment of ApoE, several sequences reported to interact with LDLRs^{158,159} were compared.⁸² In this comprehensive study, ApoE fragments were expressed as a fusion protein with a lysosomal enzyme (IDUA) in the liver in order to target this enzyme to the brain (Fig. 4).

One of the two peptides with the best performance found in the aforementioned study⁸² was the tandem dimer sequence ApoE (159–167)₂ (UniProt P02649-1) which had better transport

in a cell-based BBB model than the monomer.¹⁶⁰ This peptide was also the best-ranked BBB shuttle in a very recent comparative study¹⁵⁷ using a mouse model for another lysosomal storage disease, namely metachromic leukodystrophy. In that publication, ApoE (159–167)₂ did not show the best performance in endocytosis or in a porcine BBB cell-based model; however, it was the only shuttle to increase the *in vivo* brain delivery of arylsulphatase A (by 54%) when compared to Angiopep-2, ApoE (148–170), and ApoB. Surprisingly, this shuttle did not compete with the endogenous protein as suggested by the lack of increased transport efficiency in ApoE-knock out mice.

It is also worth highlighting the delivery of large proteins into the brain using a physical mixture of the cargoes with ApoE (151–170) in tandem with a 16-lysine sequence.¹⁶¹ However, this construct has subsequently been found to prompt a transient disruption of the BBB,¹⁶² as has been observed with other highly positively charged carriers.¹⁶³

5.3. g7-Mediated PLGA-nanoparticle delivery

Although Angiopep-2 has shown the highest versatility for nanoparticle delivery, many other shuttles have proven useful for certain kinds of nanocarriers. In particular, the g7 heptapeptide is a modified analogue of the synthetic opioid MMP-2200, in which the *N*-terminal tyrosine was exchanged for phenylalanine in order to avoid the antinociceptive effect.¹⁰⁰ It was also found that *O*-glycosylation with glucose but not xylose or lactose



favoured brain uptake. This observation, together with a remarkable selectivity for this organ and a poor permeation of a scrambled version of the peptide, indicates that transport across the BBB is due to a receptor, though not necessarily opioid.

Numerous *in vitro* and *in vivo* studies show that g7 is capable of delivering PEG-PLGA nanoparticles into the brain.¹⁶⁴ The increase in brain accumulation has been assessed by whole-animal fluorescence imaging¹⁶⁵ and also by measuring the amount of rhodamine 123 released from the nanoparticle; remarkably, 15% of the fluorophore administered intravenously was reported to reach the brain. These nanoparticles are mainly delivered to the grey matter¹⁶⁶ and their presence in the brain parenchyma has been imaged using fluorescence and transmission electron microscopy. Further evidence of the therapeutic effect of this peptide-coated nanocarriers upon intravenous injection would certainly encourage a more widespread use.

6. Rescuing the origins

In general, CPPs and peptides undergoing passive diffusion across the BBB do not provide brain selectivity. However, their high internalizing capacity can be fine-tuned or exploited in tandem with BBB shuttles in order to enhance the delivery of cargoes to the brain.

6.1. Cell-penetrating peptides

TAT is the most used CPP for brain delivery for proteins^{52,167–170} and nanoparticles.^{171–173} However, in addition to its lack of brain selectivity, very little qualitative or quantitative data are available regarding intact BBB penetration. Although some studies achieve a fast bulk brain accumulation and a few show an improved therapeutic effect, others indicate that the constructs could be trapped in brain endothelium. In this regard, one of the most perspicuous examples is the 800-fold increase in ritonavir delivery achieved two weeks after injection using TAT-coated polylactate nanoparticles.¹⁷⁴ As it could be expected from the cell-penetrating ability of TAT, nanoparticles are efficiently internalized in brain capillary endothelial cells, probably by adsorptive-mediated endocytosis, and are slowly released as indicated by the parenchyma/capillary ratio.

Nonetheless, recent studies have shown that dual-targeted liposomes with either Angiopep-2-oligoarginine, T7-TAT, THR-transportan or Tf combined with TAT, penetratin or mastoparan outperformed those with a single targeting peptide, both *in vitro* and *in vivo*.^{131,175–177} This strategy takes advantage of the penetrating capacity of CPPs by combining it with the higher selectivity of receptor ligands. It would certainly be interesting to study the effect of the double functionalization approach with more novel, potent and less toxic CPPs such as dNP2.¹⁷⁸

6.2. Passive diffusion shuttles

Passive diffusion BBB-shuttle peptides dramatically enhance the transport of drugs like baicalin, dopamine, 4-aminobutanoic acid, nipecotic acid and 5-aminolevulinic acid in a BBB cell-based model and in the parallel artificial membrane permeability

assay (PAMPA),^{30–32} which is a well-established method to measure passive diffusion.¹⁷⁹ With the aim of further enhancing the transport capacity of prolyl oligopeptidase inhibitors, diketopiperazines have been combined with a redox chemical delivery system to avoid back transport across brain endothelium.¹⁸⁰ Very recently, the chirality of phenylproline shuttle diastereomers has been shown to affect their permeability.⁶⁸ Thus, the transport of these peptides may depend on the phospholipid composition of biological membranes, thereby suggesting a route towards cell-type and even tissue selectivity. Furthermore, phenylprolines have overcome the low solubility limitations of their forerunners.

Despite the achievements described above, the applicability of BBB shuttle peptides that work through passive diffusion is still limited by the lack of selectivity and the non-negligible impact of the cargo on the efficiency of the shuttle and *vice versa*. On the side of the BBB transport capacity, this problem can be overcome by fine-tuning the peptide for each particular cargo.²⁹ In order to decrease the effect of the construct on the activity of the molecule, linkers that can be cleaved inside the brain parenchyma could be incorporated.

7. Toward protease-resistant shuttles

Most of the sequences reported for the delivery of large cargoes are linear and made of L-amino acid residues. Both of these features make peptides susceptible to degradation by proteases, a process that decreases their efficiency, especially *in vivo*. Notwithstanding, many strategies can be applied to overcome this limitation such as the use of non-natural amino acids, N-methylation, and cyclization.⁵⁰ Very recently, several publications have revealed the great potential of increasing the metabolic resistance of BBB shuttle peptides.

7.1. The *retro-enantio* approach

The *retro-enantio* or *retro-inverso* sequence of a peptide is obtained by changing the stereochemistry of all the amino acid residues (from L- to D-amino acids) and reversing the order of the sequence. In this way, the topochemical features and the structure of the peptide are often preserved despite the inversion of the amide bond, yielding highly protease-resistant analogues. It has recently been shown that this approach yields more efficient BBB shuttles.^{94,181,182} In order to illustrate this point, we will focus on the *retro-enantio* THR and ^DCDX peptides.

THR is a dodecapeptide obtained by phage display that interacts with TfR but does not compete with Tf.⁸⁸ This peptide shuttle enhances the *in vitro* and *in vivo* transport of gold NPs coated with a peptide (LPFFD) capable of binding amyloid- β in order to disrupt aggregates upon microwave irradiation.¹⁸³ TEM micrographs confirmed the presence of NPs in the parenchyma. Remarkably, it has recently been shown that the *retro-enantio* version of THR (THRre) transports a variety of cargoes with higher efficiency than the parent peptide in a cell-based BBB model and *in vivo*.¹⁸² Moreover, THRre was capable



of delivering quantum dots to the brain parenchyma as shown by two-photon intravital microscopy (Fig. 5).

In contrast to THR, ¹CDX is a peptide of natural origin. This linear fragment of snake neurotoxin candotoxin, which interacts with nAChRs, was reported following the success of RVG29.⁹³ ¹CDX increased nanoparticle accumulation in mouse brain and enhanced the survival of tumour-bearing mice. Recently, the *retro-enantio* analogue has been developed and shown to retain the capacity to interact with the same receptor.⁹⁴ Although the affinity of this analogue is 5-fold lower than the original peptide, its transport capacity is enhanced because of its superior resistance to proteases in serum and in the lysosome.

7.2. A venom-inspired peptidomimetic shuttle

The *retro-enantio* approach has proved highly efficient. However, this transformation decreases the affinity for the transporter and requires D-amino acids, thereby significantly raising production costs. Hence, it would be of interest to identify alternative sources of protease-resistant shuttles. Although the capacity of peptides found in animal venoms has already been exploited in this field, the relevance of preserving their knotted structure has been overlooked.

Apamin is a bicyclic neurotoxin that binds KCa2.2 channels, which are found in neural cells and the vasculature, and has long been known to reach the CNS.^{184,185} This peptide is highly

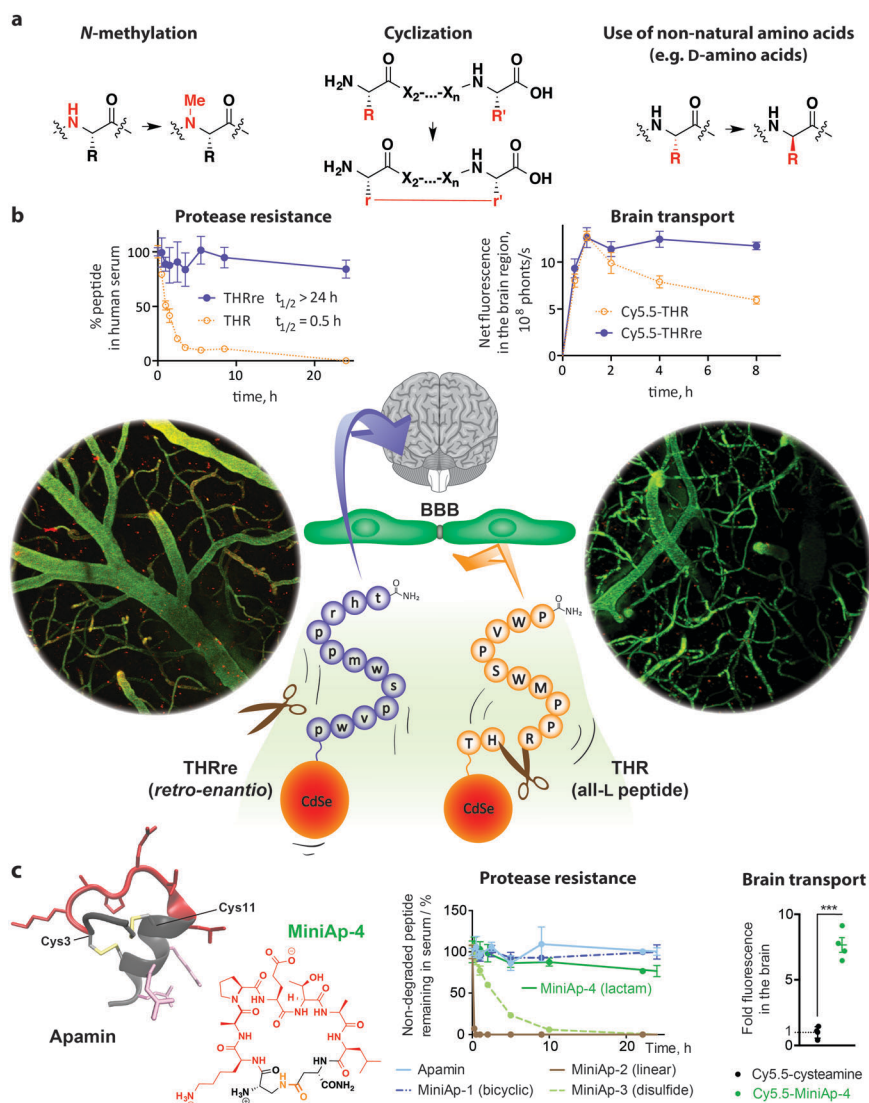


Fig. 5 Protease-resistance provides efficient BBB shuttle peptides. (a) Structural modifications to achieve protease-resistance that have been applied to BBB shuttle peptides. (b) The half-life of THR peptide is 30 min, whereas that of the *retro-enantio* version (THRre) is over 24 h.¹⁸² As a result, THRre transcytosed in a cell-based BBB model and accumulated in the brain more efficiently. Furthermore, this peptide was capable of delivering quantum dots across the BBB as shown by intracranial two-photon microscopy (bottom; capillaries are shown in green and quantum dots in red). The image is reproduced with permission from ref. 182. Copyright 2015 Wiley); this technique avoids artefacts introduced during perfusion, necropsy and tissue preparation for *ex vivo* microscopy imaging. (c) Cyclization of peptides also results in increased protease-resistance as illustrated in the case of apamin derivatives.⁹⁷ In addition, introducing non-natural elements such as substitution of a disulfide bond by a lactam bridge to produce peptidomimetics like MiniAp-4 further reinforces metabolic stability. MiniAp-4 enhanced the transport of a variety of cargoes in mice and in a human cell-based BBB model.



resistant to proteases⁹⁵ and is capable of targeting nanoparticles and proteins to the brain.⁹⁶ However, high toxicity and immunogenicity, as well as a relatively complex structure, have discouraged its extended application as a shuttle. Recently, MiniAp-4, which is a safer and minimized version of apamin cyclized through a lactam bridge, has been reported.⁹⁷ Importantly, this cyclic peptidomimetic preserves the high protease-resistance and brain targeting ability of apamin and has reduced toxicity and immunogenicity. MiniAp-4 is more permeable than the natural peptide and can transport nanoparticles and proteins in a human cell-based model of the BBB. Furthermore, this shuttle can carry a cargo across the BBB in mice and displays remarkable selectivity for the brain.

8. Conclusions and outlook

Although the BBB remains a formidable obstacle, since the Trojan horse concept was coined in the 1980s, the field of drug delivery to the brain has made remarkable progress. In the last few years, a plethora of new BBB shuttle peptides have emerged and hold great promise to overcome the limitations of the first generation of shuttles dominated by large proteins. Peptides are more affordable, easier to characterize and to link to nano-carriers or proteins. Moreover, they have lower immunogenicity and often have a reduced effect on the activity of the cargo than their larger counterparts. Furthermore, many peptide shuttles do not compete with endogenous substrates in contrast to endogenous proteins, nor stay bound to the receptor unlike some antibodies. BBB shuttle peptides have so far provided promising results in terms of brain delivery in preclinical settings. In addition, a relevant increase in the therapeutic effect has been proven in a wide variety of animal disease models, with a focus on brain tumours but also including neurodegenerative and lysosomal diseases as well as epilepsy among others.

Despite the considerable achievements described, new shuttles with higher transport capacity and selectivity are required. Approaches like phage display and natural sources of peptides that reach the CNS offer an excellent opportunity to explore the multitude of poorly characterized or still unknown routes into the brain. These strategies should be complemented with additional efforts in the characterization of the transport mechanisms and in global proteomic approaches to identify new receptors. Also, further comparative studies between shuttles and a more accurate quantification of the free drug in the brain parenchyma would enable a more efficient identification and optimization of BBB shuttles. The next generation of BBB shuttle peptides should aim for an enhanced metabolic stability, a higher transendothelial transport and an improved selectivity for the brain – even for particular regions of this organ – possibly through yet uncharacterized transcytotic pathways.

Acknowledgements

IRB Barcelona is the recipient of a Severo Ochoa Award of Excellence from MINECO (Government of Spain). We appreciate financial support from MINECO-FEDER (Bio2013-40716-R and

CTQ2013-49462-EXP), MINECO (PCIN-2015-051 Cure2DIPG), RecerCaixa-2014-Gate2Brain, Generalitat de Catalunya (XRB and 2014-SGR-521), FARA and GENEFA. B.O.-S. and M.S.-N. are grateful for “La Caixa”/IRB Barcelona and Juan de la Cierva fellowships, respectively.

References

- 1 C. J. L. Murray, T. Vos, R. Lozano, M. Naghavi, A. D. Flaxman, C. Michaud, M. Ezzati, K. Shibuya, J. A. Salomon and S. Abdalla, *et al.*, *Lancet*, 2012, **380**, 2197–2223.
- 2 A. Gustavsson, M. Svensson, F. Jacobi, C. Allgulander, J. Alonso, E. Beghi, R. Dodel, M. Ekman, C. Faravelli and L. Fratiglioni, *et al.*, *Eur. Neuropsychopharmacol.*, 2011, **21**, 718–779.
- 3 W. M. Pardridge, *J. Cereb. Blood Flow Metab.*, 2012, **32**, 1959–1972.
- 4 M. Malakoutikhah, M. Teixidó and E. Giralt, *Angew. Chem., Int. Ed.*, 2011, **50**, 7998–8014.
- 5 W. M. Pardridge, *Nat. Rev. Drug Discovery*, 2002, **1**, 131–139.
- 6 N. J. Abbott, A. A. Patabendige, D. E. Dolman, S. R. Yusof and D. J. Begley, *Neurobiol. Dis.*, 2010, **37**, 13–25.
- 7 R. Daneman and A. Prat, *Cold Spring Harbor Perspect. Biol.*, 2015, **7**, a020412.
- 8 Z. Zhao, A. R. Nelson, C. Betsholtz and B. V. Zlokovic, *Cell*, 2015, **163**, 1064–1078.
- 9 K. Nagpal, S. K. Singh and D. N. Mishra, *Expert Opin. Drug Delivery*, 2013, **10**, 927–955.
- 10 S. Mitragotri, P. A. Burke and R. Langer, *Nat. Rev. Drug Discovery*, 2014, **13**, 655–672.
- 11 A. G. de Boer and P. J. Gaillard, *Annu. Rev. Pharmacol. Toxicol.*, 2007, **47**, 323–355.
- 12 C. T. Lu, Y. Z. Zhao, H. L. Wong, J. Cai, L. Peng and X. Q. Tian, *Int. J. Nanomed.*, 2014, **9**, 2241–2257.
- 13 D. J. Wolak and R. G. Thorne, *Mol. Pharmaceutics*, 2013, **10**, 1492–1504.
- 14 B. Obermeier, R. Daneman and R. M. Ransohoff, *Nat. Med.*, 2013, **19**, 1584–1596.
- 15 J. Lichota, T. Skjørringe, L. B. Thomsen and T. Moos, *J. Neurochem.*, 2010, **113**, 1–13.
- 16 J. Rautio, K. Laine, M. Gynther and J. Savolainen, *AAPS J.*, 2008, **10**, 92–102.
- 17 T. B. Lentz, S. J. Gray and R. J. Samulski, *Neurobiol. Dis.*, 2012, **48**, 179–188.
- 18 G. F. Woodworth, G. P. Dunn, E. A. Nance, J. Hanes and H. Brem, *Front. Oncol.*, 2014, **4**, 126.
- 19 E. V. Batrakova, H. E. Gendelman and A. V. Kabanov, *Expert Opin. Drug Delivery*, 2011, **8**, 415–433.
- 20 M. Bourdenx, N. Duthel, E. Bezar and B. Dehay, *Front. Mol. Neurosci.*, 2014, **7**, 1–8.
- 21 J. Niewoehner, B. Bohrmann, L. Collin, E. Urich, H. Sade, P. Maier, P. Rueger, J. O. Stracke, W. Lau, A. C. Tissot, H. Loetscher, A. Ghosh and P. O. Freskgård, *Neuron*, 2014, **81**, 49–60.
- 22 H. Liu, W. Zhang, L. Ma, L. Fan, F. Gao, J. Ni and R. Wang, *Int. J. Pharm.*, 2014, **476**, 1–8.



- 23 B. Zhang, X. Sun, H. Mei, Y. Wang, Z. Liao, J. Chen, Q. Zhang, Y. Hu, Z. Pang and X. Jiang, *Biomaterials*, 2013, **34**, 9171–9182.
- 24 D. Guarnieri, A. Falanga, O. Muscetti, R. Tarallo, S. Fusco, M. Galdiero, S. Galdiero and P. A. Netti, *Small*, 2013, **9**, 853–862.
- 25 H. Xin, X. Sha, X. Jiang, L. Chen, K. Law, J. Gu, Y. Chen, X. Wang and X. Fang, *Biomaterials*, 2012, **33**, 1673–1681.
- 26 R. Prades, M. Teixidó and E. Giralt, in *Nanostructured Biomaterials for Overcoming Biological Barriers*, ed. M. J. Alonso and N. S. Csaba, Royal Soc Chemistry, Cambridge, 2012, pp. 364–391.
- 27 J. D. Malcor, N. Payrot, M. David, A. Faucon, K. Abouzid, G. Jacquot, N. Floquet, F. Debarbieux, G. Rougon, J. Martinez, M. Khrestchatsky, P. Vlieghe and V. Lisowski, *J. Med. Chem.*, 2012, **55**, 2227–2241.
- 28 F. C. Thomas, K. Taskar, V. Rudraraju, S. Goda, H. R. Thorsheim, J. A. Gaasch, R. K. Mittapalli, D. Palmieri, P. S. Steeg, P. R. Lockman and Q. R. Smith, *Pharm. Res.*, 2009, **26**, 2486–2494.
- 29 M. Malakoutikhah, B. Guixer, P. Arranz-Gibert, M. Teixidó and E. Giralt, *ChemMedChem*, 2014, **9**, 1594–1601.
- 30 M. Malakoutikhah, R. Prades, M. Teixidó and E. Giralt, *J. Med. Chem.*, 2010, **53**, 2354–2363.
- 31 M. Malakoutikhah, M. Teixidó and E. Giralt, *J. Med. Chem.*, 2008, **51**, 4881–4889.
- 32 M. Teixidó, E. Zurita, M. Malakoutikhah, T. Tarragó and E. Giralt, *J. Am. Chem. Soc.*, 2007, **129**, 11802–11813.
- 33 E. Soddu, G. Rassu, P. Giunchedi, B. Sarmento and E. Gavini, *Eur. J. Pharm. Sci.*, 2015, **74**, 63–76.
- 34 W. M. Pardridge, *Endocr. Rev.*, 1986, **7**, 314–330.
- 35 A. K. Kumagai, J. B. Eisenberg and W. M. Pardridge, *J. Biol. Chem.*, 1987, **262**, 15214–15219.
- 36 P. M. Friden, L. R. Walus, G. F. Musso, M. A. Taylor, B. Malfroy and R. M. Starzyk, *Proc. Natl. Acad. Sci. U. S. A.*, 1991, **88**, 4771–4775.
- 37 Y. J. Yu, Y. Zhang, M. Kenrick, K. Hoyte, W. Luk, Y. Lu, J. Atwal, J. M. Elliott, S. Prabhu, R. J. Watts and M. S. Dennis, *Sci. Transl. Med.*, 2011, **3**, 84ra44.
- 38 J. Kreuter, T. Hekmatara, S. Dreis, T. Vogel, S. Gelperina and K. Langer, *J. Controlled Release*, 2007, **118**, 54–58.
- 39 W. Pan, A. J. Kastin, T. C. Zankel, P. van Kerkhof, T. Terasaki and G. Bu, *J. Cell Sci.*, 2004, **117**, 5071–5078.
- 40 Z. M. Qian, H. Li, H. Sun and K. Ho, *Pharmacol. Rev.*, 2002, **54**, 561–587.
- 41 C. Fillebeen, L. Descamps, M. P. Dehouck, L. Fenart, M. Benaissa, G. Spik, R. Cecchelli and A. Pierce, *J. Biol. Chem.*, 1999, **274**, 7011–7017.
- 42 M. Demeule, J. Poirier, J. Jodoin, Y. Bertrand, R. R. Desrosiers, C. Dagenais, T. Nguyen, J. Lanthier, R. Gabathuler, M. Kennard, W. A. Jefferies, D. Karkan, S. Tsai, L. Fenart, R. Cecchelli and R. Beliveau, *J. Neurochem.*, 2002, **83**, 924–933.
- 43 W. A. Banks, A. J. Kastin, W. Huang, J. B. Jaspán and L. M. Maness, *Peptides*, 1996, **17**, 305–311.
- 44 W. A. Banks and R. D. Broadwell, *J. Neurochem.*, 1994, **62**, 2404–2419.
- 45 P. J. Gaillard, A. Brink and A. G. de Boer, *Int. Cong. Ser.*, 2005, **1277**, 185–198.
- 46 Y. J. Yu, J. K. Atwal, Y. Zhang, R. K. Tong, K. R. Wildsmith, C. Tan, N. Bien-Ly, M. Hersom, J. A. Maloney and W. J. Meilandt, *et al.*, *Sci. Transl. Med.*, 2014, **6**, 261ra154.
- 47 A. Abulrob, J. Zhang, J. Tanha, R. MacKenzie and D. Stanimirovic, *Int. Congr. Ser.*, 2005, **1277**, 212–223.
- 48 T. Uhlig, T. Kyprianou, F. G. Martinelli, C. A. Oppici, D. Heiligers, D. Hills, X. R. Calvo and P. Verhaert, *EuPa Open Proteomics*, 2014, **4**, 58–69.
- 49 C. J. Camacho, Y. Katsumata and D. P. Ascherman, *PLoS Comput. Biol.*, 2008, **4**, e1000231.
- 50 I. R. Kumarasinghe and V. J. Hruby, in *Peptide chemistry and drug design*, ed. B. M. Dunn, Wiley, 2015, pp. 247–264.
- 51 W. A. Banks, *Peptides*, 2015, **72**, 16–19.
- 52 S. R. Schwarze, A. Ho, A. Vocero-Akbani and S. F. Dowdy, *Science*, 1999, **285**, 1569–1572.
- 53 P. Kumar, H. Wu, J. L. McBride, K. E. Jung, M. H. Kim, B. L. Davidson, S. K. Lee, P. Shankar and N. Manjunath, *Nature*, 2007, **448**, 39–43.
- 54 M. Demeule, A. Régina, C. Ché, J. Poirier, T. Nguyen, R. Gabathuler, J. P. Castaigne and R. Béliveau, *J. Pharmacol. Exp. Ther.*, 2008, **324**, 1064–1072.
- 55 P. J. Gaillard, C. C. Appeldoorn, J. Rip, R. Dorland, S. M. van der Pol, G. Kooij, H. E. de Vries and A. Reijerkerk, *J. Controlled Release*, 2012, **164**, 364–369.
- 56 H. F. Liang, Y. C. Chen, T. F. Yang, L. W. Chang, A. J. Wang, J. M. Lu, C. H. Jian, Y. F. Lin and S. J. Liu, US7704956 B2, 2010.
- 57 F. Hervé, N. Ghinea and J. M. Scherrmann, *AAPS J.*, 2008, **10**, 455–472.
- 58 I. Martín, M. Teixidó and E. Giralt, *Curr. Pharm. Des.*, 2013, **19**, 2924–2942.
- 59 F. Madani, S. Lindberg, Ü. Langel, S. Futaki and A. Gräslund, *J. Biophys.*, 2011, **2011**, 414729.
- 60 A. D. Frankel and C. O. Pabo, *Cell*, 1988, **55**, 1189–1193.
- 61 A. Joliot, C. Pernelle, H. Deagostini-Bazin and A. Prochiantz, *Proc. Natl. Acad. Sci. U. S. A.*, 1991, **88**, 1864–1868.
- 62 C. Rousselle, P. Clair, J. M. Lefauconnier, M. Kaczorek, J. M. Scherrmann and J. Tamsamani, *Mol. Pharmacol.*, 2000, **57**, 679–686.
- 63 G. Drin, S. Cottin, E. Blanc, A. R. Rees and J. Tamsamani, *J. Biol. Chem.*, 2003, **278**, 31192–31201.
- 64 M. Pooga, M. Hallbrink, M. Zorko and U. Langel, *FASEB J.*, 1998, **12**, 67–77.
- 65 S. Futaki, T. Suzuki, W. Ohashi, T. Yagami, S. Tanaka, K. Ueda and Y. Sugiura, *J. Biol. Chem.*, 2001, **276**, 5836–5840.
- 66 S. Stalmans, N. Bracke, E. Wynendaele, B. Gevaert, K. Peremans, C. Burvenich, I. Polis and B. De Spiegeleer, *PLoS One*, 2015, **10**, e0139652.
- 67 E. G. Chikhale, K. Y. Ng, P. S. Burton and R. T. Borchardt, *Pharm. Res.*, 1994, **11**, 412–419.
- 68 P. Arranz-Gibert, B. Guixer, M. Malakoutikhah, M. Muttenthaler, F. Guzmán, M. Teixidó and E. Giralt, *J. Am. Chem. Soc.*, 2015, **137**, 7357–7364.
- 69 C. Zhan, C. Li, X. Wei, W. Lu and W. Lu, *Adv. Drug Delivery Rev.*, 2015, **90**, 101–118.



- 70 G. P. Smith, *Science*, 1985, **228**, 1315–1317.
- 71 S. Cabilly, *Mol. Biotechnol.*, 1999, **12**, 143–148.
- 72 K. K. Jain, *Drug Delivery in Central Nervous System Disorders – Technologies, Markets and Companies*, Jain PharmaBiotech, Basel, Switzerland, 2012.
- 73 B. Dehouck, L. Fenart, M. P. Dehouck, A. Pierce, G. Torpier and R. Cecchelli, *J. Cell Biol.*, 1997, **138**, 877–889.
- 74 K. R. Duffy and W. M. Pardridge, *Brain Res.*, 1987, **420**, 32–38.
- 75 H. Tu, W. Pan, L. Feucht and A. J. Kastin, *J. Cell. Physiol.*, 2007, **212**, 215–222.
- 76 G. L. Barrett, J. Trieu and T. Naim, *Regul. Pept.*, 2009, **155**, 55–61.
- 77 Y. Liu, J. Li, K. Shao, R. Huang, L. Ye, J. Lou and C. Jiang, *Biomaterials*, 2010, **31**, 5246–5257.
- 78 A. P. Sagare, R. Deane and B. V. Zlokovic, *Pharmacol. Ther.*, 2012, **136**, 94–105.
- 79 N. S. Chung and K. M. Wasan, *Adv. Drug Delivery Rev.*, 2004, **56**, 1315–1334.
- 80 M. Notarnicola, M. Linsalata, M. Caruso, A. Cavallini and A. Di Leo, *J. Gastroenterol.*, 1995, **30**, 705–709.
- 81 B. J. Spencer and I. M. Verma, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 7594–7599.
- 82 D. Wang, S. S. El-Amouri, M. Dai, C. Y. Kuan, D. Y. Hui, R. O. Brady and D. Pan, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 2999–3004.
- 83 Y. Uchida, S. Ohtsuki, Y. Katsukura, C. Ikeda, T. Suzuki, J. Kamiie and T. Terasaki, *J. Neurochem.*, 2011, **117**, 333–345.
- 84 R. Daneman, L. Zhou, D. Agalliu, J. D. Cahoy, A. Kaushal and B. A. Barres, *PLoS One*, 2010, **5**, e13741.
- 85 Y. Uchida, M. Tachikawa, W. Obuchi, Y. Hoshi, Y. Tomioka, S. Ohtsuki and T. Terasaki, *Fluids Barriers CNS*, 2013, **10**, 21.
- 86 A. R. Jones and E. V. Shusta, *Pharm. Res.*, 2007, **24**, 1759–1771.
- 87 H. Xia, B. Anderson, Q. Mao and B. L. Davidson, *J. Virol.*, 2000, **74**, 11359–11366.
- 88 J. H. Lee, J. A. Engler, J. F. Collawn and B. A. Moore, *Eur. J. Biochem.*, 2001, **268**, 2004–2012.
- 89 F. I. Staquicini, M. G. Ozawa, C. A. Moya, W. H. Driessen, E. M. Barbu, H. Nishimori, S. Soghomonyan, L. G. Flores, 2nd, X. Liang and V. Paolillo, *et al.*, *J. Clin. Invest.*, 2011, **121**, 161–173.
- 90 L. Xiang, R. Zhou, A. Fu, X. Xu, Y. Huang and C. Hu, *J. Drug Targeting*, 2011, **19**, 632–636.
- 91 A. Fu, Y. Wang, L. Zhan and R. Zhou, *Pharm. Res.*, 2012, **29**, 1562–1569.
- 92 C. Zhan, Z. Yan, C. Xie and W. Lu, *Mol. Pharmaceutics*, 2010, **7**, 1940–1947.
- 93 C. Zhan, B. Li, L. Hu, X. Wei, L. Feng, W. Fu and W. Lu, *Angew. Chem., Int. Ed.*, 2011, **50**, 5482–5485.
- 94 X. Wei, C. Zhan, Q. Shen, W. Fu, C. Xie, J. Gao, C. Peng, P. Zheng and W. Lu, *Angew. Chem., Int. Ed.*, 2015, **54**, 3023–3027.
- 95 B. Oller-Salvia, M. Teixidó and E. Giralt, *Biopolymers*, 2013, **100**, 675–686.
- 96 J. Wu, H. Jiang, Q. Bi, Q. Luo, J. Li, Y. Zhang, Z. Chen and C. Li, *Mol. Pharmaceutics*, 2014, **11**, 3210–3222.
- 97 B. Oller-Salvia, M. Sánchez-Navarro, S. Ciudad, S. Guiu, P. Arranz-Gibert, C. Garcia, R. Gomis, R. Cecchelli, J. Garcia, E. Giralt and M. Teixidó, *Angew. Chem., Int. Ed.*, 2016, **55**, 572–575.
- 98 J. Rip, L. Chen, R. Hartman, A. van den Heuvel, A. Reijerkerk, J. van Kregten, B. van der Boom, C. Appeldoorn, M. de Boer, D. Maussang, E. C. de Lange and P. J. Gaillard, *J. Drug Targeting*, 2014, **22**, 460–467.
- 99 A. K. Bachhawat, A. Thakur, J. Kaur and M. Zulkifli, *Biochim. Biophys. Acta*, 2013, **1830**, 3154–3164.
- 100 L. Costantino, F. Gandolfi, G. Tosi, F. Rivasi, M. A. Vandelli and F. Forni, *J. Controlled Release*, 2005, **108**, 84–96.
- 101 K. N. Sugahara, T. Teesalu, P. P. Karmali, V. R. Kotamraju, L. Agemy, D. R. Greenwald and E. Ruoslahti, *Science*, 2010, **328**, 1031–1035.
- 102 D. Hauenberger, J. Klominek and K. G. Sundqvist, *J. Immunol.*, 1994, **153**, 960–971.
- 103 Y. Miura, T. Takenaka, K. Toh, S. Wu, H. Nishihara, M. R. Kano, Y. Ino, T. Nomoto, Y. Matsumoto, H. Koyama, H. Cabral, N. Nishiyama and K. Kataoka, *ACS Nano*, 2013, **7**, 8583–8592.
- 104 D. E. Saslowsky, Y. M. te Welscher, D. J. Chinnapen, J. S. Wagner, J. Wan, E. Kern and W. I. Lencer, *J. Biol. Chem.*, 2013, **288**, 25804–25809.
- 105 G. Tettamanti, R. Bassi, P. Viani and L. Riboni, *Biochimie*, 2003, **85**, 423–437.
- 106 J. K. Liu, Q. Teng, M. Garrity-Moses, T. Federici, D. Tanase, M. J. Imperiale and N. M. Boulis, *Neurobiol. Dis.*, 2005, **19**, 407–418.
- 107 J. V. Georgieva, R. P. Brinkhuis, K. Stojanov, C. A. Weijers, H. Zuillhof, F. P. Rutjes, D. Hoekstra, J. C. van Hest and I. S. Zuhorn, *Angew. Chem., Int. Ed.*, 2012, **51**, 8339–8342.
- 108 K. Stojanov, J. V. Georgieva, R. P. Brinkhuis, J. C. van Hest, F. P. Rutjes, R. A. Dierckx, E. F. de Vries and I. S. Zuhorn, *Mol. Pharmaceutics*, 2012, **9**, 1620–1627.
- 109 J. Li, L. Feng, L. Fan, Y. Zha, L. Guo, Q. Zhang, J. Chen, Z. Pang, Y. Wang, X. Jiang, V. C. Yang and L. Wen, *Biomaterials*, 2011, **32**, 4943–4950.
- 110 H. Gao, J. Qian, S. Cao, Z. Yang, Z. Pang, S. Pan, L. Fan, Z. Xi, X. Jiang and Q. Zhang, *Biomaterials*, 2012, **33**, 5115–5123.
- 111 C. Zhang, X. Wan, X. Zheng, X. Shao, Q. Liu, Q. Zhang and Y. Qian, *Biomaterials*, 2014, **35**, 456–465.
- 112 T. Vitalis and R. Gabathuler, WO2014160438 A1, 2014.
- 113 M. Demeule, J. C. Currie, Y. Bertrand, C. Ché, T. Nguyen, A. Régina, R. Gabathuler, J. P. Castaigne and R. Béliveau, *J. Neurochem.*, 2008, **106**, 1534–1544.
- 114 C. Ché, G. Yang, C. Thiot, M. C. Lacoste, J. C. Currie, M. Demeule, A. Régina, R. Béliveau and J. P. Castaigne, *J. Med. Chem.*, 2010, **53**, 2814–2824.
- 115 A. Régina, M. Demeule, C. Ché, I. Lavallée, J. Poirier, R. Gabathuler, R. Béliveau and J. P. Castaigne, *Br. J. Pharmacol.*, 2008, **155**, 185–197.
- 116 M. Demeule, N. Beaudet, A. Reégina, É. Besserer-Offroy, A. Murza, P. Tétreault, K. Belleville, C. Ché, A. Larocque,



- C. Thiot, R. Béliveau, J. M. Longpré, É. Marsault, R. Leduc, J. E. Lachowicz, S. L. Gonias, J. P. Castaigne and P. Sarret, *J. Clin. Invest.*, 2014, **124**, 1199–1213.
- 117 Y. Bertrand, J. C. Currie, J. Poirier, M. Demeule, A. Abulrob, D. Fatehi, D. Stanimirovic, H. Sartelet, J. P. Castaigne and R. Béliveau, *Br. J. Cancer*, 2011, **105**, 1697–1707.
- 118 R. Gabathuler, in *Drug Delivery to the Central Nervous System*, ed. K. K. Jain, Humana Press, 2010, vol. 45, pp. 249–260.
- 119 J. Drappatz, A. Brenner, E. T. Wong, A. Eichler, D. Schiff, M. D. Groves, T. Mikkelsen, S. Rosenfeld, J. Sarantopoulos, C. A. Meyers, R. M. Fielding, K. Elian, X. Wang, B. Lawrence, M. Shing, S. Kelsey, J. P. Castaigne and P. Y. Wen, *Clin. Cancer Res.*, 2013, **19**, 1567–1576.
- 120 R. Kurzrock, N. Gabrail, C. Chandhasin, S. Moulder, C. Smith, A. Brenner, K. Sankhala, A. Mita, K. Elian, D. Bouchard and J. Sarantopoulos, *Mol. Cancer Ther.*, 2012, **11**, 308–316.
- 121 S. E. Bates, M. L. Lindenberg, C. Bryla, M. E. Burotto Pichun, N. Patronas, E. Mena Gonzalez, L. Amiri-Kordestani, T. Fojo, S. Balasubramaniam and P. L. Choyke, *J. Clin. Oncol.*, 2015, **33**(suppl. 15), 2552.
- 122 Z. Z. Yang, J. Q. Li, Z. Z. Wang, D. W. Dong and X. R. Qi, *Biomaterials*, 2014, **35**, 5226–5239.
- 123 X. Sun, Z. Pang, H. Ye, B. Qiu, L. Guo, J. Li, J. Ren, Y. Qian, Q. Zhang, J. Chen and X. Jiang, *Biomaterials*, 2012, **33**, 916–924.
- 124 J. Ren, S. Shen, D. Wang, Z. Xi, L. Guo, Z. Pang, Y. Qian, X. Sun and X. Jiang, *Biomaterials*, 2012, **33**, 3324–3333.
- 125 H. Yan, L. Wang, J. Wang, X. Weng, H. Lei, X. Wang, L. Jiang, J. Zhu, W. Lu, X. Wei and C. Li, *ACS Nano*, 2012, **6**, 410–420.
- 126 H. Yan, J. Wang, P. Yi, H. Lei, C. Zhan, C. Xie, L. Feng, J. Qian, J. Zhu, W. Lu and C. Li, *Chem. Commun.*, 2011, **47**, 8130–8132.
- 127 X. Gao, J. Qian, S. Zheng, Y. Xiong, J. Man, B. Cao, L. Wang, S. Ju and C. Li, *Pharm. Res.*, 2013, **30**, 2538–2548.
- 128 S. Huang, J. Li, L. Han, S. Liu, H. Ma, R. Huang and C. Jiang, *Biomaterials*, 2011, **32**, 6832–6838.
- 129 R. Huang, H. Ma, Y. Guo, S. Liu, Y. Kuang, K. Shao, J. Li, Y. Liu, L. Han, S. Huang, S. An, L. Ye, J. Lou and C. Jiang, *Pharm. Res.*, 2013, **30**, 2549–2559.
- 130 G. Huile, P. Shuaiqi, Y. Zhi, C. Shijie, C. Chen, J. Xinguo, S. Shun, P. Zhiqing and H. Yu, *Biomaterials*, 2011, **32**, 8669–8675.
- 131 H. Gao, S. Zhang, S. Cao, Z. Yang, Z. Pang and X. Jiang, *Mol. Pharmaceutics*, 2014, **11**, 2755–2763.
- 132 H. Xin, X. Jiang, J. Gu, X. Sha, L. Chen, K. Law, Y. Chen, X. Wang, Y. Jiang and X. Fang, *Biomaterials*, 2011, **32**, 4293–4305.
- 133 H. Xin, X. Sha, X. Jiang, W. Zhang, L. Chen and X. Fang, *Biomaterials*, 2012, **33**, 8167–8176.
- 134 J. Shen, C. Zhan, C. Xie, Q. Meng, B. Gu, C. Li, Y. Zhang and W. Lu, *J. Drug Targeting*, 2011, **19**, 197–203.
- 135 X. Ying, Y. Wang, J. Liang, J. Yue, C. Xu, L. Lu, Z. Xu, J. Gao, Y. Du and Z. Chen, *Angew. Chem., Int. Ed.*, 2014, **53**, 12436–12440.
- 136 D. Ni, J. Zhang, W. Bu, H. Xing, F. Han, Q. Xiao, Z. Yao, F. Chen, Q. He, J. Liu, S. Zhang, W. Fan, L. Zhou, W. Peng and J. Shi, *ACS Nano*, 2014, **8**, 1231–1242.
- 137 S. Ruan, M. Yuan, L. Zhang, G. Hu, J. Chen, X. Cun, Q. Zhang, Y. Yang, Q. He and H. Gao, *Biomaterials*, 2015, **37**, 425–435.
- 138 A. Regina, M. Demeule, S. Tripathy, S. Lord-Dufour, J. C. Currie, M. Iddir, B. Annabi, J. P. Castaigne and J. E. Lachowicz, *Mol. Cancer Ther.*, 2015, **14**, 129–140.
- 139 K. Shao, J. Wu, Z. Chen, S. Huang, J. Li, L. Ye, J. Lou, L. Zhu and C. Jiang, *Biomaterials*, 2012, **33**, 6898–6907.
- 140 K. Shao, R. Huang, J. Li, L. Han, L. Ye, J. Lou and C. Jiang, *J. Controlled Release*, 2010, **147**, 118–126.
- 141 A. Mdzinarishvili, V. Sutariya, P. K. Talasila, W. J. Geldenhuys and P. Sadana, *Drug Delivery Transl. Res.*, 2013, **3**, 309–317.
- 142 D. H. Lee, C. Rötger, C. C. Appeldoorn, A. Reijerkerk, W. Gladdines, P. J. Gaillard and R. A. Linker, *J. Neuroimmunol.*, 2014, **274**, 96–101.
- 143 W. Geldenhuys, T. Mbimba, T. Bui, K. Harrison and V. Sutariya, *J. Drug Targeting*, 2011, **19**, 837–845.
- 144 A. Lindqvist, J. Rip, P. J. Gaillard, S. Björkman and M. Hammarlund-Udenaes, *Mol. Pharmaceutics*, 2013, **10**, 1533–1541.
- 145 M. Rotman, M. M. Welling, A. Bunschoten, M. E. de Backer, J. Rip, R. J. Nabuurs, P. J. Gaillard, M. A. van Buchem, S. M. van der Maarel and L. van der Weerd, *J. Controlled Release*, 2015, **203**, 40–50.
- 146 S. S. Kim, C. Ye, P. Kumar, I. Chiu, S. Subramanya, H. Wu, P. Shankar and N. Manjunath, *Mol. Ther.*, 2010, **18**, 993–1001.
- 147 C. Gong, X. Li, L. Xu and Y. H. Zhang, *Biomaterials*, 2012, **33**, 3456–3463.
- 148 S. Zadran, G. Akopian, H. Zadran, J. Walsh and M. Baudry, *NeuroMol. Med.*, 2013, **15**, 74–81.
- 149 Y. Liu, Y. Hu, Y. Guo, H. Ma, J. Li and C. Jiang, *J. Controlled Release*, 2012, **163**, 203–210.
- 150 W. Hwang do, S. Son, J. Jang, H. Youn, S. Lee, D. Lee, Y. S. Lee, J. M. Jeong, W. J. Kim and D. S. Lee, *Biomaterials*, 2011, **32**, 4968–4975.
- 151 H. Huo, Y. Gao, Y. Wang, J. Zhang, Z. Y. Wang, T. Jiang and S. Wang, *J. Colloid Interface Sci.*, 2015, **447**, 8–15.
- 152 Y. Liu, R. Huang, L. Han, W. Ke, K. Shao, L. Ye, J. Lou and C. Jiang, *Biomaterials*, 2009, **30**, 4195–4202.
- 153 L. Alvarez-Erviti, Y. Seow, H. Yin, C. Betts, S. Lakhali and M. J. A. Wood, *Nat. Biotechnol.*, 2011, **29**, 341–345.
- 154 Y. Liu, Y. Guo, S. An, Y. Kuang, X. He, H. Ma, J. Li, J. Lu, N. Zhang and C. Jiang, *PLoS One*, 2013, **8**, e62905.
- 155 H. L. Wong, X. Y. Wu and R. Bendayan, *Adv. Drug Delivery Rev.*, 2012, **64**, 686–700.
- 156 N. C. Sorrentino, L. D'Orsi, I. Sambri, E. Nusco, C. Monaco, C. Spanpanato, E. Polishchuk, P. Saccone, E. De Leonibus, A. Ballabio and A. Fraldi, *EMBO Mol. Med.*, 2013, **5**, 675–690.
- 157 A. Böckenhoff, S. Cramer, P. Wölte, S. Knieling, C. Wohlenberg, V. Gieselmann, H. J. Galla and U. Matzner, *J. Neurosci.*, 2014, **34**, 3122–3129.
- 158 C. A. Dyer, D. P. Cistola, G. C. Parry and L. K. Curtiss, *J. Lipid Res.*, 1995, **36**, 80–88.



- 159 D. Clayton, I. M. Brereton, P. A. Kroon and R. Smith, *Protein Sci.*, 1999, **8**, 1797–1805.
- 160 F. Re, I. Cambianica, C. Zona, S. Sesana, M. Gregori, R. Rigolio, B. La Ferla, F. Nicotra, G. Forloni, A. Cagnotto, M. Salmona, M. Masserini and G. Sancini, *Nanomedicine*, 2011, **7**, 551–559.
- 161 G. Sarkar, G. L. Curran, E. Mahlum, T. Decklever, T. M. Wengenack, A. Blahnik, B. Hoesley, V. J. Lowe, J. F. Poduslo and R. B. Jenkins, *PLoS One*, 2011, **6**, e28881.
- 162 Y. Meng, I. Sohar, D. E. Sleat, J. R. Richardson, K. R. Reuhl, R. B. Jenkins, G. Sarkar and P. Lobel, *Mol. Ther.*, 2014, **22**, 547–553.
- 163 I. Westergren and B. B. Johansson, *Acta Physiol. Scand.*, 1993, **149**, 99–104.
- 164 G. Tosi, B. Bortot, B. Ruozi, D. Dolcetta, M. A. Vandelli, F. Forni and G. M. Severini, *Curr. Med. Chem.*, 2013, **20**, 2212–2225.
- 165 G. Tosi, L. Bondioli, B. Ruozi, L. Badiali, G. M. Severini, S. Biffi, A. De Vita, B. Bortot, D. Dolcetta, F. Forni and M. A. Vandelli, *J. Neural Transm.*, 2011, **118**, 145–153.
- 166 A. Vilella, G. Tosi, A. M. Grabrucker, B. Ruozi, D. Belletti, M. A. Vandelli, T. M. Boeckers, F. Forni and M. Zoli, *J. Controlled Release*, 2014, **174**, 195–201.
- 167 E. Kilic, G. P. Dietz, D. M. Hermann and M. Bähr, *Ann. Neurol.*, 2002, **52**, 617–622.
- 168 M. Aarts, Y. Liu, L. Liu, S. Besshoh, M. Arundine, J. W. Gurd, Y. T. Wang, M. W. Salter and M. Tymianski, *Science*, 2002, **298**, 846–850.
- 169 S. S. Elliger, C. A. Elliger, C. Lang and G. L. Watson, *Mol. Ther.*, 2002, **5**, 617–626.
- 170 E. Kilic, U. Kilic and D. M. Hermann, *CNS Drug Rev.*, 2005, **11**, 369–378.
- 171 Y. Qin, H. Chen, Q. Zhang, X. Wang, W. Yuan, R. Kuai, J. Tang, L. Zhang, Z. Zhang, Q. Zhang, J. Liu and Q. He, *Int. J. Pharm.*, 2011, **420**, 304–312.
- 172 H. Wang, K. Xu, L. Liu, J. P. Tan, Y. Chen, Y. Li, W. Fan, Z. Wei, J. Sheng, Y. Y. Yang and L. Li, *Biomaterials*, 2010, **31**, 2874–2881.
- 173 X.-H. Tian, Z.-G. Wang, H. Meng, Y.-H. Wang, W. Feng, F. Wei, Z.-C. Huang, X.-N. Lin and L. Ren, *Int. J. Nanomed.*, 2013, **8**, 865–876.
- 174 K. S. Rao, M. K. Reddy, J. L. Horning and V. Labhasetwar, *Biomaterials*, 2008, **29**, 4429–4438.
- 175 G. Sharma, A. Modgil, T. Zhong, C. Sun and J. Singh, *Pharm. Res.*, 2014, **31**, 1194–1209.
- 176 T. Zong, L. Mei, H. Gao, W. Cai, P. Zhu, K. Shi, J. Chen, Y. Wang, F. Gao and Q. He, *Mol. Pharmaceutics*, 2014, **11**, 2346–2357.
- 177 P. Youn, Y. Chen and D. Y. Furgeson, *Mol. Pharmaceutics*, 2014, **11**, 486–495.
- 178 S. Lim, W.-J. Kim, Y.-H. Kim, S. Lee, J.-H. Koo, J.-A. Lee, H. Yoon, D.-H. Kim, H.-J. Park and H.-M. Kim, *et al.*, *Nat. Commun.*, 2015, **6**, 8244.
- 179 J. Mensch, J. Oyarzabal, C. Mackie and P. Augustijns, *J. Pharm. Sci.*, 2009, **98**, 4429–4468.
- 180 M. Teixidó, E. Zurita, L. Mendieta, B. Oller-Salvia, R. Prades, T. Tarragó and E. Giralt, *Biopolymers*, 2013, **100**, 662–674.
- 181 X. Wei, C. Zhan, X. Chen, J. Hou, C. Xie and W. Lu, *Mol. Pharmaceutics*, 2014, **11**, 3261–3268.
- 182 R. Prades, B. Oller-Salvia, S. M. Schwarzmaier, J. Selva, M. Moros, M. Balbi, V. Grazú, J. M. de La Fuente, G. Egea, N. Plesnila, M. Teixidó and E. Giralt, *Angew. Chem., Int. Ed.*, 2015, **54**, 3967–3972.
- 183 R. Prades, S. Guerrero, E. Araya, C. Molina, E. Salas, E. Zurita, J. Selva, G. Egea, C. López-Iglesias, M. Teixidó, M. J. Kogan and E. Giralt, *Biomaterials*, 2012, **33**, 7194–7205.
- 184 J. P. Vincent, H. Schweitz and M. Lazdunski, *Biochemistry*, 1975, **14**, 2521–2525.
- 185 E. Habermann and K. G. Reiz, *Biochem. Z.*, 1965, **343**, 192–203.
- 186 L. Di, H. Rong and B. Feng, *J. Med. Chem.*, 2013, **56**, 2–12.
- 187 M. Guttman, J. H. Prieto, T. M. Handel, P. J. Domaille and E. A. Komives, *J. Mol. Biol.*, 2010, **398**, 306–319.
- 188 J. Spengler, J. C. Jiménez, K. Burger, E. Giralt and F. Albericio, *J. Pept. Res.*, 2005, **65**, 550–555.
- 189 R. Cecchelli, V. Berezowski, S. Lundquist, M. Culot, M. Renftel, M. P. Dehouck and L. Fenart, *Nat. Rev. Drug Discovery*, 2007, **6**, 650–661.
- 190 I. van Rooy, S. Cakir-Tascioglu, W. E. Hennink, G. Storm, R. M. Schiffelers and E. Mastrobattista, *Pharm. Res.*, 2011, **28**, 456–471.
- 191 C. D. Kuhnline Sloan, P. Nandi, T. H. Linz, J. V. Aldrich, K. L. Audus and S. M. Lunte, *Annu. Rev. Anal. Chem.*, 2012, **5**, 505–531.
- 192 J. Bicker, G. Alves, A. Fortuna and A. Falcão, *Eur. J. Pharm. Biopharm.*, 2014, **87**, 409–432.
- 193 I. Wilhelm and I. A. Krizbai, *Mol. Pharmaceutics*, 2014, **11**, 1949–1963.
- 194 U. Bickel, *NeuroRx*, 2005, **2**, 15–26.
- 195 R. Gabathuler, *Neurobiol. Dis.*, 2010, **37**, 48–57.

