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# Redox potentials of aryl derivatives from hybrid functional based first principles molecular dynamics

Xiandong Liu,<sup>\*a</sup> Jun Cheng,<sup>\*bc</sup> Xiancai Lu,<sup>a</sup> Mengjia He<sup>a</sup> and Rucheng Wang<sup>a</sup>

We report the redox potentials of a set of organic aryl molecules, including quinones, juglone, tyrosine and tryptophan, calculated using a first principles molecular dynamics (FPMD) based method. The hybrid functional HSE06 reproduces the redox potentials spanning from  $-0.25$  V to  $1.15$  V within an error of  $0.2$  V, whereas the errors with the BLYP functional are much larger (up to  $0.7$  V). It is found that the BLYP functional predicts consistently lower electron affinities/ionization potentials than HSE06 both in gas phase and in an aqueous solution. In water, the ionization potentials are significantly underestimated by BLYP due to the exaggeration of the mixing between the solute states and the valence band states of liquid water. Hybrid HSE06 markedly improves both the solute levels and water band positions, leading to accurate redox potentials. This study suggests that the current FPMD based method at the level of hybrid functionals is able to accurately compute the redox potentials of a wide spectrum of organic molecules.

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## 1. Introduction

Redox potential is a measure of the tendency of a species to gain electrons and therefore is a key thermodynamic quantity for characterizing electron transfer reactions. Redox potentials, together with acidity constants (*i.e.* the free energies of proton transfer reactions) are the basis for constructing pH-Eh diagrams, which are widely used to understand the stabilities of chemical species under various redox and pH conditions in many areas of chemistry.

Computation of redox potentials is of great interest to the quantum chemistry communities and a number of methods have been developed. Solvation effects are often described by empirical methods such as implicit solvation models<sup>1,2</sup> and the distribution of point dipoles.<sup>3</sup> QM/MM methods can further include atomic detail of solvation shells by replacing the continuum solvent by classical force field models.<sup>4</sup> Previous studies have shown that these methods can compute redox potentials accurately for many species, including inorganic compounds, organic compounds, and transition metal complexes.<sup>2</sup>

Alternatively, redox potentials can be calculated using first-principles molecular dynamics (FPMD), which treats solutes and solvents at the same quantum mechanical level and also accounts for the atomic level details of dynamical solvent effects.<sup>5</sup> These can be important in many cases, for example, for the coordination spheres yielding drastic change upon reduction/oxidation, wherein entropic contributions could be significant to free energies. Another example is that at the elevated T-P conditions relevant to the Earth's interior, the solvent effects are hard to include in the implicit solvent protocols, because the models are usually parameterized for ambient conditions.<sup>1</sup>

The FPMD based vertical energy gap method developed by the Sprik group<sup>6–8</sup> provides a feasible way to compute redox potentials and acidity constants. Extensive tests on molecular acids and metal cations indicate that acidity constants are reproduced within an error of  $2$  pK<sub>a</sub> units (approximately  $0.1$  eV) by GGA (generalized gradient approximation) functionals.<sup>9–14</sup> In contrast, redox potentials calculated by GGA are much worse, for example, the values of benzoquinones (including benzoquinone, semi-benzoquinone and hydro-benzoquinone),<sup>15</sup> tyrosine and tryptophan,<sup>6</sup> are underestimated by  $0.2$ – $0.7$  V. It has been found that GGA places the valence band of liquid water at a too high position and therefore exaggerates the mixing of the electronic states of the solute with the water valence band, leading to underestimation of the redox potential of the solute.<sup>16</sup> Hybrid functionals such as HSE06<sup>17,18</sup> improve the prediction of the band states of liquid water, which eventually leads to much better estimates of the redox potentials. This has been confirmed on some small inorganic molecules<sup>16</sup> and transition metal aqua-cations.<sup>19</sup> Very recently,

<sup>a</sup> State Key Laboratory for Mineral Deposits Research, School of Earth Sciences and Engineering, Nanjing University, Nanjing 210093, P. R. China.

E-mail: [xiandongliu@nju.edu.cn](mailto:xiandongliu@nju.edu.cn); Fax: +86 25 83686016; Tel: +86 25 89680700

<sup>b</sup> Collaborative Innovation Center of Chemistry for Energy Materials, State Key Laboratory of Physical Chemistry of Solid Surfaces, College of Chemistry and Chemical Engineering, Xiamen University, Xiamen 361005, P. R. China.  
E-mail: [chengjun@xmu.edu.cn](mailto:chengjun@xmu.edu.cn); Tel: +86 5922181570

<sup>c</sup> Department of Chemistry, University of Aberdeen, Aberdeen AB24 3UE, UK.  
E-mail: [jcheng@abdn.ac.uk](mailto:jcheng@abdn.ac.uk)



random phase approximation and a double hybrid functional have been found to compute accurate water band positions and redox potentials for the oxidizing species of OH• and Cl•.<sup>20</sup>

Redox chemistry of aryl derivatives, such as quinones, tyrosine and tryptophan, are of great importance in many research areas. In biology, tyrosyl and tryptophanyl radicals act as intermediates in the redox reactions of enzymes and quinones usually perform as the redox-active part of some cofactors.<sup>21</sup> In geochemistry, it has been widely accepted that quinones are the major redox active groups in natural organic matters (NOMs),<sup>22</sup> which, as important components of soils, take part in important superegene processes, *e.g.* the reduction and fixation of contaminants and metal cations. Independent small quinones (*e.g.* juglone, duroquinone and lawsone) and amino acids present in soils also play active roles in numerous electron transfer reactions.<sup>23</sup> Therefore, accurate prediction of their redox potentials is very helpful for elucidating the redox reaction mechanisms in biological and geochemical processes.

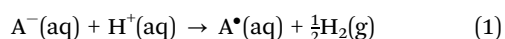
In this study, hybrid functional based FPMD has been validated in the prediction of redox potentials of five organic molecules. We revisit the model systems used in the development of the method (*i.e.* benzoquinones, tyrosine and tryptophan) and test two other quinones, juglone and duroquinone, which have negative redox potentials. The solvation structures and solvent reorganization have been investigated using FPMD simulations. The redox potentials obtained using GGA and hybrid functionals have been compared in detail by analyzing the energy levels. The accurate computation of redox potentials with hybrid functionals suggests that the current FPMD based methodology has a wide range of potential applications in redox chemistry studies.

## 2. Methodology

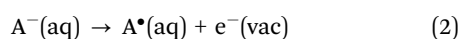
### 2.1. Free energy perturbation theory and redox potential calculation

In the early study on transition metal cations,<sup>24–28</sup> the computed redox potentials could not be directly compared with experimental measurements due to the lack of a physical potential reference. A molecular dynamics based computational standard hydrogen electrode (SHE) was developed to address this issue by Sprik *et al.*<sup>6,15</sup> Herein, we present a brief introduction to the methodology and refer the interested readers to the original papers for more detail.

The potential of a redox couple with respect to SHE corresponds to the free energy of the oxidation reaction in which the reduced state is oxidized by an aqueous proton:



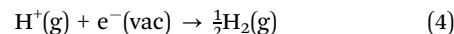
In the computational SHE protocol, eqn (1) is divided into three steps: the reversible removal of an electron from the reduced state



the desolvation of an aqueous proton



and the combination of the proton and the electron into half of a H<sub>2</sub> molecule in the gas phase:



The free energies of the reversible removal of an electron/proton (*i.e.* eqn (2) and (3)) can be calculated by using free energy perturbation theory. In this scheme, an auxiliary mapping Hamiltonian  $H_\eta$  is constructed by linearly mixing the Hamiltonian of the reactant  $H_R$  and Hamiltonian of the product  $H_P$  through the Kirkwood coupling parameter  $\eta$ ,

$$H_\eta = (1 - \eta)H_R + \eta H_P \quad (5)$$

When the coupling parameter  $\eta$  increases from 0 to 1, the Hamiltonian is transformed from the reactant to the product, *i.e.* from the reduced/protonated state to oxidized/deprotonated state for an oxidation/deprotonation reaction. The intermediate states for  $0 < \eta < 1$  correspond to the hybrids of the reactant and product states and have no physical counterparts. FPMD simulations, however, can be used to sample these mixing potential energy surfaces. The free energy change ( $\Delta A$ ) of the transformation can be obtained by integrating the ensemble average of the vertical energy gap ( $\langle \Delta E \rangle_\eta$ ) with respect to the coupling parameter,

$$\Delta A = \int_0^1 d\eta \langle \Delta E \rangle_\eta \quad (6)$$

The vertical energy gap is defined as the energy difference between the reactant and product states at fixed configurations in the FPMD trajectories.

Thus, the computed free energies of eqn (2) and (3) can be expressed in the form of thermodynamic integrals, *i.e.*, the oxidation integral of  $A^-(aq)$  ( $\Delta_{ox}A_{A^-}$ ) and the deprotonation integral of  $H_3O^+(aq)$  ( $\Delta_{dp}A_{H_3O^+}$ ), respectively. Note that in our implementation the desolvation of an aqueous proton (reaction (3)) is effectively replaced by the deprotonation of a solvated hydronium  $H_3O^+(aq)$ . Including the free energy of the gas phase reaction (4) ( $\mu_{H^+}^{g,o}$ ), the formula for computing the redox potential is

$$eU^0 = \Delta_{ox}A_{A^-} + \Delta_{dp}A_{H_3O^+} - \mu_{H^+}^{g,o} - \Delta E_{zp} \quad (7)$$

where  $e$  is the elementary charge of an electron and  $U^0$  is the redox potential *vs.* SHE.  $\Delta E_{zp}$  accounts for the zero point energy of the breaking of an O–H bond in  $H_3O^+(aq)$  and is estimated to be 0.35 eV.<sup>6</sup> The experimental value of the free energy of reaction (4) is 15.72 eV, taking  $c^0 = 1 \text{ mol L}^{-1}$  and  $p^0 = 1 \text{ bar}$  as the standard states for an aqueous solution and gas phase.<sup>29</sup> Adding the correction for converting the gas phase standard state also to  $c^0 = 1 \text{ mol L}^{-1}$ , the free energy of reaction (4) used in eqn (7) is  $\mu_{H^+}^{g,o} = 15.81 \text{ eV}$ .<sup>6</sup> It is important to note that the values of the individual thermodynamic integrals ( $\Delta_{ox}A_{A^-}$  and  $\Delta_{dp}A_{H_3O^+}$ ), which are associated with half reactions with a change of the net charge, do not bear physical meanings owing to the artificial offset in the potential reference under periodic boundary conditions (PBC). However, when combining the two in eqn (7), the potential offsets in the integrals are canceled, leading to the meaningful redox potentials *vs.* SHE. Strictly speaking, the complete cancelation is



an assumption that the artificial potential offset is dominated by the solvent water. This should be rather safe for small ions; however, there might be some residual error for bulky solutes such as the organic molecules studied in this study. In a previous study,<sup>6</sup> an empirical correction term of 0.3 eV was estimated by comparing the computed and experimental  $pK_a$ 's. However, the  $pK_a$ 's were computed with the BLYP functional and the reference integral  $\Delta_{dp}A_{H_3O^+}$  was taken as a value of 15.20 eV,<sup>6</sup> but now the recommended value is 15.35 eV and will be used in this study.<sup>9,10</sup> We, therefore, will not include this correction in computing the redox potentials of the organic molecules, and as will be observed below, the values obtained by using hybrid HSE06 are very close to experimental values.

For the oxidation reaction, the vertical energy gaps at  $\eta = 0$  and  $\eta = 1$  have physical meanings, corresponding to the vertical ionization potential of the reduced state ( $IP_{A^-}$ ) and the electron affinity of the oxidized state ( $EA_{A^\bullet}$ ), respectively. They can also be aligned with respect to the SHE by using the following formulas, which are very similar to eqn (7),<sup>7</sup>

$$EA_{A^\bullet} = \langle \Delta_{ox} E_{A^-} \rangle_{\eta=0} + \Delta_{dp}A_{H_3O^+} - \mu_{H^+}^{g,0} - \Delta E_{zp} \quad (8)$$

$$IP_{A^-} = \langle \Delta_{ox} E_{A^-} \rangle_{\eta=1} + \Delta_{dp}A_{H_3O^+} - \mu_{H^+}^{g,0} - \Delta E_{zp} \quad (9)$$

The differences between the vertical electronic energies and redox potentials are the solvent reorganization energies. Thus, the two reorganization energies,  $\lambda_R$  for the reduced state and  $\lambda_O$  in the oxidized state, are written as

$$\lambda_R = eU^0 - EA_{A^\bullet} \quad (10)$$

$$\lambda_O = IP_{A^-} - eU^0 \quad (11)$$

If the solvent response is linear, as assumed in the Marcus theory of electron transfer, the following relations hold;  $eU^0 = (EA_{A^\bullet} + IP_{A^-})/2$  and  $\lambda_R = \lambda_O$ , the deviation from which is a sign of nonlinearity in the solvent response. In particular, we will use the ratio  $\lambda_R/\lambda_O$  as a descriptor for measuring the asymmetry in the organization energies for the redox couples.

## 2.2. Model systems of organic molecules in water

Redox potential is a measure of the tendency of a species to gain electrons and therefore is a key thermodynamic quantity for characterizing electron transfer reactions. Redox potentials, together with acidity constants (*i.e.* the free energies of proton transfer reactions) are the basis for constructing pH-Eh diagrams, which are widely used to understand the stabilities of chemical species under various redox and pH conditions in many areas of chemistry. The molecular structures of the model systems are shown in Fig. 1. Benzoquinone, juglone, duroquinone, tyrosine anion and protonated tryptophan are denoted as Q, Jug, DQ, TyrO<sup>-</sup> and TrpH, respectively. The cell for all the simulations is a cubic box of the length of 12.43 Å. The number of solvent water molecules for each solute is listed in Table 1. The numbers were determined to approximately represent the density of liquid water under the ambient conditions. For Q, TyrO<sup>-</sup> and TrpH, the numbers of H<sub>2</sub>O are the same as those of previous simulations.<sup>6,15</sup>

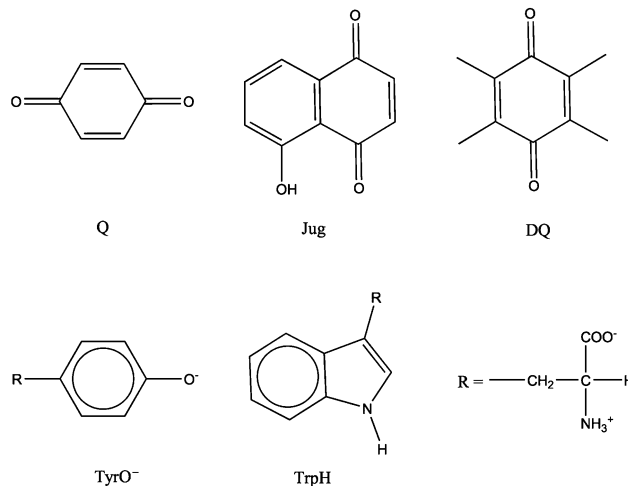


Fig. 1 Structures of the model systems. Q = benzoquinone, Jug = juglone, DQ = duroquinone, TyrO<sup>-</sup> = tyrosine anion, TrpH = protonated tryptophan.

Table 1 The numbers of water molecules used for the oxidation reactions

Oxidation	Number of H <sub>2</sub> O molecules
Q <sup>•-</sup> → Q	60
Jug <sup>•-</sup> → Jug	57
DQ <sup>•-</sup> → DQ	56
TyrO <sup>-</sup> → TyrO <sup>•</sup>	49
TrpH → TrpH <sup>•+</sup>	48

## 2.3. Computational setup

The simulations were performed using the freely available CP2K/QUICKSTEP package.<sup>30</sup> In QUICKSTEP, the electronic structures were calculated with density functional theory implemented based on a hybrid GPW (Gaussian plane wave) technique.<sup>31</sup> BLYP<sup>32,33</sup> has been the favored GGA functional for FPMD simulations of aqueous systems and was widely used in previous calculations.<sup>6,15</sup> While BLYP was used in this study for the sake of consistency, we do not expect the usage of other GGA functionals such as PBE to make any major difference in computing redox potentials. In the calculations with the hybrid functional HSE06,<sup>17,18</sup> the exact exchange under PBC was implemented using the auxiliary density matrix method.<sup>34</sup> Goedecker-Teter-Hutter (GTH) pseudopotentials were applied to represent the core electrons.<sup>35</sup> Double- $\zeta$  basis sets augmented with polarization functions were employed for H, O, N and C. The cut off for expanding electron density in the reciprocal space was set to be 280 Ry. The convergence criteria for the electronic gradient and the energy difference between final SCF cycles were set to be  $1.0 \times 10^6$  and  $1.0 \times 10^{-12}$  a.u., respectively.

Born–Oppenheimer molecular dynamics simulations were carried out with a time step of 0.5 fs. The temperature was controlled at 330 K using the Nosé–Hoover chain thermostat. The elevated temperature is to avoid the glassy behavior at the lower temperatures.<sup>36</sup> HSE06 MD runs were started from the BLYP equilibrated configurations. For each simulation, the production



run was performed for over 6.0 ps, following a prior equilibration run for at least 2.0 ps.

To better understand the difference between BLYP and HSE06, we calculated the EA of DQ and TyrO and IP of  $\text{DQ}^{\bullet-}$  and  $\text{TyrO}^-$  in the gas phase. These calculations were carried out with a cubic box of the length of 25 Å. For charged ions, the electrostatic interaction was treated using the Martyna–Tuckerman method<sup>37</sup> to effectively eliminate interactions between periodic images.

### 3. Results

#### 3.1. Hydration structures

The H-bonding structures of the solutes have been characterized in previous BLYP FPMD calculations.<sup>6,15</sup> The hydrophilic centers of quinones are the oxygen atoms. In addition to the ammonium and carboxyl groups, the indolic nitrogen and the oxygen atom are the hydrophilic centers of tryptophan and tyrosine, respectively.

Taking DQ and Q as examples, the RDF-CN (radial distribution function-coordination number) curves for water H around quinone O derived from the HSE06 simulations are shown in Fig. 2. It is clearly observed that as oxidation proceeds, the H-bonds get weaker, as discussed in the previous study.<sup>15</sup> For both  $\text{DQ}^{\bullet-}$  and  $\text{Q}^{\bullet-}$ , the H-bonds are peaked at around 1.65 Å and 1.70 Å, respectively, while for their oxidation states, the peaks shift to 1.85 Å. Similarly, for  $\text{DQ}^{\bullet-}$  and  $\text{Q}^{\bullet-}$ , the CNs are 2.1 and 2.2, respectively, while the CN is only 1.4 for DQ and Q (see the images of the DQ couple in Fig. 3). It is found that HSE06 and BLYP present similar H-bonding structures; for example, similar to HSE06, BLYP predicts the average H-bond distances for  $\text{Q}^{\bullet-}$  and Q to be 1.70 Å and 1.90 Å, respectively.<sup>15</sup>

#### 3.2. Vertical energy gaps and redox potentials

Table 2 lists the computed vertical energy gaps, thermodynamic integrals and redox potentials. All the vertical energy gaps converge within 0.1 eV in the MD simulations. This level of statistical uncertainty is consistent with the previous studies.<sup>6,15,19</sup> The statistical accuracy can be shown by the accumulative averages of the vertical energy gaps of TrpH and DQ in Fig. 4.

Fig. 5 plots the computed redox potentials against the experimental values. It is clear that BLYP underestimates all redox potentials and HSE06 significantly improves the predictions.

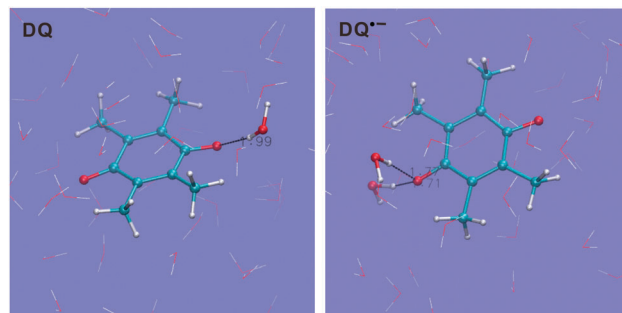


Fig. 3 Images for  $\text{DQ}^{\bullet-}$  and DQ. O = red, H = white and C = blue.

Table 2 Vertical energy gaps (eV), thermodynamic integrals (eV) and redox potentials vs. SHE (V) computed with HSE06 in comparison with the BLYP results

		$\eta = 0$ (eV)	$\eta = 0.5$ (eV)	$\eta = 1.0$ (eV)	Integral (eV)	$U_{\text{Cal}}^0$ (V)	$U_{\text{Exp}}^0$ (V)
DQ	BLYP	−0.62	0.30	0.91	0.25	−0.56	−0.24 <sup>38</sup>
	HSE06	−0.46	0.6	1.61	0.59	−0.22	
Jug	BLYP	−0.50	0.27	1.02	0.27	−0.54	−0.09 <sup>39</sup>
	HSE06	−0.22	0.63	1.50	0.63	−0.18	
Q	BLYP <sup>15</sup>	−0.33	0.45	1.40	0.48	−0.33	0.10 <sup>40</sup>
	HSE06	−0.26	0.97	1.97	0.93	0.12	
TyrO	BLYP <sup>6</sup>	0.43	1.35	1.88	1.28	0.47	0.72 <sup>21</sup>
	HSE06	0.81	1.61	2.38	1.60	0.79	
TrpH	BLYP <sup>6</sup>	1.00	1.55	2.01	1.53	0.72	1.15 <sup>21</sup>
	HSE06	1.50	2.11	2.63	2.07	1.28	

Even for the lower end, *i.e.* DQ and Jug, the errors in BLYP are over 0.3 V. HSE06 increases all values relative to BLYP. Most redox potentials are reproduced within an error of  $\sim 0.1$  V. This error margin is of the same magnitude as the statistical errors in the MD simulations (Table 3). The overall accuracy of computation of redox potentials achieved by the current FPMD method is comparable to that of implicit solvation methods.<sup>2</sup>

#### 3.3. Energy level diagram and solvent reorganization

The results of BLYP and HSE06 can be compared in more detail by analyzing the energy level diagrams. Vertical energy gaps are aligned to the SHE scale so that one can compare the coupling of the solute states with the band states of liquid water. As shown in

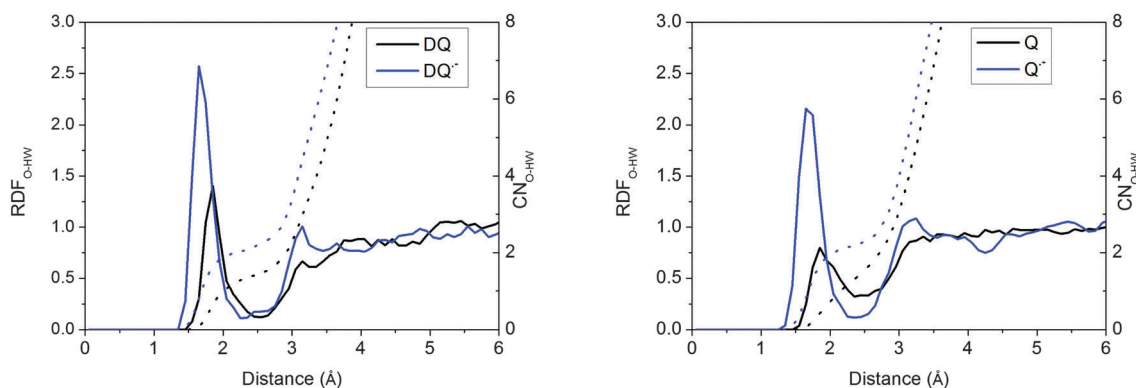


Fig. 2 RDFs (radial distribution functions) and CNs (coordination numbers) for water H around the O of quinones derived from the HSE06 simulations.





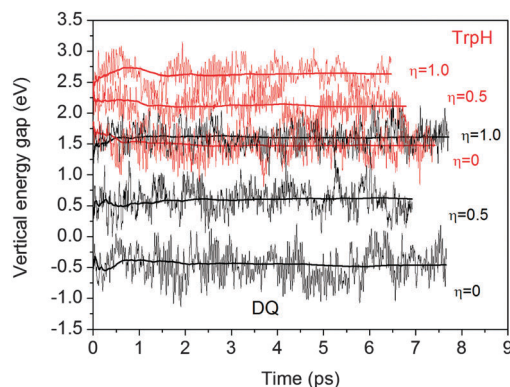


Fig. 4 Time accumulative averages of the vertical energy gaps of TrpH and DQ calculated with HSE06.

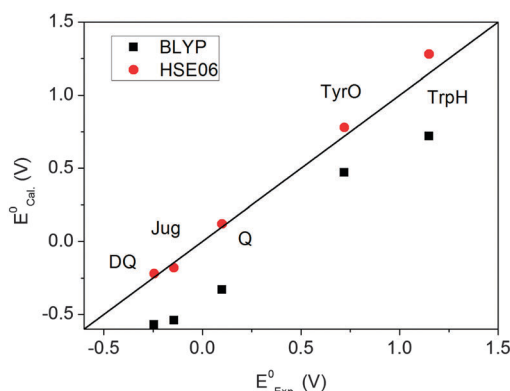


Fig. 5 Comparison between the calculated and experimental redox potentials.

**Table 3** Vertical EA and IP energies for DQ and TyrO computed by BLYP and HSE06. The numbers in parentheses are the differences between the HSE06 and the BLYP values

	BLYP (eV)	HSE06 (eV)
EA of DQ	1.18	1.52 (0.34)
EA of TyrO	1.71	1.95 (0.24)
IP of DQ <sup>•-</sup>	1.67	2.09 (0.42)
IP of TyrO <sup>-</sup>	1.93	2.21 (0.28)

the previous study, BLYP estimates the CBM (conduction band minimum) and VBM (valence band maximum) of liquid water to be  $-2.6$  V and  $2.3$  V, respectively. Compared to experimental values of  $-3.2$  V and  $5.5$  V, the BLYP VBM is very high by  $3.2$  V, in contrast to the relatively small error of  $0.6$  V in CBM.<sup>7</sup> This will cause enhanced mixing of the solute states and the valence band of liquid water, leading to very high  $-IP$  levels of the solutes and therefore very high redox levels. HSE06 improves the computed VBM of liquid water by  $1.3$  V and places the CBM at the right position.<sup>7</sup> This is also consistent with the observation from Fig. 6 that HSE06 significantly improves the  $-IP$  levels relative to those of BLYP by at least  $0.48$  V, while the differences in the  $-EA$  levels are much smaller. Therefore, the better description of the band structure of liquid water by hybrid HSE06 improves the accurate positioning of vertical and redox levels of the solutes.

As can be observed from the energetics in the gas phase (Table 3), the HSE06 results are consistently higher than the BLYP results by  $0.2$ – $0.4$  eV. Note that the EA energy of DQ calculated by HSE06,  $1.52$  eV, agrees well with the experimental value of  $1.60$  eV.<sup>41</sup> Similar underestimation by GGA has also been found in the comparison between BLYP and B3LYP in the previous publication.<sup>15</sup> These results suggest that BLYP underestimates the attachment and detachment energies of these gas phase molecules compared to those of hybrid functionals. In water, the increases in the EA energies switching from BLYP to HSE06 are  $0.16$  eV and  $0.38$  eV for DQ and TyrO, respectively (see Table 2), close to the respective differences in vacuum ( $0.34$  eV and  $0.24$  eV, respectively, from Table 3). In contrast, the increases in the IP differences are more obvious: from  $0.42$  eV and  $0.28$  eV in vacuum to  $0.7$  eV and  $0.5$  eV in water, respectively. The finding that the EA differences between BLYP and HSE06 in water and vacuum are close is consistent with the fact that the water VBM of BLYP and HSE06 differ by only  $0.5$  V. On the other hand, HSE06 significantly reduces the coupling of the HOMO of the solutes with valence band states of liquid water due to lowering of the water VBM by  $1.3$  eV compared to that using BLYP and therefore the IP differences between BLYP and HSE06 are more pronounced.<sup>16</sup>

The ratios of  $\lambda_R/\lambda_O$  (Fig. 7) calculated by HSE06 are very close to  $1.0$  for all five redox couples, within a  $15\%$  deviation. This indicates that the solvent response to oxidation/reduction of

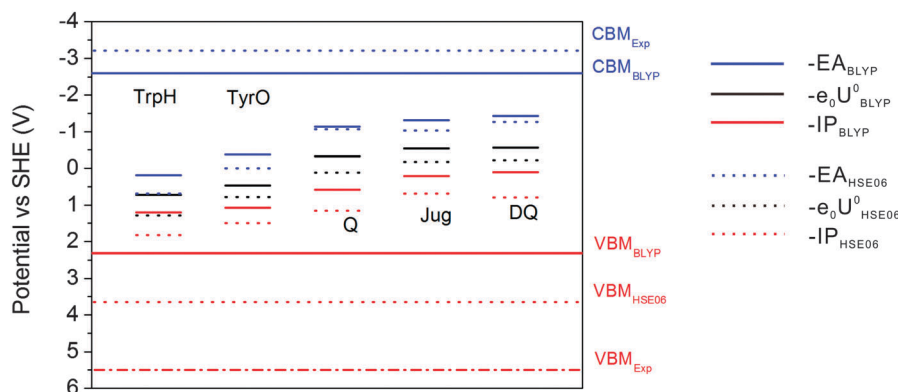


Fig. 6 Energy level diagram calculated with HSE06 and BLYP.  $CBM_{HSE06}$  is overlapped with  $CBM_{Exp}$ . Solid lines: BLYP; Dashed lines: HSE06.

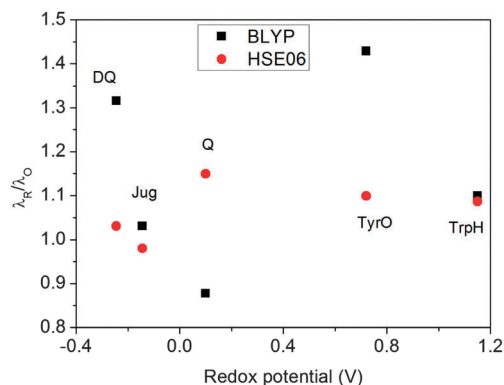


Fig. 7 The ratio of  $\lambda_R/\lambda_O$  calculated with HSE06 and BLYP functionals.

these molecules predicted by HSE06 is rather linear, consistent with the Marcus theory of electron transfer.<sup>42</sup> In contrast, the  $\lambda_R/\lambda_O$  ratios from BLYP are more scattered; Jug and TrpH deviate by 10%, while DQ and TyrO deviate by more than 30%. This asymmetry in the reorganization energies has been found previously to be much more pronounced for couples with very positive redox potentials such as  $\text{OH}^-/\text{OH}^\bullet$  and  $\text{Cl}^-/\text{Cl}^\bullet$ ; BLYP predicts  $\lambda_R/\lambda_O$  ratios to be close to 3, while HSE06 estimates are still about 1.4.<sup>8</sup>

## 4. Conclusion

This study shows that the FPMD based redox potential calculation method can accurately reproduce the redox potentials of a set of five aryl derivatives spanning from  $-0.25$  V to  $1.15$  V at the level of hybrid HSE06, whereas BLYP results are too low. Including a fraction of exact exchange, HSE06 effectively improves the vertical energy levels of these organic solutes, which are underestimated by GGA functionals both in vacuum and in water. The test models in this study cover the redox potential range of NOMs ( $-0.3$  to  $0.15$  V at pH = 7),<sup>22</sup> and the computational settings used should be able to provide accurate estimates for NOMs. Considering also the previous successes in calculating transition metal cations,<sup>19</sup> we are optimistic regarding application of this method to electron transfer reactions of transition metal–organic complexes. We hope our extensive test calculations demonstrate that equipped with hybrid functionals, the FPMD base method can be a powerful and reliable tool for investigating the redox chemistry in complex environments, including biological and geochemical systems.

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## References

- 1 J. Tomasi, B. Mennucci and R. Cammi, *Chem. Rev.*, 2005, **105**, 2999–3094.
- 2 A. V. Marenich, J. Ho, M. L. Coote, C. J. Cramer and D. G. Truhlar, *Phys. Chem. Chem. Phys.*, 2014, **16**, 15068–15106.
- 3 A. Warshel, P. K. Sharma, M. Kato and W. W. Parson, *Biochim. Biophys. Acta, Proteins Proteomics*, 2006, **1764**, 1647–1676.
- 4 L.-P. Wang and T. Van Voorhis, *J. Chem. Theory Comput.*, 2012, **8**, 610–617.
- 5 D. Marx and J. Hutter, *Ab Initio Molecular Dynamics: Basic Theory and Advanced Methods*, Cambridge University Press, Cambridge, 2009.
- 6 F. Costanzo, M. Sulpizi, R. G. Della Valle and M. Sprik, *J. Chem. Phys.*, 2011, **134**, 244508.
- 7 J. Cheng and M. Sprik, *Phys. Chem. Chem. Phys.*, 2012, **14**, 11245–11267.
- 8 J. Cheng, X. Liu, J. VandeVondele, M. Sulpizi and M. Sprik, *Acc. Chem. Res.*, 2014, **47**, 3522–3529.
- 9 M. Sulpizi and M. Sprik, *J. Phys.: Condens. Matter*, 2010, **22**, 284116.
- 10 M. Sulpizi and M. Sprik, *Phys. Chem. Chem. Phys.*, 2008, **10**, 5238–5249.
- 11 X. Liu, J. Cheng, M. Sprik and X. Lu, *J. Phys. Chem. Lett.*, 2013, **4**, 2926–2930.
- 12 X. Liu, J. Cheng, M. Sprik, X. Lu and R. Wang, *Geochim. Cosmochim. Acta*, 2013, **120**, 487–495.
- 13 X. Liu, M. He, X. Lu and R. Wang, *Chem. Geol.*, 2015, **411**, 192–199.
- 14 M. Mangold, L. Rolland, F. Costanzo, M. Sprik, M. Sulpizi and J. Blumberger, *J. Chem. Theory Comput.*, 2011, **7**, 1951–1961.
- 15 J. Cheng, M. Sulpizi and M. Sprik, *J. Chem. Phys.*, 2009, **131**, 154504.
- 16 C. Adriaanse, J. Cheng, V. Chau, M. Sulpizi, J. VandeVondele and M. Sprik, *J. Phys. Chem. Lett.*, 2012, **3**, 3411–3415.
- 17 J. Heyd, G. E. Scuseria and M. Ernzerhof, *J. Chem. Phys.*, 2003, **118**, 8207–8215.
- 18 A. V. Krukau, O. A. Vydrov, A. F. Izmaylov and G. E. Scuseria, *J. Chem. Phys.*, 2006, **125**, 224106.
- 19 X. Liu, J. Cheng and M. Sprik, *J. Phys. Chem. B*, 2015, **119**, 1152–1163.
- 20 J. Cheng and J. VandeVondele, *Phys. Rev. Lett.*, 2016, **116**, 086402.
- 21 J. Stubbe and W. A. van der Donk, *Chem. Rev.*, 1998, **98**, 705–762.
- 22 J. T. Nurmi and P. G. Tratnyek, *Environ. Sci. Technol.*, 2002, **36**, 617–624.
- 23 M. Uchimiya and A. T. Stone, *Chemosphere*, 2009, **77**, 451–458.
- 24 J. Blumberger, L. Bernasconi, I. Tavernelli, R. Vuilleumier and M. Sprik, *J. Am. Chem. Soc.*, 2004, **126**, 3928–3938.
- 25 J. Blumberger and M. Sprik, *Theor. Chem. Acc.*, 2006, **115**, 113–126.
- 26 J. Blumberger, I. Tavernelli, M. L. Klein and M. Sprik, *J. Chem. Phys.*, 2006, **124**, 64507.
- 27 Y. Tateyama, J. Blumberger, M. Sprik and I. Tavernelli, *J. Chem. Phys.*, 2005, **122**, 234505.



- 28 M. Sulpizi, S. Rauegi, J. VandeVondele, P. Carloni and M. Sprik, *J. Phys. Chem. B*, 2007, **111**, 3969–3976.
- 29 D. R. Lide, *CRC handbook of chemistry and physics*, CRC press, 2004.
- 30 J. VandeVondele, M. Krack, F. Mohamed, M. Parrinello, T. Chassaing and J. Hutter, *Comput. Phys. Commun.*, 2005, **167**, 103–128.
- 31 G. Lippert, J. Hutter and M. Parrinello, *Mol. Phys.*, 1997, **92**, 477–487.
- 32 A. D. Becke, *Phys. Rev. A: At., Mol., Opt. Phys.*, 1988, **38**, 3098–3100.
- 33 C. Lee, W. Yang and R. G. Parr, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1988, **37**, 785.
- 34 M. Guidon, J. Hutter and J. VandeVondele, *J. Chem. Theory Comput.*, 2010, **6**, 2348–2364.
- 35 S. Goedecker, M. Teter and J. Hutter, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1996, **54**, 1703.
- 36 J. VandeVondele, F. Mohamed, M. Krack, J. Hutter, M. Sprik and M. Parrinello, *J. Chem. Phys.*, 2005, **122**, 014515.
- 37 G. J. Martyna and M. E. Tuckerman, *J. Chem. Phys.*, 1999, **110**, 2810–2821.
- 38 R. F. Anderson, *Biochim. Biophys. Acta, Bioenerg.*, 1983, **722**, 158–162.
- 39 T. Mukherjee, *Radiat. Phys. Chem.*, 1987, **29**, 455–462.
- 40 M. H. V. Huynh and T. J. Meyer, *Chem. Rev.*, 2007, **107**, 5004–5064.
- 41 E. K. Fukuda and R. T. McIver Jr, *J. Am. Chem. Soc.*, 1985, **107**, 2291–2296.
- 42 R. A. Marcus and N. Sutin, *Biochim. Biophys. Acta, Rev. Bioenerg.*, 1985, **811**, 265–322.

