## ChemComm



## COMMUNICATION

**View Article Online** 



Cite this: Chem. Commun., 2016 52 12901

Received 13th September 2016, Accepted 7th October 2016

DOI: 10.1039/c6cc07496k

www.rsc.org/chemcomm

## A 'photorelease, catch and photorelease' strategy for bioconjugation utilizing a p-hydroxyphenacyl group†

D. Madea, T. Slanina\* and P. Klán\*

A bioorthogonal 'catch and photorelease' strategy, which combines alkyne-azide cycloaddition between p-hydroxyphenacyl azide and alkyne derivatives to form a 1,2,3-triazole adduct and subsequent photochemical release of the triazole moiety via a photo-Favorskii rearrangement, is introduced. The first step can also involve photorelease of a strained alkyne and its Cu-free click reaction with azide.

Bioconjugation is a strategy to form covalent links between two molecules of biological interest, which is commonly used in nucleic acid research, pharmaceutical chemistry or microbiology. While the first procedures used non-specific approaches, such as ionic interactions with negatively charged DNA<sup>5</sup> or reactions of nucleophilic amines, carboxylates and thiols with electrophiles, modern methods rely on targeted functionalization, such as biotin-streptavidin conjugates, Staudinger ligation 10 and click reactions.4

Many bioconjugation strategies are based on bioorthogonal reactions which allow for chemical transformations of molecules without the need to protect reactive functional groups. They are used for fluorescent labeling, 11,12 cell detection and many other applications. 14 The most common types of bioorthogonal click reactions are cycloadditions, such as Cu(1)-catalyzed alkyne-azide 1,3-dipolar cycloadditions (CuAAC) or Cu-free click reactions of azides or nitrones with strained alkynes, and inverse electron-demand Diels-Alder reaction of 1,2,4,5-tetrazines with strained alkynes or alkenes. 15-17 Photochemically triggered bioorthogonal reactions, which enable spatial and temporal control over the bioconjugation and can be utilized in site-specific functionalizations, are based on photolabile diazirines, 18 tetrazoles, 19-21 and cyclopropenones.<sup>22</sup>

The concept of 'catch and release', involving selective binding of a species from the reaction media, followed by its controlled

Department of Chemistry and RECETOX, Faculty of Science, Masaryk University, Kamenice 5, 625 00, Brno, Czech Republic. E-mail: slanina.tomas@seznam.cz, klan@sci.muni.cz

liberation, has been used, for example, in combinatorial and solid-phase synthesis of complex natural products<sup>23</sup> and biologically active molecules, 24 DNA nanomachine 25 applications, supramolecular host-guest chemistry26 or resin purification methods.27 The majority of these methods are based on nonphotochemical processes. Nevertheless, involvement of a photochemical step (release) in this strategy has been demonstrated on photocleavable polymers, 28 dendrimers, 29 and liposomes. 30

A reversible version of click reactions would be highly demanded in the regulation and reversible on-off switching of biological processes, advanced surface modifications, or solidphase synthesis. However, the chemical stability of the products of click reactions makes the reverse process very difficult. Several attempts for reversible bioorthogonal reactions have been reported. Aside from the retracted scientific report, 31 only a few 'catch and release' strategies suitable for biological applications have been reported. Catch and release DNA decoys are rare examples of nonnatural DNA probes that capture and dissociate from DNA-binding proteins upon irradiation of the 4-nitroindole linker.<sup>32</sup> Such a strategy has been demonstrated earlier by Porter and coworkers who developed a reagent containing azido and biotin groups separated by a photocleavable linker based on a benzoin photoremovable protecting group for isolation of protein adducts of lipid-derived electrophiles using streptavidin beads.<sup>33</sup> In addition, Popik and coworkers have recently developed a reversible derivatization technique for peptides and proteins based on the reaction of photochemically generated 2-napthoquinone-3-methides.34

Photoremovable protecting groups (PPGs) allow for the precise spatial and temporal release of biologically active molecules.<sup>35</sup> The 4-hydroxyphenacyl photoremovable (pHP) protecting group has been shown to liberate rapidly  $(k_{\rm obs} = (7-100) \times 10^8 \, {\rm s}^{-1})$  and efficiently ( $\Phi = 0.1-1.0$ ) a wide range of leaving groups. <sup>36-40</sup>

In this work, we present a 'catch and photorelease' strategy which combines alkyne-azide 1,3-dipolar cycloaddition between p-hydroxyphenacyl azide 1 and alkyne 2 derivatives to form a 1,2,3-triazole linkage in adduct 3 in the first ('click' or 'catch') step, and subsequent selective irreversible photochemical

<sup>†</sup> Electronic supplementary information (ESI) available: Materials and methods; spectroscopy; synthesis and compound characterization; and NMR and HRMS and optical spectra. See DOI: 10.1039/c6cc07496k

Communication ChemComm

$$\begin{array}{c} \text{Catch} \\ \text{N}_3 \\ \text{R}_1 \\ \text{N}_3 \\ \text{R}_2 \\ \text{Cool} \\ \text{R}_1 \\ \text{R}_3 \\ \text{Photorelease} \\ \text{N} \\ \text{R}_3 \\ \text{Photorelease} \\ \text{N} \\ \text{R}_4 \\ \text{R}_5 \\ \text{R}_5 \\ \text{R}_7 \\ \text{R}_7 \\ \text{R}_8 \\ \text{R}_9 \\ \text{R}_9 \\ \text{R}_9 \\ \text{R}_9 \\ \text{R}_9 \\ \text{R}_9 \\ \text{R}_1 \\ \text{R}_1 \\ \text{R}_2 \\ \text{R}_1 \\ \text{R}_2 \\ \text{R}_3 \\ \text{R}_1 \\ \text{R}_2 \\ \text{R}_3 \\ \text{R}_1 \\ \text{R}_2 \\ \text{R}_3 \\ \text{R}_4 \\ \text{R}_1 \\ \text{R}_2 \\ \text{R}_1 \\ \text{R}_2 \\ \text{R}_3 \\ \text{R}_4 \\ \text{R}_4 \\ \text{R}_5 \\ \text{R}_5 \\ \text{R}_6 \\ \text{R}_7 \\ \text{R}_8 \\ \text{R}_8$$

Scheme 1 A "catch and photorelease" strategy.

release of 1-*H*-1,2,3-triazole 4 and 4-hydroxyphenylacetic acid 5 moieties via a photo-Favorskii rearrangement<sup>41,42</sup> (Scheme 1). In addition, the first step can involve either a Cu(i)-catalyzed or Cu-free alkyne–azide process; a strained alkyne for the latter approach can be generated from the corresponding cyclopropenone via photochemical decarbonylation. <sup>22</sup> We show that these strategies can facilitate efficient bioorthogonal sequential connection and splitting of two (bio)molecules.

Two *p*HP triazole derivatives **3a** and **b** were synthesized by a CuAAC click ('catch') approach from *p*HP azide (**1a** and **b**) and acetylene (**2a** and **b**) derivatives using CuSO<sub>4</sub> and sodium ascorbate in water in 52% and 38% isolated chemical yields, respectively (Scheme 2 and ESI†).<sup>43</sup> The *O*-methylglycine group in the 3-position of **1b** represents a stable amide linkage between the photoactive *p*HP group and a peptide chain.

The second step of this strategy (Scheme 1) involves photochemical release of a triazole moiety from the pHP cage. Irradiation of 3a in CD<sub>3</sub>CN/D<sub>2</sub>O (1:1, v/v) at  $\lambda$  = 313 nm to >99% conversion gave 4-phenyl-1-H-1,2,3-triazole (4a) as the major photoproduct in quantitative yield and 4-hydroxyphenylacetic acid (5a, 80%) and 4-hydroxybenzyl alcohol (6a, 20%) as side-products, identified by comparison of their NMR spectra with those reported before  $^{44,45}$  and using authentic compounds (Scheme 3; Fig. S1, ESI†). Acid 5a is a characteristic product of the photo-Favorskii rearrangement;  $^{46,47}$  but benzyl alcohol (such as 6a) also often accompanies the photolysis of pHP derivatives.  $^{41}$  The major side-photoproducts 5a and 6a are water-soluble and have negligible molar absorption coefficients at 313 nm; thus they do not act as internal optical filters.

The necessary role of water in the pHP photodegradation mechanism $^{36,41,48}$  predetermines this transformation for

HO
$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{3}$$

$$R_{3}$$

$$R_{3}$$

$$R_{3}$$

$$R_{3}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{2}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{1}$$

$$R_{2}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{5}$$

$$R_{5}$$

$$R_{6}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{6}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{6}$$

$$R_{7}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{6}$$

$$R_{7}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{6}$$

$$R_{7}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{6}$$

$$R_{7}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{3}$$

$$R_{4}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{8}$$

$$R_{8}$$

$$R_{9}$$

$$R_{1}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{3}$$

$$R_{4}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{8}$$

$$R_{9}$$

$$R_{1}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R_{4}$$

$$R_{3}$$

$$R_{4}$$

$$R_{5}$$

$$R_{5}$$

$$R_{7}$$

$$R_{8}$$

$$R_{8}$$

$$R_{9}$$

$$R_{1}$$

$$R_{1}$$

$$R_{2}$$

$$R_{3}$$

$$R$$

Scheme 2 Formation of triazole derivatives in the 'catch' step.

HO 
$$R_1$$
  $R_2$   $R_3$   $R_3$   $R_4$   $R_5$   $R_5$   $R_5$   $R_5$   $R_5$   $R_6$   $R_7$   $R_8$   $R_8$   $R_8$   $R_9$   $R$ 

Scheme 3 Release of 1,2,3-triazoles from the pHP PPG 3.

biocompatible aqueous solutions. Acetonitrile in  $CD_3CN/D_2O$  (1:1, v/v) mixtures was used to increase the solubility of the systems, and it resulted in the formation of considerable amounts of the second byproduct **6**, which would not be formed in pure water.<sup>41</sup>

Compound 3b represents a moiety that can interlink the photoactive pHP group with another molecule via a peptide chain. Upon irradiation, this peptide tether, which could bear another molecule or a functional group of interest, remains on the photoproduct(s) formed from the pHP chromophore. The *m*-substitution of the *p*HP group by an amido group was chosen because, unlike substituents in the p- and o-positions, 47 m-substituents do not significantly affect the photophysical properties of pHP.<sup>49</sup> Exhaustive irradiation of 3b in CD<sub>3</sub>CN/D<sub>2</sub>O (1:1, v/v) at  $\lambda = 313$  nm led to the formation of free triazole **4b** in quantitative yield and compounds 5b ( $\approx 25\%$ ) and 6b ( $\approx 20\%$ ) as the major photo-Favorskii rearrangement products (Fig. S2, ESI†). In addition, p-hydroxybenzaldehyde derivative 7 was identified as another side product (Fig. S41, ESI†). Because this compound appeared only after prolonged irradiation, we assume that it is a secondary product formed by 6b oxidation in the presence of oxygen in air.

To increase the quantum yield of photorelease, two modified click adducts  $3\mathbf{c}$  and  $3\mathbf{d}$  were synthesized using a SPAAC (strain-promoted azide-alkyne cycloaddition) and a CuAAC procedure, respectively (Scheme 4 and ESI†). Compound  $3\mathbf{c}$  possesses two electron withdrawing fluorine atoms in the near proximity of the triazole core, which lower its  $pK_a$  value and make it a better leaving group. Indeed, the  $\Phi_{\text{dis}}$  value of  $3\mathbf{c}$  was enhanced by a factor of  $\approx 30$  compared to that of  $3\mathbf{a}$  (Table 1). Because the triazoles  $3\mathbf{a}$ ,  $3\mathbf{b}$  and  $3\mathbf{d}$  were synthesized by CuAAC in the presence of Cu<sup>I</sup> ions, which are toxic due to the generation of reactive oxygen species,  $^{51}$  the non-catalyzed click (SPAAC)

Table 1 Disappearance quantum yields of pHP derivatives 3<sup>a</sup>

ChemComm

pHP derivative	$\Phi_{ m dis}$ /%	
$3a^b$	$0.09\pm0.02$	
$3c^b$ $3d^b$	$2.9\pm0.4$	
$3\mathbf{d}^b$	$83 \pm 4$	
20°	$0.20 \pm 0.05$	

<sup>a</sup> Irradiated at  $\lambda = (313 \pm 1.5)$  nm (3a, 3d) or (320 ± 10) nm (3c, 3e); 4-hydroxyphenacyl fluoride<sup>39</sup> was used as an actinometer. <sup>b</sup> In a mixture of acetonitrile (10%) and acetate buffer (pH = 5.0). In a mixture of acetonitrile (20%) and acetate buffer (pH = 5.0).

Photochemistry of 3c and d.

synthesis of the DIFO-derivative<sup>52</sup> 3c represents a more biocompatible method.

Because controlling the  $pK_a$  values of triazoles is structurally limited, we adopted a different approach. The poor leaving ability of alcohols, phenols or amines can be enhanced by connecting them to the chromophore through a carbonate 53,54 or carbamate<sup>55</sup> linker. Here we modified the triazole moiety through a methylcarbonyloxy linker in 3d, where the leaving group is a carboxylate; thus the  $pK_a$  value of its conjugate acid is approximately 4. The  $\Phi_{\rm dis}$  value of 0.83, which is 3 orders of magnitude higher than that of 3a, is within the values found for other pHP esters  $(\Phi_{\rm dis} = 0.4 - 0.94)^{38}$  and represents a significant improvement for any future applications. A measure of the photorelease (the uncaging cross section), the product of the quantum yield and the decadic molar absorption coefficient,  $\Phi \varepsilon (\lambda_{\rm irr})$ , <sup>35</sup> for **3d** at 313 nm ( $\varepsilon_{313} = 2.73 \times 10^3 \ {\rm M}^{-1} \ {\rm cm}^{-1}$ ) is remarkably large (2.27  $\times$  10 $^3$  M $^{-1}$  cm $^{-1}$ ).

We further modified the first step of the 'catch and photorelease' strategy by introducing an angle-strained cycloalkyne which allows for efficient Cu-free click chemistry via facile 1,3-dipolar cycloaddition (SPAAC). 17,56,57 Such a reactive alkyne can be efficiently photochemically produced in situ from its cyclopropenone precursor, which is an elegant solution to spatial and temporal control of the release.<sup>22,58</sup> In this work, cyclopropenone 8 was synthesized by a 2-step procedure in 27% overall yield (Scheme 5 and ESI†) using a LiNK metalation-coupling procedure.<sup>59</sup> This is a significant improvement when compared to a previously reported 4-step synthesis<sup>60</sup> of cyclopropenone 8. It has been shown that this compound photodecarbonylates with a quantum yield of 0.33 to give a strained alkyne 9, which

'Release, catch and release' strategy

can undergo non-photochemical SPAAC reaction with azides.<sup>22</sup> Thus, a solution of 8 and pHP azide (1a; 1 equiv.,  $c \approx 5 \text{ mg mL}^{-1}$ ) in acetonitrile was irradiated at a wavelength of 350 nm, at which only 8 absorbs (Fig. S6, ESI†). The generated alkyne 9 underwent in situ non-photochemical SPAAC reaction with 1a to form 3e (quantitative yield determined by <sup>1</sup>H NMR; 76% isolated yield). Upon irradiation at 313 nm, this pHP derivative was converted quantitatively into triazole 4e and p-hydroxybenzyl derivatives 5a and 6a, which could bear a peptide tether to connect the released moiety with a biomolecule, via the photo-Favorskii rearrangement. The disappearance quantum yield of 3e was higher than that of 3a because 4e is a better leaving group compared to 4a.

Our 'photorelease, catch and photorelease' strategy is a onepot multistep procedure, which is solely controlled by light, and does not require any other reagents besides water. In addition, both photochemical steps are orthogonal. The pHP group does not absorb at a wavelength of 350 nm (Fig. S39, ESI†) used for generation of strained alkyne 9; thus cycloadduct 3e remains unreacted in the solution during the first irradiation step. The pHP chromophore transformations rely on irradiation at wavelengths in the UV region ( $< \approx 320$  nm), which may limit their bioapplications. However, 2-photon excitation could overcome this issue, as was recently successfully demonstrated by Givens and coworkers on pHP-caged diethyl phosphate and ATP using a 550 nm laser.40

In conclusion, we synthesized a series of substituted pHP azides and alkynes which can be interlinked using an alkyneazide 1,3-dipolar cycloaddition protocol and subsequently be released upon irradiation with UV-light via a photo-Favorskii rearrangement. We show that the reactions can be accomplished in aqueous media favorable for biological applications and that the linker can be used to connect/disconnect two (bio)molecules. A rational design of the leaving group (triazole) structure led to the development of a linker that cleaves upon irradiation with near-unity quantum yield. The second generation of this strategy encompasses chromatically orthogonal photoproduction of a strained alkyne which allows for Cu-free click chemistry. We propose that the systems could be used in bioconjugation or drug-delivery applications.

Communication ChemComm

Support for this work was provided by the Czech Science Foundation (GA13-25775S) and by the Czech Ministry of Education, Youth and Sports (LO1214 and LM2015051).

## References

- 1 J. Kalia and R. T. Raines, Curr. Org. Chem., 2010, 14, 138-147.
- 2 R. Sunasee and R. Narain, Chemistry of Bioconjugates, John Wiley & Sons, Inc., 2014, pp. 1-75.
- 3 F. M. Veronese and M. Morpurgo, Farmaco, 1999, 54, 497-516.
- 4 Q. Wang, T. R. Chan, R. Hilgraf, V. V. Fokin, K. B. Sharpless and M. G. Finn, J. Am. Chem. Soc., 2003, 125, 3192-3193.
- 5 B. S. Gaylord, A. J. Heeger and G. C. Bazan, Proc. Natl. Acad. Sci. U. S. A., 2002, 99, 10954-10957.
- 6 A. Varadarajan, R. M. Sharkey, D. M. Goldenberg and M. F. Hawthorne, Bioconjugate Chem., 1991, 2, 102-110.
- 7 K. A. Totaro, X. Liao, K. Bhattacharya, J. I. Finneman, J. B. Sperry, M. A. Massa, J. Thorn, S. V. Ho and B. L. Pentelute, Bioconjugate Chem., 2016, 27, 994-1004.
- 8 S. D. Fontaine, R. Reid, L. Robinson, G. W. Ashley and D. V. Santi, Bioconjugate Chem., 2015, 26, 145-152.
- 9 M. Howarth, D. J. F. Chinnapen, K. Gerrow, P. C. Dorrestein, M. R. Grandy, N. L. Kelleher, A. El-Husseini and A. Y. Ting, Nat. Methods, 2006, 3, 267-273.
- 10 J. A. Prescher, D. H. Dube and C. R. Bertozzi, Nature, 2004, 430, 873-877.
- 11 J. R. G. Navarro, G. Conzatti, Y. Yu, A. B. Fall, R. Mathew, M. Edén and L. Bergström, Biomacromolecules, 2015, 16, 1293-1300.
- 12 J. M. Baskin and C. R. Bertozzi, *QSAR Comb. Sci.*, 2007, 26, 1211–1219.
- J. B. Haun, N. K. Devaraj, S. A. Hilderbrand, H. Lee and R. Weissleder, Nat. Nanotechnol., 2010, 5, 660-665.
- 14 O. Boutureira and G. J. L. Bernardes, Chem. Rev., 2015, 115, 2174-2195.
- 15 K. Lang and J. W. Chin, ACS Chem. Biol., 2014, 9, 16-20.
- 16 D. M. Patterson, L. A. Nazarova and J. A. Prescher, ACS Chem. Biol., 2014, 9, 592-605.
- J. C. Jewett and C. R. Bertozzi, Chem. Soc. Rev., 2010, 39, 1272-1279.
- 18 A. L. MacKinnon and J. Taunton, Current Protocols in Chemical Biology, John Wiley & Sons, Inc., 2009.
- W. Song, Y. Wang, J. Qu, M. M. Madden and Q. Lin, Angew. Chem., Int. Ed., 2008, 47, 2832-2835.
- 20 W. Song, Y. Wang, Z. Yu, C. I. R. Vera, J. Qu and Q. Lin, ACS Chem. Biol., 2010, 5, 875-885.
- 21 P. An, Z. Yu and Q. Lin, Chem. Commun., 2013, 49, 9920.
- 22 A. A. Poloukhtine, N. E. Mbua, M. A. Wolfert, G.-J. Boons and V. V. Popik, J. Am. Chem. Soc., 2009, 131, 15769-15776.
- 23 I. R. Baxendale, J. Deeley, C. M. Griffiths-Jones, S. V. Ley, S. Saaby and G. K. Tranmer, Chem. Commun., 2006, 2566.
- 24 M. C. Pirrung, L. N. Tumey, A. L. McClerren and C. R. H. Raetz, J. Am. Chem. Soc., 2003, 125, 1575-1586.
- 25 E. M. McConnell, R. Bolzon, P. Mezin, G. Frahm, M. Johnston and M. C. DeRosa, Bioconjugate Chem., 2016, 27, 1493–1499.
- 26 N. Kishi, M. Akita, M. Kamiya, S. Hayashi, H.-F. Hsu and M. Yoshizawa, J. Am. Chem. Soc., 2013, 135, 12976-12979.
- T. Fujii, N. Kato, I. Iwakura, Y. Manabe and M. Ueda, Chem. Lett., 2008, 37, 52-53.
- 28 M. D. Green, A. A. Foster, C. T. Greco, R. Roy, R. M. Lehr, I. I. I. T. H.
- Epps and M. O. Sullivan, Polym. Chem., 2014, 5, 5535-5541. 29 A. Sharma, B. König and N. Jayaraman, New J. Chem., 2014, 38, 3358.
- 30 S. J. Leung and M. Romanowski, Adv. Mater., 2012, 24, 6380-6383.

- 31 The report 'Unclicking the click: mechanically facilitated 1,3-dipolar cycloreversions' by Bielawski and coworkers (Science, 2011, 333, 1606-1609) has been retracted by Science.
- 32 N. B. Struntz and D. A. Harki, ACS Chem. Biol., 2016, 11, 1631-1638.
- 33 H. Y. H. Kim, K. A. Tallman, D. C. Liebler and N. A. Porter, Mol. Cell. Proteomics, 2009, 8, 2080-2089.
- 34 S. Arumugam, J. Guo, N. E. Mbua, F. Friscourt, N. Lin, E. Nekongo, G.-J. Boons and V. V. Popik, Chem. Sci., 2014, 5, 1591.
- 35 P. Klan, T. Solomek, C. G. Bochet, A. Blanc, R. Givens, M. Rubina, V. Popik, A. Kostikov and J. Wirz, Chem. Rev., 2013, 113, 119-191.
- 36 R. S. Givens, A. Jung, C.-H. Park, J. Weber and W. Bartlett, J. Am. Chem. Soc., 1997, 119, 8369-8370.
- 37 R. S. Givens, J. F. W. Weber, P. G. Conrad, G. Orosz, S. L. Donahue and S. A. Thayer, J. Am. Chem. Soc., 2000, 122, 2687-2697.
- 38 R. S. Givens, M. Rubina and J. Wirz, Photochem. Photobiol. Sci., 2012, 11, 472.
- 39 T. Slanina, P. Sebej, A. Heckel, R. S. Givens and P. Klan, Org. Lett., 2015, 17, 4814-4817.
- 40 A. L. Houk, R. S. Givens and C. G. Elles, J. Phys. Chem. B, 2016, 120, 3178-3186.
- 41 R. S. Givens, D. Heger, B. Hellrung, Y. Kamdzhilov, M. Mac, P. G. Conrad, E. Cope, J. I. Lee, J. F. Mata-Segreda, R. L. Schowen and J. Wirz, J. Am. Chem. Soc., 2008, 130, 3307-3309.
- 42 R. S. Givens, M. Rubina and K. F. Stensrud, J. Org. Chem., 2013, 78, 1709-1717.
- 43 The yields were diminished upon purification; the reaction conversion was complete and no other products were found.
- 44 Y. Y. Kuang, K. Baakrishnan, V. Gandhi and X. H. Peng, J. Am. Chem. Soc., 2011, 133, 19278-19281.
- 45 O. Cloarec, A. Campbell, L. H. Tseng, U. Braumann, M. Spraul, G. Scarfe, R. Weaver and J. K. Nicholson, Anal. Chem., 2007, 79, 3304-3311.
- 46 K. Stensrud, J. Noh, K. Kandler, J. Wirz, D. Heger and R. S. Givens, J. Org. Chem., 2009, 74, 5219-5227.
- 47 P. Sebej, B. H. Lim, B. S. Park, R. S. Givens and P. Klan, Org. Lett., 2011, 13, 644-647.
- 48 T. Solomek, D. Heger, B. P. Ngoy, R. S. Givens and P. Klan, J. Am. Chem. Soc., 2013, 135, 15209-15215.
- 49 R. S. Givens, K. Stensrud, P. G. Conrad, A. L. Yousef, C. Perera, S. N. Senadheera, D. Heger and J. Wirz, Can. J. Chem., 2011, 89,
- 50 A. C. Tomé, in Science of Synthesis, 13: Hetarenes and Related Ring Systems, ed. R. C. Storr and T. L. Gilchrist, Georg Thieme Verlag, Stuttgart, 2004, vol. 13, pp. 415-602.
- 51 V. Hong, N. F. Steinmetz, M. Manchester and M. G. Finn, Bioconjugate Chem., 2010, 21, 1912-1916.
- 52 J. M. Baskin, J. A. Prescher, S. T. Laughlin, N. J. Agard, P. V. Chang, I. A. Miller, A. Lo, J. A. Codelli and C. R. Bertozzi, Proc. Natl. Acad. Sci. U. S. A., 2007, 104, 16793-16797.
- 53 A. Banerjee, K. Lee and D. E. Falvey, *Tetrahedron*, 1999, 55, 12699–12710.
- 54 J. Literak, J. Wirz and P. Klan, Photochem. Photobiol. Sci., 2005, 4,
- 55 L. Kammari, L. Plistil, J. Wirz and P. Klan, Photochem. Photobiol. Sci., 2007, 6, 50-56.
- 56 M. F. Debets, J. C. M. van Hest and F. P. J. T. Rutjes, Org. Biomol. Chem., 2013, 11, 6439.
- 57 R. Huisgen, Angew. Chem., Int. Ed. Engl., 1963, 2, 565-598.
- 58 A. Poloukhtine and V. V. Popik, J. Phys. Chem. A, 2006, 110, 1749-1757.
- 59 M. Blangetti, P. Fleming and D. F. O'Shea, J. Org. Chem., 2012, 77, 2870-2877.
- 60 F. R. Fischer and C. Nuckolls, Angew. Chem., Int. Ed., 2010, 49, 7257-7260.