



Cite this: *Chem. Commun.*, 2016, 52, 8612

Received 6th April 2016,
Accepted 14th June 2016

DOI: 10.1039/c6cc02879a

www.rsc.org/chemcomm

Incorporation and visualization of azido-functionalized *N*-oleoyl serinol in Jurkat cells, mouse brain astrocytes, 3T3 fibroblasts and human brain microvascular endothelial cells†

T. Walter,^{‡a} L. Collenburg,^{‡b} L. Japtok,^c B. Kleuser,^c S. Schneider-Schaulies,^b N. Müller,^b J. Becam,^d A. Schubert-Unkmeir,^d J. N. Kong,^e E. Bieberich^e and J. Seibel^{*a}

The synthesis and biological evaluation of azido-*N*-oleoyl serinol is reported. It mimicks biofunctional lipid ceramides and has shown to be capable of click reactions for cell membrane imaging in Jurkat and human brain microvascular endothelial cells.

Ceramide **1** regulates physiological functions including apoptosis, cell-growth, -differentiation, -migration and adhesion and has been implicated¹ in cancer, neurodegeneration, diabetes, microbial pathogenesis, obesity, and inflammation.^{2,3} Bieberich *et al.* described *N*-oleoyl serinol **2** (S18) as a ceramide mimicking molecule (see Fig. 1).^{4,5} The validity of this suggestion is reflected in the fact that it induces apoptosis in rapidly dividing neuroblastoma cells, but not in resting or differentiated cells.⁴ Also, **2** activates atypical protein kinase C (aPKC)⁴ and induces formation of aPKC associated complexes with polarity proteins in stem cell experiments.⁶ The similarity of S18 to ceramide is also represented by the fact that a highly specific antibody against ceramide, also developed by Bieberich *et al.*, binds to S18.⁷ aPKC has been also evaluated successfully as a target for small molecule drug candidates against pancreatic cancer cells.⁸ Here we present preliminary data on the synthesis and biological studies of **3a** and **3b** azido analogues of S18. Ceramide analogs with fluorescent tags proved to be useful for the study of dynamics and visualizing membrane architecture. However, the suitability of such lipids is limited to its ability to behave similar to its unmodified counterpart. Bioorthogonal chemical reporter strategies have addressed this

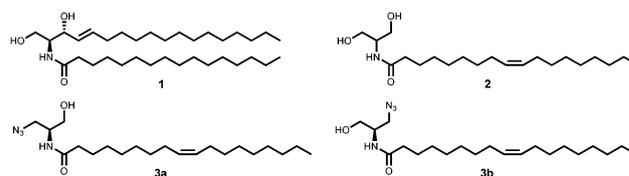
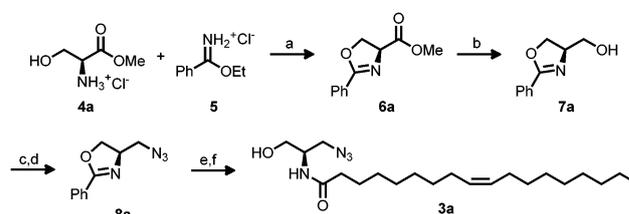


Fig. 1 Structure of C16-ceramide **1**, *N*-oleoyl serinol **2** and its azido-modified analogs **3a**, **3b** used in this work.

issue and successfully applied, *i.e.* the alkyne-azide “click” chemistry. In this context, we aimed to apply click chemistry to *N*-oleoyl serinol mimicking ceramide by introducing an azide group and examining if substitution of one serinol-hydroxyl group by an azide will preserve its biological capabilities. For the synthesis of **3** we used a modified protocol of Hawkins *et al.*^{9,10} with an enantiomer excess (ee) of 92 for **3a** and 95 for **3b**. For detailed information of the synthesis see Scheme 1 and ESI.†

Recently the Golgi apparatus has been imaged using a clicked ceramide analogue by 3D confocal and STED microscopy in HeLa cells.¹¹ Building on those studies, we first examined if the azido-analogues **3a** and **3b** of *N*-oleoyl serinol **2** (S18) shows cell toxicity in Jurkat T cells. Compound **3b** proved to be almost non toxic at a



Scheme 1 Synthesis of azide modified *R* *N*-oleoyl serinol **3a** from *L*-serine methyl ester hydrochloride **4a** or analogue with *D*-serine methyl ester hydrochloride **4b**. Reagents and conditions: (a) NET_3 , CH_2Cl_2 , rt, 24 h, 84% (**6a**), 83% (**6b**). (b) DIBAL-H, thf, 0 °C \rightarrow rt, 24 h, 74% (**7a**), 94% (**7b**). (c) MsCl , NET_3 , DCM, 0 °C, 1 h. (d) dmf, 60 °C, 16 h 80% over two steps (**8a**), 86% over two steps (**8b**). (e) 4 N HCl, 107 °C, 16 h. (f) Oleic acid, DCC, NHS, DIPEA, dmf, rt, 24 h 47% over two steps (**3a**), 73% over two steps (**3b**).

^a Institute for Organic Chemistry, Julius-Maximilians University Würzburg, Am Hubland C1, Würzburg, Germany. E-mail: seibel@chemie.uni-wuerzburg.de

^b Institute of Virology and Immunobiology, Versbacher Str. 7, Würzburg, Germany

^c Institute of Nutritional Science, University of Potsdam, Arthur-Scheunert Allee 114-116, Nuthetal, Potsdam, Germany

^d Institute of Hygiene and Microbiology, Josef-Schneider-Straße 2/E1, Würzburg, Germany

^e Department of Neuroscience and Regenerative Medicine, Medical College of Georgia, Augusta University, 1120 15th Street, Augusta, GA 30912, USA

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c6cc02879a

‡ The authors contributed equally to this work.



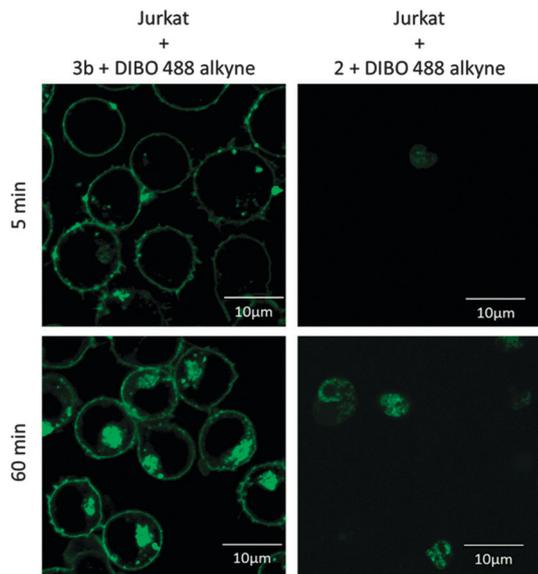


Fig. 2 Jurkat T cells were incubated with 25 μM **3b** (left panels) or **2** (right panels) in HBSS for 30 min at RT. After washing, the cells were exposed to 20 μM DIBO 488 dye and the click reaction was performed for 5 min at RT (upper panels) or for 60 min at 37 $^{\circ}\text{C}$ (lower panels). Cells were washed three times with HBSS and kept at rt until microscopic analysis.

concentration of 25 μM (Fig. S1 and S2, ESI[†]), comparable to the Golgi marker NBD-C6-ceramide.¹² We next aimed to establish if **3b** would be incorporated into the target T cells. DIBO488 dye was coupled under copper-free conditions to **3b** 5 min following feeding of Jurkat cells and its accumulation at the plasma membrane was analyzed over time by flow cytometry.

While NBD-C6-ceramide included as control only slightly peaked after 25 min, this was more pronounced for **3b** which continued to accumulate at the cell surface (see Fig. 2). As revealed by mass spectrometry analyses, 70% (pmol) of **3b** were incorporated into Jurkat cells over time. In contrast to that seen in Jurkat cells, incorporation of **3b** into unstimulated primary T cells was very low. The differential uptake was also observed upon supplementation of the labeling reaction by F-pluronic, which generally increased uptake of the azide-lipids.

To analyze whether its incorporation pattern would be compatible with that of ceramide, subcellular distribution of **3b** was monitored over time in living cells. **3b** efficiently accumulated at the plasma membrane and in the Golgi-compartment. Thus **3b** behaves similar to the Golgi marker NBD-C6-ceramide indicating that it might truly represent a ceramide. Incorporation of the click-labeled compounds appeared to be specific as labeling intensities were dose-dependent and unconjugated, but not conjugated dye diffused into the cytosol. In addition to staining the plasma membrane, clicked **3b** accumulated in an intracellular compartment also targeted by the Golgi-marker NBD-C6-ceramide (see Fig. S6 and S7, ESI[†]). Although both ceramide **1** and azido *N*-oleoyl serinol **3** insert into membranes in the similar way, they can have very different biological functions.

In addition to colocalization studies with NBD-C6 ceramide, we also tested colocalization of **3b** with intracellular structures labeled with an antibody raised against ceramide. We used primary cultured

astrocytes because they were previously shown to enrich ceramide in Golgi and other vesicular compartments.¹³ Fig. 4 shows that there the anti-ceramide antibody labeled compartments that were also positive for **3b**. Consistent with previous results obtained with anti-ceramide antibody and NBD-C6 ceramide labeling of the Golgi, these compartments partially colabeled with an antibody against GM130, a marker for *cis*-Golgi (see Fig. S7, ESI[†]). In a third approach we used human brain microvascular endothelial cells (HBMEC) as serinol acceptors. HBMEC are unique EC representatives as they have continuous intercellular tight junctions (TJs), which control movement of molecules through the EC layer. The EC-pericyte interactions lead to induction of TJ formation. Under inflammatory conditions, pericytes stimulate immune response by production of cytokines.¹⁴ Also it has been shown that *Neisseria meningitidis*, which targets brain ECs, induces the formation of ceramide-enriched platforms on HBMEC to favor bacterial uptake.^{15,16}

One of the fundamental biological functions of ceramides is that they can induce apoptotic cell death.^{17,18} HBMEC were incubated with unmodified *N*-oleoyl serinol **2** and the enantiomeric enriched compounds **3a** and **3b** and cell death were determined. Annexin V binding and propidium iodide (PI) staining enabled to discriminate between early apoptotic, late apoptotic and necrotic cell death. Treatment of HBMEC with 25 μM for 1 h did not result in any toxic effects (Fig. S2A, ESI[†]), however, incubation overnight (16 h) with 10 μM or 50 μM resulted in an increase of late apoptotic and in particular necrotic cells (Fig. S2B, ESI[†]). **3a** and **3b** were comparable with unmodified *N*-oleoyl serinol **2**. They are not toxic below concentrations of 50 μM (see Fig. S2, ESI[†]). Bieberich *et al.* observed that *N*-oleoyl serinol **2** induces apoptosis only in rapidly dividing neuroblastoma cells. Here **3** also induced apoptosis at similar concentrations. **3b** incorporation in HBMEC was efficiently (see Fig. 3). While **3b** is found in the membrane after 5 min

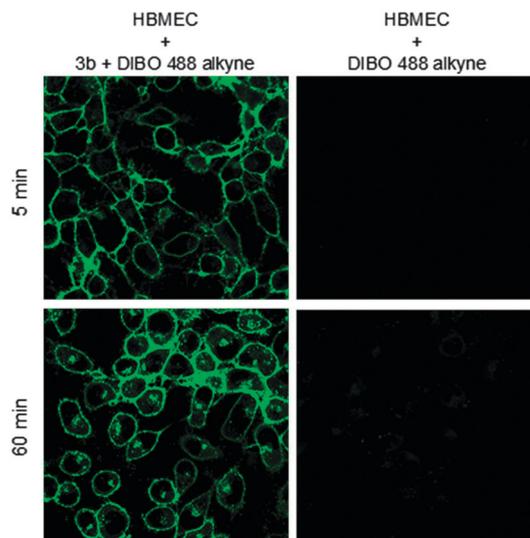


Fig. 3 HBMEC were treated with 25 μM **3b** (left panels) in RPMI medium for 5 min at RT or were left untreated (right panels). After washing, the cells were exposed to 25 μM DIBO 488 alkyne and click reaction was performed for 5 min at RT (upper panels) or for 60 min at 37 $^{\circ}\text{C}$ (lower panels). Cells were washed three times with PBS and kept at RT until microscopic analysis.



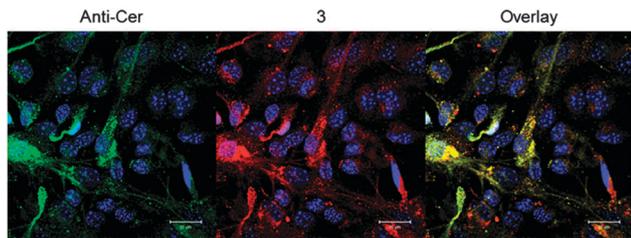


Fig. 4 Primary cultured astrocytes were incubated with 5 μM each **3a** and **b** for 1 h, washed with PBS, and then click reaction performed with 5 μM Alexa 546 DIBO alkyne for 20 min at RT. After washing, cells were fixed with *p*-formaldehyde/glutaraldehyde and permeabilized under mild conditions (0.2% Triton X-100/PBS, 5 min, RT). Immunocytochemistry and co-labeling was performed with anti-ceramide rabbit IgG and Alexa 488 anti-rabbit IgG.

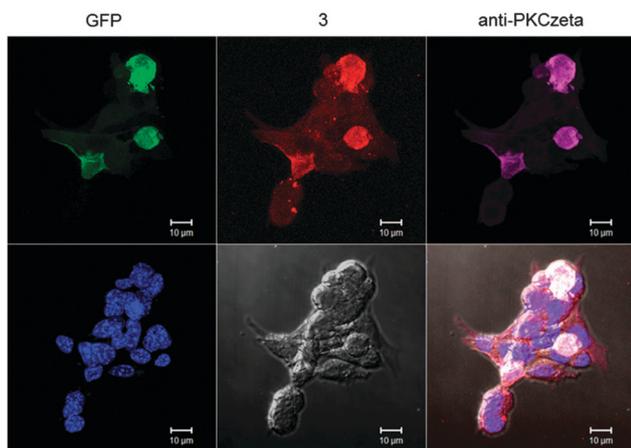


Fig. 5 3T3 fibroblasts were transfected with human PKC ζ -GFP and 24 h post-transfection, incubated for 1 h with 5 μM each **3a** and **3b**. Click reaction was performed with 5 μM Alexa 546 DIBO alkyne for 20 min at RT. After washing, cells were fixed with *p*-formaldehyde/glutaraldehyde and permeabilized under mild conditions (0.2% Triton X-100/PBS, 5 min, RT). Immunocytochemistry and co-labeling was performed with anti-PKC ζ rabbit IgG and Alexa 647 anti-rabbit IgG.

exclusively, after 60 min it is also transported in the Golgi apparatus (see Fig. 2, lower panels). We hypothesized that **3** may thus be useful in evaluation of PKC ζ -mediated signal cascades. To test this hypothesis we analyzed potential interaction of the ceramide analog with PKC ζ . We expressed the fusion protein human PKC ζ -GFP in 3T3 fibroblasts and performed incubation and click reaction with a mixture of **3a** and **b**. Fig. 5 shows that GFP fluorescence as well as anti-PKC ζ antibody colocalized with **3a/b**, suggesting that the ceramide analog interacts with PKC ζ . These results indicate that **3** is useful to study the function of ceramide and its interaction with target proteins such as PKC ζ for neural cell development and cell polarity. On this basis further anti-cancer strategies include the inhibition of the protein kinase C signaling pathways. For the understanding of the bacterial uptake process and the role of ceramide-enriched platforms on HBMEC these ceramide analogues might be a good tool for studying Par6 and aPKCs interactions. The Par6–aPKC

interaction is necessary for the establishment and maintenance of the cell polarity complex in non-transformed cells.¹⁹ Moreover it has been shown that infection of brain ECs with *N. meningitidis* result in recruitment of the polarity complex Par3/Par6/Pkc ζ underneath the attached bacterial microcolonies, which is followed by the delocalization of junctional proteins from the intercellular junctions.²⁰

In summary we have demonstrated the incorporation and visualization of azido-functionalized *N*-oleoyl serinol by bioorthogonal click chemistry in Jurkat cells and human brain microvascular endothelial cells. We have also shown that apoptosis of the HBMEC can be titrated. It should be noted that more than half of the mitotic glia cells and neurons die during fetal brain development by apoptosis. The stained ceramide analogue can be localized. Thus it might be a helpful tool to investigate sphingosine-1-phosphate and ceramide, and their metabolites roles in endothelial barrier function, which may be a further advancement in this field.

This study was funded through the Deutsche Forschungsgemeinschaft (RU2123) and NIH grants R01 R01AG034389 and R56NS095215 (to E. B.)

Notes and references

- 1 Y. A. Hannun and L. M. Obeid, *Nat. Rev. Mol. Cell Biol.*, 2008, **9**, 139–150.
- 2 Y. H. Zeidan and Y. A. Hannun, *Trends Mol. Med.*, 2007, **13**, 327–336.
- 3 D. Wu, Z. Ren, M. Pae, W. Guo, X. Cui, A. H. Merrill and S. N. Meydani, *J. Immunol.*, 2007, **179**, 4829–4839.
- 4 E. Bieberich, T. Kawaguchi and R. K. Yu, *J. Biol. Chem.*, 2000, **275**, 177–181.
- 5 E. Bieberich, B. Hu, J. Silva, S. MacKinnon, R. K. Yu, H. Fillmore, W. C. Broaddus and R. M. Ottenbrite, *Cancer Lett.*, 2002, **181**, 55–64.
- 6 G. Wang, J. Silva, K. Krishnamurthy, E. Tran, B. G. Condie and E. Bieberich, *J. Biol. Chem.*, 2005, **280**, 26415–26424.
- 7 K. Krishnamurthy, S. Dasgupta and E. Bieberich, *J. Lipid Res.*, 2007, **48**, 968–975.
- 8 A. M. Butler, M. L. Scotti Buzhardt, E. Erdogan, S. Li, K. S. Inman, A. P. Fields and N. R. Murray, *Oncotargets Ther.*, 2015, **6**, 15297–15310.
- 9 L. D. Hawkins and S. T. Ishizaka, *US Pat.*, 20030153532, 2004.
- 10 L. D. Hawkins and S. T. Ishizaka, *WO Pat.*, 2003099195, 2003.
- 11 R. S. Erdmann, H. Takakura, A. D. Thompson, F. Rivera-Molina, E. S. Allgeyer, J. Bewersdorf, D. Toomre and A. Schepartz, *Angew. Chem.*, 2014, **53**, 10242–10246.
- 12 N. G. Lipsky and R. E. Pagano, *Science*, 1985, **228**, 745–747.
- 13 G. Wang, M. Dinkins, Q. He, G. Zhu, C. Poirier, A. Campbell, M. Mayer-Proschel and E. Bieberich, *J. Biol. Chem.*, 2012, **287**, 21384–21395.
- 14 B. Obermeier, R. Daneman and R. M. Ransohoff, *Nat. Med.*, 2013, **19**, 1584–1596.
- 15 A. Simonis, S. Hebling, E. Gulbins, S. Schneider-Schaulies and A. Schubert-Unkmeir, *PLoS Pathog.*, 2014, **10**, e1004160.
- 16 A. Unkmeir, K. Latsch, G. Dietrich, E. Wintermeyer, B. Schinke, S. Schwender, K. S. Kim, M. Eigenthaler and M. Frosch, *Mol. Microbiol.*, 2002, **46**, 933–946.
- 17 L. M. Obeid, C. M. Linardic, L. A. Karolak and Y. A. Hannun, *Science*, 1993, **259**, 1769–1771.
- 18 O. Cuivillier, G. Pirianov, B. Kleuser, P. G. Vanek, O. A. Coso, S. Gutkind and S. Spiegel, *Nature*, 1996, **381**, 800–803.
- 19 Y. Hirano, S. Yoshinaga, R. Takeya, N. N. Suzuki, M. Horiuchi, M. Kohjima, H. Sumimoto and F. Inagaki, *J. Biol. Chem.*, 2005, **280**, 9653–9661.
- 20 M. Coureuil, G. Mikaty, F. Miller, H. Lecuyer, C. Bernard, S. Bourdoulous, G. Dumenil, R. M. Mege, B. B. Weksler, I. A. Romero, P. O. Couraud and X. Nassif, *Science*, 2009, **325**, 83–87.

