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Thioether bond formation by SPASM domain radical SAM enzymes: C α H-atom abstraction in subtilisin A biosynthesis†

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AlbA is a radical SAM enzyme catalyzing the formation of three unusual thioether bonds in the antibiotic subtilisin A. We demonstrate here that AlbA catalyzes direct C α H-atom abstraction and likely contains three essential [4Fe–4S] centers. This leads us to propose novel mechanistic perspectives for thioether bond catalysis by radical SAM enzymes.

Subtilisin A is a bacteriocin produced by *Bacillus subtilis* targeting several pathogens such as *Listeria monocytogenes*.¹ This bacteriocin is the founding member of sactipeptides, a diverse group of ribosomally synthesized and post-translationally modified peptides (RiPPs) characterized by the presence of unusual sulfur to α -carbon thioether bridges (Fig. 1a). Since the discovery of subtilisin A, which contains two Cys-to-Phe and one Cys-to-Thr linkages,² several sactipeptides have been identified including the sporulation killing factor of *Bacillus subtilis*³ and two bacteriocins produced by *Bacillus thuringiensis* called thuricin CD⁴ and thurincin H,⁵ respectively. However, mechanistic details regarding the enzymes catalyzing these reactions are still lacking notably because of the high insolubility of their peptide substrates.²

The biosynthesis of subtilisin A is dependent upon a 43-amino-acid peptide precursor (SboA) and the AlbA enzyme.¹ Recently, it has been demonstrated that AlbA is a radical SAM (rSAM) enzyme catalyzing the formation of the three thioether bridges in subtilisin A providing the first biochemical information on the biosynthesis of sactipeptides.⁶ AlbA belongs to the SPASM-domain group of rSAM enzymes⁷ which catalyze peptide or protein post-translational modifications and are characterized by the presence of a specific protein domain containing

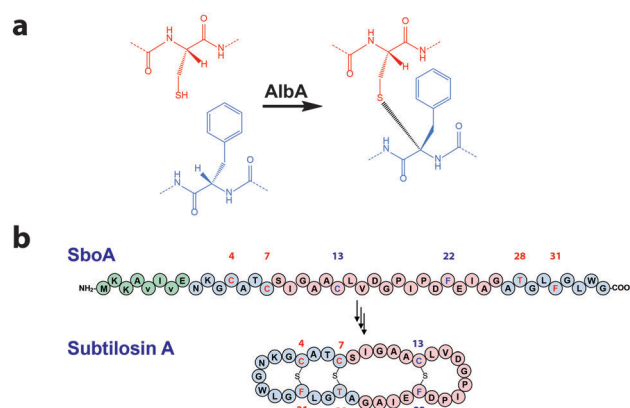


Fig. 1 (a) Reaction catalyzed by AlbA. (b) Sequence of the pro-peptide SboA and structure of the mature subtilisin A. Numbers indicate the position of the residues involved in thioether bonds (i.e. **4**, **7** & **13**: Cys; **22** & **31**: Phe; **28**: Thr). Amino acids involved in the L-configured thioether bond are labeled in blue (i.e. L-Phe **22**) and those involved in D-configured bond are labeled in red (i.e. D-Phe **31** and D-Thr **28**). Green circles indicate the leader peptide. Red circles correspond to the sequence of **SboA**₈₋₂₇ and blue circles correspond to **SboA**₂₇₋₆ (see Fig. 2).

[4Fe–4S] centers.^{6,8–10} However, to date, the function of the SPASM-domain is still unclear.⁷

To probe the mechanism of AlbA, we rationally designed synthetic peptides based on the sequence of the mature subtilisin A (Fig. 1b & Fig. S1, ESI†). These two peptides, **SboA**₈₋₂₇ and **SboA**₂₇₋₆, encompassed the full sequence of subtilisin A and were designed to allow the formation of only one thioether bond in order to dissect catalysis. The sequence of **SboA**₈₋₂₇ corresponded to residues 8–27 while **SboA**₂₇₋₆ contained the residues 27–35 covalently linked to residues 1–6 by an amide bond (Fig. 1b, 2a and b).

After anaerobic reconstitution (Fig. S2, ESI†), AlbA activity was assayed in the presence of **SboA**₈₋₂₇ and S-adenosyl-L-methionine (SAM) under reducing conditions. As shown, AlbA efficiently cleaved SAM, although no substrate modification was evidenced, even after an extended incubation time

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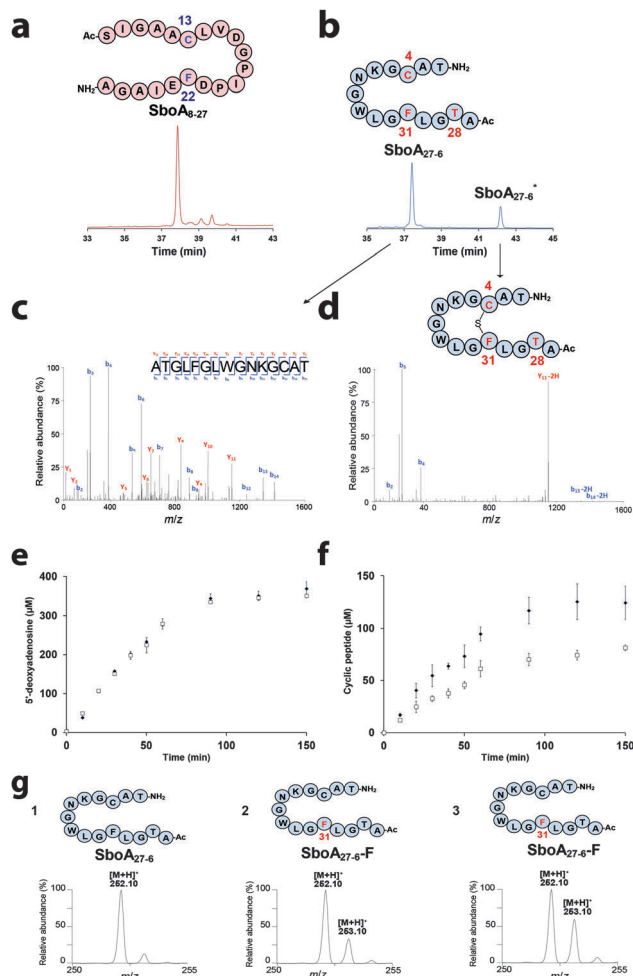


Fig. 2 AlBA activity on synthetic peptides. HPLC analysis of AlBA reaction in the presence of (a) **SboA₈₋₂₇** or (b) **SboA₂₇₋₆** after 120 min (UV detection at 215 nm). LC-MS² analysis of **SboA₂₇₋₆** (c) and the cyclic peptide **SboA₂₇₋₆*** (d) produced by AlBA. Relevant fragments of the *b* and *y* series are indicated (see Fig. S5 and Table S2, ESI[†] for complete assignment). Time course for the production of 5'-dA (e) and cyclic peptide (f) by AlBA in the presence of **SboA₂₇₋₆** (■) or the deuterated substrate, **SboA₂₇₋₆-F** (□). LC-MS analysis of 5'-dA produced by AlBA (g) in the presence of (1) **SboA₂₇₋₆**, (2) **SboA₂₇₋₆-F** using sodium dithionite (DT) or (3) **SboA₂₇₋₆-F** using flavodoxin/flavodoxin reductase/NADPH (Fldx/Fpr) as a reducing system. AlBA (20 μM) was incubated in the presence of SAM (1 mM), peptide substrate (1 mM) and sodium dithionite (2 mM) or Fldx/Fpr.

(Fig. 2a, Fig. S3 & Table S1, ESI[†]). In contrast, with **SboA₂₇₋₆**, a product (**SboA₂₇₋₆***) with a retention time of 42.15 min was formed along with 5'-deoxyadenosine (5'-dA) (Fig. 2b & Fig. S3, ESI[†]). LC-MS² analysis showed that this new peptide had a mass shift of -2 Da (Fig. S4 and S5, ESI[†]) and contained a covalent bond between Cys⁴ and Phe³¹ as in authentic subtilisin A² (Fig. 1b, 2c, d and Table S2, ESI[†]). This result was consistent with AlBA catalyzing the formation of a thioether bond on **SboA₂₇₋₆**. Kinetic analysis showed that AlBA exhibited a specific activity of 1.59 ± 0.1 nmol min⁻¹ mg⁻¹ and 4.72 ± 0.02 nmol min⁻¹ mg⁻¹ for the production of the cyclic peptide and 5'-dA, respectively (Fig. 2e and f). In two hours ~ 120 μM of a cyclic peptide was produced, a result similar to the one

estimated using **SboA** as the substrate.⁶ Unexpectedly, the short peptide **SboA₂₇₋₆**, despite being deprived of the leader peptide, proved to be an efficient substrate suitable to dissect enzyme catalysis.

In order to demonstrate that AlBA catalyzes substrate H-atom abstraction and to identify the target of the 5'-dA[•] radical, we synthesized two **SboA₂₇₋₆** derivatives: **SboA₂₇₋₆-F** and **SboA₂₇₋₆-C** containing a per-deuterated phenylalanine or a cysteine (β,β -D₂), respectively. With **SboA₂₇₋₆-F**, we obtained a cyclic peptide with a mass shift of -3 Da corresponding to the loss of a H-atom and a D-atom. LC-MS² confirmed the formation of a thioether bond between Cys⁴ and the deuterated Phe³¹ (Fig. S6, ESI[†]). In the presence of **SboA₂₇₋₆-F**, AlBA exhibited a specific activity of 0.96 ± 0.01 nmol min⁻¹ mg⁻¹ for thioether bond formation and a SAM cleavage activity of 4.66 ± 0.07 nmol min⁻¹ mg⁻¹ (Fig. 2e and f).

Apparent from these kinetic experiments, a decoupling between SAM cleavage and product formation occurred with a ratio between 5'-dA and the cyclic peptide of ~ 2.9 . This ratio increased to 5 in the presence of the deuterated substrate. Comparing the reaction rate for cyclic peptide production using the unlabeled or the deuterated substrate, we determined a kinetic isotope effect of ~ 1.7 (Fig. 2e and f). This behavior mirrored what has been reported for another SPASM-domain enzyme, the anaerobic Sulfatase-Maturing Enzyme (anSME)⁸⁻¹⁰ when the amino acid residue targeted by this enzyme was specifically deuterated.¹¹

To definitively determine if Phe³¹ was the target of the 5'-dA[•] radical, we analyzed the 5'-dA produced in reaction using LC-MS (Fig. 2g). As shown, $\sim 20\%$ deuterium was incorporated into the 5'-dA produced when **SboA₂₇₋₆-F** was used as the substrate (Fig. 2g trace 2). This result was in-line with the 5-to-1 ratio measured between 5'-dA and cyclic peptide production. To improve the coupling between both reactions, we substituted sodium dithionite by the physiological reducing system: flavodoxin/flavodoxin reductase/NADPH.^{12,13} Under these conditions, deuterium incorporation increased up to 50% (Fig. 2g trace 3). In contrast, with **SboA₂₇₋₆-C** (labeled on Cys⁴), no labeling was introduced into the 5'-dA produced, even when the physiological reducing system was used (Fig. S7, ESI[†]). This result excluded possible hydride shifts between residues during catalysis. Altogether, these labeling experiments strongly support that AlBA catalyzes direct C_α H-atom abstraction on Phe³¹ and it is the rate determining step of the reaction.

In addition of the rSAM center, AlBA was hypothesized to contain a second [4Fe-4S] center coordinated by three vicinal cysteine residues (*i.e.* Cys⁴⁰⁸, Cys⁴¹⁴ and Cys⁴¹⁷) with a critical role for substrate interaction.⁶ However, the crystal structure of anSME demonstrated that the [4Fe-4S] centers coordination in the SPASM-domain involves remote cysteine residues and that [4Fe-4S] centers do not coordinate the enzyme substrate.¹⁴ Furthermore, structural predictions positioned the SPASM [4Fe-4S] center of AlBA too far away from the enzyme active site to allow direct substrate interaction.⁷ Using a 3D structural model (see methods in the ESI[†]), we identified 7 cysteine residues in the AlBA SPASM-domain which perfectly superimposed and



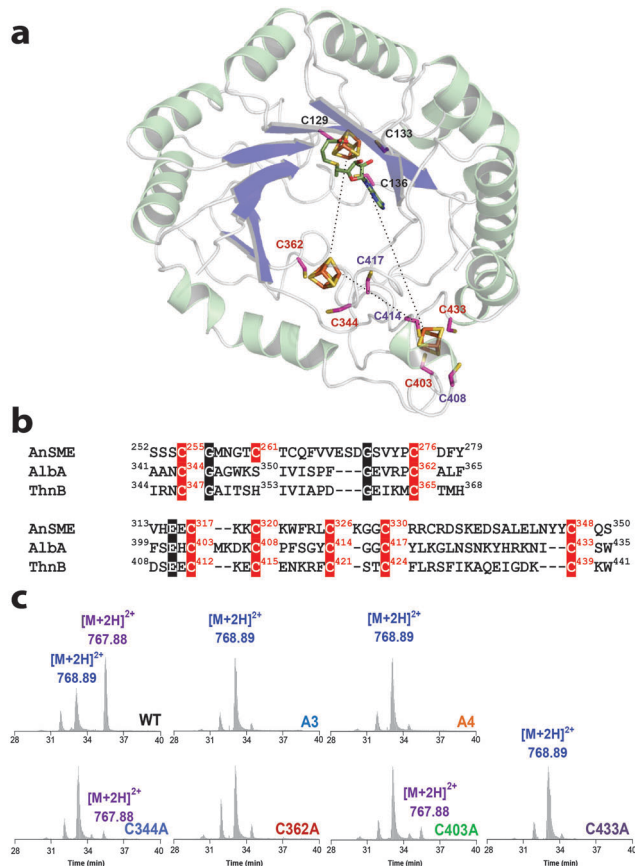


Fig. 3 Role of the Alba SPASM-domain. (a) Structural model of Alba with three iron-sulfur centers. The cysteine residues involved in the coordination of [4Fe-4S] centers are depicted in pink. Numbers in purple indicate cysteine residues from the SPASM-domain previously mutated.⁶ (b) Sequence alignment of the rSAM enzymes: anSME, Alba and ThnB. Highlighted in red, the anSME SPASM-domain cysteine residues involved in the coordination of [4Fe-4S] centers and the corresponding residues in Alba and ThnB. In black are highlighted other strictly conserved residues. (c) Activity of Alba wild-type (WT) and cysteine mutants: **A3**, **A4**, **C344A**, **C362A**, **C403A** and **C433A** measured using LC-MS. The **SboA**₂₇₋₆ peptide has an m/z value of $[M + 2H]^{2+} = 768.89$ and the cyclic peptide **SboA**₂₇₋₆* has an m/z value of $[M + 2H]^{2+} = 767.88$. Each protein (20 μ M) was incubated under anaerobic and reducing conditions in the presence of SAM (1 mM), peptide substrate (1 mM) and sodium dithionite (2 mM).

aligned with anSME (Fig. 3a and b & Fig. S8, ESI[†]). This model predicted that not only Cys⁴⁰⁸, Cys⁴¹⁴ and Cys⁴¹⁷ but also Cys³⁴⁴, Cys³⁶², Cys⁴⁰³ and Cys⁴³³ are involved in [4Fe-4S] center coordination.⁶ Hence, not one but two [4Fe-4S] centers were predicted in the SPASM-domain of Alba. To probe this hypothesis, we generated the following single mutants: **C344A**, **C362A**, **C403A** and **C433A**. In addition, we also produced one mutant lacking these 4 cysteine residues (Mutant **A4**) and, as a control, a 6th mutant lacking the radical SAM binding motif CxxCxxC (Mutant **A3**) (Fig. S9, ESI[†]).

The UV-visible spectra of the reconstituted **C344A**, **C362A**, **C403A** and **C433A** mutants were identical to the wild-type enzyme while the **A3** and **A4** mutants exhibited a decrease in absorbance in the region between 320 and 420 nm, indicating partial iron-sulfur loading (Fig. S10, ESI[†]). Assayed in the

presence of SAM and the **SboA**₂₇₋₆ substrate, all mutants, except the **A3**, cleaved SAM into 5'-dA (Fig. S11, ESI[†]). However, they were all impaired for thioether bond formation with only the **C344A** and **C403A** mutants producing detectable levels of a cyclic peptide ($[M + 2H]^{2+}$: 767.88) (Fig. 3c & Fig. S11, ESI[†]). The 7 conserved cysteine residues are thus critical for Alba activity.

In addition to subtilosin A, recent studies on the sporulation killing factor,¹⁵ thuricin CD⁴ and thurincin H⁵ have shown that rSAM enzymes are responsible for the formation of sulfur to α -carbon thioether bridges. These rSAM enzymes belong either to the SPASM- (Alba & ThnB) or twitch-domain (SkfB & TrnCD) groups.⁷ Interestingly, Alba and ThnB despite having no significant homology (<25% identity) possess the 7 conserved cysteine residues (Fig. 3b). For all these enzymes, it has been speculated that they catalyze C α H-atom abstraction on a diverse range of amino acid residues; however, no experimental proof has been provided thus far.

Our results support that Alba, and likely all known rSAM enzymes involved in sactipeptide thioether bond formation, catalyzes C α H-atom abstraction. Similarly to what was demonstrated for anSME,⁹ H-atom abstraction is the rate determining step of the reaction. Our study also supports that Alba contains two SPASM [4Fe-4S] centers coordinated by 7 cysteine residues. While it remains to be determined whether they have partial or full ligation, sequence alignment (Fig. 3b) showed that Cys²⁶¹ in anSME aligns with Ser³⁵⁰ in Alba. Interestingly, a Ser residue has been recently reported to be involved in the coordination of the auxiliary [4Fe-4S] center in the rSAM enzyme LipA.¹⁶

Remarkably, the 3 thioether bonds catalyzed by Alba have different (L/D) stereochemistries. In the case of lanthipeptides, which are by far the most well-studied thioether bond-containing peptides, it was elegantly demonstrated that substrate flexibility only governs thioether bond configuration.¹⁷ Furthermore, we showed here that even single mutations, which likely alter the redox properties of the [4Fe-4S] centers, are sufficient to dramatically alter enzyme activity. Collectively, these data support a role as an electron conduit for the auxiliary [4Fe-4S] centers rather than a role in substrate coordination which appears inconsistent with the formation of thioether bonds with both stereochemistry.^{10,17}

In conclusion, we propose a novel mechanism for the catalysis of thioether bond formation by rSAM enzymes (Fig. 4). Following SAM cleavage, the 5'-dA radical abstracts a C α H-atom on Phe (or Thr), a process thermodynamically favorable considering the C α -H-atom bond dissociation energies of 348.9 kJ mol⁻¹ and 345.1 kJ mol⁻¹ for Phe and Thr respectively.¹⁸ Among rSAM enzymes, the pyruvate formate lyase activase (PFL-AE)¹⁹ and the ribonucleotide reductase activase (RNR-AE)²⁰ have been shown to catalyze C α H-atom abstraction of a glycine residue. However, because of steric interactions between the Phe side chain and the peptide backbone, Alba generates a less stable tertiary C α carbon-centered radical than the secondary glycy radical produced by PFL-AE²¹ or RNA-AE.²⁰ This carbon-centered radical intermediate will have a high propensity to abstract H-atoms and thus cannot react directly with cysteine to form a thioether bond. Hence, it has been



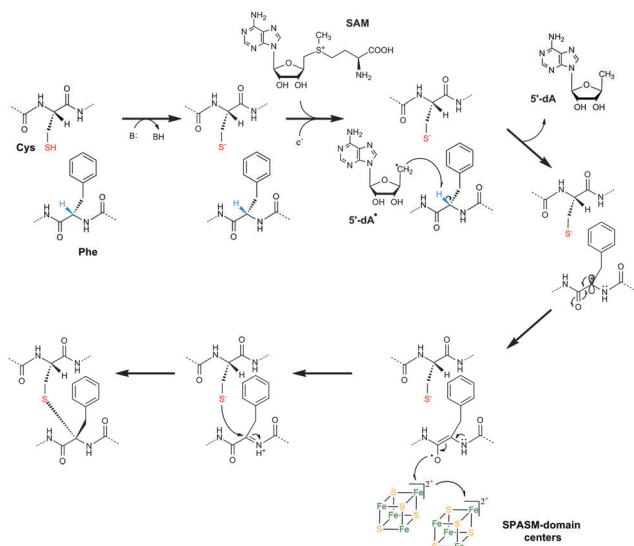


Fig. 4 Proposed mechanism for AlbA.

proposed that the substrate interacts first through its cysteine residues with a SPASM [4Fe-4S] center,⁶ a process presumed to allow direct addition of the C_α carbon-centered radical to the S-atom. However, as detailed above, AlbA exhibits strong biochemical and structural analogies with anSME. This latter enzyme was shown to catalyze C_β H-atom abstraction⁹ leading to the formation of a C-S bond between the C_β-atom and the deprotonated thiol group of its cysteine substrate.²² Based on structural and biochemical data, it is established that the SPASM [4Fe-4S] centers of anSME fulfill a key redox function which remains to be fully elucidated.^{14,22} In the case of AlbA, our data support a similar mechanism. After H-atom abstraction and loss of the C_α-atom stereochemistry (of Phe or Thr), the radical intermediate likely oxidizes to an *N*-acyliminium ion (Fig. 4). This intermediate can be readily trapped by the nucleophilic thiolate group of a remote cysteine leading to the formation of the C-S bond. In support of this hypothesis, the chemical synthesis of cyclic peptides containing thioether bonds has been shown to involve *N*-acyliminium intermediates.²³ Depending on the attack on the *N*-acyliminium intermediate, *L*- or *D*-configured bonds are formed, guided by substrate flexibility.¹⁷ In this mechanistic hypothesis, the SPASM [4Fe-4S] centers of AlbA are keys for the oxidation of the radical intermediate as proposed for anSME^{9,24} and for the formation of the *N*-acyliminium intermediate. This mechanism represents a novel redox alternative to the formation of thioether bonds in natural products.

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Notes and references

- 1 G. Zheng, R. Hehn and P. Zuber, *J. Bacteriol.*, 2000, **182**, 3266–3273.
- 2 K. E. Kawulka, T. Sprules, C. M. Diaper, R. M. Whittall, R. T. McKay, P. Mercier, P. Zuber and J. C. Vederas, *Biochemistry*, 2004, **43**, 3385–3395.
- 3 J. E. Gonzalez-Pastor, E. C. Hobbs and R. Losick, *Science*, 2003, **301**, 510–513.
- 4 M. C. Rea, C. S. Sit, E. Clayton, P. M. O'Connor, R. M. Whittall, J. Zheng, J. C. Vederas, R. P. Ross and C. Hill, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 9352–9357.
- 5 B. M. Wiecekowsky, J. D. Hegemann, A. Mielcarek, L. Boss, O. Burghaus and M. A. Marahiel, *FEBS Lett.*, 2015, **589**, 1802–1806.
- 6 L. Flühe, T. A. Knappe, M. J. Gattner, A. Schäfer, O. Burghaus, U. Linne and M. A. Marahiel, *Nat. Chem. Biol.*, 2012, **8**, 350–357.
- 7 T. A. Grell, P. J. Goldman and C. L. Drennan, *J. Biol. Chem.*, 2015, **290**, 3964–3971.
- 8 A. Benjdia, J. Leprince, A. Guillot, H. Vaudry, S. Rabot and O. Berteau, *J. Am. Chem. Soc.*, 2007, **129**, 3462–3463.
- 9 A. Benjdia, J. Leprince, C. Sandstrom, H. Vaudry and O. Berteau, *J. Am. Chem. Soc.*, 2009, **131**, 8348–8349.
- 10 A. Benjdia, S. Subramanian, J. Leprince, H. Vaudry, M. K. Johnson and O. Berteau, *J. Biol. Chem.*, 2008, **283**, 17815–17826.
- 11 O. Berteau, A. Guillot, A. Benjdia and S. Rabot, *J. Biol. Chem.*, 2006, **281**, 22464–22470.
- 12 L. Decamps, B. Philmus, A. Benjdia, R. White, T. P. Begley and O. Berteau, *J. Am. Chem. Soc.*, 2012, **134**, 18173–18176.
- 13 B. Philmus, L. Decamps, O. Berteau and T. P. Begley, *J. Am. Chem. Soc.*, 2015, **137**, 5406–5413.
- 14 P. J. Goldman, T. L. Grove, L. A. Sites, M. I. McLaughlin, S. J. Booker and C. L. Drennan, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 8519–8524.
- 15 L. Fluhe, O. Burghaus, B. M. Wiecekowsky, T. W. Giessen, U. Linne and M. A. Marahiel, *J. Am. Chem. Soc.*, 2013, **135**, 959–962.
- 16 J. E. Harmer, M. J. Hiscox, P. C. Dinis, S. J. Fox, A. Iliopoulos, J. E. Hussey, J. Sandy, F. T. Van Beek, J. W. Essex and P. L. Roach, *Biochem. J.*, 2014, **464**, 123–133.
- 17 W. Tang, G. Jimenez-Oses, K. N. Houk and W. A. van der Donk, *Nat. Chem.*, 2015, **7**, 57–64.
- 18 B. N. Moore and R. R. Julian, *Phys. Chem. Chem. Phys.*, 2012, **14**, 3148–3154.
- 19 J. Knappe, F. A. Neugebauer, H. P. Blaschkowski and M. Ganzler, *Proc. Natl. Acad. Sci. U. S. A.*, 1984, **81**, 1332–1335.
- 20 X. Sun, S. Ollagnier, P. P. Schmidt, M. Atta, E. Mulliez, L. Lepape, R. Eliasson, A. Graslund, M. Fontecave, P. Reichard and B. M. Sjöberg, *J. Biol. Chem.*, 1996, **271**, 6827–6831.
- 21 A. F. Wagner, M. Frey, F. A. Neugebauer, W. Schafer and J. Knappe, *Proc. Natl. Acad. Sci. U. S. A.*, 1992, **89**, 996–1000.
- 22 A. Benjdia, S. Subramanian, J. Leprince, H. Vaudry, M. K. Johnson and O. Berteau, *FEBS J.*, 2010, **277**, 1906–1920.
- 23 T. E. Nielsen, S. Le Qument and M. Meldal, *Org. Lett.*, 2005, **7**, 3601–3604.
- 24 A. Benjdia, G. Deho, S. Rabot and O. Berteau, *FEBS Lett.*, 2007, **581**, 1009–1014.

