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Co-adsorption of peptide amphiphile V<sub>6</sub>K and conventional surfactants SDS and C<sub>12</sub>TAB at the solid/water interface<sup>†</sup>

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Recent research has reported many attractive benefits from short peptide amphiphiles. A practical route for them to enter the real world of applications is through formulation with conventional surfactants. This study reports the co-adsorption of the surfactant-like peptide, V6K, with conventional anionic and cationic surfactants at the solid/water interface. The time-dependant adsorption behaviour was examined using spectroscopic ellipsometry whilst adsorbed layer composition and structural distribution of the components were investigated by neutron reflection with the use of hydrogen/deuterium labelling of the surfactant molecules. Both binary (surfactant/peptide mixtures) and sequential (peptide followed by surfactant) adsorption have been studied. It was found that at the hydrophilic SiO2/water interface, the peptide was able to form a stable, flat, defected bilayer structure however both the structure and adsorbed amount were highly dependent on the initial peptide concentration. This consequently affected surfactant adsorption. In the presence of a pre-adsorbed peptide layer anionic sodium dodecyl sulfate (SDS) could readily co-adsorb at the interface; however, cationic dodecyl trimethyl ammonium bromide (C12TAB) could not co-adsorb due to the same charge character. However on a trimethoxy octyl silane (C<sub>8</sub>) coated hydrophobic surface, V<sub>6</sub>K formed a monolayer, and subsequent exposure to cationic and anionic surfactants both led to some co-adsorption at the interface. In binary surfactant/peptide mixtures, it was found that adsorption was dependent on the molar ratio of the surfactant and peptide. For SDS mixtures below molar unity and concentrations below CMC for C12TAB, V6K was able to dominate adsorption at the interface. Above molar unity, no adsorption was detected for SDS/V<sub>6</sub>K mixtures. In contrast, C<sub>12</sub>TAB gradually replaced the peptide and became dominant at the interface. These results thus elucidate the adsorption behaviour of  $V_6K$ , which was found to dominate interfacial adsorption but its exact adsorbed amount and distribution were affected by interfacial hydrophobicity and interactions with conventional surfactants.

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# 1. Introduction

The great versatility of short designed peptide materials lies in the ability to either alter amino acid sequences of known peptide structures found in nature or create completely novel sequences. *De novo* designed short peptides are biocompatible, biodegradable and have found applications in a wide range of areas including tissue engineering, drug delivery, membrane protein stabilisation, cosmetics and skin care.<sup>1,2</sup> To further explore their potential applications, it is important to reveal not only their interfacial behaviour alone but also investigate

their interactions with other molecules, in particular, conventional surfactants.

One of the important classes of designed peptides are peptide amphiphiles which are designed to mimic the basic structural features of common surfactants. They generally bear distinct hydrophilic and hydrophobic moieties and thus share many common functions to conventional surfactants such as sodium dodecyl sulfate (SDS) and dodecyl trimethyl ammonium bromide (C<sub>12</sub>TAB).<sup>3</sup> However the amino acid sequence which builds up a peptide surfactant renders them far more complex than simple alkyl chain surfactants. The resulting increased functionality of these peptides is appreciable in their antimicrobial and antitumor activity.<sup>4–6</sup> In addition their surfactant-like amphiphilicity is attractive to a wide range of biotechnological applications including regenerative medicine and drug delivery systems.<sup>7-10</sup> Thus peptide amphiphiles are promising alternatives to synthetic surfactants derived from petrochemical sources as a more environmentally responsible option showing strong sustainability, degradability and biocompatibility.<sup>11,12</sup>

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We have previously reported the self-assembling properties and surface adsorption dynamics of a class of short designed peptides ( $V_m K_n$ , where m = 3-6 and n = 1-3) at the solid/liquid interface.<sup>13–15</sup> It was found that changes in salt, pH and peptide concentration all have significant effects on their interfacial adsorption.<sup>15</sup> At the silicon oxide/water interface V<sub>6</sub>K was found to be the most surface active peptide with the highest adsorption amount. It reached the steady-state adsorption faster than V<sub>6</sub>K<sub>2</sub> and V<sub>3</sub>K. Both V<sub>6</sub>K and V<sub>6</sub>K<sub>2</sub> were capable of self-assembling into nanostructures at the hydrophilic silica/water interface. Over high peptide concentrations, it was found that V<sub>6</sub>K formed a peptide bilayer that also incorporated some defects and peptide stacks or vesicles. The bilayer arrangement is also common to many other surfactants such as C12E6.16,17 Over lower peptide concentrations, V<sub>6</sub>K was shown to adsorb onto the silicon oxide interface and formed distinct flat cylindrical micellar structures.<sup>13</sup> Once adsorbed, the V<sub>6</sub>K layers could not be removed easily by rinsing, showing great stability across a wide range of pH values and peptide concentrations.<sup>14</sup> Short peptides that demonstrate good adsorption dynamics are attractive to the personal care industry as they can be formulated in skin care products.<sup>18-20</sup> Similarly they are also attractive in the pharmaceutical industry to help drug solubilisation.<sup>21</sup> To realize the potential of such peptides in formulations, it is important to rationalise their basic interfacial behaviour in the presence of other conventional surfactants such as SDS and C12 TAB which have been extensively studied as they are relevant to numerous applications including biotechnology, oil recovery, detergency, and personal care.<sup>22</sup>

Adsorption at the solid/liquid interface is generally a result of a complex combination of forces including hydrophobic interactions, electrostatic interactions and hydrogen bonding.<sup>22-24</sup> Extensive research has been carried out to study adsorption from surfactant mixtures and surfactant/polymer mixtures.<sup>25-29</sup> Recent research has also focused on adsorption involving biosurfactants such as rhamnolipids and proteins such as hydrophobin and their interactions with conventional surfactants.<sup>11,30-32</sup> Due to the lack of understanding of interfacial behaviour the commercial use of short peptides or other biosurfactants is still at an early stage. Thus detailed studies are required to explore their basic adsorption behaviour when mixed with conventional surfactants in different manners.

This paper aims to investigate how a surfactant-like peptide,  $V_6K$ , interacts with conventional surfactants, SDS and  $C_{12}TAB$ , at the solid/water interface by performing selective spectroscopic ellipsometry and neutron reflection measurements and elucidate the main interactions taking place in order to maximize understanding for potential applications.

### 2. Materials and method

#### Materials

The peptide was synthesized using the Solid Phase Peptide Synthesis (SPPS) procedure with C-terminal amides attached and N-terminal acetylated. The peptide was then purified by Gel Permeation Chromatography (GPC) twice, giving the final purity >95%. The peptide solution was made freshly by dissolving 5 mg peptide in 5.0 ml ultrahigh quality (UHQ) water (Purelab UHQ, Vivendi Water Systems Ltd) and subsequent serial dilution to the desired concentration. The peptide solutions were adjusted to pH 7 using minimal amounts of either HCl or NaOH. The critical aggregation concentration (CAC) for V<sub>6</sub>K was determined to be 0.15 mM, from conductance measurements in pure water and by fluorescence probe measurements.<sup>13,14</sup> SDS and C<sub>12</sub>TAB were purchased respectively from Lancaster and Sigma, UK. Both of them were purified by recrystallization more than 3 times, in ethanol + water for SDS and in acetone + absolute ethanol for C<sub>12</sub>TAB, till the surface tension around their CMC showed no minimum.<sup>33</sup>

#### Substrates

Silicon wafers were purchased from Compart Technology Ltd, UK. The silicon surface has a native oxide layer and bears silanol (Si–OH) groups and when in contact with water or moisture, it becomes very hydrophilic. Prior to each experiment the silicon wafers were cleaned by Piranha treatment (95% H<sub>2</sub>SO<sub>4</sub>/30% H<sub>2</sub>O<sub>2</sub> = 3:1 at 90 °C for 1 min) before wash by 5% Decon90 solution (from Decon Laboratory, UK), and followed by copious rinsing with UHQ water. To mimic the hydrophobic nature of different surfaces, silica surfaces were modified with C<sub>8</sub> hydrocarbon (trimethoxy octyl silane) as described in our previous work.<sup>34,35</sup> It is the model hydrophobic surface that has been most widely used for adsorption. It is useful at this stage to establish their basic interfacial behaviour using model substrate surface so that results can be compared with adsorption at more complex interfaces.

#### Spectroscopic ellipsometry

Measurements were determined using a Jobin-Yvon UVISEL spectroscopic ellipsometer. The SE measurements were performed over a wavelength range between 300 and 600 nm. A liquid cell with fused quartz windows was used to enable the SE measurement at the solid/liquid interface with the incident light beam at 70°. The experimental data were analysed using a software called DeltaPsi II developed by Jobin-Yvon. The ellipsometer measured the change in the polarization state of light reflected from the surface of the sample. By studying the changes in the state of polarization, information about layer thickness and refractive index was revealed through the simultaneous analysis of two ellipsometric angles  $\psi$ and  $\Delta$ . The changes in amplitude and phase of polarization of the light after reflection were determined in two components, the plane of reflection (p-plane), and that perpendicular to it (s-plane). The sample ellipticity, F, is defined as the ratio of the Fresnel coefficients of the p and s planes ( $R_p$  and  $R_s$ ) and is expressed as<sup>36</sup>

$$\rho = \frac{R_{\rm p}}{R_{\rm s}} = \tan \psi e^{i\Delta} \tag{1}$$

The refractive index  $n_{\rm f}$  and the coupled thickness  $\tau_{\rm f}$  were subsequently calculated by the software using eqn (1). The surface adsorbed amount  $\Gamma$  (mg m<sup>-2</sup>) of the sample is finally calculated from  $n_{\rm f}$  and  $\tau_{\rm f}$  (in Å) through eqn (2),

$$\Gamma = \frac{\tau_{\rm f}(n_{\rm f} - n_0)}{{\rm d}n/{\rm d}c} \tag{2}$$

where  $n_0$  is the refractive index of the buffer, dn/dc stands for the change of refractive index against solution concentration and a value of 0.18 ml g<sup>-1</sup> was used in this work.<sup>34</sup>

#### Neutron reflection (NR)

Measurements were carried out on SURF at RAL, Oxford, UK, using a neutron beam of wavelength 0.5 to 6.5 Å. The silicon (111) blocks used were polished by Crystran Ltd, UK and treated with Piranha solution at 90 °C for 1 min. Solution samples (2 ml) were filled into the lumen cell made by clamping a Perspex trough against the polished face of a silicon block with dimensions of  $6 \times 5 \times 1.2$  cm<sup>3</sup>. The sample cell was mounted on a goniometer stage controlled by the computer terminals. The neutron beam entered the small face of the silicon block, was reflected from the solid/solution interface and exited from the opposite end of the small face. The neutron beam was collimated by two sets of horizontal and vertical slits placed before the sample cell, creating a typical beam illuminated area around  $4 \times 3$  cm<sup>2</sup>. Each reflectivity experiment was carried out at three incidence angles of 0.35, 0.8 and  $1.8^{\circ}$  and the resulting reflectivity profiles combined to cover a wave vector ( $\kappa$ ) between 0.012 and 0.5  $Å^{-1}$ . Reflectivity profiles below the critical angle were theoretically equal to unity and all the data measured were scaled accordingly. Constant background was subtracted using the average reflectivity between 0.3 and 0.5  $Å^{-1}$ . The background was found to be typically around  $2 \times 10^{-6}$  in D<sub>2</sub>O.

Model fitting of neutron reflection data has been extensively used to quantitatively analyse information regarding thickness and composition of adsorbed layers and has been found to be ideally suited to investigate multi component mixed layers adsorbed at an interface.<sup>37-40</sup> The Motofit package was used for the data fitting.<sup>41</sup> When fitting the data a structural model was assumed and the interface was divided into a suitable number of uniform sublayers. A model with minimum number of sublayers is preferred as it reduces the complexity of the system and overfitting of the data. Reflectivity from the model layers is calculated using the optical matrix formulism this is then compared with the measured reflectivity and the structural parameters (mainly thickness  $(\tau)$  and scattering length density  $(\rho)$ ) are modified in a least-square iteration until a good fit is obtained. For a two component adsorption, such as surfactant and peptide in water, the volume fraction and scattering length density of a layer can be expressed using eqn (3) and (4)

$$\rho = \rho_{\rm p} \varphi_{\rm p} + \rho_{\rm s} \varphi_{\rm s} + \rho_{\rm w} \varphi_{\rm w} \tag{3}$$

$$\varphi_{\rm p} + \varphi_{\rm s} + \varphi_{\rm w} = 1 \tag{4}$$

where  $\rho$  is the total scattering length density of a layer and  $\rho_{\rm p}$ ,  $\rho_{\rm s}$ , and  $\rho_{\rm w}$  are the known individual scattering length densities of peptide, surfactant and water, and  $\varphi_{\rm p}$ ,  $\varphi_{\rm s}$  and  $\varphi_{\rm w}$ , are the respective unknown volume fractions of the components found in the layer which must add up to one. However, since there are three unknowns, the volume fraction of each component cannot be solved. In order to solve the equation, the adsorption experiment is repeated using a deuterated version of the surfactant as an isotopic contrast to yield two independent

equations for eqn (3) allowing the volume fraction for both  
components to be calculated. Fitting two different isotopic  
compositions to one structural model also significantly reduces  
the possible ambiguity in the interpretation of the data. Once  
the volume fractions have been calculated the surface area per  
molecule (
$$A$$
) can be calculated using eqn (5)

$$A = \frac{V_{\rm p}}{\tau \phi_{\rm p}} \tag{5}$$

where  $V_{\rm p}$  is the volume of the peptide. The surface excess ( $\Gamma$ ) can then be calculated as

$$\Gamma = \frac{M_{\rm W}}{6.02\rm A} \tag{6}$$

where  $M_{\rm W}$  is the molecular weight of the component.

### Results and discussion

#### (A). SDS and V<sub>6</sub>K system

Adsorption of SDS/V<sub>6</sub>K mixed solution at the solid/liquid interface. The effect of different molar ratios of SDS on the time dependent adsorption of the V<sub>6</sub>K peptide was explored using SE (Fig. 1). The peptide itself readily adsorbed onto the silicon oxide surface and reached a stable plateau within 10 minutes with a maximum adsorbed amount of 3.2 mg m<sup>-2</sup>. Addition of SDS significantly changed the adsorption dynamics at the solid/ liquid interface. At a molar ratio of 0.5:1 (SDS/V<sub>6</sub>K), the adsorption reached the same plateau as V6K alone but the process was slower, and as the ratio was further increased the adsorption dynamics slowed down further. It took over 100 min to reach a lower plateau, at a ratio of 0.78:1, with a final surface adsorbed amount of 2.8 mg  $m^{-2}$ . When the ratio increased to unity and above, only a negligible amount (less than  $0.2 \text{ mg m}^{-2}$ ) of adsorption was detected. The SE results indicated that peptide adsorption was strongly inhibited by the increasing molar ratio of SDS in the solution. As the molar ratio of SDS/V6K is also equal to the charge ratio, the adsorption dynamics in Fig. 1 is explained by the charge neutralization of cationic V6K molecules



**Fig. 1** The adsorption of V<sub>6</sub>K (×) peptide and co-adsorption of SDS and V<sub>6</sub>K at the silica/water interface with the molar ratio of SDS:V<sub>6</sub>K at 0.5:1 (□), 0.78:1 (Δ), 1:1 (◊), 3.9:1 (\*), and 7.8:1 (○). V<sub>6</sub>K was fixed at 100 µg ml<sup>-1</sup>, pH 7.

by SDS in the solution. At and above unity, the amount of free peptide was minimal, hence adsorption was negligible. Below molar unity, SDS was not enough to neutralize all the peptide molecules in the solution, thus excess peptides drove the surface adsorption. The adsorbed amount and dynamic process both reduced and slowed down as the ratio was increased, similar to the SE adsorption curves at concentrations below  $100 \ \mu g \ ml^{-1}$  reported in our previous studies.<sup>14</sup>

SE results alone cannot reveal how much SDS contributed to the total adsorbed amount and whether the presence of SDS in the mixture altered the self-assembling properties of V<sub>6</sub>K at the solid/ liquid interface. Therefore, NR was carried out for the adsorption of SDS/V<sub>6</sub>K (0.78/1) at pH 7, the reflectivity profiles are shown in Fig. 2A for both hydrogenated SDS (h-SDS) and deuterated SDS (d-SDS). The fitted data for the curves along with the calculated volume fraction and adsorbed amounts for each layer are shown in Table 1.



**Fig. 2** (A) Reflectivity profiles for h-SDS/V<sub>6</sub>K (0.78/1) mixture ( $\Box$ ) and d-SDS/V<sub>6</sub>K (0.78/1) ( $\diamond$ ) at the SiO<sub>2</sub> interface in D<sub>2</sub>O pH 7. Solid lines through the data points correspond to the model fits for the corresponding measured reflectivity data points. (B) Schematic diagram showing the arrangement of V<sub>6</sub>K molecules (blue), SDS (red), at SDS/V<sub>6</sub>K = 0.78/1.

Due to the electrostatic repulsion SDS does not adsorb directly onto the hydrophilic SiO<sub>2</sub> surface. However, the presence of cationic V6K peptides at the SiO2 interface can aid the subsequent adsorption of SDS through increased hydrophobic interaction and charge reversal of the surface. Careful evaluation and analysis of the NR data revealed that three distinct layers were found at the interface (Fig. 2B). The innermost layer was 15 Å thick and densely packed, above it a more diffuse 13 Å laver followed by an even more diffuse SDS layer, as shown by the volume fraction values in Table 1. The peptide structures in the innermost layer were reminiscent of the V<sub>6</sub>K structures adsorbed at 20  $\mu g$  ml<sup>-1</sup> (discussed later). The thickness of the outermost SDS layer (38 Å) suggested the formation of surface micelles or lamellar structures which have frequently been reported to occur on positively charged hydrophilic surfaces.<sup>27,42-44</sup> Since the outermost layer was predominantly occupied by SDS and D2O, the layer was essentially invisible and cannot be detected by the d-SDS run as both d-SDS and D<sub>2</sub>O have similar SLDs. Co-adsorption of SDS was likely aided by a combination of hydrophobic/electrostatic interactions with excess V<sub>6</sub>K peptides which had readily adsorbed at the interface. The adsorbed SDS reduced the electrostatic repulsion between peptides and allowed for a tight packing of V<sub>6</sub>K in the inner leaflet of the peptide bilayer. Overall the total adsorbed amount measured by NR, 2.6 mg  $m^{-2}$ , closely matched ellipsometry results, but V6K was revealed to have contributed only 1.8 mg m<sup>-2</sup> (2.3 × 10<sup>-3</sup> mM m<sup>-2</sup>) to the total amount, significantly less peptide compared to adsorption at the same peptide concentration without surfactant. On the other hand, adsorption from the binary mixture significantly aided SDS adsorption and resulted in 0.8 mg m<sup>-2</sup> (2.9  $\times$  10<sup>-3</sup> mM m<sup>-2</sup>) at the interface.

Interaction of SDS with the pre-adsorbed V<sub>6</sub>K peptide. Short surfactant-like peptides have a tendency to self-assemble at the solid/liquid interface. Experiments conducted by Han *et al.* highlighted the presence of two distinct plateaus in the adsorption isotherms for V<sub>6</sub>K corresponding to different structural arrangements of the molecules at the interface.<sup>13</sup> The effect of SDS addition on pre-adsorbed V<sub>6</sub>K peptide layers was investigated at peptide concentrations of 20 and 100 µg ml<sup>-1</sup> which correspond to two different peptide self-assembled structures.

Analysis of the reflectivity curves (Fig. 3A and Table 2) from the pre-adsorbed peptide layer, at 20  $\mu$ g ml<sup>-1</sup>, supports the formation of flattened V<sub>6</sub>K cylindrical micelles. The adsorbed peptide layer was very reproducible and had a surface excess of 1.4 mg m<sup>-2</sup> after 20 minutes adsorption at pH 7, matching

Table 1 Structural parameters obtained from best fits of neutron reflection data shown in Fig. 2 for the co-adsorption of SDS/V <sub>6</sub> K in D <sub>2</sub> O pH 7								
Sample/contrast	Fitted thickness ±2 Å	Fitted SLD $\pm 0.1  imes 10^{-6}$ /Å $^{-2}$	$\begin{array}{l} \text{Sample SLD (V_6K/SDS)} \\ \pm 0.01  \times  10^{-6}/\text{\AA}^{-2} \end{array}$	Volume fraction $(V_6K/SDS) \pm 0.005$	$\Gamma$ (V <sub>6</sub> K/SDS) $\pm$ 0.1 mg m <sup>-2</sup>			
h-SDS/V <sub>6</sub> K (0.78/1)/D <sub>2</sub> O	15 13 38	2.1 4.6 5.6	1.71/0.37 1.71/0.37 1.71/0.37	0.815/0.080 0.255/0.095 0/0.115	$egin{array}{c} 1.4/0.1 \\ 0.4/0.2 \\ 0/0.5 \\ arGamma_{ m Total} 2.6 \end{array}$			
d-SDS/V <sub>6</sub> K (0.78/1)/D <sub>2</sub> O	15 13	2.6 5.2	1.71/6.72 1.71/6.72	0.815/0.080 0.255/0.095	$1.4/0.1 \\ 0.4/0.2 \\ \Gamma_{ m Total} \ 2.1$			



**Fig. 3** (A) Reflectivity profiles for 20  $\mu$ g ml<sup>-1</sup> V<sub>6</sub>K pH 7 ( $\Box$ ), 20  $\mu$ g ml<sup>-1</sup> V<sub>6</sub>K pH 7 + 4 mM d-SDS ( $\Diamond$ ) and 20  $\mu$ g ml<sup>-1</sup> V<sub>6</sub>K pH 7 + 4 mM h-SDS ( $\Delta$ ). Solid lines through the data points correspond to the best fits for the corresponding reflectivity data points. (B) Schematic diagram showing the arrangement of pre-adsorbed V<sub>6</sub>K peptides at 20  $\mu$ g ml<sup>-1</sup> (blue) and adsorption of SDS (red) at the SiO<sub>2</sub>/D<sub>2</sub>O interface at pH 7.

adsorption results from ellipsometry measurements reported by Pan *et al.*<sup>14</sup> The thickness of the peptide layer was found to be 19 Å with a volume fraction of 0.66.

Following the initial adsorption, the peptide solution was carefully rinsed out with  $D_2O$  (peptide desorption was negligible) and 4 mM h-SDS solution was added. As expected, the addition of h-SDS resulted in an increased layer thickness at the interface.

To investigate whether SDS adsorption caused any changes to the initial structural arrangement of the pre-adsorbed peptide layer and quantify adsorption, the experiment was repeated using d-SDS (Table 2). Addition of 4 mM d-SDS revealed no significant changes in the pre-adsorbed V<sub>6</sub>K layer, suggesting that SDS adsorption was limited to the surface of the peptide structures. If SDS had strongly interfered with the pre-adsorbed peptide structures (*e.g.* disruption), the d-SDS run would reveal a significant change to the layer SLD and or its thickness. However, in this case, both the subsequent addition of d-SDS and peptide only runs could be fitted with the same model, indicating that SDS displaced  $D_2O$  and adsorbed onto the peptide layer. On the other hand the changes in SLD between the h-SDS and d-SDS run (Table 2) were used to determine the adsorbed amounts at the interface. Volume fraction values show less than 10% penetration of SDS into the pre-adsorbed peptide layer with a surface excess amounting to 0.20 mg m<sup>-2</sup> ( $0.7 \times 10^{-3}$  mM m<sup>-2</sup>). A higher proportion of the adsorbed SDS, 0.4 mg m<sup>-2</sup> ( $1.5 \times 10^{-3}$  mM m<sup>-2</sup>) formed a distinct 16 Å thick layer on top of the peptide structures. Fig. 3B shows a cross-sectional diagram of the proposed arrangement of pre-adsorbed peptide, highlighting a compact flattened micellar structure with gaps between individual structures and SDS adsorbed around and on top.

Similar to ionic surfactant adsorption onto oppositely charged hydrophilic surfaces; individual surfactant molecules are likely to have initiated adsorption onto the peptide structures with their negatively charged heads. Subsequent adsorption of SDS molecules would be through hydrophobic interactions with the alkyl tails of SDS molecules already adsorbed or hydrophobic patches on the peptide structures. SDS molecules extending from the peptide structures in such a manner would indeed stretch to around 16 Å.<sup>44</sup>

At 100  $\mu$ g ml<sup>-1</sup>, the resulting adsorbed V<sub>6</sub>K layer could withstand UHQ water rinse and a series of rinses with SDS concentrations ranging from 0.1 mM to 4 mM (Fig. S1 in the ESI†). Initially, during the UHQ water rinse, minimal amounts (less than 0.2 mg m<sup>-2</sup>) of loosely adsorbed peptide were removed leaving a very stable and firmly absorbed peptide layer. Subsequent SDS rinses with increasing SDS concentration, up to 4 mM, resulted in increasing surface adsorbed amounts measured by SE at the solid interface.

NR data (Fig. 4A) confirmed the presence of a bilayer structure of  $V_6K$  at the SiO<sub>2</sub>/water interface. The pre-adsorbed peptide formed three distinctive layers (Table 3) detectable due to the ordering of the lysine heads and valine tails, the latter having a lower SLD than lysine. Due to irregularities in the bilayer structure, the three layers could not be modelled to have perfectly defined K–V–K (lysine head–valine core–lysine head) layers, instead, the peptide layer was fitted by incorporating at least one V in the head region, hence KV–V–KV was used. Similar structure arrangement was found in our previous study of the interfacial structure of  $V_6K_2$  peptide at the SiO<sub>2</sub>/water interface.<sup>15</sup>

Table 2 Structural parameters obtained from the best fits as shown in Fig. 3 for 20 minute pre-adsorbed  $V_6K$  at 20  $\mu$ g ml<sup>-1</sup>, followed by addition of 4 mM SDS

Sample/contrast	Fitted thickness ±2 Å	Fitted SLD $\pm 0.1 \times 10^{-6}$ /Å <sup>-2</sup>	Sample SLD (V <sub>6</sub> K/SDS) $\pm 0.01 \times 10^{-6}$ /Å <sup>-2</sup>	Volume fraction $(V_6K/SDS) \pm 0.005$	$egin{array}{l} \Gamma \ (V_6 { m K/SDS}) \ \pm 0.1 \ { m mg \ m}^{-2} \end{array}$
$20 \ \mu g \ ml^{-1} \ V_6 K/D_2 O$	19	3.3	1.71/—	0.660/—	1.4/ $\Gamma_{ m Total}$ 1.4
20 $\mu g m l^{-1} V_6 K$ + 4 mM d-SDS/D <sub>2</sub> O	19	3.3	1.71/6.72	0.660/0.095	1.4/0.2 $\Gamma_{ m Total}$ 1.6
$20 \ \mu g \ m l^{-1} \ V_6 K + 4 \ m M \ h\text{-SDS/D}_2 O$	19 16	2.7 5.0	1.71/0.37 —/0.37	0.660/0.095 0/0.225	1.4/0.2 /0.4 $\Gamma_{ m Total}$ 2.0

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**Fig. 4** (A) Reflectivity profiles for: 100 µg ml<sup>-1</sup> V<sub>6</sub>K pH 7 ( $\Box$ ), 100 µg ml<sup>-1</sup> V<sub>6</sub>K pH 7 + 4 mM d-SDS ( $\diamond$ ) and 100 µg ml<sup>-1</sup> V<sub>6</sub>K pH 7 + 4 mM h-SDS ( $\Delta$ ). Solid lines through the data points correspond to the model fits for the corresponding reflectivity data points. (B) Schematic diagram showing the arrangement of pre-adsorbed 100 µg ml<sup>-1</sup> V<sub>6</sub>K peptides (blue) and the effect of SDS (red) on its arrangement at the SiO<sub>2</sub>/D<sub>2</sub>O interface at pH 7.

The valine core had a thickness of 15 Å and SLD of 2.7  $\times$   $10^{-6}$ Å $^{-2}$  whilst the inner and outer layer were both 9 Å thick with an SLD of 4.7  $\times$   $10^{-6}$ Å $^{-2}$ . This corresponds to the peptide molecules being tightly packed with a highly interdigitated valine tail core. The total thickness of the pre-adsorbed bilayer was 33 Å, which is in good agreement with the extended length of a V<sub>6</sub>K molecule around 2.5–3 nm. The total surface excess value calculated from the VK–V–VK layer was slightly lower than ellipsometry results (3.1 mg m $^{-2}$ ) shown in Fig. S1 in ESI† but is in agreement with the values found in the literature ranging between 2.5 and 3.0 mg m $^{-2}$ .

Addition of 4 mM h-SDS solution after careful rinsing of the peptide solution caused a significant change in the reflectivity

curves (Fig. 4A) and fitted SLD (Table 3) confirming that SDS strongly interacted with the pre-adsorbed peptide layer. Measurements using d-SDS also showed a change in the reflectivity curve indicating a change had occurred to the pre-adsorbed peptide layer upon SDS addition. Careful evaluation of the reflectivity curves revealed that SDS had penetrated into the outer peptide layer (schematic diagram shown in Fig. 4B). The surfactant was able to insert itself in the spaces between adjacent lysine heads by displacing D<sub>2</sub>O. However, SDS was not able to penetrate further into the bilayer valine core to a significantly appreciable amount. There was no additional SDS adsorbed on top of the peptide bilayer, suggesting that the adsorbed SDS was enough to cause an overall charge neutralization of the peptide surface and limit any further SDS adsorption. The total SDS adsorbed amount was 0.5 mg m<sup>-2</sup> (1.6  $\times$  10<sup>-3</sup> mM m<sup>-2</sup>) and is comparable to the surfactant adsorbed amount found on the top layer of the 20  $\mu$ g ml<sup>-1</sup> V<sub>6</sub>K run.

Surface effect. It was found that surface charge and hydrophobicity had a strong impact on the adsorption of V<sub>6</sub>K. As noted earlier adsorption of 100 µg ml<sup>-1</sup> V<sub>6</sub>K resulted in a very stable bilayer. When the surface substrate was changed to a hydrophobic C<sub>8</sub> surface the total peptide adsorption was less than half of the adsorption on the  $SiO_2$  surface, only 1 mg m<sup>-2</sup> (data shown in Fig. S2 in the ESI<sup>+</sup>). From the surface excess results, the area per molecule was found to be 130  $\text{\AA}^2$  indicating that the peptide molecules formed a flat monolayer on the C<sub>8</sub> surface. At the SiO<sub>2</sub> surface, the adsorption driving force was the electrostatic attraction between the anionic surface and the cationic peptide, whilst at the C8 surface, the adsorption was driven by the hydrophobic interaction between the C8 and valine tail, analogous to cationic surfactant adsorption.<sup>17</sup> This led to a change in the packing of the peptide and a bilayer structure could not be formed.

Subsequent addition of SDS solution (up to 4 mM) onto the peptide monolayer resulted in an additional adsorbed amount of 1.2 mg m<sup>-2</sup> (Fig. S2 in the ESI<sup>†</sup>). This is significantly higher than the SDS adsorption onto the peptide bilayer found at the hydrophilic SiO<sub>2</sub> interface of only 0.5 mg m<sup>-2</sup> under the same conditions with the same peptide concentration. The increased

Table 3 Structural parameters obtained from model best fits of neutron reflection data shown in Fig. 4 for the adsorption of 4 mM SDS onto preadsorbed  $V_6$ K in  $D_2$ O pH 7

Sample/contrast	Fitted tl	nickness ±2 Å	Fitted SLD $\pm 0.1  imes 10^{-6}$ /Å <sup>-2</sup>	$\begin{array}{l} \text{Sample SLD} \left(\text{V}_6\text{K/SDS}\right) \\ \pm 0.01 \times 10^{-6} / \text{\AA}^{-2} \end{array}$	Volume fraction $(V_6K/SDS) \pm 0.005$	$\Gamma$ (V <sub>6</sub> K/SDS) ±0.1 mg m <sup>-2</sup>
100 $\mu g \ ml^{-1} \ V_6 K/D_2 O$	KV V KV	9 15 9	4.7 2.7 4.7	2.13/— 1.54/— 2.13/—	0.390/— 0.760/— 0.390/—	0.5/— 1.3/— 0.5/— Γ <sub>Total</sub> 2.3
100 $\mu$ g ml <sup>-1</sup> V <sub>6</sub> K + 4 mM d-SDS/D <sub>2</sub> O	KV V KV	9 15 9	4.7 2.7 4.9	2.13/6.72 1.54/6.72 2.13/6.72	0.390/0 0.760/0 0.390/0.420	0.5/0 1.3/0 0.5/0.5 Γ <sub>Total</sub> 2.8
100 µg ml <sup>-1</sup> V <sub>6</sub> K + 4 mM h-SDS/D <sub>2</sub> O	KV V KV	9 15 9	4.7 2.7 2.2	2.13/0.37 1.54/0.37 2.13/0.37	0.390/0 0.760/0 0.390/0.420	0.5/0 1.3/0 0.5/0.5 Γ <sub>Total</sub> 2.8

adsorption of SDS is consistent with the peptide being arranged flat on the hydrophobic surface. In this arrangement, the hydrophobic valine tails of the peptide are more exposed allowing strong hydrophobic interactions with the SDS alkyl chains.

#### (B). C<sub>12</sub>TAB and V<sub>6</sub>K system

 $C_{12}TAB/V_6K$  mixed solution adsorption at the solid/liquid interface.  $C_{12}TAB$  was selected to investigate the effect of cationic surfactants on the adsorption of  $V_6K$  peptide and to draw comparison with the SDS/V<sub>6</sub>K system. The co-adsorption of  $C_{12}TAB$  and  $V_6K$  at varying molar ratios, as well as the adsorption of pure peptide and pure  $C_{12}TAB$  at equivalent molar concentrations, is shown in Fig. 5. The adsorption dynamics of  $C_{12}TAB$ and  $V_6K$  at a ratio of 1:1 was similar to the pure peptide but had a slightly lower surface excess. Increasing the ratio to 20:1 caused the final plateau to reduce further with a noticeable slowing of the adsorption process. A further increase in the ratio to 94:1 resulted in a significant drop of the surface excess. At 94:1,  $C_{12}TAB$  had reached its CMC and was highly in excess. Indeed, both the surface excess and the adsorption dynamics closely resembled the adsorption of pure  $C_{12}TAB$  at 12 mM (CMC).

The SE results show both the surfactant and the peptide competing for adsorption at the interface. At the highest ratio (94:1) the adsorption was dominated by  $C_{12}$ TAB, limiting total adsorption to under 1.3 mg m<sup>-2</sup>. However, it is not clear from the SE results alone how the two components coexisted and arranged themselves at the interface.

NR was used to probe the arrangement of the two molecules at a molar ratio of 20:1. Fig. 6A shows neutron reflection data and model fits for the adsorption of  $C_{12}TAB$  and  $d-C_{12}TAB$  with  $V_6K$  in  $D_2O$  pH 7. Analysis of the data revealed a bilayer structure retained by the  $V_6K$  peptide but had significant amounts of  $C_{12}TAB$  incorporated within it (Table 4). The inner layer was dominated by  $V_6K$  and only 0.1 mg m<sup>-2</sup> ( $0.3 \times 10^{-3}$  mM m<sup>-2</sup>) of  $C_{12}TAB$  present. The bilayer core was densely packed and housed the largest amount of  $C_{12}TAB$ , 0.3 mg m<sup>-2</sup> ( $1.0 \times 10^{-3}$  mM m<sup>-2</sup>), and the outer layer had no surfactant present as shown in the



**Fig. 5** The adsorption isotherm of V<sub>6</sub>K at 100  $\mu$ g ml<sup>-1</sup> (×) and C<sub>12</sub>TAB at 0.128 mM ( $\bigcirc$ ); 2.63 mM (\*); 12 mM ( $\square$ ). The co-adsorption was at the molar ratio of C<sub>12</sub>TAB and V<sub>6</sub>K of 1:1 ( $\diamondsuit$ ), 20:1 ( $\Delta$ ) and 94:1 (+). V<sub>6</sub>K was fixed at 100  $\mu$ g ml<sup>-1</sup> (0.128 mM), all at pH 7.



**Fig. 6** (A) Reflectivity profiles for:  $C_{12}TAB/V_6K$  (20/1) mixture ( $\Box$ ) and  $d-C_{12}TAB/V_6K$  (20/1) ( $\diamond$ ). Solid lines through the data points correspond to the model fits for the corresponding reflectivity data points. (B) Schematic diagram showing the arrangement of V<sub>6</sub>K molecules (blue),  $C_{12}TAB$  (green) at  $C_{12}TAB/V_6K = 20/1$ , at the SiO<sub>2</sub> interface in D<sub>2</sub>O pH 7.

schematic diagram in Fig. 6B. Peptide molecules adsorbed faster and in higher amount than  $C_{12}TAB$ , allowing the peptide to form a bilayer and in the process trapping  $C_{12}TAB$  within its core. Following the formation of the bilayer, electrostatic repulsion stopped additional adsorption onto the co-adsorbed layer. This supports the SE results which suggest competitive adsorption of  $C_{12}TAB$  at the interface with increasing amounts of surfactant at increasing molar ratio.

Interaction of  $C_{12}TAB$  with pre-adsorbed  $V_6K$  peptide. Following the adsorption of  $V_6K$ , at 100 µg ml<sup>-1</sup>, the adsorbed layer was rinsed with UHQ water. The layer was subsequently subjected to a series of concentrations of  $C_{12}TAB$  solution ranging from 0.1 mM to 4 mM (Fig. S3 in the ESI†). As it was outlined earlier for the SDS adsorption results, there was minimal peptide loss during the UHQ rinse, however, in contrast to the SDS system, no increase of surface excess was observed when  $C_{12}TAB$  solutions were added. The preadsorbed peptide layer prevented any  $C_{12}TAB$  adsorbing onto the surface.

**Surface effect.** Addition of up to 4 mM  $C_{12}$ TAB onto preadsorbed V<sub>6</sub>K on the hydrophobic C<sub>8</sub> (Fig. S4 in the ESI<sup>†</sup>) surface yielded a maximum increase of less than 0.4 mg m<sup>-2</sup> ( $1.3 \times 10^{-3}$  mM m<sup>-2</sup>). Similar to the SDS system, the increase in total surface excess upon C<sub>12</sub>TAB addition was attributed to the hydrophobic interaction between the exposed hydrophobic valine tails and the hydrocarbon tails of C<sub>12</sub>TAB. This highlights the strength of hydrophobic interactions which enabled the C<sub>12</sub>TAB to adsorb onto the surface even in the presence of alike charges. However, the adsorption of C<sub>12</sub>TAB was less than that of SDS 1.2 mg m<sup>-2</sup> ( $4.2 \times 10^{-3}$  mM m<sup>-2</sup>).

Table 4 Structural parameters obtained from model best fits of neutron reflection data shown in Fig. 6 for the co-adsorption of C<sub>12</sub>TAB/V<sub>6</sub>K in D<sub>2</sub>O at pH 7

Sample/contrast	Fitted thickness $\pm 2$ Å		Fitted SLD $\pm 0.1  imes 10^{-6}$ /Å $^{-2}$	$\begin{array}{l} \text{Sample SLD}\left(V_{6}\text{K/C}_{12}\text{TAB}\right) \\ \pm 0.01\times10^{-6}/\text{\AA}^{-2} \end{array}$	Volume fraction $(V_6K/C_{12}TAB) \pm 0.005$	$ \begin{array}{c} \Gamma \ (V_6 K/C_{12} TAB) \\ \pm 0.1 \ \mathrm{mg \ m}^{-2} \end{array} $
C <sub>12</sub> TAB/V <sub>6</sub> K (20/1)/D <sub>2</sub> O	KV V KV	9 15 9	4.4 1.4 4.6	2.13/-0.24 1.54/-0.24 2.13/-0.24	0.375/0.055 0.750/0.205 0.385/0.020	0.5/0.1 1.3/0.3 0.5/0.0 Γ <sub>Total</sub> 2.7
d-C <sub>12</sub> TAB/V <sub>6</sub> K (20/1)/D <sub>2</sub> O	KV V KV	9 15 9	4.7 2.5 4.7	2.13/5.13 1.54/5.13 2.13/5.13	0.375/0.055 0.750/0.205 0.385/0.020	0.5/0.1 1.3/0.3 0.5/0.0 $\Gamma_{\rm Total}$ 2.7

## 4. Conclusions

In this study, spectroscopic ellipsometry and neutron reflection were used to evaluate the adsorption behaviour of the V6K peptide and its interaction with conventional surfactants SDS and C12TAB. When the peptide was pre-adsorbed it was able to form a stable bilayer with different coverage and stability. At the highest concentration of 100  $\mu$ g ml<sup>-1</sup>, the bilayer formed was very stable against rinsing by UHQ water. The positively charged lysine heads exposed on preadsorbed peptide layers facilitated SDS adsorption but hampered C12TAB adsorption. When SDS and V6K are mixed together in solution, adsorption showed a strong dependence on the molar ratio of the two components. SDS has a neutralizing effect on adsorption and when the molar unity was reached no adsorption was detected. Below molar unity, excess V6K aided SDS adsorption onto the surface. This behaviour is analogous to several polymer and polyelectrolyte systems which adsorb and cause charge reversal of the SiO<sub>2</sub> surface and increase hydrophobic interactions thus aiding SDS adsorption.<sup>26,27,30,42</sup> On the other hand, when C<sub>12</sub>TAB is combined with V<sub>6</sub>K in solution, the adsorption became competitive. Above surfactant CMC, V6K was stopped from adsorbing at the interface but at lower molar ratios V6K was able to adsorb at the interface faster than C12TAB and could coexist with C12TAB at the interface. These results are also consistent with other studies involving cationic surfactant mixtures.<sup>30,42</sup> Overall, the results indicate that V6K was able to adsorb faster at the solid/liquid interface with the exact amount depending on the ratio and other factors. The fast adsorption and stability of V<sub>6</sub>K in the presence of surfactants shows the potential for the peptide to be used as a biosurfactant at the solid/liquid interface.

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