



Cite this: *Soft Matter*, 2015,
11, 7579

Received 29th April 2015,
Accepted 10th August 2015

DOI: 10.1039/c5sm01032b

www.rsc.org/softmatter

Hyperelastic models for hydration of cellular tissue†

R. G. M. van der Sman

In this paper we present hyperelastic models for swelling elastic shells, due to pressurization of the internal cavity. These shells serve as model systems for cells having cell walls, as can be found in bacteria, plants and fungi. The pressurized internal cavity represents the cell vacuole with intact membrane at a certain turgor pressure, and the elastic shell represents the hydrated cell wall. At pressurization the elastic shell undergoes inhomogeneous deformation. Its deformation is governed by a strain energy function. Using the scaling law of Cloizeaux for the osmotic pressure, we obtain approximate analytical expressions of the cell volume *versus* turgor pressure – which are quite comparable to numerical solutions of the problem. Subsequently, we have simulated the swelling of shells – where the cell wall material is embedded with microfibrils, leading to strain hardening and anisotropic cell expansion. The purpose of our investigations is to elucidate the contribution of cell membrane integrity and turgor to the water holding capacity (hydration) of plant foods. We conclude with a discussion of the impact of this work on the hydration of food material, and other fields like plant science and the soft matter physics of responsive gels.

1 Introduction

Understanding of the hydration of cellular tissue is of importance in various fields of science. In plant science this subject is named water relations, which governs phenomena like water transport, water stress, growth and mechanical stability.¹ In food science the subject is named water holding capacity, which governs sensorial traits like juiciness and hardness.^{2,3} In biotechnological fermentations the role of water in cells is important, because many relevant biochemical processes require that the organisms is in the state of full turgor.⁴

In the recent years we have been investigating theories explaining the water holding capacity (WHC) of foods. Before our investigations, there has hardly been any understanding on the physics governing the water holding of foods.⁵ Moreover, there is no well-defined experimental method to measure it. Consequently, WHC is still regarded as an ill-defined concept. We have defined the WHC as the amount of moisture retained by a food subject to a defined mechanical load.^{6–8} We have drawn an analogy with polymer gels, and we have shown that the classical Flory–Rehner theory,⁹ describing the swelling of polymer gels, applies equally to food materials.^{10,11} However, for plant food materials having cellular tissue, the theory is incomplete, as it only applies to gel-like parts of the cellular tissue: the cell wall material and cytoplasm.

In this paper we extend our theory to account for the interaction between cell wall, and the osmotic pressure generated by solutes in vacuole and the integrity of the cell membrane. The osmotic pressure induces uptake of water, inflates the cell volume and deforms the cell wall. The cell wall is an elastic network of biopolymers, and its deformation generates the turgor pressure, which balances the osmotic pressure in homeostasis.¹

While in food science the effect of turgor pressure is hardly regarded in the water holding capacity, the reverse holds for plant science. In the science of water relations of plants the main focus is on turgor,¹ but the possible water retention by cell wall material is largely ignored.^{12–15} The contribution of turgor pressure to WHC is quite evident if the cell membrane is damaged *via* freezing. This loss of turgor leads to drip loss upon thawing of the frozen tissue.^{16,17} The contribution of the cell wall to the water retention in plants is not directly evident, and consequently it is some debate in literature.¹⁸ Hence, below we discuss this point in more detail.

In plant science there are a few studies, which do take into account the water retention of the cell wall.¹⁹ Here one distinguishes water in the symplast and the apoplast separately, which are also named free and bound water. These studies have found that in leafy vegetables like spinach and lettuce, the relative amount of water (RWC) in the apoplast is in the range of 10–40%.²⁰ Similar values are found for leaves of various grain cultivars.^{15,18} Under conditions where the cell wall is less stretched as under drought stress conditions, the relative contribution of water retention by the apoplast becomes more significant.^{18,21} In cactus the apoplast water is even reaching 50%, and thus helps

Agrotechnology and Food Sciences Group, Wageningen University & Research, the Netherlands. E-mail: ruud.vandersman@wur.nl

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c5sm01032b



considerably in its survival under dry conditions.¹⁵ Yet for other foods like berry and citrus fruits, which are meant to be juicy, the RWC is estimated to be 5% at maximum. Their juiciness is thought to be controlled by the relative cell wall thickness.²² But, in apples one has found RWC = 10%.²³

Most models in plant science on mechanical behaviour of cells consider the cell wall only as a thin elastic shell.²⁴ It is recognized that cell wall thickness and initial radial stretch of the inflated cell are difficult to determine. Also, mature fruits cells have undergone large expansion, and consequently the cell wall are largely stretched.^{25,26} The debate is not made easy by the inconsistency between experimental methods to determine RWC. While one method determines bound water by the biopolymer matrix after drying, the other method extrapolates it from turgor pressure/cell volume curve.¹⁸ This inconsistency is probably caused by the fact that the bound water measurement also includes water bound to the cytoplasm. Furthermore, it is assumed that amount of bound water is independent of the inflation of the cell (*i.e.* stretching of the cell wall material). Hence, the resolution of our question in how far turgor pressure influences water retention in foods, will also help the resolution of the debated question in plant science in how far water in apoplast contribute to water retention in plants.

To resolve these questions, we will perform a theoretical analysis of the contributions of both the turgor and the hydrated cell wall to the total water holding capacity of cellular tissue. The cell wall material will be viewed as a hydrogel, which undergoes large deformation to reach full turgor. *Via* the deformation the hydration state of the cell wall will change. Because of the large deformation of the cell wall, it is required to describe that with hyperelastic models.^{27–29} These kinds of models have been applied earlier to cell walls,^{30–34} but their swelling properties have been disregarded.

Only recently models have been developed taking into account of the interaction between swelling and large deformation of gels in the fields of responsive hydrogels,^{35–37} controlled release,^{38–41} and swelling induced cavitation in gels.^{42–44} In these models the mixing free energy has been described with Flory–Huggins as in Flory–Rehner theory. This theory can easily be extended for polyelectrolyte gels, *via* inclusion of an extra term in the free energy describing the electrostatic interactions.⁴⁵ These models also bear similarity with models considering biological elastic media which grow due to synthesis of polymers.^{46–48} Swelling rubbers is often taken as a model for the growth of biological tissue.^{49–51}

In these hyperelastic models the cellular structure can be represented by a (spherical or cylindrical) shell of hydrogel material, surrounding a liquid-like region having an elevated internal pressure.^{30–34} Because the similarity of cell tissue with liquid-filled foams,^{24,52} we assume that cell models like developed by Amon and Denson can readily applied to represent the behaviour cell tissue.⁵³ Like in foams, the cells in tissues have a hexagonal shape – but which can be approximated by spherical shells having stress free boundary conditions at the outer boundary.⁵⁴ More advanced unit cell models accounting for the hexagonal shape are developed.⁵⁵ Further, in some cases

like mature fruit the cells do have spherical shape. For these foods, cell wall hydration during ripening has been observed.²⁶

The pressure within the shell represents the osmotic pressure generated by the solutes in the vacuole. To describe the swelling behaviour of the cell wall, we require a description of the osmotic pressure of the hydrogel shell (which is different from the osmotic pressure inside the cell). For the osmotic pressure we will make use of the scaling law of Cloizeaux,^{56,57} which we have recently shown to hold for a wide range of biopolymer networks.⁵⁸ The scaling law renders similar predictions as the Flory–Rehner theory, but it is more useful in this case, because it has a compact formulation, and renders approximate analytical solutions for the cell volume and hydration state of the swellable elastic shells as function of the internal pressure – as shown below.

In the fields of responsive gels and tissue growth, the elastic behaviour is often described by the simple Neo-Hookean model. However, cell walls have often a composite structure, where microfibrils are embedded in a polysaccharide gel.^{33,34,59–61} This structure leads to more complex mechanical behaviour involving strain hardening, and anisotropic deformation.^{62–64} Hyperelastic models for composite biomaterials with fibrils have largely been developed for arteries.⁶⁵ But, here again swelling of the gel matrix has been ignored. But, building on these models of arteries, descriptions of swelling elastic tubes with fibers has been developed.^{66,67} However, swelling has been imposed instead of derived from a description of the osmotic pressure of the gel matrix. Moreover, the swelling is assumed to be homogeneous. Cell walls account often more than 30% of the dry matter of cellular tissue,⁶⁸ and due to the inhomogeneous deformation, the swelling can also be expected to be inhomogeneous.

In this paper we focus on cells with spherical or cylindrical geometry, as a simple representation of the tissues of vegetables and mushrooms, whose water holding capacity we have investigated earlier.^{6,8} In these foods the osmotic pressure is generated by solutes in large vacuoles, which are enclosed by their own membrane.²⁵ In between the vacuole, and the cell wall, there is cytoplasm. Because their macromolecular nature, we will include the cytoplasm into the hyperelastic shell. This is also inline with the practice in plant science, where one distinguishes osmotic water and bound water.

First, we will derive models describing the swelling of these cells as function of internal pressure, assuming a Neo-Hookean strain energy function and the Cloizeaux scaling law for the osmotic pressure. This allows the formulation of approximate analytical solutions to the problem, which are in good agreement with numerical solutions. Commonly, it is difficult to obtain analytical solutions for inhomogeneous deformations of gels due to the strong non-linearity of the large deformations.⁶⁹

Subsequently, we extend these models towards strain hardening, and anisotropic deformation – which approaches more the realistic behaviour of vegetable and mushroom tissue. Based on our findings from the simulations, we conclude with a discussion on the impact of the cellular structure on the hydration of cellular tissue, *i.e.* the water holding capacity of plant food materials. Because of the novelty of several elements in the



context of WHC of foods, we discuss them in the ESI,[†] for the non-informed reader – in a more simplified setting. It comprises of (1) the description of hyper-elastic models of homogeneous swelling of hydrogels, using the scaling law for the osmotic pressure, (2) the description of inhomogeneous deformation of elastic shells without swelling, and with the Neo-Hookean strain energy function, (3) a parameter study of various angles of microfibrils, and (4) the numerical solution of hyperelastic models incorporating swelling for spherical and cylindrical geometries. The second part largely deals with similar models as introduced by Chaplain,³⁴ though he does not go into depth of the details. Hence, this part of the ESI,[†] must be viewed as a tutorial, introducing the non-informed (food) scientist to these types of models.

2 Inflation of swellable elastic shells

The swelling of hydrogel shells is described *via* thermodynamics using a free energy functional as in the Flory–Rehner theory. The free energy functional has two independent contributions: (1) the mixing free energy, and (2) the strain energy function. The strain energy function W accounts for the contribution of the elastic deformation. It is described in terms of the stretching of the polymers λ_i in the three principal directions.

It is important for the discussion below to distinguish two states. In the first state the polymers are in the relaxed state, *i.e.* where they are unstretched $\lambda_i = 1$. This so-called relaxed state is taken as the reference state for the strain energy function W , which will have zero value in the reference state. If $\lambda_i > 1$ the polymers are stretched *via* tension, while they are compressed if $\lambda_i < 1$. In compression the polymer will behave more like a collapsed random coil. In the relaxed state the polymer volume fraction has the value ϕ_{ref} .

The second state is the stress-free state. The total stress σ is a resultant of both the strain energy function and the mixing free energy. In the stress-free state both contributions just equals out, rendering $\sigma = 0$. For biopolymer gels holds that the reference state does not equal the stress-free state.^{58,70} The interaction of the hydrophilic biopolymers with water induces an osmotic pressure, which makes the gel to swell. Hence, in the stress-free state, the biopolymers will always be stretched: $\lambda_i > 1$. For random gels the swelling in the stress-free state is isotropic, and all three principal stretches are equal: $\lambda_i = \lambda$. The polymer volume fraction of the gel in this stress-free state ϕ_0 appears to hold a universal relation: $\phi_0 = 2/3\phi$.^{58,70}

In our models below we will always assume that polymer and water are incompressible. Hence, we have the following relation between stretching and polymer volume fraction:

$$\lambda_1 \lambda_2 \lambda_3 = \lambda^3 = \frac{\phi_{\text{ref}}}{\phi} \quad (1)$$

Below, we will refer it as the incompressibility condition.

In this section we assume the Neo-Hookean model for the elastic deformation of the hydrogels, with the following strain energy function:

$$W(\lambda) = \frac{1}{2}G \sum_i (\lambda_i^2 - 1) \quad (2)$$

G is the elastic modulus. The mechanical stress is derived from the strain energy function as follows:^{35–37}

$$\sigma_{ii} = \tilde{\phi} \lambda_i \frac{\partial W}{\partial \lambda_i} - \Pi_{\text{mix}} \quad (3)$$

Π_{mix} is the osmotic pressure, derived from the Cloizeaux scaling, which is described in more detail in the ESI.[†] At equilibrium the mechanical stress is zero everywhere in the gel. At equilibrium the polymer volume fraction is ϕ_0 , which will be different from ϕ_{ref} . Because of the use of the scaling law, it is convenient to define the reduce polymer fraction $\tilde{\phi} = \phi/\phi_{\text{ref}}$. The osmotic pressure is described by:

$$\Pi_{\text{mix}} = \alpha G \tilde{\phi}^\beta \quad (4)$$

For biopolymer gels it holds that $\beta = 9/4$.⁵⁷

The physical meaning of ϕ_{ref} for polymer gels is still controversial.^{71,72} Originally, it is said that ϕ_{ref} is the polymer volume fraction at the moment the polymers are crosslinked to form a network. Synthetic polymers are often crosslinked in absence of the solvent, and it is often taken that the dry state is the reference state, *i.e.* $\phi_{\text{ref}} = 1$.^{73,74} However, for biopolymer gels the network formation often happens with solvent present. Furthermore, there are physical bonds contributing to elastic properties of the gel, which are mediated *via* the solvent such as hydrogen bonding.⁵⁸ Hence, it holds that $\phi_{\text{ref}} \ll 1$, as is shown for biopolymer gels in our previous paper.⁵⁸ Other models for synthetic hydrogels also take this assumption. It is said that the reference state is defined by the θ -temperature, where the conformation of polymers in between crosslinks are closest to unperturbed Gaussian chains.^{75,76} In our scaling theory⁵⁸ ϕ_{ref} is fully defined by the degree of crosslinking, and thus it has a clear physical meaning. Hence, in contrast to the original Flory–Rehner theory the scaling theory has only one model parameter (the crosslink density or equivalently ϕ_{ref} , ϕ_0 or elastic modulus G in a defined state). Hence, in presence of a good solvent we prefer to use the scaling law over the Flory–Rehner theory. However, the scaling law does not cope yet with changes in swelling properties due to heating leading to protein denaturation or gel collapse, or shift in pH or ionic strength. In the models below, we will assume that the environmental conditions will be constant.

Via the models we will investigate the inflation and hydration of the elastic hydrogel shells due to internal pressure p_{int} . The shell represents the wall of a cell. The shell is immersed in a liquid bath of pure water – with zero osmotic pressure. The inner surface of the shell is impermeable to solutes, representing the cell membrane. *Via* increase of solute content of the vacuole, the cell can increase the osmotic pressure and swell.

This problem is solved with the condition of mechanical equilibrium $\nabla \cdot \sigma = 0$, formulated in spherical and cylindrical coordinates. It is subject to the following boundary conditions: $\sigma_{rr}(r=b) = 0$, and $\sigma_{rr}(r=a) = -p_{\text{int}}$. Here, σ_{rr} is the stress in radial direction. The cell membrane is positioned at $r = a$, and the cell wall is in the region $a < r < b$. Due to the inflation the radii a and b will change. The extension of the radii will be described with respect to the reference state, and we have defined $\Lambda_a = a/A$, and $\Lambda_b = b/B$, with A and B the radii in the reference state.



2.1 Spherical shell

For spherical geometries the principal directions for mechanical deformation are the radial direction, and the two orthogonal circumferential directions. The principal stretches are indicated with λ_r and λ_θ for the radial and circumferential direction respectively. The elastic shell is divided into control volumes. In the reference (relaxed) state the control volume has a radius of R , a thickness of dR and polymer volume fraction of ϕ_{ref} . In the swollen, deformed state, the control volume has radius r , thickness dr , and polymer volume fraction is ϕ . The radial stretching is defined as $\lambda_r = dr/dR$, and the circumferential stretching is defined as $\lambda_\theta = r/R$. Hence, from the incompressibility of polymer and solvent follows the condition:⁴³

$$\tilde{\phi} = \frac{\phi}{\phi_{\text{ref}}} = \frac{1}{\lambda_\theta^2 \lambda_r} \quad (5)$$

The condition for mechanical equilibrium in spherical coordinates is:⁴³

$$\frac{d\sigma_{rr}}{dr} + 2\frac{\sigma_{rr} - \sigma_{\theta\theta}}{r} = 0 \quad (6)$$

σ_{rr} is the stress in radial direction, and $\sigma_{\theta\theta}$ is the hoop stress.

The mechanical stresses are derived from the free energy functional:³⁵

$$\begin{aligned} \sigma_{\theta\theta} &= \frac{1}{\lambda_\theta \lambda_r} \frac{\partial W}{\partial \lambda_\theta} - \Pi_{\text{mix}} \\ &= \lambda_\theta \tilde{\phi} \frac{\partial W}{\partial \lambda_\theta} - \Pi_{\text{mix}} \\ \sigma_{rr} &= \frac{1}{\lambda_\theta^2} \frac{\partial W}{\partial \lambda_r} - \Pi_{\text{mix}} \\ &= \lambda_r \tilde{\phi} \frac{\partial W}{\partial \lambda_r} - \Pi_{\text{mix}} \end{aligned} \quad (7)$$

For the last lines we have used the incompressibility condition, eqn (5).

The difference in hoop and radial stresses is:

$$\sigma_{\theta\theta} - \sigma_{rr} = \tilde{\phi} \left[\lambda_\theta \frac{\partial W}{\partial \lambda_\theta} - \lambda_r \frac{\partial W}{\partial \lambda_r} \right] G \left(\tilde{\phi} \lambda_\theta^2 - \tilde{\phi}^{-1} \lambda_\theta^{-4} \right) \quad (8)$$

With the last line following after substitution of the Neo-Hookean strain energy function. It is an expression having only λ_θ as a variable. It is custom to define $\lambda = \lambda_\theta$.

Hence, the condition for mechanical equilibrium becomes:

$$d\sigma_{rr} = 2G \left(\tilde{\phi} \lambda^2 - \frac{1}{\tilde{\phi} \lambda^4} \right) \frac{dr}{r} \quad (9)$$

We need a relation between dr and $d\lambda$ to solve the condition for mechanical equilibrium. Pence and Tsai propose a conversion of the integration variable *via* the total derivative of $\lambda = r/R$, with use of the incompressibility condition:⁴³

$$\frac{dr}{r} = \frac{d\lambda}{\lambda \left(\tilde{\phi} - \lambda^3 \right)} \quad (10)$$

After replacing dr/r we obtain:

$$d\sigma_{rr} = 2G \left(\tilde{\phi} \lambda^2 - \frac{1}{\tilde{\phi} \lambda^4} \right) \frac{1}{\lambda - \tilde{\phi} \lambda^4} d\lambda \quad (11)$$

The local value of $\tilde{\phi}$ is determined by the condition of local chemical equilibrium.

As a first approximation, we can assume that for thin to moderately thick shells the swelling is uniform *cf.* ref. 43 Upon integration of the condition for mechanical equilibrium, we obtain an analytical expression:

$$\frac{2(p_{\text{int}} - p_{\text{ext}})}{G} = \frac{1 + 4\tilde{\phi} \Lambda_a^3}{\tilde{\phi} \Lambda_a^4} - \frac{1 + 4\tilde{\phi} \Lambda_b^3}{\tilde{\phi} \Lambda_b^4} \quad (12)$$

which is very similar to the expression for the inflation of an elastomeric shell, as derived in the ESI.† The above expression differs due to the presence of the swelling factor $\tilde{\phi}$. This implies that even for thin cell walls, with RWC \approx 5%, it will still be important to include the water retention in the apoplast for correct prediction of mechanical behaviour.

An approximation for $\tilde{\phi}$ follows from the boundary condition at $r = b$, *i.e.* $\sigma_{rr}(b) = 0$. Using eqn (7) and (4), we obtain:

$$\tilde{\phi}_b = (\alpha \Lambda_b^4)^{-1/(\beta+1)} \quad (13)$$

We will compare the above analytical expressions with numerical simulation, with the solution method as described in the ESI.† A schematic of the computed deformation occurring during inflation is shown in Fig. 1. For clarity of the figure, we have computed the deformation of a cell with a rather thick cell wall, and a large pressurization with $\Lambda_b = 1.75$. In the same figure we have also depicted the cell in the stress-free state. The coordinates are rescaled by the cell radius in the stress-free state, R_0 . Hence, the inner side of the cell wall is positioned at $r/R_0 = 1.0$. As said we

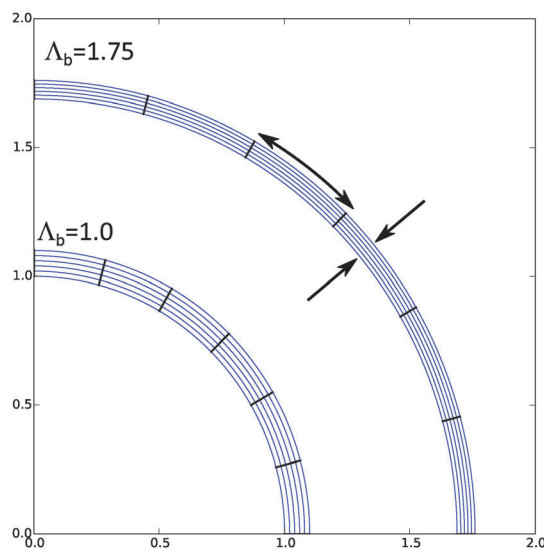


Fig. 1 Schematic of the deformation of cell wall during pressurization. We have depicted the mesh of the stress-free cell wall (with $\Lambda_b = 1.0$) and the mesh of the inflated cell wall (with $\Lambda_b = 1.75$) – as is computed from the numerical model. Observe the compression in radial direction ($\lambda_r < 1$), and the extension in the hoop direction ($\lambda_\theta > 1$).



have taken a thick cell wall, with the outer radius at $B/R_0 = 1.1$. Observe from the mesh that in the stress-free state the cell wall has uniform deformation, and uniform polymer volume fraction.

Inflated cells show large inhomogeneity in the deformation and hydration, as is shown in Fig. 2. There we have performed computation for the same cell as above, but now with $A_b = 1.4$. Hence, for hydration of cell walls, it is indeed important to take non-homogeneous deformation into account.

We have performed a series of computations for a range of deformations $1.0 < A_b < 1.4$, and cell wall thickness $B/R_0 = 1.1$. We compare these computations on the basis of inflation pressure, $\Delta p/G$, versus cell volume V/V_0 , the hydration at the outer radius of the cell wall ϕ_b as function of cell size r/R_0 . These relations can be compared to the analytical solutions for the p - V relation, eqn (12), and for the polymer volume fraction at the outer radius, eqn (13). Results are shown in Fig. 3 and 4. As one can observe the numerical solution quite follows the analytical solution, except for high inflation of the cell. As shown in Fig. 5, we have also computed the relative amount of water in the cell wall (RWC), which decreases from 25% to 15%. The decrease of the polymer volume fraction $\langle \phi \rangle$ does show that the absolute amount of water is increasing, but not as fast as in the inflating cell (Fig. 4).

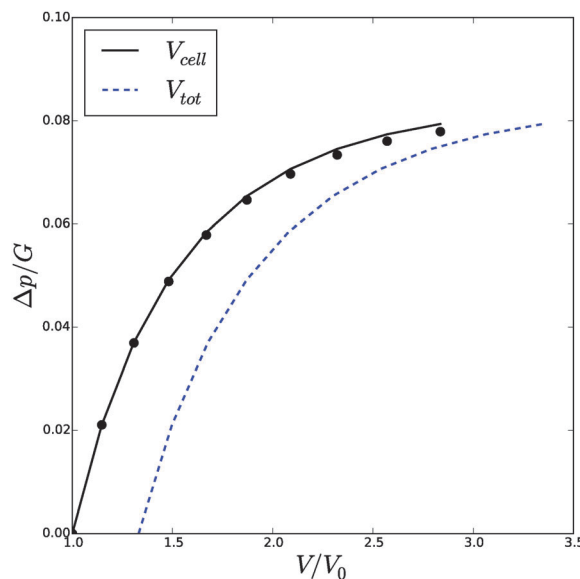


Fig. 3 Cell volume, V_{cell} and total volume of a spherical cell and its cell wall, V_{tot} , as function of turgor pressure Δp . The volumes are scaling with the cell volume in the stress free state, V_0 , and the pressure with the elastic modulus G of the cell in the reference state.

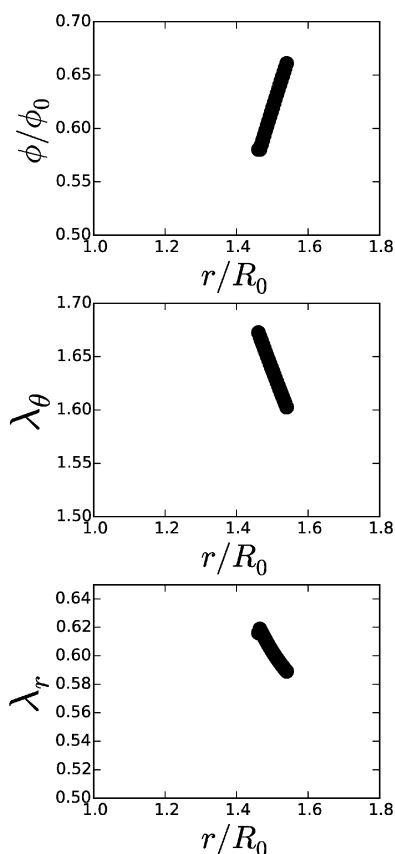


Fig. 2 Local deformation and hydration of an inflated cell wall, as computed for a thick cell wall (outer radius in stress-free state is $B/R_0 = 1.1$) and large deformation ($A_b = 1.4$). Observe the non-homogeneous deformation, with compression in radial direction and extension in hoop direction. The hydration is even more inhomogeneously distributed along the radial direction than the mesh deformation.

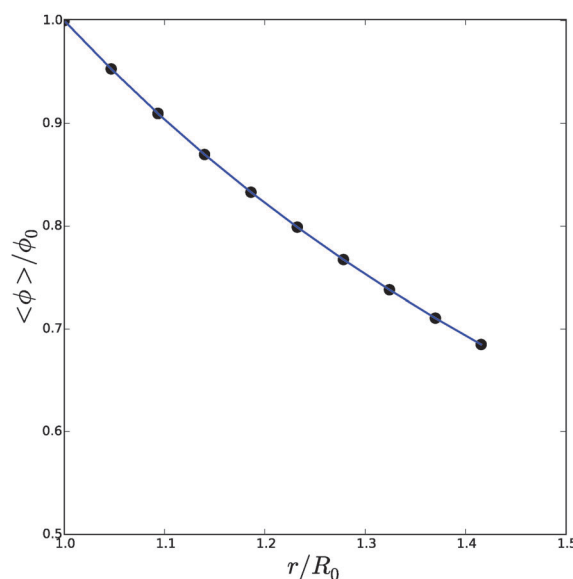


Fig. 4 Average polymer volume fraction, $\langle \phi \rangle / \phi_0$, as function of cell size r/R_0 , as follows from the numerical solution (symbols) and compared to the analytical solution for ϕ_b cf. eqn (13).

2.2 Cylindrical shells

For cylindrical geometries the principal directions for mechanical deformation are the radial direction, the circumferential direction, and the axial direction. The principal stretches are indicated with λ_r , λ_θ and λ_z for the radial, circumferential and axial direction respectively. For sake of simplicity we first like to investigate the two-dimensional problem with a constant axial stress, cf. ref. 42. The axial stretch is assumed equal to that of the zero stress state: $\lambda_z = A_z = \phi_0^{-1/3}$. The solution method follows similar lines as the spherical case.



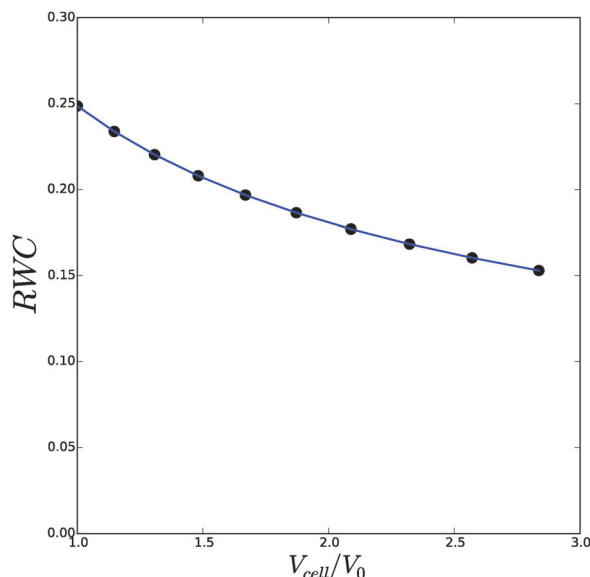


Fig. 5 The relative cell wall water content (RWC) as function of the spherical cell volume V_{cell}/V_0 . Symbols follow from the numerical solution, and the line follows from the analytical solution.

The incompressibility condition for cylinders is:⁴²

$$\tilde{\phi}\lambda_\theta\lambda_r\lambda_z = 1 \quad (14)$$

The condition for mechanical equilibrium in cylindrical coordinates is:

$$\frac{d\sigma_{rr}}{dr} + \frac{\sigma_{rr} - \sigma_{\theta\theta}}{r} = 0 \quad (15)$$

The expressions for the mechanical stresses $\sigma_{\theta\theta}$ and σ_{rr} are identical to those for the swelling spherical shell, eqn (7).

Using the condition of incompressibility, eqn (14), the stress difference becomes:

$$\sigma_{\theta\theta} - \sigma_{rr} = G\left(\tilde{\phi}\lambda^2 - \frac{1}{\tilde{\phi}\lambda^2 A_z^2}\right) \quad (16)$$

Hence, the condition for mechanical equilibrium becomes:

$$d\sigma_{rr} = G\left(\tilde{\phi}\lambda^2 - \frac{1}{\tilde{\phi}\lambda^2 A_z^2}\right)\frac{dr}{r} \quad (17)$$

We require the relation between dr and $d\lambda$:⁴²

$$\frac{dr}{r} = \frac{d\lambda}{\lambda - \tilde{\phi}A_z\lambda^3} \quad (18)$$

After replacing dr/r we obtain:

$$d\sigma_{rr} = G\left(\tilde{\phi}\lambda^2 - \frac{1}{\tilde{\phi}\lambda^2 A_z^2}\right)\frac{d\lambda}{\lambda - \tilde{\phi}A_z\lambda^3} \quad (19)$$

This differential equation can not be solved, because A_z is still unknown. The axial stretch is determined by the force applied in that direction. We investigate the case of zero axial force. The force balance reads:⁷⁷

$$F_z = \int_a^b \sigma_{zz} 2\pi r dr - \Delta p \pi a^2 = 0 \quad (20)$$

Following the Neo-Hookean model, the stress in axial direction is:

$$\sigma_{zz} = \tilde{\phi}\lambda_z \frac{\partial W}{\partial \lambda_z} - \Pi_{\text{mix}} = \tilde{\phi}GA_z^2 - \alpha G\tilde{\phi}^\beta \quad (21)$$

We assume that the axial stretch is uniform over the whole cylinder. Furthermore, if we assume uniform swelling, the force balance reads:

$$\frac{\Delta p}{G} = \left(\tilde{\phi}A_z^2 - \alpha\tilde{\phi}^\beta\right)\left((A_b/A_a)^2 - 1\right) \quad (22)$$

An approximate analytical solution for the pressure difference over the shell is obtained if uniform swelling is assumed.⁴³ Upon integration, we obtain:

$$\frac{\Delta p}{G} = \frac{1}{2\tilde{\phi}A_z^2 A_a^2} - \frac{1}{2\tilde{\phi}A_z^2 A_b^2} - \frac{\ln(A_a)}{\tilde{\phi}A_z} + \frac{\ln(A_b)}{\tilde{\phi}A_z} \quad (23)$$

which is very similar to the analytical expression for the elastomeric cylindrical shell, shown in the ESI,[†] except for the swelling factor $\tilde{\phi}$. We have also found this for the spherical case.

An approximate solution for the volume fraction $\tilde{\phi}$, follows from the boundary condition at the outer surface, $\tilde{\phi}_{rr}(r=b)=0$:

$$\tilde{\phi} \approx \tilde{\phi}_b = (\alpha A_b^2 A_z^2)^{-1/(\beta+1)} \quad (24)$$

The three above equations can be solved simultaneously to render solutions for $\tilde{\phi}$, Δp and A_a for any given inflation of the cylindrical shell A_b . We will compare these analytical expressions with the numerical solution, which is computed with the method described in the ESI.[†] We solve the problem for a range of stretches of the outer surface $1 < A_b < 1.6$, and for a cell wall thickness of 5% of the cell radius. In Fig. 6 and 7 we have compared the numerical solutions with the analytical approximations for respectively (a) the distribution of the polymer

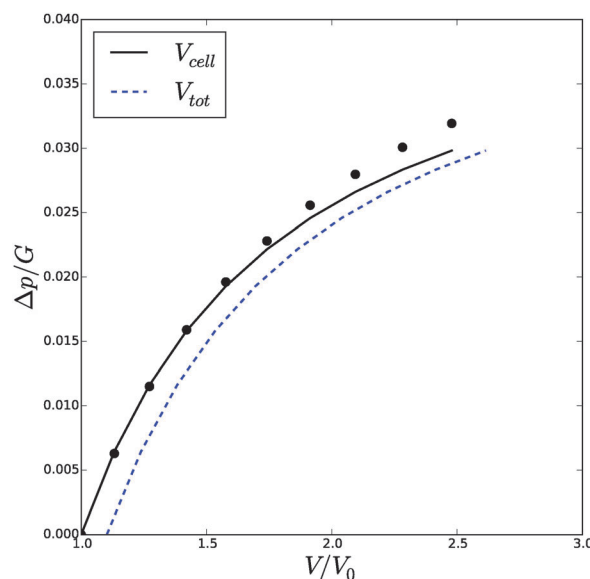


Fig. 6 Volume of the cell V_{cell} and the total volume, V_{tot} , as function of the turgor pressure in the cell Δp for an inflated cylindrical shell. The symbols indicate the analytical solution of the pressure–volume relation, eqn (23), assuming a homogeneous swelling and axial stretch of the gel.



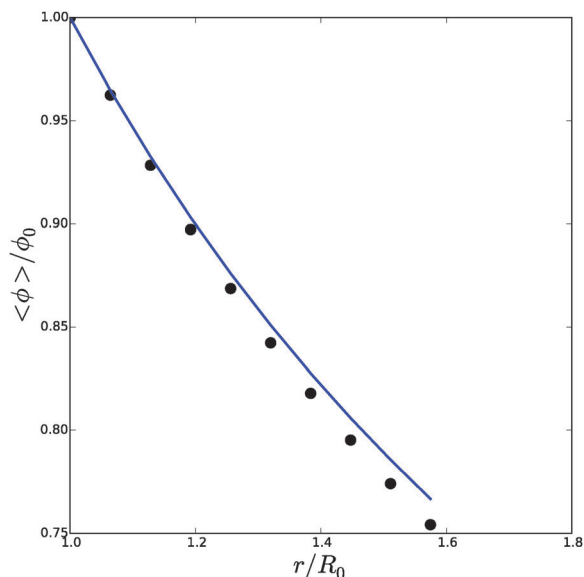


Fig. 7 Average polymer volume fraction at the outer radius of an inflated cylindrical shell, as function of cell size r/R_0 . Symbols indicate the numerical solution, and the solid line is the analytical solution for ϕ_b , cf. eqn (24).

fraction, (b) the turgor pressure – cell volume relation. The numerical results are comparable with the analytical results, except for high stretching of the cell wall. Deviations are due to the assumptions of homogeneous swelling in the analytical solutions. Comparing these figures with those for the spherical case, we observe little difference in the general trends – but there is a stronger deviation of the numerical solutions from the analytical solutions. The deviation of the average polymer fraction $\langle \phi \rangle$ with that at the outer radius ϕ_b is of course a good indication of the inhomogeneity of deformation and hydration.

3 Advanced models

Real cell walls are composite materials showing more complex mechanical behaviour, such as strain hardening and anisotropic deformation, due to presence of aligned microfibrils. In this section we investigate these kinds of mechanical behaviour. Strain hardening will be investigated for spherical cells, and anisotropic swelling for cylindrical cells.

The modelling of this complex behaviour requires more elaborate strain energy functions, formulated in terms of the tensor invariants of the Cauchy tensor $\mathbf{C} = \mathbf{F}^T \mathbf{F}$, with \mathbf{F} the deformation tensor, indicating the deformation of control volume elements. In absence of rotational deformation, the deformation tensor is a diagonal tensor containing the principal stretches λ_i , which are its eigenvalues. The eigenvalues of the Cauchy tensor are λ_i^2 . The tensor invariants of the Cauchy tensor are:

$$\begin{aligned} I_1 &= \text{tr}(\mathbf{C}) = \lambda_1^2 + \lambda_2^2 + \lambda_3^2 \\ I_2 &= \frac{1}{2}[\text{tr}(\mathbf{C})^2 - \text{tr}(\mathbf{C}^2)] = \lambda_1^2 \lambda_2^2 + \lambda_2^2 \lambda_3^2 + \lambda_3^2 \lambda_1^2 \\ I_3 &= \det(\mathbf{C}) = \det(\mathbf{J})^2 = \lambda_1^2 \lambda_2^2 \lambda_3^2 \end{aligned} \quad (25)$$

For an incompressible material holds $\det(\mathbf{J}) = 1$.

3.1 Strain hardening

The first model incorporating strain hardening has been formulated by Fung,^{62,63} where

$$W = \frac{G}{2k} \exp[k(I_1 - 3)] \quad (26)$$

Indeed recently, it is found that many biopolymer networks made with collagen, actin or fibrin are shown to exhibit strain hardening, which is described by the above strain energy function.⁶⁴ However, the Fung model does not account for swelling. We modify the expression because $I_1 \neq 3$ at zero stress conditions. As $\phi_0 \neq \phi_{\text{ref}}$ the polymers will be stretched with $\lambda_i^3 = \Lambda_0^3 = 1/\tilde{\phi}_0$. Hence, at zero stress conditions it holds $I_1 = I_{1,0} = 3\Lambda_0^2$. A more proper expression of the strain energy function for strain hardening of swelling materials is:

$$W = \frac{G}{2k} \exp[k(I_1/I_{1,0} - 1)] \quad (27)$$

A similar transformation of models of non-swelling materials into models of swelling materials has been proposed earlier by Treloar,²⁸ by Gent⁷⁴ and Pence and coworkers.^{42,43,66}

Recall that for swelling spherical shells holds $\lambda_\theta^2 \lambda_r = 1/\tilde{\phi}$, and hence

$$I_1 = (2\lambda^2 + \tilde{\phi}^{-2}\lambda^{-4} - 3) \quad (28)$$

We compute $\sigma_{\theta\theta} - \sigma_{rr}$ via:

$$\begin{aligned} \sigma_{\theta\theta} - \sigma_{rr} &= \tilde{\phi} \frac{\partial W}{\partial I_1} \left(\lambda_\theta \frac{\partial I_1}{\partial \lambda_\theta} - \lambda_r \frac{\partial I_1}{\partial \lambda_r} \right) \\ &= \tilde{\phi} G \exp[k_2(I_1/I_{1,0} - 1)] \left(\lambda^2 - \frac{1}{\tilde{\phi}^2 \lambda^4} \right) \end{aligned} \quad (29)$$

and the condition for mechanical equilibrium becomes:

$$d\sigma_{rr} = 2G \left(\tilde{\phi} \lambda^2 - \tilde{\phi}^{-1} \lambda^{-4} \right) \exp[k_2(I_1/I_{1,0} - 1)] \frac{dr}{r} \quad (30)$$

Compared to the Neo-Hookean model, the condition for mechanical equilibrium is multiplied with just a factor $\exp[k_2(I_1/I_{1,0} - 1)]$. Using a Taylor expansion, an approximate analytical expression can be obtained. Below, we show only the numerical solution – where we focus on the pressure volume relation.

The numerical results concerning the p - V relation are shown in Fig. 8, which are compared to experimental data for cells of bell pepper leaves. The experimental data has been fitted by estimating $V_0/V_{\text{max}} = 3$ and $G = 100$ kPa, using an assumed ratio of $k/G = 5$, and $B/R_0 = 1.06$. V_{max} is the maximal cell volume at full turgor. One can observe that the model now follows the experimental trend. At large strains (V/V_0) further deformation of the cell requires an increasingly larger turgor pressure. This behaviour is quite opposite to inflating Neo-Hookean spherical shells, whose behaviour we have shown in Fig. 3. There, deformation at large strains requires increasingly smaller increments of turgor pressure. The latter behaviour is also well known for rubber balloons, but is absent in biological cells, thanks to the presence of microfibrils. For spherical cells the microfibrils are randomly oriented, leading to strain hardening, but isotropic deformation.



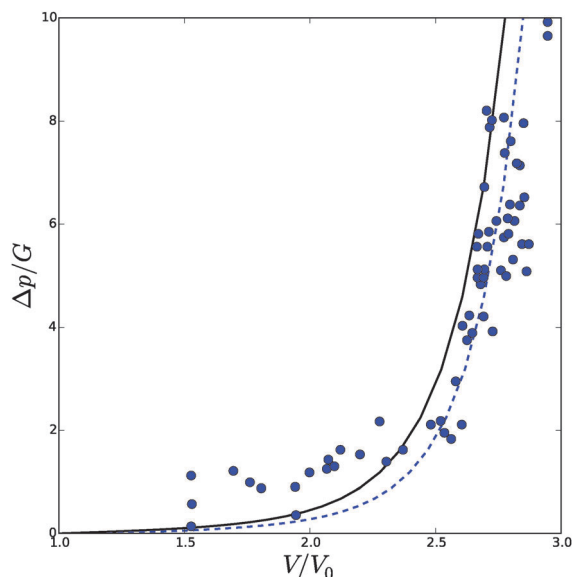


Fig. 8 Total volume of the cell V_{tot} as function of the turgor pressure in a spherical cell p , with strain hardening cell wall material. Experimental data (symbols) hold for pepper,⁶⁰ and simulation data (line) hold for the modified Fung model eqn (27), with $k/G = 5$. The experimental data are rescaled with estimates of $V_0/V_{\text{max}} = 3$ and $G = 100$ kPa, which fitted the data.

Viewing the debate in plant science on the amount of bound water, it is instructive to analyse the relative water content of the cell wall as function of the inflation. The change in RWC following the previous simulation are shown in Fig. 9. We observe large differences in RWC between hydrated and elastomeric shells at low values of inflation. RWC of the elastomeric shell varies only between 2% and 5%, while the RWC of the hydrated

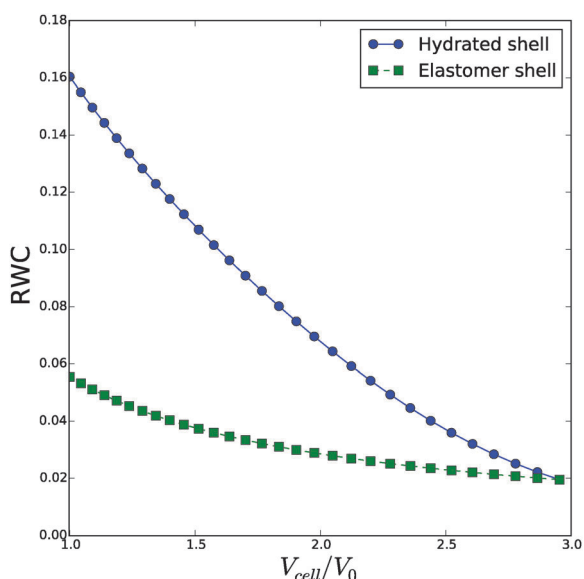


Fig. 9 Relative water content (RWC) in the cell wall as a function of cell volume V_{cell}/V_0 , computed for a hydrated and elastomeric shell. The elastomeric shell does not absorb water and thus retain a constant volume, which is taken equal to that of the hydrated shell at maximal inflation ($V_{\text{cell}}/V_0 = 3$).

shell varies from 16% to 2%. The change of RWC of the strain hardening, hydrated shell with inflation is opposite to the change of a hydrated Neo-Hookean shell, where hydration increases with inflation. In real biological tissue the strain hardening prevents this phenomenon. The consequences of this behaviour will be discussed in the next section, after we have considered the effect of fibrils.

3.2 Anisotropic deformation

We consider anisotropic deformation due to oriented fibrils in cylindrical cells. This cell model is representative for hyphen of fungi. Particular, we take the situation of the fruiting body of the *Agaricus bisporus* mushroom, which we have investigated earlier.⁸ In these hyphen chitin microfibrils are particularly oriented in the circumferential direction, while in mature cells we find also β -glucans, which predominantly oriented along the cylinder-axis.⁶¹ Effectively, we assume that the fibers have an orientation of $\psi_0 \approx \pi/4$.

Strain energy functions for composite materials with oriented microfibrils have been developed in particular for arteries.^{65,78} The strain energy function has two contributions, $W = W_{\text{iso}} + W_{\text{aniso}}$. The first part describes the isotropic deformation of the gel-like matrix, $W_{\text{iso}} = W_{\text{iso}}(I_1, I_2, I_3)$, while the second part describes the anisotropic deformation, which is a function of other invariants I_4, I_5, I_6 . These higher order invariants measure the stretch in the particular fibre direction $t_0 = (\cos \psi_0, \sin \psi_0)$, with ψ_0 the fibre direction with respect to the cylinder rotational axis. The invariant is defined as:⁷⁹

$$I_4 = t_0 \cdot C t_0 = \lambda_{\theta}^2 \cos^2 \psi_0 + \lambda_z^2 \sin^2 \psi_0 \quad (31)$$

We investigate models, with $W_{\text{iso}} = W_{\text{iso}}(I_1)$ and $W_{\text{aniso}} = W_{\text{aniso}}(I_4)$, cf. ref. 65:

$$W_{\text{iso}} = \frac{1}{2} G (I_1 - 3) \quad (32)$$

$$W_{\text{aniso}} = \frac{k_1}{2k_2} \left(\exp \left[k_2 (I_4/I_{4,0} - 1)^2 \right] - 1 \right)$$

with $I_{4,0} = \tilde{\phi}_0^{-2/3}$. Similar to the previous section we impose that at $\phi = \phi_0$ the material is stress free.

The principle stresses are equal to:

$$\sigma_{\theta\theta} = \tilde{\phi} G \lambda_{\theta}^2 - \Pi_{\text{mix}} + \tilde{\phi} \lambda_{\theta}^2 \frac{k_1}{k_2} \exp \left[k_2 (I_4/I_{4,0})^2 \right] \cos^2 \psi_0 \left(\frac{I_4}{I_{4,0}} - 1 \right) \quad (33)$$

$$\begin{aligned} \sigma_{rr} &= \tilde{\phi} G \lambda_r^2 - \Pi_{\text{mix}} \\ \sigma_{zz} &= \tilde{\phi} G \lambda_z^2 - \Pi_{\text{mix}} + \tilde{\phi} \lambda_z^2 \frac{k_1}{k_2} \exp \left[k_2 (I_4/I_{4,0})^2 \right] \sin^2 \psi_0 \left(\frac{I_4}{I_{4,0}} - 1 \right) \end{aligned} \quad (34)$$

The numerical solution procedure follows largely that of isotropic swelling, cylindrical shells. Computations are performed for $k_1/G = 1$, $k_2/G = 2.9$, and $B = 1.05R_0$. Results of numerical simulation are shown in Fig. 10–12. From Fig. 11 we observe that the stretching is indeed anisotropic. The axial



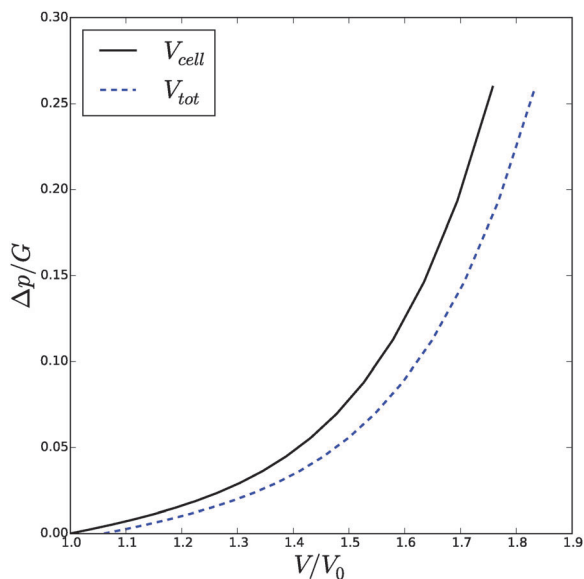


Fig. 10 Volume of the cell V_{cell} and the total volume of cell and cell wall, V_{tot} , as function of the turgor pressure in the cell Δp for a swelling cylindrical shell reinforced with fibers.

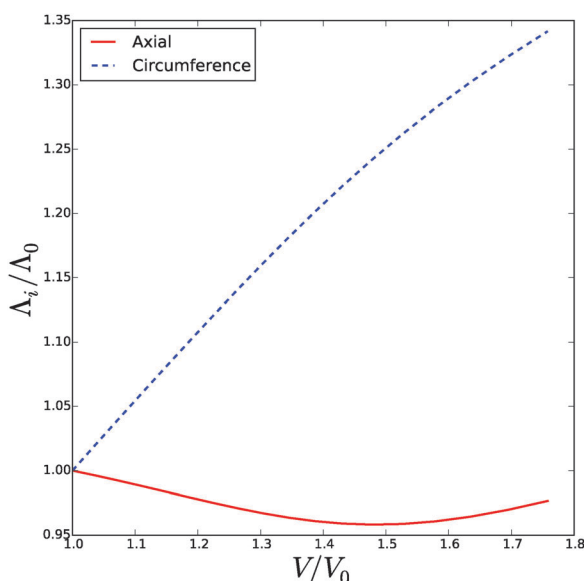


Fig. 11 Axial stretch Λ_z and radial stretch Λ_r at $r = a$ of a swelling cylindrical shell reinforced with fibers, as function of the cell volume. The orientation angle of the fibers is $\theta \approx \pi/4$ leading to restricted expansion in radial direction, in comparison to the axial direction.

stretch is quite limited, while the circumferential stretch increases more or less linear with the cell volume. At large expansions of the cell volume, the radial distribution of the polymer fraction ϕ , becomes significantly inhomogeneous. The pressure–volume curve is qualitatively similar to that of bell pepper. However, the decrease of RWC is not as dramatically as for bell pepper. This may depend on the particular values we have chosen.

It is interesting to note in simulations with small fiber angles $\psi_0 < \pi/8$ (with data shown in ESI[†]), the behaviour is

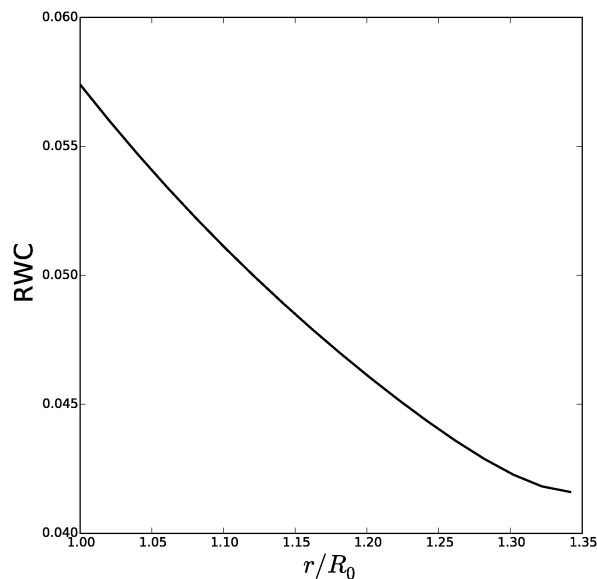


Fig. 12 Relative water content (RWC) of the cell wall during inflation of cylindrical cell with fibers.

quite different, where the cell predominantly expands in the axial direction. This fiber configuration definitely promotes the growth of hyphen in axial direction, as has been observed experimentally.⁶¹

4 Impact of cellular structure on water holding capacity of plant food materials

The hydration behaviour of most plant food materials will be in line with Fig. 8. At harvest the cells are expected to be in full turgor. At full turgor the polymers the microfibrils in the cell wall are in tension, and render the strain hardening. A small, but not-negligible part of the water held in the tissue is held by the cell wall material. Because of the highly stretched state of the biopolymer network, with $\phi \ll \phi_0$, it is far less hydrated than in the stress free state. The behaviour of mushrooms, having cylindrical cells with oriented microfibrils, is more complicated. Though, the pressure–volume relation is similar to that of plant foods with more spherical cells. Its different behaviour is more apparent in the stretching in the axial direction, which extends during strain hardening – a reflection of the propensity of the hyphens to grow in axial direction. Due to the axial extension the change in RWC is much less pronounced as for spherical cells.

Many foods will be processed prior to consumption, *via* cutting, heating or freezing. This processing makes that the cell membrane loses its integrity. Consequently, solutes from the vacuole will diffuse out of the cell, and the osmotic and turgor pressure are lost. The cell wall will return to the zero-stress state. In this state the cell volume V_0 is much less than in the full turgor state. However, in the stress-free state the cell wall is more hydrated than in the full turgor state, see Fig. 8 and 12.



Furthermore, the solutes from the vacuole will diffuse into the cell wall. Hence, they will improve the hydration of the cell wall material *via* increase of the water activity.^{7,8} Consequently, a significant part of the water liberated by the loss of cell volume, will be resorbed by the unstressed cell wall material and solutes diffused into the cell wall material. After loosing the cell membrane integrity, the water held inside the cell will be held in pores in between the plasmolyzed cell and cell wall. This water is held by capillary forces, and is easily lost *via* gravity or *via* the mechanical load during oral processing.

Often, the net effect of processing is that the water holding will be less than for the fresh food, having intact cell membranes. In practice, this loss of water holding is noticed *via* the water loss (drip) after thawing of frozen plant foods.¹⁷ The effect of cooking is more complicated because proteins denature and crosslinks in the cell wall material will be hydrolysed. Proteins denaturation will lower water holding,¹¹ while hydrolysis of crosslinks will increase it. At temperatures above 80 degrees there is thermal hydrolysis of pectin, which will lower the number of crosslinks in the polymer network – which will increase the water holding. However, if the heating is performed slowly, enzymes like PME (Pectin-Methyl-Esterase) will be activated around 50 degrees, which will increase the crosslinks if calcium is present.⁸⁰

It appears that freezing is more appropriate to study the effect of the cellular structure on the water holding capacity.⁸¹ However, during freezing the cell walls of several cells will be collapsed on top of each other (due to the growing ice crystals).⁸² During compression new crosslinks will be formed. This becomes apparent due to the inability of the cells to reswell completely during rehydration after freeze-drying.⁸³ A more appropriate method will be electroporation of the cell membranes as with the Pulsed Electric Field method, provided that the temperature rise due to Joule heating is limited.⁸⁴

Also for plant science it is important to note the ability of the cell wall material to retain more water in situations, where the cell wall is less stressed than at full turgor. This situation occurs during drought stress. During loss of turgor the amount of symplasmatic water decreases, but a significant part of it can be resorbed by the apoplast because of its enhanced hydration. Our findings show that the hypothesis in plant science that bound water is independent of cell volume should be reconsidered.

Summarizing, this study implies that in models for water relations in plants one needs account for the water held in the apoplast. The models similar to those presented in this paper can be used for this purpose.

5 Conclusions

In this paper we have presented hyper-elastic models for the investigation of the hydration of cellular tissue. We have used pressurized, spherical and cylindrical shells as model systems for walled cells, as can be found in plants and fungi. The pressurized internal cavity represent the cell bounded by the cell wall, that is impermeable to solutes inside the vacuole. The internal pressure represents the turgor pressure. The shell

represents the polymeric material of the cell: the cytoplasm and cell wall. The shell has been modelled as a hydrogel and a composite material with microfibrils. We have solved these models numerically, and obtained how the cell volume changes as function of the internal pressure.

Furthermore, the hydration of the cell wall changes with cell volume. The composite character of the cell wall leads to strain hardening, and anisotropic deformation in case of cylindrical cells. Strain hardening of the cell wall becomes apparent if the cell is in the full turgor state. For cells in this state, it requires a much larger increase of pressure to obtain cell volume change, than compared with cells in the zero stress state. Anisotropic deformation is induced to microfibrils with a preferred orientation.

We have interpreted the results for the impact of the cellular structure on the water holding capacity of plant food materials. From the comparison of the cell volume and the hydration of the cell wall of cells in the zero stress state and the full turgor state, we conclude that the cellular structure, and particularly the integrity of the cell membrane has a significant impact on the water holding capacity. This aspect has hardly been acknowledged in food science, dealing with water holding capacity. The theory we are developing for water holding capacity of foods, definitely has to be extended to account for the cellular structure. The presented hyperelastic models provide the proper framework for that.

The found results have significant impact for other scientific fields. One such field is in plant science, dealing with water relations. There, the hydration of cell wall material is hardly acknowledged – but our results show that the cell wall (or rather apoplast) gives a significant contribution to the hydration of the tissue.¹⁵ Its significance is even larger under drought stress. Moreover, mucilage-like apoplast in cactus stems and plant roots have clearly a function to attract and store water, almost the similar amount as stored in the cells.^{85,86} Furthermore, water transport in plants is mainly *via* the apoplast (the connected cell walls of all cells in the tissue), and is enhanced if the cells are dehydrated. Another field is the post harvest biology, where one has found that cell wall hydration and swelling are highly relevant processes for of fruit ripening.^{26,87} Next to ripening, the softening of fruit is also controlled by moisture loss *via* evaporation. Recently, the water transport problem in fruit are approached with hyperelastic models, but which neglect the water holding capacity and swelling/shrinkage of the cell wall material.^{88–90} These models can be extended like the models presented in this paper. The multiscale simulation approach for the water transport in fruit, extended with more appropriate microscale models as presented here, is also the proper framework for modelling drying and shrinkage of any food material, based on cellular tissue.

The model system used, of an pressurized elastic, hydrated shell is also usefull for bubble expansion during bread baking, and snacks frying.^{91–93} The internal pressure is generated by the gas inside the bubble. The expansion will be restrained by the visco-elasticity of the matrix, which can be regarded as a hydrated biopolymer network. One often associates strain hardening to the gluten matrix in bread dough, which is important for the stability of bubbles during proofing.⁹⁴ The presented models



are well suited for these problems, if extended for the viscous properties of the matrix.

For the field of responsive gels the presented model can also mean a step forward. The current models in this field are largely based on the Flory–Huggins theory for the osmotic pressure, Π_{mix} , and the Neo-Hookean strain energy function.^{95–97} But in more advanced models, incorporating strain hardening or anisotropic deformation due to fibrils, swelling has not been taken into account. This paper shows examples how to extend that. Furthermore, the use of the expression of the osmotic pressure from the Cloizeaux scaling law could benefit the field, because (1) it allows for more analytical solutions given simple geometries – instead of analytical Taylor series expansions,⁹⁸ and (2) it has less model parameters. One should note that the Cloizeaux scaling is only applicable in the semi-dilute regime. However, for (biopolymeric) hydrogels in the concentrated regime also the Flory–Huggins theory must be modified *via* making the interaction parameter dependent on the polymer volume fraction – to account for effects of hydrogen bonding.^{58,99}

References

- 1 M. T. Tyree and P. G. Jarvis, Water in tissues and cells, *Physiological plant ecology II*, Springer, 1982, pp. 35–77.
- 2 A. S. Szczesniak and R. Ilker, The meaning of textural characteristics-juiciness in plant foodstuffs, *J. Texture Stud.*, 1988, **19**(1), 61–78.
- 3 E. Huff-Lonergan and S. M. Lonergan, Mechanisms of water-holding capacity of meat: the role of postmortem biochemical and structural changes, *Meat Sci.*, 2005, **71**(1), 194–204.
- 4 P. Gervais and P. Molin, The role of water in solid-state fermentation, *Biochem. Eng. J.*, 2003, **13**(2), 85–101.
- 5 R. G. M. van der Sman, Modeling cooking of chicken meat in industrial tunnel ovens with the flory-rehner theory, *Meat Sci.*, 2013, **95**(4), 940–957.
- 6 R. G. M. van der Sman, E. Paudel, A. Voda and S. Khalloufi, Hydration properties of vegetable foods explained by flory-rehner theory, *Food Res. Int.*, 2013, **54**(1), 804–811.
- 7 R. G. M. van der Sman, F. J. Vergeldt, H. Van As, G. Van Dalen, A. Voda and J. P. M. Van Duynhoven, Multiphysics pore-scale model for the rehydration of porous foods, *Innovative Food Sci. Emerging Technol.*, 2014, **24**, 69–79.
- 8 E. Paudel, R. M. Boom and R. G. M. van der Sman, Change in water-holding capacity in mushroom with temperature analyzed by flory-rehner theory, *Food Bioprocess Technol.*, 2014, 1–11.
- 9 P. J. Flory and J. Rehner Jr, Statistical mechanics of cross-linked polymer networks ii. swelling, *J. Chem. Phys.*, 1943, **11**(11), 521–526.
- 10 R. G. M. Van der Sman, Moisture transport during cooking of meat: an analysis based on flory-rehner theory, *Meat Sci.*, 2007, **76**(4), 730–738.
- 11 R. G. M. van der Sman, Thermodynamics of meat proteins, *Food Hydrocolloids*, 2012, **27**(2), 529–535.
- 12 H. H. Wiebe, Matric potential of several plant tissues and biocolloids, *Plant Physiol.*, 1966, **41**(9), 1439–1442.
- 13 J. H. McClendon, The balance of forces generated by the water potential in the cell-wall-matrix—a model, *Am. J. Bot.*, 1981, 1263–1268.
- 14 S. P. Hardegree, Derivation of plant cell wall water content by examination of the water-holding capacity of membrane-disrupted tissues, *J. Exp. Bot.*, 1989, **40**(10), 1099–1104.
- 15 I. F. Wardlaw, Viewpoint: consideration of apoplastic water in plant organs: a reminder, *Funct. Plant Biol.*, 2005, **32**(6), 561–569.
- 16 A. R. Rahman, W. L. Henning and D. E. Westcott, Histological and physical changes in carrots as affected by blanching, cooking, freezing, freeze drying and compression, *J. Food Sci.*, 1971, **36**(3), 500–502.
- 17 S. Phothiset and S. Charoenrein, Effects of freezing and thawing on texture, microstructure and cell wall composition changes in papaya tissues, *J. Sci. Food Agric.*, 2014, **94**(2), 189–196.
- 18 A. Rascio, G. Nicastro, E. Carlino and N. Di Fonzo, Differences for bound water content as estimated by pressure–volume and adsorption isotherm curves, *Plant Sci.*, 2005, **169**(2), 395–401.
- 19 M. Santakumari and G. A. Berkowitz, Protoplast volume: water potential relationship and bound water fraction in spinach leaves, *Plant Physiol.*, 1989, **91**(1), 13–18.
- 20 M. Barg, M. V. Agüero, A. Yommi and S. I. Roura, Evolution of plant water status indices during butterhead lettuce growth and its impact on post-storage quality, *J. Sci. Food Agric.*, 2009, **89**(3), 422–429.
- 21 S. Maatallah, M. E. Ghanem, A. Albouchi, E. Bizid and S. Lutts, A greenhouse investigation of responses to different water stress regimes of laurus nobilis trees from two climatic regions, *J. Arid Environ.*, 2010, **74**(3), 327–337.
- 22 F. R. Harker, M. G. H. Stec, I. C. Hallett and C. L. Bennett, Texture of parenchymatous plant tissue: a comparison between tensile and other instrumental and sensory measurements of tissue strength and juiciness, *Postharvest Biol. Technol.*, 1997, **11**(2), 63–72.
- 23 M. U. H. Joardder, R. J. Brown, C. Kumar and M. A. Karim, Effect of cell wall properties on porosity and shrinkage of dried apple, *Int. J. Food Prop.*, 2015, **18**(10), 2327–2337.
- 24 D. M. Bruce, Mathematical modelling of the cellular mechanics of plants, *Philos. Trans. R. Soc. London, Ser. B*, 2003, **358**(1437), 1437–1444.
- 25 R. Ilker and A. S. Szczesniak, Structural and chemical bases for texture of plant foodstuffs, *J. Texture Stud.*, 1990, **21**(1), 1–36.
- 26 R. J. Redgwell, E. MacRae, I. Hallett, M. Fischer, J. Perry and R. Harker, In vivo and in vitro swelling of cell walls during fruit ripening, *Planta*, 1997, **203**(2), 162–173.
- 27 L. R. G. Treloar, The elasticity and related properties of rubbers, *Rep. Prog. Phys.*, 1973, **36**(7), 755.
- 28 L. R. G. Treloar, *The physics of rubber elasticity*, Oxford University Press, 1975.
- 29 L. R. G. Treloar, Swelling of bonded-rubber cylinders, *Polymer*, 1976, **17**(2), 142–146.
- 30 D. R. P. Hettiaratchi and J. R. O'callaghan, A membrane model of plant cell extension, *J. Theor. Biol.*, 1974, **45**(2), 459–465.



- 31 H. I. Wu and P. J. H. Sharpe, Stomatal mechanics ii*: material properties of guard cell walls, *Plant, Cell Environ.*, 1979, **2**(3), 235–244.
- 32 H. Wu, R. D. Spence, P. J. H. Sharpe and J. D. Gieschl, Cell wall elasticity: I. a critique of the bulk elastic modulus approach and an analysis using polymer elastic principles, *Plant, Cell Environ.*, 1985, **8**(8), 563–570.
- 33 D. R. P. Hettiaratchi and J. R. O'Callaghan, Structural mechanics of plant cells, *J. Theor. Biol.*, 1978, **74**(2), 235–257.
- 34 M. A. J. Chaplain, The strain energy function of an ideal plant cell wall, *J. Theor. Biol.*, 1993, **163**(1), 77–97.
- 35 W. Hong, Z. Liu and Z. Suo, Inhomogeneous swelling of a gel in equilibrium with a solvent and mechanical load, *Int. J. Solids Struct.*, 2009, **46**(17), 3282–3289.
- 36 W. Hong, X. Zhao and Z. Suo, Large deformation and electrochemistry of polyelectrolyte gels, *J. Mech. Phys. Solids*, 2010, **58**(4), 558–577.
- 37 S. Cai and Z. Suo, Mechanics and chemical thermodynamics of phase transition in temperature-sensitive hydrogels, *J. Mech. Phys. Solids*, 2011, **59**(11), 2259–2278.
- 38 J. Klier and N. A. Peppas, Anomalous penetrant transport in glassy polymers: 4. stresses in partially swollen polymers, *Polymer*, 1987, **28**(11), 1851–1859.
- 39 R. S. Harland, J. Klier and N. A. Peppas, Thermodynamic models for swelling of heterogeneous networks, *J. Appl. Polym. Sci.*, 1990, **41**(1–2), 249–265.
- 40 N. A. Peppas and J. Klier, Controlled release by using poly (methacrylic acid-g-ethylene glycol) hydrogels, *J. Controlled Release*, 1991, **16**(1), 203–214.
- 41 B. Amsden, A model for osmotic pressure driven release from cylindrical rubbery polymer matrices, *J. Controlled Release*, 2003, **93**(3), 249–258.
- 42 T. J. Pence and H. Tsai, Swelling-induced microchannel formation in nonlinear elasticity, *IMA journal of applied mathematics*, 2005, **70**(1), 173–189.
- 43 T. J. Pence and H. Tsai, Swelling-induced cavitation of elastic spheres, *Math. Mech. Solids*, 2006, **11**(5), 527–551.
- 44 F. P. Duda, A. C. Souza and E. Fried, Solvent uptake and cavitation, *J. Mech. Phys. Solids*, 2011, **59**(11), 2341–2354.
- 45 R. Marcombe, S. Cai, W. Hong, X. Zhao, Y. Lapusta and Z. Suo, A theory of constrained swelling of a pH-sensitive hydrogel, *Soft Matter*, 2010, **6**(4), 784–793.
- 46 A. Goriely and M. Tabor, Biomechanical models of hyphal growth in actinomycetes, *J. Theor. Biol.*, 2003, **222**(2), 211–218.
- 47 M. B. Amar and A. Goriely, Growth and instability in elastic tissues, *J. Mech. Phys. Solids*, 2005, **53**(10), 2284–2319.
- 48 A. Goriely, D. E. Moulton and R. Vandiver, Elastic cavitation, tube hollowing, and differential growth in plants and biological tissues, *EPL*, 2010, **91**(1), 18001.
- 49 Y.-P. Cao, B. Li and X.-Q. Feng, Surface wrinkling and folding of core-shell soft cylinders, *Soft Matter*, 2012, **8**(2), 556–562.
- 50 B. Li, G.-K. Xu and X.-Q. Feng, Tissue-growth model for the swelling analysis of core-shell hydrogels, *Soft Mater.*, 2013, **11**(2), 117–124.
- 51 Z. Liu, S. Swaddiwudhipong and W. Hong, Pattern formation in plants via instability theory of hydrogels, *Soft Matter*, 2013, **9**(2), 577–587.
- 52 M. Warner, B. L. Thiel and A. M. Donald, The elasticity and failure of fluid-filled cellular solids: theory and experiment, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**(4), 1370–1375.
- 53 M. Amon and C. D. Denson, A study of the dynamics of foam growth: analysis of the growth of closely spaced spherical bubbles, *Polym. Eng. Sci.*, 1984, **24**(13), 1026–1034.
- 54 H. X. Zhu and J. R. Melrose, A mechanics model for the compression of plant and vegetative tissues, *J. Theor. Biol.*, 2003, **221**(1), 89–101.
- 55 S. L. Everitt, O. G. Harlen and H. J. Wilson, Bubble growth in a two-dimensional viscoelastic foam, *J. Non-Newtonian Fluid Mech.*, 2006, **137**(1), 46–59.
- 56 J. D. Cloizeaux, The lagrangian theory of polymer solutions at intermediate concentrations, *J. Phys.*, 1975, **36**(4), 281–291.
- 57 P.-G. De Gennes, *Scaling concepts in polymer physics*, Cornell university press, 1979.
- 58 R. G. M. van der Sman, Biopolymer gel swelling analysed with scaling laws and flory-rehner theory, *Food Hydrocolloids*, 2015, **48**, 94–101.
- 59 G. O. Michalenko, H. R. Hohl and D. Rast, Chemistry and architecture of the mycelial wall of agaricus bisporus, *J. Gen. Microbiol.*, 1976, **92**(2), 251–262.
- 60 H.-i. Wu, R. D. Spence and P. J. H. Sharpe, Plant cell wall elasticity ii: polymer elastic properties of the microfibrils, *J. Theor. Biol.*, 1988, **133**(2), 239–253.
- 61 P. C. Mol, C. A. Vermeulen and J. G. H. Wessels, Diffuse extension of hyphae in stipes of agaricus bisporus may be based on a unique wall structure, *Mycol. Res.*, 1990, **94**(4), 480–488.
- 62 Y. C. Fung, K. Fronek and P. Patitucci, Pseudoelasticity of arteries and the choice of its mathematical expression, *Am. J. Physiol.: Heart Circ. Physiol.*, 1979, **237**(5), H620–H631.
- 63 Y.-C. Fung, *Biomechanics*, Springer, 1990.
- 64 K. A. Erk, K. J. Henderson and K. R. Shull, Strain stiffening in synthetic and biopolymer networks, *Biomacromolecules*, 2010, **11**(5), 1358–1363.
- 65 G. A. Holzapfel, T. C. Gasser and R. W. Ogden, A new constitutive framework for arterial wall mechanics and a comparative study of material models, *Journal of elasticity and the physical science of solids*, 2000, **61**(1–3), 1–48.
- 66 H. Demirkoparan and T. J. Pence, Swelling of an internally pressurized nonlinearly elastic tube with fiber reinforcing, *Int. J. Solids Struct.*, 2007, **44**(11), 4009–4029.
- 67 H. Demirkoparan and T. J. Pence, Torsional swelling of a hyperelastic tube with helically wound reinforcement, *J. Elast.*, 2008, **92**(1), 61–90.
- 68 T. M. Beecher, N. Magan and K. S. Burton, Water potentials and soluble carbohydrate concentrations in tissues of freshly harvested and stored mushrooms (agaricus bisporus), *Postharvest Biol. Technol.*, 2001, **22**(2), 121–131.
- 69 H.-H. Dai and Z. Song, Some analytical formulas for the equilibrium states of a swollen hydrogel shell, *Soft Matter*, 2011, **7**(18), 8473–8483.



- 70 H. Hinsken and W. Borchard, Continuous swelling pressure equilibria of the system κ -carrageenan/water, *Colloid Polym. Sci.*, 1995, **273**(10), 913–925.
- 71 A. R. Khokhlov, Swelling and collapse of polymer networks, *Polymer*, 1980, **21**(4), 376–380.
- 72 M. Quesada-Pérez, J. A. Maroto-Centeno, J. Forcada and R. Hidalgo-Alvarez, Gel swelling theories: the classical formalism and recent approaches, *Soft Matter*, 2011, **7**(22), 10536–10547.
- 73 J. Li, Y. Hu, J. J. Vlassak and Z. Suo, Experimental determination of equations of state for ideal elastomeric gels, *Soft Matter*, 2012, **8**(31), 8121–8128.
- 74 J. Li, Z. Suo and J. J. Vlassak, A model of ideal elastomeric gels for polyelectrolyte gels, *Soft Matter*, 2014, **10**(15), 2582–2590.
- 75 T. Hino and J. M. Prausnitz, Swelling equilibria for heterogeneous polyacrylamide gels, *J. Appl. Polym. Sci.*, 1996, **62**(10), 1635–1640.
- 76 M. Shibayama, Y. Shirotani, H. Hirose and S. Nomura, Simple scaling rules on swollen and shrunken polymer gels, *Macromolecules*, 1997, **30**(23), 7307–7312.
- 77 M. A. Zulliger, P. Frédez, K. Hayashi and N. Stergiopoulos, A strain energy function for arteries accounting for wall composition and structure, *J. Biomech.*, 2004, **37**(7), 989–1000.
- 78 Y. Zhu, X. Y. Luo and R. W. Ogden, Nonlinear axisymmetric deformations of an elastic tube under external pressure, *Eur. J. Mech. A Solids*, 2010, **29**(2), 216–229.
- 79 W. G. Li, J. Going, N. A. Hill and X. Y. Luo, Breaking analysis of artificial elastic tubes and human artery, *Int. J. Appl. Mech. Eng.*, 2013, **5**, 1350024.
- 80 H. Kunzek, R. Kabbert and D. Gloyna, Aspects of material science in food processing: changes in plant cell walls of fruits and vegetables, *Z. Lebensm.-Unters. -Forsch. A*, 1999, **208**(4), 233–250.
- 81 F. Gómez and I. Sjöholm, Applying biochemical and physiological principles in the industrial freezing of vegetables: a case study on carrots, *Trends Food Sci. Technol.*, 2004, **15**(1), 39–43.
- 82 A. Voda, N. Homan, M. Witek, A. Duijster, G. van Dalen, R. van der Sman, J. Nijssse, L. van Vliet, H. Van As and J. van Duynhoven, The impact of freeze-drying on microstructure and rehydration properties of carrot, *Food Res. Int.*, 2012, **49**(2), 687–693.
- 83 F. J. Vergeldt, G. van Dalen, A. J. Duijster, A. Voda, S. Khalloufi, L. J. van Vliet, H. Van As, J. P. M. van Duynhoven and R. G. M. van der Sman, Rehydration kinetics of freeze-dried carrots, *Innovative Food Sci. Emerging Technol.*, 2014, **24**, 40–47.
- 84 M. Bazhal, N. Lebovka and E. Vorobiev, Optimisation of pulsed electric field strength for electroporation of vegetable tissues, *Biosyst. Eng.*, 2003, **86**(3), 339–345.
- 85 P. S. Nobel, J. Cavelier and J. L. Andrade, Mucilage in cacti: its apoplastic capacitance, associated solutes, and influence on tissue 5, *J. Exp. Bot.*, 1992, **43**(5), 641–648.
- 86 M. E. McCully and J. S. Boyer, The expansion of maize root-cap mucilage during hydration. 3. changes in water potential and water content, *Physiol. Plant.*, 1997, **99**(1), 169–177.
- 87 C. Frenkel and T. G. Hartman, Decrease in fruit moisture content heralds and might launch the onset of ripening processes, *J. Food Sci.*, 2012, **77**(10), S365–S376.
- 88 W. A. Aregawi, T. Defraeye, P. Verboven, E. Herremans, G. De Roeck and B. M. Nicolai, Modeling of coupled water transport and large deformation during dehydration of apple tissue, *Food Bioprocess Technol.*, 2013, **6**(8), 1963–1978.
- 89 S. W. Fanta, M. K. Abera, W. A. Aregawi, Q. T. Ho, P. Verboven, J. Carmeliet and B. M. Nicolai, Microscale modeling of coupled water transport and mechanical deformation of fruit tissue during dehydration, *J. Food Eng.*, 2014, **124**, 86–96.
- 90 W. A. Aregawi, M. K. Abera, S. W. Fanta, P. Verboven and B. Nicolai, Prediction of water loss and viscoelastic deformation of apple tissue using a multiscale model, *J. Phys.: Condens. Matter*, 2014, **26**(46), 464111.
- 91 P. Babin, G. D. Valle, H. Chiron, P. Cloetens, J. Hoszowska, P. Pernot, A. L. Réguerre, L. Salvo and R. Dendievel, Fast x-ray tomography analysis of bubble growth and foam setting during breadmaking, *J. Cereal Sci.*, 2006, **43**(3), 393–397.
- 92 R. G. M. van der Sman and J. Broeze, Effects of salt on the expansion of starchy snacks: a multiscale analysis, *Food Funct.*, 2014, **5**(12), 3076–3082.
- 93 R. G. M. van der Sman and J. Broeze, Multiscale analysis of structure development in expanded starch snacks, *J. Phys.: Condens. Matter*, 2014, **26**(46), 464103.
- 94 B. J. Dobraszczyk and C. A. Roberts, Strain hardening and dough gas cell-wall failure in biaxial extension, *J. Cereal Sci.*, 1994, **20**(3), 265–274.
- 95 S. Cai, D. Chen, Z. Suo and R. C. Hayward, Creasing instability of elastomer films, *Soft Matter*, 2012, **8**(5), 1301–1304.
- 96 D. Okumura, T. Kuwayama and N. Ohno, Effect of geometrical imperfections on swelling-induced buckling patterns in gel films with a square lattice of holes, *Int. J. Solids Struct.*, 2014, **51**(1), 154–163.
- 97 A. D. Drozdov, Equilibrium swelling of core-shell composite microgels, *Meccanica*, 2015, 1–14.
- 98 X. Chen and H.-H. Dai, Asymptotic solutions and new insights for cylinder and core-shell polymer gels in a solvent, *Soft Matter*, 2013, **9**(36), 8664–8677.
- 99 R. G. M. van der Sman and M. B. J. Meinders, Prediction of the state diagram of starch water mixtures using the Flory-Huggins free volume theory, *Soft Matter*, 2011, **7**(2), 429–442.

